

Natural Compounds and Autophagy Modulation

Subjects: [Pharmacology & Pharmacy](#) | [Biochemistry & Molecular Biology](#)

Contributor: Md. Ataur Rahman , MD. Hasanur Rahman , Md. Shahadat Hossain , Partha Biswas , Rokibul Islam , Md Jamal Uddin , Md. Habibur Rahman , Hyewhon Rhim , Md Ataur Rahman

Autophagy is a vacuolar, lysosomal degradation pathway for injured and damaged protein molecules and organelles in eukaryotic cells, which is controlled by nutrients and stress responses. Dysregulation of cellular autophagy may lead to various diseases such as neurodegenerative disease, obesity, cardiovascular disease, diabetes, and malignancies. Recently, natural compounds have come to attention for being able to modulate the autophagy pathway in cancer prevention, although the prospective role of autophagy in cancer treatment is very complex and not yet clearly elucidated. Numerous synthetic chemicals have been identified that modulate autophagy and are favorable candidates for cancer treatment, but they have adverse side effects. Therefore, different phytochemicals, which include natural compounds and their derivatives, have attracted significant attention for use as autophagy modulators in cancer treatment with minimal side effects.

[autophagy](#)[cancer](#)[phytochemical](#)[natural compound](#)[treatment](#)

1. Introduction

Autophagy is triggered by the entrapment of abnormal intracellular proteins, invading microorganisms, and damaged organelles with the formation of double-layer autophagosomes, in the presence of nutrient stress, injury, and fasting ^{[1][2]}. Along with cellular energy stability, autophagy contributes to the control of cellular quality, and destruction of abnormal proteins and damaged organelles ^[3]. As a result, the origin and development of many diseases, such as neurodegenerative disorders, cancer, and autoimmune diseases, can be explained by defective autophagy ^[4]. Multiple data states a connection between the variety of diet and feedback to cancer medication ^[5]. It has been established that sufficient consumption of dietary and medicinal plant-derived biochemical compounds lowers the frequency of cancer mortality. This occurs due to the activation or balancing of various cellular oncogenes. In different human disorders, including cancer, there is documented evidence of reduction of autophagic regulation. Several lines of evidence have shown that cancer plays a critical role in inhibiting or augmenting autophagic pathways ^[6]. In addition, this dual role of autophagy has a significant therapeutic benefit against cancer ^[7]. The present report has established that natural product-derived bioactive molecules, along with lysosomal inhibitors, including chloroquine (CQ) and hydroxychloroquine (HCQ) are important regulators of autophagy signaling ^[8]. In addition, these proactive molecules can regulate the process of autophagy both in vitro and in vivo by involving the enzymes, transcription factors, and various intracellular communication pathways ^[9]. The autophagy stimulation or prohibition process is vastly complicated and regulated, so it needs to be extensively

explored. However, emerging evidence has implicated that extreme or damaged autophagy might lead to a unique type of cell death known as autophagic cell death [10]. Conversely, the proposal of using natural compounds as anti-cancer and autophagy-modulating agents requires a better understanding of their cellular and molecular mechanisms. The mechanism of action needs to be further addressed.

In this review, we compiled the cellular and molecular mechanisms involved in the dual role of autophagy, as a tumor-suppressing as well as a tumor-augmenting phenomenon in cancer. Current improvements in the use of natural bioactive compounds for cancer treatment by targeting autophagic action have been consistently analyzed [11]. Given the vital role of autophagy in cancer prevention and treatment, we closely reviewed different plant-derived biomolecules that may play a role in modulating the autophagy-linked signaling pathways as well as contribute towards competent treatment strategies against specific molecular aspects in cancer. In particular, administration of naturally derived compounds have been considered to regulate autophagy modulation, which might help improve our understanding of the mechanisms of natural compounds in the management and treatment of autophagy-linked diseases and cancers.

2. Mechanisms of Autophagy Signaling Pathway

Autophagy is a sensitive and tightly mannered process that has multiple steps [12]. Proteins crucial for autophagy were first reported in yeast and are termed as autophagy-related (Atg) proteins [13]. Multiple sequential molecular events take place to initiate the autophagy signaling cascade. First, an Atg1/unc51-like kinase (ULK) complex kinase regulates the induction, and the BECN1/class III PI3K complex initiates the nucleation of vesicles [14]. In addition, ubiquitin-like conjugation systems (Atg12 and Atg8/LC3B) control vesicle elongation as well as retrieval of lipids mediated by transmembrane Atg9 and associated proteins, which involve other Atg proteins [15]. Lysosome-associated membrane protein 2 (LAMP2) and RAS-related protein-7 (RAB7A) participate in the fusion between lysosomes and autophagosome formation [16]. Finally, vesicle breakdown and degradation by lysosomal hydrolases produce recyclable products [17] (Figure 1). Microtubule-associated protein 1 light chain 3-beta (MAP1LC3B) and Atg5-Atg12 complex pathways, which are ubiquitin-like pathways, are regulatory for vesicle elongation and are encoded by the yeast Atg8 pathway, which has a human homologue [18]. Following the autophagy response, human Atg4 cysteine protease cleaves LC3B protein and thereby produces the LC3B-I isoform [19]. Activated LC3B-I isoform causes the activation of Atg7 protein through LC3B-I and is transported to the Atg3 before formation of the LC3B-II isoform by the phosphatidylethanolamine (PE) conjugation with the carboxyl glycine of LC3B-I protein [20]. As a consequence, elevated synthesis and processing of LC3B shows promise and has been mentioned as a key marker of autophagy [20]. In the degradation of matured autophagosomes through the enzymatic action of lysosomal enzymes, small molecules are produced that are used for de novo protein synthesis, helping in survival and maintenance of homeostasis [21].

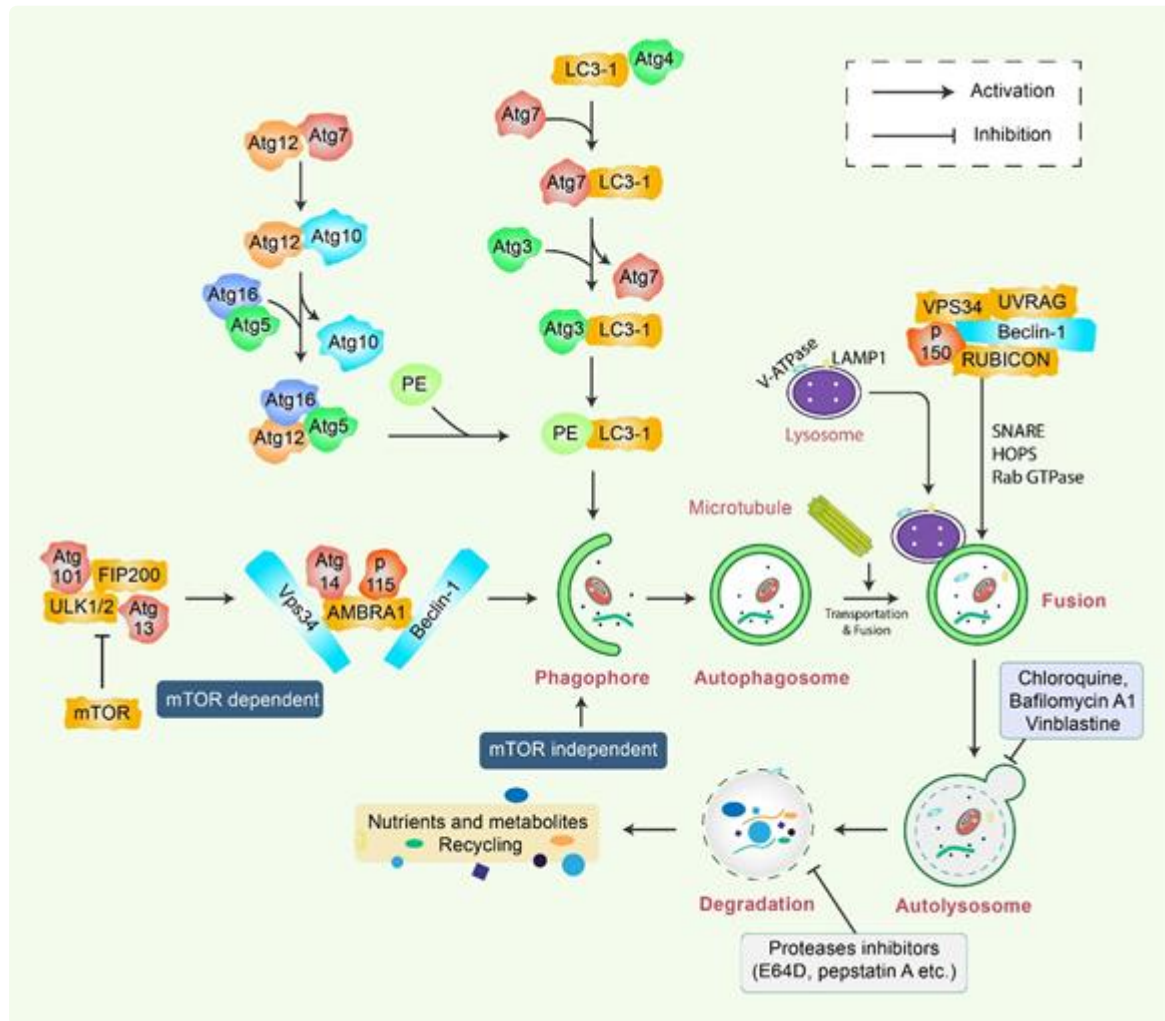


Figure 1. Molecular mechanism of autophagy signaling. Autophagy is generally initiated by a deficiency of growth factors or nutrients that trigger AMPK or mTOR inhibition. This stimulates FIP200 and Atg13 associated ULK complex. After that phosphorylation of Beclin-1, it leads to the activation of VPS34, which further initiates the formation of phagophore. Conjugation of Atg5-Atg12 encompasses Atg7 as well as Atg10 to form an Atg12-Atg5-Atg16 complex that also stimulates phagophores formation. Atg5, in addition to Atg12, makes an Atg16 complex, which acts as an E3-function concerning LC3-PE assembly (LC3-II). This complex likewise initiates the formation of a phagophore. Specifically, LC3-II is an autophagy marker that is ultimately interrupted via autolysosomes. Maturation of autophagosome leads to fusion with lysosomes in association with several lysosomal proteins, finally leading to degradation of cargo in addition to recycling of metabolites and nutrients.

3. Natural Compound Triggers in Autophagy Modulation

A single therapeutic strategy and a combination modality can both be favorable for cancer treatment. The various strategies include radiation therapy, immune or genetic therapy, and chemotherapy, all of which create new diversions and avenues in the field of cancer treatment with adequate levels of safety and accuracy [22][23]. Natural compounds are effective in the positive regulation of anticancer activity, as they inhibit cellular proliferation, adjust the oxidative stress response, and manipulate autophagic signaling and response [24]. This entire response of

natural compounds is dependent on the cell type. The role of several natural compounds that modulate different pathways of autophagy signaling is illustrated in Figure 2.

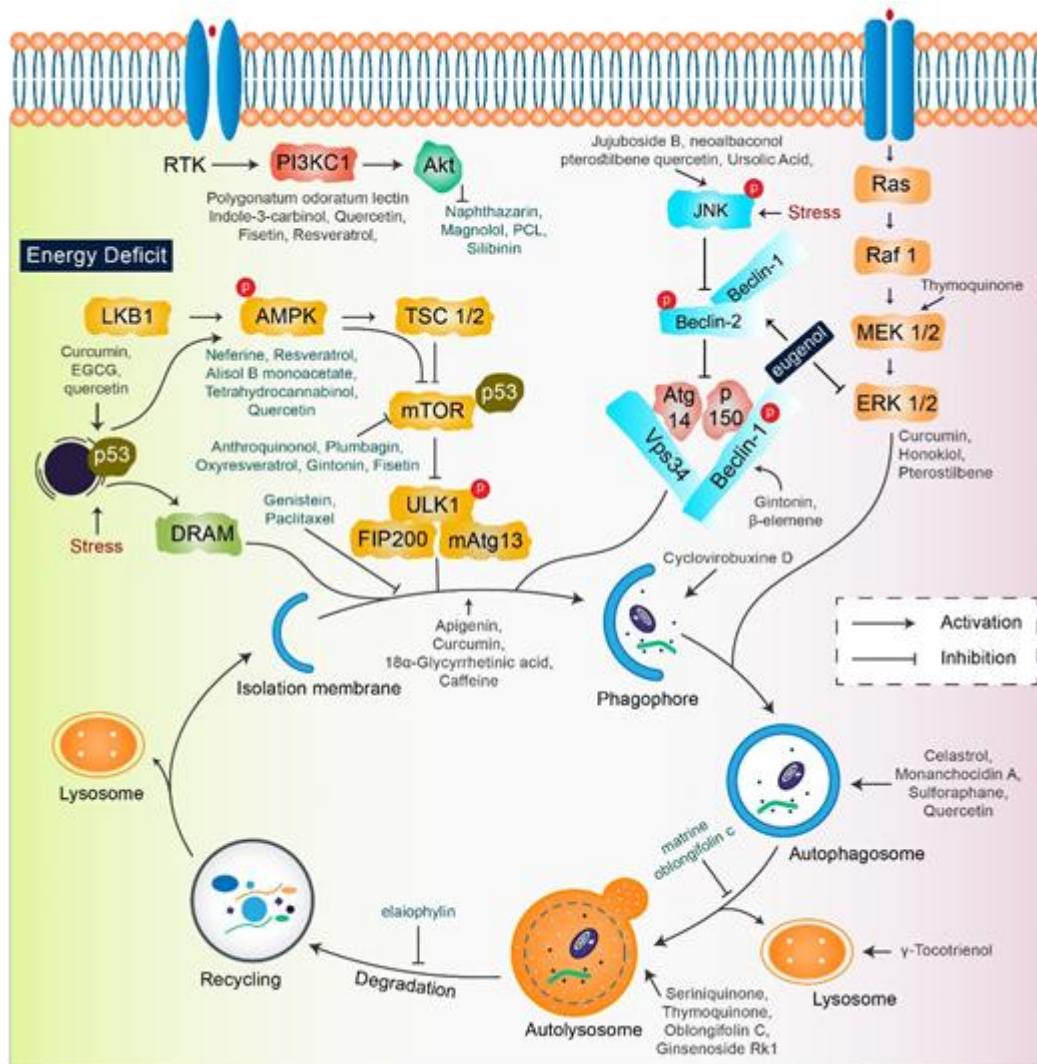


Figure 2. Key autophagy pathway and its regulators (natural compounds) for treating diverse cancers. This figure is modified from Wang et al. [25]. Natural compounds activate autophagy by several autophagic signaling targets and show a close relationship with cancer development and control. Multiple signaling pathways are activated or inhibited by natural compounds to modulate cancer cells.

3.1. Inhibition of Autophagy by Natural Compounds for Cancer Therapy

Apigenin: Apigenin, a flavonoid found in vegetables and fruits, can initiate tumor cell death through autophagy and apoptosis [26]. In combination with retinamide, *N*-(4-hydroxyphenyl), apigenin suppresses autophagy and increases apoptosis in human neuroblastoma (NB) cells [27].

Genistein: Genistein, a nutritive natural compound, has been documented to repress autophagy and induce apoptosis of human colon cancer HT-29 cells alone with downregulation of the PI3K/Akt signaling pathway [28]. Another mechanism of action of genistein is that it also exerts an antiproliferative effect on ovarian cancer cells

through autophagy as well as apoptosis [28]. Genistein augments miR-451 expression and improves isoproterenol-mediated cardiac hypertrophy [29]. Furthermore, genistein increases miR-1469 expression and prevents Mcl-1 expression in laryngeal cancer [30].

Indole-3-carbinol: Indole-3-carbinol, a natural compound collected from cruciferous vegetables, usually inhibits the PI3K/Akt pathway along with blockage of autophagy signaling in human colon cancer HT-29 cells [28].

Plumbagin: Plumbagin is obtained from *Plumbago zeylandica* L root and has a prompt antiproliferative activity through autophagic cell death, mediated via inhibition of AKT/mTOR pathway in human cervical cancer cells [31].

3.2. Induction of Autophagy by Natural Compounds as Potential Cancer Therapy

Antroquinonol: Antroquinonol, produced from *Antrodia Camphorata*, shows anti-tumor activity in various cancer cells [32]. Antroquinonol has been demonstrated to exhibit inhibitory effects on non-small cell lung cancer (NSCLC), as well as human pancreatic cancer, PANC-1 and ASPC-1, via the PI3K/Akt/mTOR pathway [33]. The anti-proliferative effect of antroquinonol is related to apoptosis, autophagic cell death, and rapid senescence of the pancreatic cells [33]. Several studies have demonstrated that antroquinonol can be introduced for the treatment of neoplasms [34]. A proposed clinical trial in metastatic pancreatic carcinoma is aimed at evaluating the anticancer activity of antroquinonol in combination with nab-PTX and gemcitabine [35].

18 α -Glycyrrhetic acid: 18 α -Glycyrrhetic acid (18-GA), a known gap-junction inhibitor, has been shown to demonstrate anticancer properties in human NB cells. 18-GA has been shown to induce autophagy as well as apoptosis. Autophagy increases Atg5, Atg7, and LC3II along with degradation of p62 (Figure 3). Furthermore, 18-GA, along with the autophagy inhibitor CQ, induces significant cell death in NB cells. Moreover, the Bcl-2/Beclin-1 interaction in addition to the cleavage of Beclin-1 has been revealed by 18-GA in autophagy and apoptosis induced cell death. Caspase-3 siRNA and pan-caspase inhibitor treatment have been demonstrated to prevent 18-GA-induced cellular cytotoxicity, indicating that caspase-mediated apoptosis induction was observed in human NB cells .

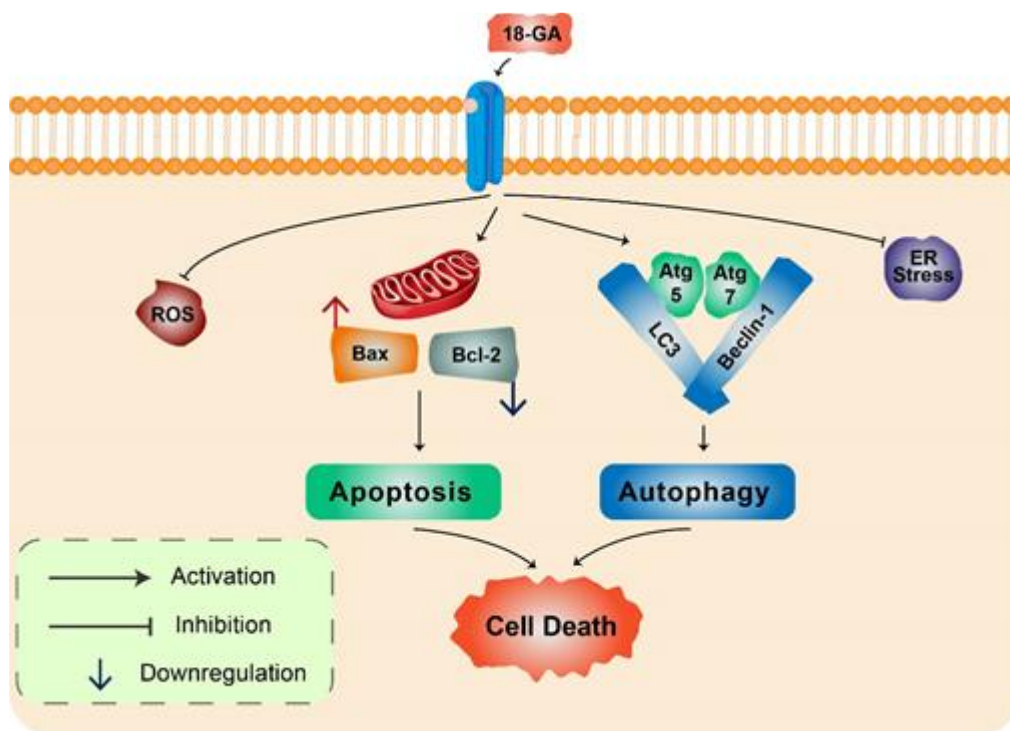


Figure 3. 18 α -Glycyrrhetic acid-mediated cell death in human neuroblastoma cells. 18-GA induces mitochondria-mediated apoptosis to alter mitochondrial membrane potential in a significant change in with Bax/Bcl-2 ratio. Autophagy-related proteins Atg5/7, Beclin-1, LC3, and p62 were activated during 18-GA-mediated autophagic cell death. Beclin-1 contributes to autophagy in addition to apoptosis induction in 18-GA-mediated cell death. Additionally, ROS and ER stress showed no changes during 18-GA treatment in SH-SY5Y and B103.

Celastrol: Celastrol, a popular natural medicinal compound triperine secreted from *Tripterygium wilfordii* Hook, is a polyubiquitinated aggregate that degrades the autophagy substrate p62 in the human glioblastoma (GBM) cancer cells [36]. Celastrol-mediated cytotoxicity was not sensitized by adjustment with autophagy inhibitors. Paraptosis-like cytoplasmic vacuolization can be induced by celastrol and has been associated with autophagy and the initiation of apoptosis in PC-3, A549, and HeLa cancer cells [37]. Celastrol can be used to treat fever, joint pain, and edema without causing any side effects [38]. Recently, many research studies have identified celastrol as a neuroprotective agent via a collaborative drug screen that can be used for the treatment of many neurodegenerative diseases.

Monanchocidin A: An active novel alkaloid, monanchocidin A, has been recently isolated from *Monanchora pulchra*, a marine sponge [39]. Monanchocidin can initiate autophagy and lysosomal membrane permeabilization in tumor cells [39]. Many studies have demonstrated that monanchocidin A can inhibit human urogenital cancers, including germ cell tumors, through autophagy signaling [40].

Paclitaxel: PTX, a natural medicinal compound isolated from *Taxus brevifolia* Nutt, shows a vast range of effect against various types of cancer such as breast, endometrial, bladder, and cervical cancer, by influencing the autophagy pathway [41]. The US Food and Drug Administration (FDA) has approved PTX for the treatment of ovarian cancer and early stage breast cancer. The anticancer activity of PTX is increased when PTX is combined

with cisplatin or carboplatin, and the combined compound can be used for the early treatment of advanced ovarian cancer [42]. It has been shown that PTX can inhibit the initial stage of autophagy, along with apoptosis, under normoxic and hypoxic conditions in human breast cancer cells [43]. PTX-initiated apoptosis was combined with the initiation of autophagy in A549 lung cancer cells. Treatment with PTX also initiated acidic vesicular organelle formation, Beclin-1, Atg5, and LC3 expression, and the prevention of autophagy proceeds via PTX-mediated cell death [44][45]. Autophagy can promote PTX-mediated cell death. In human breast cancer cells, PTX prevents autophagy via an individual mechanism based on the cell-cycle phase [45].

Gintonin: Gintonin (GT), a novel ginseng-derived exogenous ligand of lysophosphatidic acid (LPA) receptors, has shown to induce autophagy in cortical astrocytes [46]. GT intensely augmented the autophagy marker LC3 via the G protein-coupled LPA receptor-mediated pathway. However, GT-mediated autophagy was considerably decreased via autophagy inhibition 3-MA as well as Beclin-1, Atg5, and Atg7 gene knockdown [46]. Notably, pretreatment with a lysosomotropic agent, Baf A1 and E-64d/peps A, GT significantly improved LC3-II levels in addition to the formation of LC3 puncta (Figure 4). Furthermore, GT treatment improved autophagic flux, which influenced lysosome-associated membrane protein 1 (LAMP1) in addition to degradation of the autophagy substrate ubiquitinated p62/SQSTM1 protein in mouse cortical astrocytes [46].

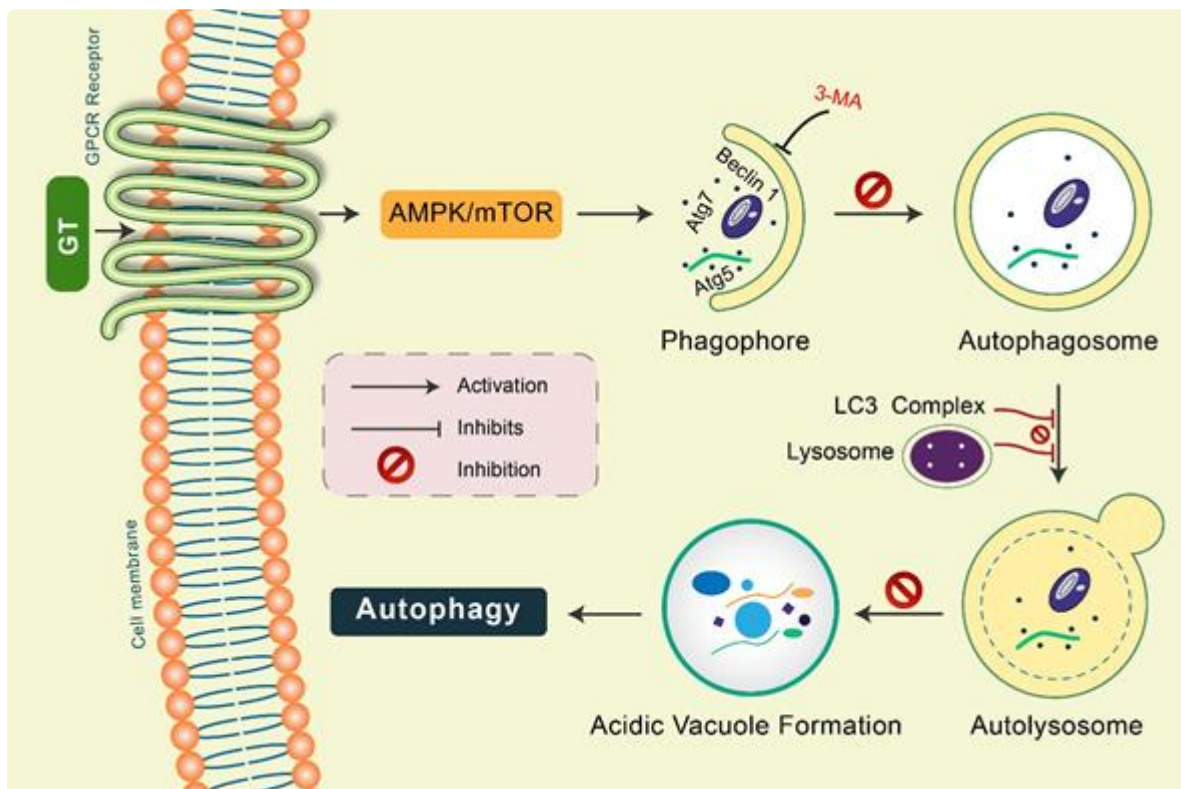


Figure 4. Gintonin (GT) activates autophagy in mouse cortical astrocytes. GT-mediated autophagy was dependent on GPCR-LPA receptors. Pretreatment with an LPA receptor antagonist, Ki16425, considerably decreased GT-mediated autophagy, while LPA agonist pretreatment, LPA 18:1, augmented autophagy in astrocytes. Autophagy initiation proteins Atg5, Atg7, and Beclin-1 were found to be activated in GT-mediated autophagy in astrocytes. Inhibition of autophagy by 3-MA, GT-mediated autophagy was prevented. Pretreated with E-64d/pepstatin A and

bafilomycin A1, GT additionally improved LC3 puncta formation indicating enhanced autophagic flux in cortical astrocytes. GT treatment increased acidic vacuole formation along with p62 protein accumulation during this autophagy process.

β-elemene: β-elemene, derived from *Rhizoma Curcumae*, is a natural terpenoid bioactive compound that shows strong activity against various types of cancers. In the human body, β-elemene works against the PI3K/Akt/mTOR/p70S6K pathway that induces autophagy and apoptosis of human NSCLCA549 cells [46]. As β-elemene increases the antitumor effect, autophagy is prevented by the interception with chloroquine [46]. In human renal-cell carcinoma 786-0 cells, β-elemene is used for the inhibition of the MAPK/ERK and P13K/Akt/mTOR signaling pathways to initiate defensive autophagy and apoptosis [47].

Caffeine: In HeLa cells, the Akt/mTOR/p70S6K pathway initiates autophagy and apoptotic cell death under the influence of caffeine that is routinely consumed in beverages [48].

Curcumin: Curcumin is a non-toxic toxic polyphenol with yellow pigment isolated from the rhizome of *Curcuma longa* L [49]. Curcumin-associated autophagy is considered to be a signal for cellular death in various types of cancer cells in the human body. Nevertheless, in the human body, curcumin can induce cellular differentiation and cellular survival by regulating the activities of AKT-AMPK-mediated autophagy induction [50]. Thus, it prevents the growth of malignant glioma cells in vitro and in vivo. Curcumin is used to induce non-apoptotic autophagy cell death in U87-MG and U373-MG malignant glioma cells [51]. This activity is related to the G2/M phase cell cycle attack, the prevention of the ribosomal S6 protein kinase pathway with the ERK1/ERK2 activation pathway [51]. Curcumin is responsible for Atg apoptosis in mesothelioma and k562 long-standing myelogenous leukemia cells by AKT/mTOR and NF-κB signaling pathways [52]. Curcumin can be used as an effective therapeutic compound in cancer management and is well endured, as confirmed by preclinical data in clinical experiments. Curcumin has been found to suppress TGF-β expression, such as p-SMAD2, TGF-β3, MMP-13, and NF-κB tumor-promoting factors as well as metastatic active adhesion molecules, such as intercellular adhesion molecule 1 (ICAM-1) and β1-integrin in stromal fibroblasts and colorectal cancer cells [53]. In addition, MiR-1246 has been shown to be involved in curcumin-induced radiosensitizing actions on bladder cancer cells by targeting p53 translation [54].

Tetrahydrocurcumin: Tetrahydrocurcumin (THC) is one of the strongest bioactive natural metabolite derived from curcumin. THC effectively shows strong antioxidant, anticancer, and cardioprotective properties [55]. The action of THC on autophagy was assessed by the decrease in the PI3K/Akt-mTOR and MAPK signaling pathways and initiation of caspase-7 mediated apoptosis [56].

Oxyresveratrol: Oxyresveratrol (Oxy R), 4-[(E)-2-(3,5-dihydroxyphenyl) ethenyl] benzene-1,3-diol/ 2,3,4,5-tetrahydroxy-trans-stilbene found in *Morus alba*, has been shown to activate autophagy and apoptotic cell death in NB cells (Figure 5). Oxy R-mediated autophagic cells were independent of apoptotic cell death. The PI3K/AKT/mTOR and p38 MAPK pathways control Oxy R-induced cell death in SH-SY5Y human NB cells.

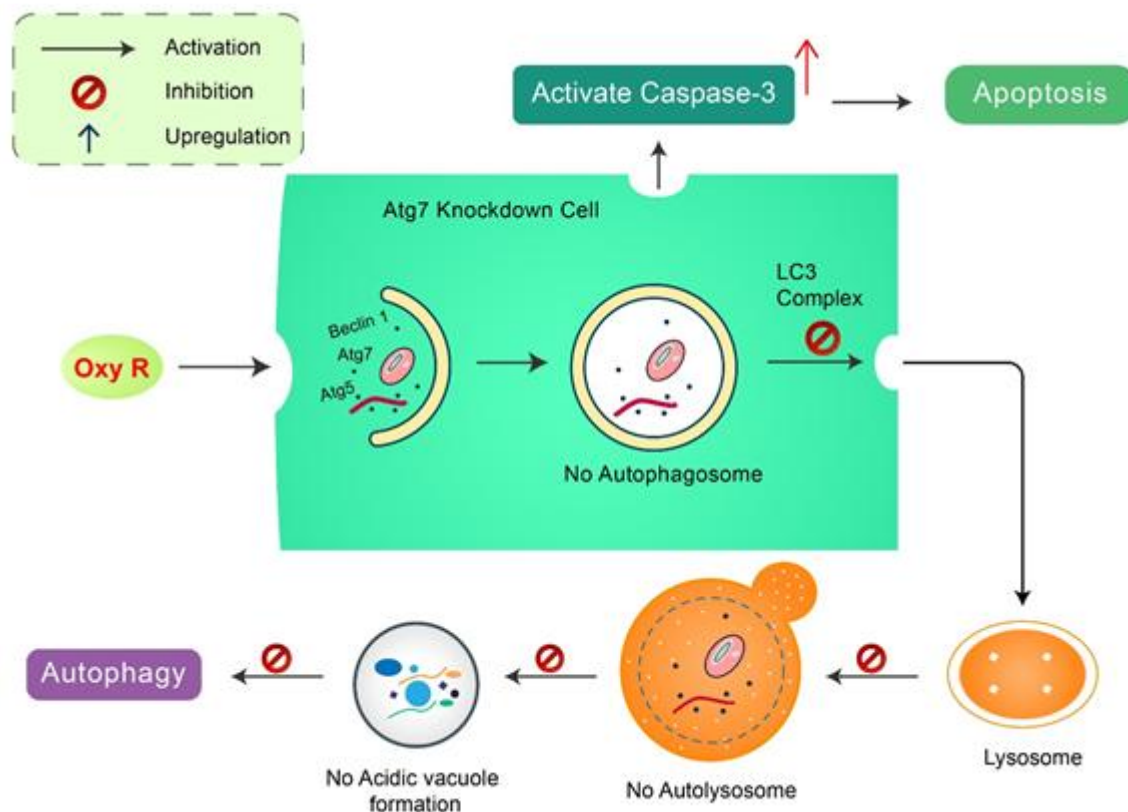


Figure 5. Natural compound oxyresveratrol (Oxy R) stimulates apoptosis and autophagy-dependent cell death. Oxy R activates autophagy through autophagy initiation Atg7 and Beclin 1 dependent pathways. Atg7 knockdown prevents autophagy, but activates apoptotic cell death in SH-SY5Y cells. Blockage of autophagy via 3-MA, Oxy R inhibits autophagic cell death. When apoptosis signaling is inhibited by Z-DEVD-FMK caspase-3 inhibitor, Oxy R exhibits autophagic cell death.

Epigallocatechin-3-gallate: The activity of cisplatin and oxaliplatin-initiated autophagy was promoted by EGCG in HT-29 and DLD-1 colorectal cancer cells, as identified by the multiplication of LC3B-II protein and autophagosome organization [57]. EGCG initiated autophagy as well as apoptosis in SSC-4 squamous cell carcinoma in combination with the upregulation of FAS, BAK, BAD, IGF-IR, WNTII, and ZEB1 proteins along with downregulation of MYC, TP53, and CASP8 proteins [58]. The formation of autophagosomes is increased by EGCG. In hepatic cells, the processes of lysosomal acidification and autophagic flux are also increased by EGCG in vitro, as well as in vivo [59]. Remarkably, luteolin and EGCG combination repressed TGF- β -mediated demonstration of myofibroblast phenotypes by reducing RhoA as well as ERK activation [60]. EGCG suggestively repressed rat hepatic stellate cells proliferation through hindering expression of PDGF- β receptor and tyrosine phosphorylation [61]. Furthermore, it has been found that EGCG attenuates uric acid-mediated NRK-49 F cell injury by up-regulating miR-9, and successively by JAK-STAT as well as NF- κ B signaling pathway activation [101]. EGCG reduces carcinoma cell growth probably by regulating miRNA expression, and may be a potential beneficial target for the prevention of cancers.

Fisetin: Fisetin (3,7,3',4'-tetrahydroxyflavone), a flavonol and a member of flavonoid polyphenols [62], is usually isolated from many vegetables and fruits [63]. Fisetin can function as an inhibitor of the expression of PI3K/Akt/mTOR pathways and regulate autophagy in prostate cancer and human NSCLC cells [64][65]. Fisetin has been found to inhibit the mTOR pathway and induce autophagy in human prostate cancer cells [66]. In addition, the effects of fisetin on autophagy are cell-type specific because fisetin inhibits autophagy in MCF-7 breast cancer cells and induces caspase-7-mediated apoptosis [67].

γ-Tocotrienol: Tocotrienols and the isoforms of vitamin E are differentiated on the basis of their antioxidant, anti-inflammatory, and anticancer properties. γ-tocotrienol induces endoplasmic inflammation and autophagy-associated cell death [68]. Pretreatment with the autophagy inhibitors 3-MA or Baf1 prevented the γ-tocotrienol-initiated cytotoxicity [69]. Many other studies have demonstrated that in mouse mammary cancer cells, the attachment of γ-tocotrienol with oridonin effectively initiated both autophagic and apoptotic effects [70]. The potency of autophagy cellular markers was significantly enhanced by the attachment of γ-tocotrienol with oridonin [71]. Simultaneously, this potency also improved the transformation of LC3-I to LC3-II, Atg3, Atg7, Beclin-1, Atg5-Atg12, LAMP-1, and cathepsin-D.

Geraniol: Fruits, vegetables, and cereal grains contain geraniol, a secondary metabolite, that in human PC-3 prostate cancer cells was shown to induce apoptosis and Atg cell death [72].

Seriniquinone: Seriniquinone is a medicinal compound derived from a marine bacterium of the genus *Serinicoccus*. It exhibits effective anti-proliferative activity against melanoma cell lines by activation of autophagocytosis [73].

Thymoquinone: Thymoquinone (TQ) is a key bioactive natural product isolated from black cumin, *Nigella sativa* L. TQ is related to increased volumes of autophagic vacuoles, as well as LC3 proteins, and enhances the combination of autophagosomes in the head and neck squamous carcinoma cells [74]. TQ effectively reduced the growth rate of GBM cells in association with the lysosomal inhibitor CQ, mostly causing apoptosis, and initiating autophagy [75]. LC3-II and p62 proteins are enhanced by the expression of TQ [75]. It was also shown that TQ prevented the growth of irinotecan-inhibited LoVo colon cancer cells by primarily activating apoptosis before autophagy [76]. Activation of the p38 and JNK MAP kinase pathways is related to TQ, which causes autophagic cell death [76]. TQ-induced caspase and autophagic cell death were also related to mitochondrial outward membrane penetrability [75].

Magnolol: Magnolol, isolated from *Magnolia officinalis*, exhibited anti-cancer and anti-tumor activity by directing autophagy and apoptosis signaling pathways [77]. However, magnolol promotes autophagy-mediated cell death in human non-small lung cancer H460 cells at high concentrations [78]. Additionally, in SGC-7901 human gastric adenocarcinoma cells, magnolol has been shown to repress PI3K/Akt pathway and encourage death by inducing autophagy [79].

Polyphenol mulberry extract: Mulberry is derived from *Morus Alba* leaf, which modulates the autophagy AMPK/PI3K/Akt pathway [80]. In Hep3B human hepatocellular carcinoma cells, mulberry extract modulates

autophagic or apoptotic cell death by the activation of the p53-dependent pathway [81]. It has been found that in NB cells, *Morus alba* root extract initiates apoptosis through FOXO-Caspase-3 dependent pathway [82].

Oblongifolin C: Oblongifolin C (OC), isolated from *Garcinia yunnanensis* Hu, is a strong autophagy inhibitor [83]. Treatment with OC resulted in an augmented number of autophagosomes as well as reduced SQSTM1/p62 and degradation [84]. OC showed anticancer effectiveness by improved staining of LC3 puncta, SQSTM1, and cleaved CASP-3, along with decreased expression levels of lysosomal cathepsins in an in vivo xenograft mouse model [83].

Naphthazarin: Naphthazarin repressed the Akt/PI3K pathway by activating autophagy and apoptosis pathways and by inducing A549 lung cancer cell death [85].

Sulforaphane: Sulforaphane is derived from cruciferous vegetables, such as cauliflower, cabbage, broccoli, and hoary weed [86]. In human prostate cancer PC-3 cells, sulforaphane has been shown to prompt autophagosome formation as well as acidic vesicular organelle formation [87]. Sulforaphane augmented the protein levels of LC3B-I in addition to encouraging LC3B-II processing. Exposure to sulforaphane in tumor cells displayed LC3B-II puncta related to autophagosome formation [88]. It also disrupted BECN1/BCL-2 interaction, causing autophagy initiation through the release of BECN1. Phosphorylated AKT-Ser473 levels decreased by sulforaphane in addition to simultaneous treatment with autophagy inhibitors, 3-MA or CQ, and inhibited tumor cell growth and proliferation [89]. Sulforaphane initiation decreases oxidative stress in addition to misfolded protein accumulation [90].

Mollugin: A bioactive phytochemical, mollugin, sequestered from *Rubia cordifolia* L., showed anticancer potential against numerous cancer cells [91]. Additional studies confirmed that the mTOR and ERK pathways are involved in mollugin-prompted autophagy in addition to apoptosis [92].

Jujuboside B: Jujuboside B is a saponin found in the seeds of *Zizyphus jujubavar* Spinose. It increased autophagy and apoptosis in human HCT-116 and AGS gastric adenocarcinoma cells in vitro and successfully repressed tumor growth in a xenograft model of nude mice in vivo [93]. Furthermore, jujuboside B activated p38/JNK-mediated autophagy induction through cytoplasmic vacuole formation in addition to LC3-I/II conversion [93].

Cyclovirobuxine D: Cyclovirobuxine D (CVB-D) has been shown to activate autophagy in the human MCF-7 breast cancer cell line through the addition of autophagosomes as well as raised LC3 puncta, along with augmented conversion of LC3-I to LC3-II [94]. However, CVB-D-mediated autophagy induction and cell viability were blocked by 3-MA [94].

Polygonatum odoratum lectin: Polygonatum odoratum lectin (POL) has been shown to reveal apoptosis-inducing and anti-proliferative properties in a wide range of cancer cells [95]. In human breast cancer NSCLC A549 and MCF-7 cells, POL prompted both autophagy and apoptosis [96]. POL stimulated apoptosis by preventing the AKT/NF-κB pathway, whereas augmented autophagy through AKT-mTOR pathway suppression [96].

Resveratrol: Resveratrol, a polyphenol compound derived from berries, red grapes, and peanuts, has been shown to facilitate cell death in a diverse variety of cancer cells through autophagy, apoptosis, and necrosis [97][98]. In

ovarian cancer cells, resveratrol has been found to activate autophagic cell death [99]. However, resveratrol inhibited NF- κ B stimulation in association with augmented permeability of lysosomal in cervical cancer cells, demonstrating autophagic cell death [100]. Additionally, in HL-60 promyelocytic leukemia cells, it has been described that resveratrol-prompted apoptosis increased LC3-II levels in addition to dependency on mTOR pathway [101]. Meanwhile, resveratrol-mediated apoptosis was related to reduced p53 and AMPK/mTOR autophagy signaling pathways in renal carcinoma cells [102]. The antiproliferative action of resveratrol has been found in liver myofibroblasts by preventing PDGF signaling and decreasing EGF-dependent DNA synthesis [103]. Additionally, in retinal endothelial cells, resveratrol has been found to increase miR15a expression under conditions of high glucose levels and decreased insulin signaling [104].

Quercetin: Quercetin, a bioflavonoid, is generally found in fruits, beverages, and vegetables. In vitro and in vivo studies have shown that it exhibits antiproliferative activities in numerous tumors conventionally related to its antioxidant properties [105].

Rottlerin: Rottlerin, a natural product sequestered from *Mallotus philippinensis*, repressed PI3K/mTOR signaling in human pancreatic cancer stem cells, and activated autophagy-mediated apoptotic cell death [106].

Angelicin: Angelicin, a psoralen, is derived from *Angelica polymorpha* and has been shown to induce apoptosis [107] [108] and autophagy [109]. Angelicin increased autophagy-related proteins Atg3, Atg7 and Atg12-5 with phosphorylation of mTOR [110][111].

Ursolic acid: A triterpenoid phytochemical, ursolic acid (UA), has been found to activate autophagy by inducing LC3 in addition to p62 protein accumulation [111]. In HCT15 cells, UA repressed growth by modulating autophagy in the JNK pathway [112]. However, UA induced cytotoxicity in addition to repressing concentration-dependent TC-1 cervical cancer cells by controlling Atg5 and LC3-II, which demonstrated its anticancer activity [150]. Furthermore, in MCF7 breast cancer cells, UA activated autophagy under ER stress [113]. In PC3 prostate cancer cells, UA-induced autophagy was facilitated by the Akt/mTOR and Beclin-1 pathways [114].

Polygonatum Cyrtonea Lectin: Polygonatum Cyrtonea Lectin (PCL) inhibits the PI3K-Akt pathway in addition to prompting autophagy and apoptosis in cancer cells [115]. Although PCL significantly prevented the growth of A375 human melanoma cells, normal melanocytes were not affected [116]. Additionally, PCL has been found to stimulate both autophagy and apoptosis in A375 cells [117].

Silibinin: Silibinin, derived from *Silybum marianum*, has been shown to induce autophagic HT1080 cell death in human fibrosarcoma through diverse mechanisms, including inhibition of MEK/ERK and PI3K/Akt pathways [118].

Plant extract induces autophagy: Several plant-derived extracts from *Dioscorea nipponica* Makino, *Melandrium firmum*, marine algae, and natural flavonoids have been found to induce apoptosis and autophagic properties [119] [120][121][122]. *Saussurea lappa* ethanol extract has been investigated to activate apoptosis in NB [123] and autophagy in LNCaP prostate cancer cells [124].

Ginsenoside Rk1: Ginsenoside Rk1, an NMDA receptor inhibitor [125], exhibited antitumor and autophagy modulation activities in HepG2 cells. Rk1-prompted autophagy was recognized through LC3-I to LC3-II conversion, which incorporated lysosomes into the autolysosome process [126]. However, the autophagy inhibitor, Beclin 1 siRNA or bafilomycin A1, and Rk1 combination boosted autophagic activities in HepG2 [126].

Climacostol: In mouse B16-F10 melanoma cells, autophagosomes accumulate with dysfunctional autophagic degradation. However, climacostol provides mechanistic insights and promotes autophagosome turnover through the p53-AMPK pathway and activated p53 protein levels [127].

References

1. Rahman, M.A.; Rhim, H. Therapeutic implication of autophagy in neurodegenerative diseases. *BMB Rep.* 2017, 50, 345–354, doi:10.5483/bmbrep.2017.50.7.069.
2. Corti, O.; Blomgren, K.; Poletti, A.; Beart, P.M. Autophagy in neurodegeneration: New insights underpinning therapy for neurological diseases. *J. Neurochem.* 2020, e15002, doi:10.1111/jnc.15002.
3. Uddin, M.S.; Stachowiak, A.; Al Mamun, A.; Tzvetkov, N.T.; Takeda, S.; Atanasov, A.G.; Bergantin, L.B.; Abdel-Daim, M.M.; Stankiewicz, A.M. Autophagy and Alzheimer's Disease: From Molecular Mechanisms to Therapeutic Implications. *Front. Aging Neurosci.* 2018, 10, doi:10.3389/fnagi.2018.00004.
4. Arroyo, D.S.; Gaviglio, E.A.; Ramos, J.M.P.; Bussi, C.; Rodriguez-Galan, M.C.; Iribarren, P. Autophagy in inflammation, infection, neurodegeneration and cancer. *Int. Immunopharmacol.* 2014, 18, 55–65, doi:10.1016/j.intimp.2013.11.001.
5. Perry, R.J.; Shulman, G.I. Mechanistic Links between Obesity, Insulin, and Cancer. *Trends Cancer* 2020, 6, 75–78, doi:10.1016/j.trecan.2019.12.003.
6. Ling, Y.; Perez-Soler, R. Disruption of autophagic and autolysosomal signaling pathways leads to synergistic augmentation of erlotinib-induced apoptosis in wild type EGFR human non-small cell lung cancer cell lines. *EJC Suppl.* 2010, 8, 183–183, doi:10.1016/S1359-6349(10)72289-8.
7. Towers, C.G.; Wodetzki, D.; Thorburn, A. Autophagy and cancer: Modulation of cell death pathways and cancer cell adaptations. *J. Cell Biol.* 2020, 219, doi:10.1083/jcb.201909033.
8. Manic, G.; Obrist, F.; Kroemer, G.; Vitale, I.; Galluzzi, L. Chloroquine and hydroxychloroquine for cancer therapy. *Mol. Cell. Oncol.* 2014, 1, e29911, doi:10.4161/mco.29911.
9. Khandia, R.; Dadar, M.; Munjal, A.; Dhama, K.; Karthik, K.; Tiwari, R.; Yatoo, M.I.; Iqbal, H.M.N.; Singh, K.P.; Joshi, S.K.; et al. A Comprehensive Review of Autophagy and Its Various Roles in

- Infectious, Non-Infectious, and Lifestyle Diseases: Current Knowledge and Prospects for Disease Prevention, Novel Drug Design, and Therapy. *Cells* 2019, 8, doi:10.3390/cells8070674.
10. Bialik, S.; Dasari, S.K.; Kimchi, A. Autophagy-dependent cell death—Where, how and why a cell eats itself to death. *J. Cell Sci.* 2018, 131, doi:10.1242/jcs.215152.
 11. Lagoa, R.; Silva, J.; Rodrigues, J.R.; Bishayee, A. Advances in phytochemical delivery systems for improved anticancer activity. *Biotechnol. Adv.* 2020, 38, doi:10.1016/j.biotechadv.2019.04.004.
 12. Tavakol, S.; Ashrafizadeh, M.; Deng, S.; Azarian, M.; Abdoli, A.; Motavaf, M.; Poormoghadam, D.; Khanbabaie, H.; Afshar, E.G.; Mandegary, A.; et al. Autophagy Modulators: Mechanistic Aspects and Drug Delivery Systems. *Biomolecules* 2019, 9, doi:10.3390/biom9100530.
 13. Hirata, E.; Ohya, Y.; Suzuki, K. Atg4 plays an important role in efficient expansion of autophagic isolation membranes by cleaving lipidated Atg8 in *Saccharomyces cerevisiae*. *PLoS ONE* 2017, 12, doi:10.1371/journal.pone.0181047.
 14. Chen, Y.; He, J.; Tian, M.; Zhang, S.Y.; Guo, M.R.; Kasimu, R.; Wang, J.H.; Ouyang, L. UNC51-like kinase 1, autophagic regulator and cancer therapeutic target. *Cell Proliferat.* 2014, 47, 494–505, doi:10.1111/cpr.12145.
 15. Rahman, M.A.; Rahman, M.R.; Zaman, T.; Uddin, M.S.; Islam, R.; Abdel-Daim, M.M.; Rhim, H. Emerging Potential of Naturally Occurring Autophagy Modulators Against Neurodegeneration. *Curr. Pharm. Des.* 2020, 26, 772–779, doi:10.2174/1381612826666200107142541.
 16. Nakamura, S.; Yoshimori, T. New insights into autophagosome-lysosome fusion. *J. Cell Sci.* 2017, 130, 1209–1216, doi:10.1242/jcs.196352.
 17. Schulze, H.; Kolter, T.; Sandhoff, K. Principles of lysosomal membrane degradation Cellular topology and biochemistry of lysosomal lipid degradation. *BBA-Mol. Cell Res.* 2009, 1793, 674–683, doi:10.1016/j.bbamcr.2008.09.020.
 18. Shpilka, T.; Weidberg, H.; Pietrokovski, S.; Elazar, Z. Atg8: An autophagy-related ubiquitin-like protein family. *Genome Biol.* 2011, 12, doi:10.1186/gb-2011-12-7-226.
 19. Agrotis, A.; von Chamier, L.; Oliver, H.; Kiso, K.; Singh, T.; Ketteler, R. Human ATG4 autophagy proteases counteract attachment of ubiquitin-like LC3/GABARAP proteins to other cellular proteins. *J. Biol. Chem.* 2019, 294, 12610–12621, doi:10.1074/jbc.AC119.009977.
 20. Lystad, A.H.; Simonsen, A. Mechanisms and Pathophysiological Roles of the ATG8 Conjugation Machinery. *Cells* 2019, 8, doi:10.3390/cells8090973.
 21. Ward, C.; Martinez-Lopez, N.; Otten, E.G.; Carroll, B.; Maetzel, D.; Singh, R.; Sarkar, S.; Korolchuk, V.I. Autophagy, lipophagy and lysosomal lipid storage disorders. *BBA-Mol. Cell Biol. Lipids* 2016, 1861, 269–284, doi:10.1016/j.bbalip.2016.01.006.

22. Rahman, M.A.; Rahman, M.S.; Uddin, M.J.; Mamum-Or-Rashid, A.N.M.; Pang, M.G.; Rhim, H. Emerging risk of environmental factors: Insight mechanisms of Alzheimer's diseases. *Environ. Sci. Pollut. Res.* 2020, 10.1007/s11356-020-08243-z, doi:10.1007/s11356-020-08243-z.
23. Chen, H.H.W.; Kuo, M.T. Improving radiotherapy in cancer treatment: Promises and challenges. *Oncotarget* 2017, 8, 62742–62758, doi:10.18632/oncotarget.18409.
24. Luo, H.; Vong, C.T.; Chen, H.B.; Gao, Y.; Lyu, P.; Qiu, L.; Zhao, M.M.; Liu, Q.; Cheng, Z.H.; Zou, J.; et al. Naturally occurring anti-cancer compounds: Shining from Chinese herbal medicine. *Chin. Med.* 2019, 14, doi:10.1186/s13020-019-0270-9.
25. Wang, P.Q.; Zhu, L.J.; Sun, D.J.; Gan, F.H.; Gao, S.Y.; Yin, Y.Y.; Chen, L.X. Natural products as modulator of autophagy with potential clinical prospects. *Apoptosis* 2017, 22, 325–356, doi:10.1007/s10495-016-1335-1.
26. Choi, E.J.; Kim, G.H. Apigenin Induces Apoptosis through a Mitochondria/Caspase-Pathway in Human Breast Cancer MDA-MB-453 Cells. *J. Clin. Biochem. Nutr.* 2009, 44, 260–265, doi:10.3164/jcbn.08-230.
27. Mohan, N.; Banik, N.L.; Ray, S.K. Combination of N-(4-hydroxyphenyl) retinamide and apigenin suppressed starvation-induced autophagy and promoted apoptosis in malignant neuroblastoma cells. *Neurosci. Lett.* 2011, 502, 24–29, doi:10.1016/j.neulet.2011.07.016.
28. Nakamura, Y.; Yogosawa, S.; Izutani, Y.; Watanabe, H.; Otsuji, E.; Sakai, T. A combination of indol-3-carbinol and genistein synergistically induces apoptosis in human colon cancer HT-29 cells by inhibiting Akt phosphorylation and progression of autophagy. *Mol. Cancer* 2009, 8, 100, doi:10.1186/1476-4598-8-100.
29. Gan, M.L.; Zheng, T.; Shen, L.Y.; Tan, Y.; Fan, Y.; Shuai, S.R.; Bai, L.; Li, X.W.; Wang, J.Y.; Zhang, S.H.; et al. Genistein reverses isoproterenol-induced cardiac hypertrophy by regulating miR-451/TIMP2. *Biomed. Pharmacother.* 2019, 112, doi:10.1016/j.biopha.2019.108618.
30. Ma, C.H.; Zhang, Y.X.; Tang, L.H.; Yang, X.J.; Cui, W.M.; Han, C.C.; Ji, W.Y. MicroRNA-1469, a p53-responsive microRNA promotes Genistein induced apoptosis by targeting Mcl1 in human laryngeal cancer cells. *Biomed. Pharmacother.* 2018, 106, 665–671, doi:10.1016/j.biopha.2018.07.005.
31. Kuo, P.L.; Hsu, Y.L.; Cho, C.Y. Plumbagin induces G2-M arrest and autophagy by inhibiting the AKT/mammalian target of rapamycin pathway in breast cancer cells. *Mol. Cancer Ther.* 2006, 5, 3209–3221, doi:10.1158/1535-7163.MCT-06-0478.
32. Thiagarajan, V.; Tsai, M.J.; Weng, C.F. Antroquinonol Targets FAK-Signaling Pathway Suppressed Cell Migration, Invasion, and Tumor Growth of C6 Glioma. *PLoS ONE* 2015, 10, e0141285, doi:10.1371/journal.pone.0141285.

33. Yu, C.C.; Chiang, P.C.; Lu, P.H.; Kuo, M.T.; Wen, W.C.; Chen, P.N.; Guh, J.H. Antroquinonol, a natural ubiquinone derivative, induces a cross talk between apoptosis, autophagy and senescence in human pancreatic carcinoma cells. *J. Nutr. Biochem.* 2012, 23, 900–907, doi:10.1016/j.jnutbio.2011.04.015.
34. Chiang, P.C.; Lin, S.C.; Pan, S.L.; Kuo, C.H.; Tsai, I.L.; Kuo, M.T.; Wen, W.C.; Chen, P.; Guh, J.H. Antroquinonol displays anticancer potential against human hepatocellular carcinoma cells: A crucial role of AMPK and mTOR pathways. *Biochem. Pharmacol.* 2010, 79, 162–171, doi:10.1016/j.bcp.2009.08.022.
35. Chiou, J.F.; Wu, A.T.H.; Wang, W.T.; Kuo, T.H.; Gelovani, J.G.; Lin, I.H.; Wu, C.H.; Chiu, W.T.; Deng, W.P. A Preclinical Evaluation of Antrodia camphorata Alcohol Extracts in the Treatment of Non-Small Cell Lung Cancer Using Non-Invasive Molecular Imaging. *Evid.-Based Complement. Altern. Med.* 2011, 10.1093/ecam/nep228, 1-12, doi:10.1093/ecam/nep228.
36. Kannaiyan, R.; Manu, K.A.; Chen, L.X.; Li, F.; Rajendran, P.; Subramaniam, A.; Lam, P.; Kumar, A.P.; Sethi, G. Celastrol inhibits tumor cell proliferation and promotes apoptosis through the activation of c-Jun N-terminal kinase and suppression of PI3 K/Akt signaling pathways. *Apoptosis* 2011, 16, 1028–1041, doi:10.1007/s10495-011-0629-6.
37. Wang, W.B.; Feng, L.X.; Yue, Q.X.; Wu, W.Y.; Guan, S.H.; Jiang, B.H.; Yang, M.; Liu, X.; Guo, D.A. Paraptosis accompanied by autophagy and apoptosis was induced by celastrol, a natural compound with influence on proteasome, ER stress and Hsp90. *J. Cell. Physiol.* 2012, 227, 2196–2206, doi:10.1002/jcp.22956.
38. Li, H.; Zhang, Y.Y.; Tan, H.W.; Jia, Y.F.; Li, D. Therapeutic effect of tripterine on adjuvant arthritis in rats. *J. Ethnopharmacol.* 2008, 118, 479–484, doi:10.1016/j.jep.2008.05.028.
39. Dyshlovoy, S.A.; Hauschild, J.; Amann, K.; Tabakmakher, K.M.; Venz, S.; Walther, R.; Guzii, A.G.; Makarieva, T.N.; Shubina, L.K.; Fedorov, S.N.; et al. Marine alkaloid Monanchocidin a overcomes drug resistance by induction of autophagy and lysosomal membrane permeabilization. *Oncotarget* 2015, 6, 17328–17341, doi:10.18632/oncotarget.4175.
40. Dyshlovoy, S.A.; Venz, S.; Hauschild, J.; Tabakmakher, K.M.; Otte, K.; Madanchi, R.; Walther, R.; Guzii, A.G.; Makarieva, T.N.; Shubina, L.K.; et al. Anti-migratory activity of marine alkaloid monanchocidin A—Proteomics-based discovery and confirmation. *Proteomics* 2016, 16, 1590–1603, doi:10.1002/pmic.201500334.
41. McGuire, W.P.; Blessing, J.A.; Moore, D.; Lentz, S.S.; Photopulos, G. Paclitaxel has moderate activity in squamous cervix cancer: A gynecologic oncology group study. *J. Clin. Oncol.* 1996, 14, 792–795, doi:10.1200/Jco.1996.14.3.792.
42. Kampan, N.C.; Madondo, M.T.; McNally, O.M.; Quinn, M.; Plebanski, M. Paclitaxel and Its Evolving Role in the Management of Ovarian Cancer. *Biomed. Res. Int.* 2015, 2015, 413076, doi:10.1155/2015/413076.

43. Qiu, Y.; Li, P.; Ji, C.Y. Cell Death Conversion under Hypoxic Condition in Tumor Development and Therapy. *Int. J. Mol. Sci.* 2015, 16, 25536–25551, doi:10.3390/ijms161025536.
44. Lee, Y.; Na, J.; Lee, M.S.; Cha, E.Y.; Sul, J.Y.; Park, J.B.; Lee, J.S. Combination of pristimerin and paclitaxel additively induces autophagy in human breast cancer cells via ERK1/2 regulation. *Mol. Med. Rep.* 2018, 18, 4281–4288, doi:10.3892/mmr.2018.9488.
45. Veldhoen, R.A.; Banman, S.L.; Hemmerling, D.R.; Odsen, R.; Simmen, T.; Simmonds, A.J.; Underhill, D.A.; Goping, I.S. The chemotherapeutic agent paclitaxel inhibits autophagy through two distinct mechanisms that regulate apoptosis. *Oncogene* 2013, 32, 736–746, doi:10.1038/onc.2012.92.
46. Liu, J.; Hu, X.J.; Jin, B.; Qu, X.J.; Hou, K.Z.; Liu, Y.P. ss-Elementene induces apoptosis as well as protective autophagy in human non-small-cell lung cancer A549 cells. *J. Pharm. Pharmacol.* 2012, 64, 146–153, doi:10.1111/j.2042-7158.2011.01371.x.
47. Zhan, Y.H.; Liu, J.; Qu, X.J.; Hou, K.Z.; Wang, K.F.; Liu, Y.P.; Wu, B. beta-Elementene Induces Apoptosis in Human Renal-cell Carcinoma 786-0 Cells through Inhibition of MAPK/ERK and PI3K/Akt/mTOR Signalling Pathways. *Asian Pac. J. Cancer P* 2012, 13, 2739–2744, doi:10.7314/Apjcp.2012.13.6.2739.
48. Saiki, S.; Sasazawa, Y.; Imamichi, Y.; Kawajiri, S.; Fujimaki, T.; Tanida, I.; Kobayashi, H.; Sato, F.; Sato, S.; Ishikawa, K.I.; et al. Caffeine induces apoptosis by enhancement of autophagy via PI3K/Akt/mTOR/p70S6K inhibition. *Autophagy* 2011, 7, 176–187, doi:10.4161/auto.7.2.14074.
49. Lopez-Lazaro, M. Anticancer and carcinogenic properties of curcumin: Considerations for its clinical development as a cancer chemopreventive and chemotherapeutic agent. *Mol. Nutr. Food Res.* 2008, 52, S103-S127, doi:10.1002/mnfr.200700238.
50. Aoki, H.; Takada, Y.; Kondo, S.; Sawaya, R.; Aggarwal, B.B.; Kondo, Y. Evidence that curcumin suppresses the growth of malignant gliomas in vitro and in vivo through induction of autophagy: Role of Akt and extracellular signal-regulated kinase signaling pathways. *Mol. Pharmacol.* 2007, 72, 29–39, doi:10.1124/mol.106.033167.
51. Klinger, N.V.; Mittal, S. Therapeutic Potential of Curcumin for the Treatment of Brain Tumors. *Oxid. Med. Cell. Longev.* 2016, 2016, 9324085, doi:10.1155/2016/9324085.
52. Masuelli, L.; Benvenuto, M.; Di Stefano, E.; Mattera, R.; Fantini, M.; De Feudis, G.; De Smaele, E.; Tresoldi, I.; Giganti, M.G.; Modesti, A.; et al. Curcumin blocks autophagy and activates apoptosis of malignant mesothelioma cell lines and increases the survival of mice intraperitoneally transplanted with a malignant mesothelioma cell line. *Oncotarget* 2017, 8, 34405–34422, doi:10.18632/oncotarget.14907.
53. Buhrmann, C.; Kraehe, P.; Lueders, C.; Shayan, P.; Goel, A.; Shakibaei, M. Curcumin Suppresses Crosstalk between Colon Cancer Stem Cells and Stromal Fibroblasts in the Tumor

- Microenvironment: Potential Role of EMT. *PLoS ONE* 2014, 9, doi:10.1371/journal.pone.0107514.
54. Xu, R.; Li, H.B.; Wu, S.Q.; Qu, J.; Yuan, H.Y.; Zhou, Y.G.; Lu, Q. MicroRNA-1246 regulates the radio-sensitizing effect of curcumin in bladder cancer cells via activating P53. *Int. Urol. Nephrol.* 2019, 51, 1771–1779, doi:10.1007/s11255-019-02210-5.
 55. Aggarwal, B.B.; Deb, L.; Prasad, S. Curcumin Differs from Tetrahydrocurcumin for Molecular Targets, Signaling Pathways and Cellular Responses. *Molecules* 2015, 20, 185–205, doi:10.3390/molecules20010185.
 56. Wu, J.C.; Lai, C.S.; Badmaev, V.; Nagabhushanam, K.; Ho, C.T.; Pan, M.H. Tetrahydrocurcumin, a major metabolite of curcumin, induced autophagic cell death through coordinative modulation of PI3K/Akt-mTOR and MAPK signaling pathways in human leukemia HL-60 cells. *Mol. Nutr. Food Res.* 2011, 55, 1646–1654, doi:10.1002/mnfr.201100454.
 57. Hu, F.; Wei, F.; Wang, Y.L.; Wu, B.B.; Fang, Y.; Xiong, B. EGCG synergizes the therapeutic effect of cisplatin and oxaliplatin through autophagic pathway in human colorectal cancer cells. *J. Pharmacol. Sci.* 2015, 128, 27–34, doi:10.1016/j.jphs.2015.04.003.
 58. Irimie, A.I.; Braicu, C.; Zanoaga, O.; Pileczki, V.; Gherman, C.; Berindan-Neagoe, I.; Campian, R.S. Epigallocatechin-3-gallate suppresses cell proliferation and promotes apoptosis and autophagy in oral cancer SSC-4 cells. *Oncotargets Ther.* 2015, 8, 461–470, doi:10.2147/Ott.S78358.
 59. Zhou, J.; Farah, B.L.; Sinha, R.A.; Wu, Y.; Singh, B.K. Epigallocatechin-3-Gallate (EGCG), a Green Tea Polyphenol, Stimulates Hepatic Autophagy and Lipid Clearance (vol 9, e87161, 2014). *PLoS ONE* 2014, 9, doi:10.1371/journal.pone.0096884.
 60. Gray, A.L.; Stephens, C.A.; Bigelow, R.L.H.; Coleman, D.T.; Cardelli, J.A. The Polyphenols (-)-Epigallocatechin-3-Gallate and Luteolin Synergistically Inhibit TGF-beta-Induced Myofibroblast Phenotypes through RhoA and ERK Inhibition. *PLoS ONE* 2014, 9, e109208, doi:10.1371/journal.pone.0109208.
 61. Chen, A.P.; Zhang, L. The antioxidant (-)-epigallocatechin-3-gallate inhibits rat hepatic stellate cell proliferation in vitro by blocking the tyrosine phosphorylation and reducing the gene expression of platelet-derived growth factor-beta receptor. *J. Biol. Chem.* 2003, 278, 23381–23389, doi:10.1074/jbc.M212042200.
 62. Chen, L.L.; Xu, Y. Epigallocatechin gallate attenuates uric acid-induced injury in rat renal interstitial fibroblasts NRK-49F by up-regulation of miR-9. *Eur. Rev. Med. Pharmacol.* 2018, 22, 7458–7469.
 63. Adhami, V.M.; Syed, D.N.; Khan, N.; Mukhtar, H. Dietary flavonoid fisetin: A novel dual inhibitor of PI3K/Akt and mTOR for prostate cancer management. *Biochem. Pharmacol.* 2012, 84, 1277–1281, doi:10.1016/j.bcp.2012.07.012.

64. Sun, X.; Ma, X.M.; Li, Q.W.; Yang, Y.; Xu, X.L.; Sun, J.Q.; Yu, M.W.; Cao, K.X.; Yang, L.; Yang, G.W.; et al. Anti-cancer effects of fisetin on mammary carcinoma cells via regulation of the PI3K/Akt/mTOR pathway: In vitro and in vivo studies. *Int. J. Mol. Med.* 2018, 42, 811–820, doi:10.3892/ijmm.2018.3654.
65. Khan, N.; Afaq, F.; Khusro, F.H.; Adhami, V.M.; Suh, Y.; Mukhtar, H. Dual inhibition of phosphatidylinositol 3-kinase/Akt and mammalian target of rapamycin signaling in human nonsmall cell lung cancer cells by a dietary flavonoid fisetin. *Int. J. Cancer* 2012, 130, 1695–1705, doi:10.1002/ijc.26178.
66. Suh, Y.; Afaq, F.; Khan, N.; Johnson, J.J.; Khusro, F.H.; Mukhtar, H. Fisetin induces autophagic cell death through suppression of mTOR signaling pathway in prostate cancer cells. *Carcinogenesis* 2010, 31, 1424–1433, doi:10.1093/carcin/bgq115.
67. Yang, P.M.; Tseng, H.H.; Peng, C.W.; Chen, W.S.; Chiu, S.J. Dietary flavonoid fisetin targets caspase-3-deficient human breast cancer MCF-7 cells by induction of caspase-7-associated apoptosis and inhibition of autophagy. *Int. J. Oncol.* 2012, 40, 469–478, doi:10.3892/ijo.2011.1203.
68. Tiwari, R.V.; Parajuli, P.; Sylvester, P.W. gamma-Tocotrienol-induced endoplasmic reticulum stress and autophagy act concurrently to promote breast cancer cell death. *Biochem. Cell. Biol.* 2015, 93, 306–320, doi:10.1139/bcb-2014-0123.
69. Deng, S.; Shanmugam, M.K.; Kumar, A.P.; Yap, C.T.; Sethi, G.; Bishayee, A. Targeting autophagy using natural compounds for cancer prevention and therapy. *Cancer-Am. Cancer Soc.* 2019, 125, 1228–1246, doi:10.1002/cncr.31978.
70. Jiang, Q.; Rao, X.Y.; Kim, C.Y.; Freiser, H.; Zhang, Q.B.; Jiang, Z.Y.; Li, G.L. Gamma-tocotrienol induces apoptosis and autophagy in prostate cancer cells by increasing intracellular dihydrosphingosine and dihydroceramide. *Int. J. Cancer* 2012, 130, 685–693, doi:10.1002/ijc.26054.
71. Tiwari, R.V.; Parajuli, P.; Sylvester, P.W. gamma-Tocotrienol-induced autophagy in malignant mammary cancer cells. *Exp. Biol. Med.* 2014, 239, 33–44, doi:10.1177/1535370213511022.
72. Kim, S.H.; Park, E.J.; Lee, C.R.; Chun, J.N.; Cho, N.H.; Kim, I.G.; Lee, S.; Kim, T.W.; Park, H.H.; So, I.; et al. Geraniol induces cooperative interaction of apoptosis and autophagy to elicit cell death in PC-3 prostate cancer cells. *Int. J. Oncol.* 2012, 40, 1683–1690, doi:10.3892/ijo.2011.1318.
73. Trzoss, L.; Fukuda, T.; Costa-Lotufo, L.V.; Jimenez, P.; La Clair, J.J.; Fenical, W. Seriniquinone, a selective anticancer agent, induces cell death by autophagocytosis, targeting the cancer-protective protein dermcidin. *Proc. Natl. Acad. Sci. USA* 2014, 111, 14687–14692, doi:10.1073/pnas.1410932111.

74. Chu, S.C.; Hsieh, Y.S.; Yu, C.C.; Lai, Y.Y.; Chen, P.N. Thymoquinone Induces Cell Death in Human Squamous Carcinoma Cells via Caspase Activation-Dependent Apoptosis and LC3-II Activation-Dependent Autophagy. *PLoS ONE* 2014, 9, doi:10.1371/journal.pone.0101579.
75. Racoma, I.O.; Meisen, W.H.; Wang, Q.E.; Kaur, B.; Wani, A.A. Thymoquinone Inhibits Autophagy and Induces Cathepsin-Mediated, Caspase-Independent Cell Death in Glioblastoma Cells. *PLoS ONE* 2013, 8, doi:10.1371/journal.pone.0072882.
76. Chen, M.C.; Lee, N.H.; Hsu, H.H.; Ho, T.J.; Tu, C.C.; Hsieh, D.J.Y.; Lin, Y.M.; Chen, L.M.; Kuo, W.W.; Huang, C.Y. Thymoquinone Induces Caspase-Independent, Autophagic Cell Death in CPT-11-Resistant LoVo Colon Cancer via Mitochondrial Dysfunction and Activation of JNK and p38. *J. Agric. Food Chem.* 2015, 63, 1540–1546, doi:10.1021/jf5054063.
77. Xu, H.L.; Tang, W.; Du, G.H.; Kokudo, N. Targeting apoptosis pathways in cancer with magnolol and honokiol, bioactive constituents of the bark of *Magnolia officinalis*. *Drug Discov. Ther.* 2011, 5, 202–210, doi:10.5582/ddt.2011.v5.5.202.
78. Li, H.B.; Yi, X.; Gao, J.M.; Ying, X.X.; Guan, H.Q.; Li, J.C. Magnolol-induced H460 cells death via autophagy but not apoptosis. *Arch. Pharm. Res.* 2007, 30, 1566–1574, doi:10.1007/Bf02977326.
79. Rasul, A.; Yu, B.; Khan, M.; Zhang, K.; Iqbal, F.; Ma, T.H.; Yang, H. Magnolol, a natural compound, induces apoptosis of SGC-7901 human gastric adenocarcinoma cells via the mitochondrial and PI3K/Akt signaling pathways. *Int. J. Oncol.* 2012, 40, 1153–1161, doi:10.3892/ijo.2011.1277.
80. Bae, U.J.; Jung, E.S.; Jung, S.J.; Chae, S.W.; Park, B.H. Mulberry leaf extract displays antidiabetic activity in db/db mice via Akt and AMP-activated protein kinase phosphorylation. *Food Nutr. Res.* 2018, 62, doi:10.29219/fnr.v62.1473.
81. Cheng, K.C.; Wang, C.J.; Chang, Y.C.; Hung, T.W.; Lai, C.J.; Kuo, C.W.; Huang, H.P. Mulberry fruits extracts induce apoptosis and autophagy of liver cancer cell and prevent hepatocarcinogenesis in vivo. *J. Food Drug Anal.* 2020, 28, 84–93, doi:10.1016/j.jfda.2019.06.002.
82. Kwon, Y.H.; Bishayee, K.; Rahman, M.A.; Hong, J.S.; Lim, S.S.; Huh, S.O. *Morus alba* Accumulates Reactive Oxygen Species to Initiate Apoptosis via FOXO-Caspase 3-Dependent Pathway in Neuroblastoma Cells. *Mol. Cells* 2015, 38, 630–637, doi:10.14348/molcells.2015.0030.
83. Lao, Y.Z.; Wan, G.; Liu, Z.Y.; Wang, X.Y.; Ruan, P.; Xu, W.; Xu, D.Q.; Xie, W.D.; Zhang, Y.; Xu, H.X.; et al. The natural compound oblongifolin C inhibits autophagic flux and enhances antitumor efficacy of nutrient deprivation. *Autophagy* 2014, 10, 736–749, doi:10.4161/auto.28034.
84. Wu, M.; Lao, Y.Z.; Tan, H.S.; Lu, G.; Ren, Y.; Zheng, Z.Q.; Yi, J.; Fu, W.W.; Shen, H.M.; Xu, H.X. Oblongifolin C suppresses lysosomal function independently of TFEB nuclear translocation. *Acta*

- Pharmacol. Sin. 2019, 40, 929–937, doi:10.1038/s41401-018-0167-7.
85. Acharya, B.R.; Bhattacharyya, S.; Choudhury, D.; Chakrabarti, G. The microtubule depolymerizing agent naphthazarin induces both apoptosis and autophagy in A549 lung cancer cells. *Apoptosis* 2011, 16, 924–939, doi:10.1007/s10495-011-0613-1.
86. Herr, I.; Lozanovski, V.; Houben, P.; Schemmer, P.; Buchler, M.W. Sulforaphane and related mustard oils in focus of cancer prevention and therapy. *Wien. Med. Wochenschr.* 2013, 163, 80–88, doi:10.1007/s10354-012-0163-3.
87. Herman-Antosiewicz, A.; Johnson, D.E.; Singh, S.V. Sulforaphane causes autophagy to inhibit release of cytochrome C and apoptosis in human prostate cancer cells. *Cancer Res.* 2006, 66, 5828–5835, doi:10.1158/0008-5472.CAN-06-0139.
88. Zheng, Z.; Lin, K.; Hu, Y.; Zhou, Y.; Ding, X.; Wang, Y.; Wu, W. Sulforaphane metabolites inhibit migration and invasion via microtubule-mediated Claudins dysfunction or inhibition of autolysosome formation in human non-small cell lung cancer cells. *Cell Death Dis.* 2019, 10, 259, doi:10.1038/s41419-019-1489-1.
89. Chaudhuri, D.; Orsulic, S.; Ashok, B.T. Antiproliferative activity of sulforaphane in Akt-overexpressing ovarian cancer cells. *Mol. Cancer Ther.* 2007, 6, 334–345, doi:10.1158/1535-7163.MCT-06-0404.
90. Uddin, M.S.; Mamun, A.A.; Jakaria, M.; Thangapandian, S.; Ahmad, J.; Rahman, M.A.; Mathew, B.; Abdel-Daim, M.M.; Aleya, L. Emerging promise of sulforaphane-mediated Nrf2 signaling cascade against neurological disorders. *Sci. Total Environ.* 2020, 707, 135624, doi:10.1016/j.scitotenv.2019.135624.
91. Kim, S.M.; Park, H.S.; Jun, D.Y.; Woo, H.J.; Woo, M.H.; Yang, C.H.; Kim, Y.H. Mollugin induces apoptosis in human Jurkat T cells through endoplasmic reticulum stress-mediated activation of JNK and caspase-12 and subsequent activation of mitochondria-dependent caspase cascade regulated by Bcl-xL. *Toxicol. Appl. Pharm.* 2009, 241, 210–220, doi:10.1016/j.taap.2009.08.024.
92. Zhang, L.; Wang, H.D.; Zhu, J.H.; Xu, J.G.; Ding, K. Mollugin induces tumor cell apoptosis and autophagy via the PI3K/AKT/mTOR/p70S6K and ERK signaling pathways. *Biochem. Biophys. Res. Commun.* 2014, 450, 247–254, doi:10.1016/j.bbrc.2014.05.101.
93. Xu, M.Y.; Lee, S.Y.; Kang, S.S.; Kim, Y.S. Antitumor Activity of Jujuboside B and the Underlying Mechanism via Induction of Apoptosis and Autophagy. *J. Nat. Prod.* 2014, 77, 370–376, doi:10.1021/np401022g.
94. Lu, J.; Sun, D.P.; Gao, S.; Gao, Y.; Ye, J.T.; Liu, P.Q. Cyclovirobuxine D Induces Autophagy-Associated Cell Death via the Akt/mTOR Pathway in MCF-7 Human Breast Cancer Cells. *J. Pharmacol. Sci.* 2014, 125, 74–82, doi:10.1254/jphs.14013FP.

95. Ouyang, L.; Chen, Y.; Wang, X.Y.; Lu, R.F.; Zhang, S.Y.; Tian, M.; Xie, T.; Liu, B.; He, G. Polygonatum odoratum lectin induces apoptosis and autophagy via targeting EGFR-mediated Ras-Raf-MEK-ERK pathway in human MCF-7 breast cancer cells. *Phytomedicine* 2014, 21, 1658–1665, doi:10.1016/j.phymed.2014.08.002.
96. Li, C.Y.; Chen, J.; Lu, B.M.; Shi, Z.; Wang, H.L.; Zhang, B.; Zhao, K.L.; Qi, W.; Bao, J.K.; Wang, Y. Molecular Switch Role of Akt in Polygonatum odoratum Lectin-Induced Apoptosis and Autophagy in Human Non-Small Cell Lung Cancer A549 Cells. *PLoS ONE* 2014, 9, e101526, doi:10.1371/journal.pone.0101526.
97. Jiang, H.; Zhang, L.; Kuo, J.; Kuo, K.; Gautam, S.C.; Groc, L.; Rodriguez, A.I.; Koubi, D.; Hunter, T.J.; Corcoran, G.B.; et al. Resveratrol-induced apoptotic death in human U251 glioma cells. *Mol. Cancer Ther.* 2005, 4, 554–561, doi:10.1158/1535-7163.Mct-04-0056.
98. Rahman, M.A.; Kim, N.H.; Kim, S.H.; Oh, S.M.; Huh, S.O. Antiproliferative and Cytotoxic Effects of Resveratrol in Mitochondria-Mediated Apoptosis in Rat B103 Neuroblastoma Cells. *Korean J. Physiol. Pharm.* 2012, 16, 321–326, doi:10.4196/kjpp.2012.16.5.321.
99. Lang, F.F.; Qin, Z.Y.; Li, F.; Zhang, H.L.; Fang, Z.H.; Hao, E.K. Apoptotic Cell Death Induced by Resveratrol Is Partially Mediated by the Autophagy Pathway in Human Ovarian Cancer Cells. *PLoS ONE* 2015, 10, doi:10.1371/journal.pone.0129196.
100. Garg, T.; Yadav, V.K. Effects of Resveratrol as an anticancer agent. A Systematic Review and Meta-Analysis. *Indian J. Pharmacol.* 2013, 45, S206–S206.
101. Gong, C.H.; Xia, H.L. Resveratrol suppresses melanoma growth by promoting autophagy through inhibiting the PI3K/AKT/mTOR signaling pathway. *Exp. Ther. Med.* 2020, 19, 1878–1886, doi:10.3892/etm.2019.8359.
102. Liu, Q.J.; Fang, Q.; Ji, S.Q.; Han, Z.X.; Cheng, W.L.; Zhang, H.J. Resveratrol-mediated apoptosis in renal cell carcinoma via the p53/AMP-activated protein kinase/mammalian target of rapamycin autophagy signaling pathway. *Mol. Med. Rep.* 2018, 17, 502–508, doi:10.3892/mmr.2017.7868.
103. Godichaud, S.; Si-Tayeb, K.; Auge, N.; Desmouliere, A.; Balabaud, C.; Payrastre, B.; Negre-Salvayre, A.; Rosenbaum, J. The grape-derived polyphenol resveratrol differentially affects epidermal and platelet-derived growth factor signaling in human liver myofibroblasts. *Int. J. Biochem. Cell Biol.* 2006, 38, 629–637, doi:10.1016/j.biocel.2005.11.001.
104. Jiang, Y.D.; Liu, L.; Steinle, J.J. miRNA15a regulates insulin signal transduction in the retinal vasculature. *Cell. Signal.* 2018, 44, 28–32, doi:10.1016/j.cellsig.2018.01.016.
105. Anand David, A.V.; Arulmoli, R.; Parasuraman, S. Overviews of Biological Importance of Quercetin: A Bioactive Flavonoid. *Pharmacogn. Rev.* 2016, 10, 84–89, doi:10.4103/0973-7847.194044.

106. Singh, B.N.; Kumar, D.; Shankar, S.; Srivastava, R.K. Rottlerin induces autophagy which leads to apoptotic cell death through inhibition of PI3K/Akt/mTOR pathway in human pancreatic cancer stem cells. *Biochem. Pharmacol.* 2012, 84, 1154–1163, doi:10.1016/j.bcp.2012.08.007.
107. Rahman, M.A.; Kim, N.H.; Yang, H.; Huh, S.O. Angelicin induces apoptosis through intrinsic caspase-dependent pathway in human SH-SY5Y neuroblastoma cells. *Mol. Cell. Biochem.* 2012, 369, 95–104, doi:10.1007/s11010-012-1372-1.
108. Rahman, M.A.; Bishayee, K.; Huh, S.O. Angelica polymorpha Maxim Induces Apoptosis of Human SH-SY5Y Neuroblastoma Cells by Regulating an Intrinsic Caspase Pathway. *Mol. Cells* 2016, 39, 119–128, doi:10.14348/molcells.2016.2232.
109. Wang, Y.R.; Chen, Y.Q.; Chen, X.D.; Liang, Y.; Yang, D.P.; Dong, J.; Yang, N.; Liang, Z.Q. Angelicin inhibits the malignant behaviours of human cervical cancer potentially via inhibiting autophagy. *Exp. Ther. Med.* 2019, 18, 3365–3374, doi:10.3892/etm.2019.7985.
110. Uddin, M.S.; Rahman, M.A.; Kabir, M.T.; Behl, T.; Mathew, B.; Perveen, A.; Barreto, G.E.; Bin-Jumah, M.N.; Abdel-Daim, M.M.; Ashraf, G.M. Multifarious roles of mTOR signaling in cognitive aging and cerebrovascular dysfunction of Alzheimer's disease. *IUBMB Life* 2020, 10.1002/iub.2324, doi:10.1002/iub.2324.
111. Leng, S.; Hao, Y.; Du, D.; Xie, S.; Hong, L.; Gu, H.; Zhu, X.; Zhang, J.; Fan, D.; Kung, H.F. Ursolic acid promotes cancer cell death by inducing Atg5-dependent autophagy. *Int. J. Cancer* 2013, 133, 2781–2790, doi:10.1002/ijc.28301.
112. Xavier, C.P.R.; Lima, C.F.; Pedro, D.F.N.; Wilson, J.M.; Kristiansen, K.; Pereira-Wilson, C. Ursolic acid induces cell death and modulates autophagy through JNK pathway in apoptosis-resistant colorectal cancer cells. *J. Nutr. Biochem.* 2013, 24, 706–712, doi:10.1016/j.jnutbio.2012.04.004.
113. Zhao, C.; Yin, S.T.; Dong, Y.H.; Guo, X.; Fan, L.H.; Ye, M.; Hu, H.B. Autophagy-dependent EIF2AK3 activation compromises ursolic acid-induced apoptosis through upregulation of MCL1 in MCF-7 human breast cancer cells. *Autophagy* 2013, 9, 196–207, doi:10.4161/auto.22805.
114. Shin, S.W.; Park, J.W. Autophagy inhibition enhances ursolic acid-induced apoptosis in PC3 cells. *Free Radic. Biol. Med.* 2012, 53, S116-S116, doi:10.1016/j.freeradbiomed.2012.08.242.
115. Wang, S.Y.; Yu, Q.J.; Bao, J.K.; Liu, B. Polygonatum cyrtonema lectin, a potential antineoplastic drug targeting programmed cell death pathways. *Biochem. Biophys Res. Commun.* 2011, 406, 497–500, doi:10.1016/j.bbrc.2011.02.049.
116. Liu, B.; Cheng, Y.; Bian, H.J.; Bao, J.K. Molecular mechanisms of Polygonatum cyrtonema lectin-induced apoptosis and autophagy in cancer cells. *Autophagy* 2009, 5, 253–255, doi:10.4161/auto.5.2.7561.
117. Liu, B.; Cheng, Y.; Zhang, B.; Bian, H.J.; Bao, J.K. Polygonatum cyrtonema lectin induces apoptosis and autophagy in human melanoma A375 cells through a mitochondria-mediated ROS-

- p38-p53 pathway. *Cancer Lett.* 2009, 275, 54–60, doi:10.1016/j.canlet.2008.09.042.
118. Duan, W.J.; Li, Q.S.; Xia, M.Y.; Tashiro, S.I.; Onodera, S.; Ikejima, T. Silibinin Activated p53 and Induced Autophagic Death in Human Fibrosarcoma HT1080 Cells via Reactive Oxygen Species-p38 and c-Jun N-Terminal Kinase Pathways. *Biol. Pharm. Bull.* 2011, 34, 47–53, doi:10.1248/bpb.34.47.
 119. Rahman, M.A.; Yang, H.; Kim, N.H.; Huh, S.O. Induction of apoptosis by *Dioscorea nipponica* Makino extracts in human SH-SY5Y neuroblastoma cells via mitochondria-mediated pathway. *Anim. Cells Syst.* 2014, 18, 41–51, doi:10.1080/19768354.2014.880372.
 120. Rahman, M.A.; Yang, H.; Lim, S.S.; Huh, S.O. Apoptotic Effects of *Melandryum firmum* Root Extracts in Human SH-SY5Y Neuroblastoma Cells. *Exp. Neurobiol.* 2013, 22, 208–213, doi:10.5607/en.2013.22.3.208.
 121. Bassham, D.C.; Crespo, J.L. Autophagy in plants and algae. *Front. Plant Sci.* 2014, 5, doi:10.3389/fpls.2014.00679.
 122. Uddin, M.S.; Mamun, A.A.; Rahman, M.A.; Kabir, M.T.; Alkahtani, S.; Alanazi, I.S.; Perveen, A.; Ashraf, G.M.; Bin-Jumah, M.N.; Abdel-Daim, M.M. Exploring the Promise of Flavonoids to Combat Neuropathic Pain: From Molecular Mechanisms to Therapeutic Implications. *Front. Neurosci.* 2020, 14, 478, doi:10.3389/fnins.2020.00478.
 123. Rahman, M.A.; Hong, J.S.; Huh, S.O. Antiproliferative properties of *Saussurea lappa* Clarke root extract in SH-SY5Y neuroblastoma cells via intrinsic apoptotic pathway. *Anim. Cells Syst.* 2015, 19, 119–126, doi:10.1080/19768354.2015.1008041.
 124. Tian, X.; Song, H.S.; Cho, Y.M.; Park, B.; Song, Y.J.; Jang, S.; Kang, S.C. Anticancer effect of *Saussurea lappa* extract via dual control of apoptosis and autophagy in prostate cancer cells. *Medicine* 2017, 96, doi:10.1097/MD.00000000000007606.
 125. Ryoo, N.; Rahman, M.A.; Hwang, H.; Ko, S.K.; Nah, S.Y.; Kim, H.C.; Rhim, H. Ginsenoside Rk1 is a novel inhibitor of NMDA receptors in cultured rat hippocampal neurons. *J. Ginseng Res.* 2020, 44, 490–495, doi:10.1016/j.jgr.2019.04.002.
 126. Ko, H.; Kim, Y.J.; Park, J.S.; Park, J.H.; Yang, H.O. Autophagy Inhibition Enhances Apoptosis Induced by Ginsenoside Rk1 in Hepatocellular Carcinoma Cells. *Biosci. Biotechnol. Biochem.* 2009, 73, 2183–2189, doi:10.1271/bbb.90250.
 127. Zecchini, S.; Serafini, F.P.; Catalani, E.; Giovarelli, M.; Coazzoli, M.; Di Renzo, I.; De Palma, C.; Perrotta, C.; Clementi, E.; Buonanno, F.; et al. Dysfunctional autophagy induced by the pro-apoptotic natural compound climacostol in tumour cells. *Cell Death Dis.* 2018, 10, doi:10.1038/s41419-018-1254-x.

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