

Litter Modification of Forest Fluxes

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Litter can modify GHG fluxes often in gas-specific ways, although there are also common mechanisms underlying its effect, which are regulated by the environmental conditions, forest management and climate change factors.

litter

forest soil

greenhouse gases

CO₂

CH₄

climate change

forest management

1. Introduction

Forest ecosystems are a critical component of the global carbon (C) budget through their ability to sequester and retain large amounts of CO₂ ^[1]. An elucidation of the functioning of forest ecosystems, including their contribution to greenhouse gas (GHG) exchange, is crucial for the development of adaptation and mitigation strategies. In Europe, forests cover about 35% of the total land area, corresponding to 227 million ha with a decreasing share of coniferous (46%), broad leaved (37%), and mixed (17%) tree species ^[2].

Forest soils influence the GHG balance, with carbon dioxide (CO₂) and methane (CH₄) as key elements of the global C cycle. Of the three major GHGs the CO₂ flux is quantitatively often the most important and its concentration has increased by 1.9 ppm yr⁻¹ in the last 10 years ^[3]. The biogenic sources of CO₂ efflux from soil are root respiration, rhizomicrobial respiration, priming-related effects, and basal respiration associated with the microbial decomposition of organic matter in soils ^[4]. Forest ecosystems also significantly contribute to the increase in CO₂ emissions through forest fires, deforestation ^{[5][6][7]}, and CO₂ release by soil microorganisms colonizing dead trees ^[7]. For many forests, soil CH₄ is another important GHG with a global warming potential (GWP) 28 times greater than that of CO₂ ^[8]. Soils may be a source or sink for CH₄ depending on the balance between CH₄ production (methanogenesis in an anaerobic environment) and CH₄ oxidation (methanotrophy in aerobic conditions) ^{[9][10]}. The process of methanotrophy has significant mitigating potential since methanotrophs can contribute to a reduction in atmospheric CH₄ (high affinity methanotrophy) on the one hand, but, on the other hand, can also oxidize the higher soil CH₄ concentrations before this reaches the atmosphere (low affinity methanotrophy) ^[11]. Methanotrophy in forest soils is of particular importance, as these soils show high activity compared to soils from other ecosystems due to the dominance of high affinity methanotrophs ^[12].

In addition to soil, the surface litter layer can make an important contribution to C and nutrient cycling in forest ecosystems ^[13], changing the soil microclimate ^{[14][15]} and affecting soil microbiota. Forest litter is a layer of dead plant material present on the soil surface ^[16], which may be a source of nutrients and energy for soil

microorganisms [17] but can also act as a bidirectional (from the atmosphere to the soil profile and vice versa) barrier to gas diffusion [3][18][19]. The presence of litter may modify soil–atmosphere fluxes of GHGs through different mechanisms. Due to the predicted increase in both atmospheric CO₂ concentration and litter fall, the importance of the litter layer as a source of C is likely to rise [20][21], as would any indirect effects associated, for instance, with litter acting as a barrier to gas diffusion. Additionally, management practices in forests, e.g., cultivation or extensive deforestation, often result in enhanced litter fall combined with soil mixing, which accelerates its decomposition and may affect CO₂ emissions [1][13][22][23][24][25][26]. Thus, the litter layer could be used as an indicator of the likely amount of trace gas emissions, such as CO₂, from the forest soil [18]. In terms of CH₄ oxidation, it has been reported that litter is more important in regulating CH₄ uptake from soil than from roots [27]. The effect of litter on CH₄ consumption by forests soils has been documented to be strongly dependent on the hydrological conditions [18][19][28]. Moreover, the regulation of soil processes and the litter layer itself may be a source or sink of GHG's [29][30][31], although this has received little attention [32].

Although litter can have a major impact on the C and GHG balance in forest ecosystems, this has not always been fully recognized. In this review we summarize the available information on the effects of litter on CO₂ emissions and CH₄ uptake in forest soils, including forest-specific impacts, environmental drivers, quantification, the influence of human activity, and the likely effects of climate change. Based on recent research, we identify a number of knowledge gaps, and directions for future research are highlighted for a better understanding of the relationship between litter and soil–atmosphere GHG (CO₂, CH₄) exchange, as part of the C cycle.

2. Litter as a Controller of GHG (CO₂, CH₄) Fluxes

Litter can modify GHG fluxes in different gas-specific ways, although there are also common mechanisms of its effect, regulated by climatic conditions, forest management and climate change factors, summarized in **Figure 1**.

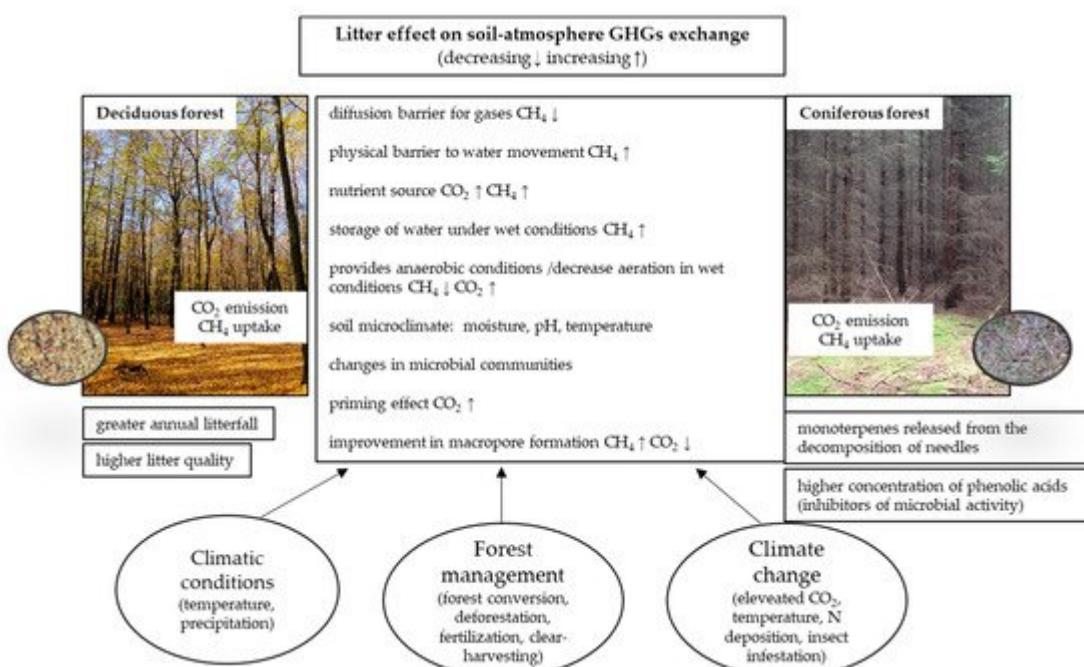


Figure 1. Diagram showing general drivers of litter effects on CO₂ emissions and CH₄ uptake, including regulating factors.

However, none of these effects are mutually exclusive and a litter-related increase in C substrate availability, for instance, could occur in concert with an alteration in the microbial communities of the soil and/or a lower soil temperature, as indicated in **Table 1**.

Table 1. The main drivers underlying the effects of litter on CO₂ emission in different forest soils from different regions based on field studies.

The Main Driver	Forest Type	Dominant Tree Species	Tree Age [Years]	Tree Height [m]	DBH [cm]	Tree Density [Trees/ha]	Litter Input [g/m ² /year]	MAT [°C]	MAP [mm]	Soil Type	Soil Texture (Sand/Silt/Clay [%])	Soil Temperature [°C]	Soil Moisture [%]	Effect of Litter on CO ₂ Fluxes	Landscape Type	Location	Ref.
Litter as a source of nutrient for microbes	Plantation	<i>T. grandis</i> (92%)	~10	n/a	10.72 ± 2.1	429	n/a	n/a	1598	n/a	n/a	28.78 ± 1.75	10.60 ± 2.42	Increased by 14.4% **	n/a	Jharkhand, Eastern India	[33]
	Plantation	<i>Eucalyptus</i> sp.	3	12	n/a	700	n/a	25	1200	Arenosol (FAO)	Sandy	~24–33	n/a	Increased **	coastal	Pointe Noire, Southwestern Congo	[30]
regrowth forest		<i>L. pubescens</i> , <i>M. sylvatica</i> , <i>V. guianensis</i> , <i>C. scrobiculata</i> (all species represent 71% of all stems in the stand)	12	4.9 ± 0.4	n/a	21,300	n/a	24–27	2539 ± 280	Dystrophic Yellow Latosol Stony Phase I (Brazilian Classification), Sombriustox (U.S. Soil Taxonomy)	Sandy clay loam (74/6/20)	n/a	n/a	Increased by 28% **	n/a	Northern Brazil (1°19' S, 47°57' W)	[34]
	Plantation	<i>Ac. mangium</i>	8	23.6	22.5	n/a	20–270 (fresh litter); 780–1130 (decayed litter); 1050–1160 (fresh + decayed litter) in wet and dry season	27.3	2750	Acrisols (WRB 1998)	n/a	n/a	55.5–66.3% WFPS	Increased *	Undulating topography (upper and lower plateau, upper and foot slope)	South Sumatra (3°52'40" S, 103°58'40" E) Indonesia	[35]
Pine forest		<i>P. massoniana</i>	30	5	n/a	2600	n/a	17.8	1785	Ferric Acrisols (USDA soil taxonomy)	Loamy clay (21/43/36)	24.2	60.3% WFPS	Increased by 24–32% *	Hilly region	Yingtan, Jiangxi Province, China	[31]

The Main Driver	Forest Type	Dominant Tree Species	Tree Age [Years]	Tree Height [m]	DBH [cm]	Tree Density [Trees/ha]	Litter Input [g/m ² /year]	MAT [°C]	MAP [mm]	Soil Type	Soil Texture (Sand/Silt/Clay [%])	Soil Temperature [°C]	Soil Moisture [%]	Effect of Litter on CO ₂ Fluxes	Landscape Type	Location	Ref.
Southeastern China (28°15' N, 116°55' E)																	
Sclerophyll forest	<i>Cr. alba</i> , <i>Q. saponaria</i> , <i>P. boldus</i> , <i>L. caerulea</i>	n/a	5.06 ± 0.87	6.51 ± 1.39	2600 ± 978	314 ± 30	n/a	503	Pachic Humixerepts	Sandy (62.4/26.4/11.2)	n/a	n/a	Increased by 21.2–33% **	Top slope (<4% slope)	Central Chile (34°7' S, 71°11'18" W)	[36]	
Mixed pine-broadleaf forest	<i>Cs. chinensis</i> (50.9%), <i>S. superba</i> , <i>P. massoniana</i>	100	n/a	n/a	n/a	861	22.3	1680	Ultisol (USDA soil taxonomy)	Lateritic	17.5–24.0	20.8–27.8	Increased by 33–38% **	n/a			
Pine forest	<i>P. massoniana</i> Lamb. (90%)	50	n/a	n/a	n/a	356	22.3	1680	Ultisol (USDA soil taxonomy)	Lateritic	18.2–24.8	17.1–20.9	Increased by 37–42% **	n/a		Dinghushan Biosphere Reserver, Southern China (23°0'9"21" N–23°1'30" N, 112°30'39" E–112°33'41" E)	[13]
Monsoon evergreen broadleaf forest	<i>Cs. chinensis</i> ; <i>Cr. chinensis</i> , <i>S. superba</i> , <i>Cr. concinna</i> , <i>Ap. yunnanensis</i> , <i>Ac. acuminatissima</i> , <i>G. subaequalis</i> (all these species represent >60% of the community biomass)	>400	n/a	n/a	n/a	849	22.3	1680	Ultisol (USDA soil taxonomy)	Lateritic	16.2–23.1	26.4–28.7	Increased by 29–35% **	n/a			
Enhancement of anaerobic conditions by litter	Plantation	<i>Pl. orientalis</i>	n/a	n/a	n/a	n/a	n/a	15.7	834	Yellow brown soil	Silty clay (11/41/48)	16.31 ± 1.05	12.34 ± 0.80	Increased by 18.84% *	n/a	Danjiangkou Reservoir, Central China (32°45' N, 111°13' E)	[37]
Soil moisture retention by litter	Mediterranean oak woodland	<i>Qr. agrifolia</i>	n/a	n/a	n/a	n/a	n/a	19	180	n/a	Gravelly loam	n/a	n/a	Increased by 34.2–44.8% ***	n/a	Santa Monica Mountains, California (34°0'53" N, 118°39'26" W) USA	[38]

The Main Driver	Forest Type	Dominant Tree Species	Tree Age [Years]	Tree Height [m]	DBH [cm]	Tree Density [Trees/ha]	Litter Input [g/m ² /year]	MAT [°C]	MAP [mm]	Soil Type	Soil Texture (Sand/Silt/Clay [%])	Soil Temperature [°C]	Soil Moisture [%]	Effect of Litter on CO ₂ Fluxes	Landscape Type	Location	Ref.
	Montane cloud forest	<i>Clusiaceae, Cunoniaceae, Myrsinaceae, Rosaceae, Clethraceae</i> families	n/a	n/a	n/a	n/a	n/a	12.5	n/a	n/a	Acidic	n/a	n/a	No effect **	n/a	Peruvian Andes (13°11'28" S, 71°35'24" E) [39]	
	Pine forest	<i>P. massoniana</i>	50	n/a	n/a	n/a	356	22.3	1680	Ultisol (USDA soil taxonomy)	Lateritic	18.2–24.8	17.1–20.9	Increased by 37–42% **	n/a	Dinghushan Biosphere Reserver, Southern China (23°09'21" N–23°11'30" N, 112°30'39" E–112°33'41" E) [43]	
	Mixed deciduous forest	<i>Ar. rubrum, Qr. rubra</i>	n/a	n/a	n/a	n/a	n/a	8.5	1050	Typic Dystrochrept	Fine sandy loam	n/a	n/a	Increased **	n/a	Harvard Forest, Petersham, Massachusetts (42°32' N, 72°11' W) USA [40]	
	Old-growth semideciduous tropical forest	n/a	n/a	n/a	>35	n/a	n/a	n/a	>2000	n/a	n/a	n/a	n/a	Increased **	Flat plateau (the planalto)	Pará, Northern Brazil (3°0'37" S, 54°3'45" W) [41]	
Priming effect	Old-growth moist lowland tropical forest	n/a	n/a	n/a	n/a	n/a	n/a	27	2600	Oxisol	n/a	n/a	n/a	Increased by 20% **	n/a	Gigante Peninsula, central Panama (9°06' N, 79°54' W) [42]	
	Undisturbed old-growth forest	<i>Ts. heterophylla, Ps. menziesii</i>	n/a	n/a	n/a	n/a	n/a	8.7	2370	Typic Hapludands	Coarse loamy	9.5	29	Increased **	n/a	H.J. Andrews Experimental Forest, Oregon (44°13' N, 122°13' W) USA [23]	
	Undisturbed old-growth forest	<i>P. menziesii, T. heterophylla</i>	n/a	n/a	n/a	n/a	n/a	8.7	2370	Typic Hapludands	Coarse loamy (13% clay)	9.5	29	Increased by 19% and 58% **	n/a	H.J. Andrews Experimental Forest, Oregon [43]	

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(44°15' N, 122°10' W) USA																	
Temperate deciduous forest		<i>Q. petraea</i> (70%), <i>Cp. betulus</i> (30%)	100–150	n/a	n/a	n/a	n/a	10.7	680	Gleyic Luvisol (WRB 2006)	Loam (41.9/38.8/19.3)	2.7 ± 0.5	20.4 ± 0.6	Increased **	n/a	Barbeau National Forest, Northern Central France (48°29' N, 02°47' E)	[20]
Mixed deciduous temperate woodland		<i>Ar. pseudoplatanus</i> , <i>Fr. excelsior</i>	n/a	n/a	n/a	n/a	n/a	10	714	Stagni-vertic Cambisol (FAO/WRB)	Clay loam	n/a	n/a	Increased by 30% **	n/a	Wytham Woods, Oxfordshire (51°43'42" N, 1°19'42" W) UK	[44]
Semi-deciduous lowland tropical forest		<i>Arecales</i> , <i>Burseraceae</i> , <i>Olacaceae</i> families	200	n/a	≥10	n/a	n/a	27	2600	Clay-rich Oxisol	n/a	n/a	n/a	Increased by 10% **	n/a	Gigante Peninsula, central Panama (9°06' N, 79°54' W)	[44]
Litter can act as an insulating layer that also buffers the effects of variations in light, temperature and irradiation	Temperate deciduous forest	<i>Qt. petraeae-ceris</i> community	n/a	n/a	n/a	n/a	2930	10.7	615.6	Brown forest soil, Cambisol (FAO)	n/a	9.94	25.4	Reduction ****	n/a	Bük Mountains, Northeastern Hungary (47°55' N, 20°26' E)	[21]
	Temperate deciduous forest	<i>Qt. petraeae-ceris</i> community	n/a	n/a	n/a	n/a	2754 ± 206 kg C ha ⁻¹ yr ⁻¹	10.8	599	Cambisol	n/a	11.4 ± 0.93–16.1 ± 0.78	12.8 ± 0.78–28.4 ± 1.39% v/v (soil)	Reduction ****	n/a	Bük Mountains, Northeastern Hungary (47°55' N, 20°26' E)	[45]

T.—Tectona; Ac.—Acacia; L.—Lacistema; M.—Myrcia; V.—Vismia; C.—Cupania; P.—Pinus; Pc.—Picea; Cr.—Cryptocarya; Cs.—Castanopsis; Q—Quillaja; Pe.—Peumus; L.—Lithraea; S.—Schima; Ap.—Aporosa; Ac.—Acmena; G.—Gironniera; Pl.—Placycladus; Qr—Quercus; Ar.—Acer; Ts.—Tsuga; Cp.—Carpinus; Fr.—Fraxinus; Qt.—Quercetum; Ps.—Pseudotsuga; DBH—diameter at breast height; n/a—data not available; Methods: * gas chromatography (GC); ** infrared gas analyzer (IRGA); *** quantum cascade laser (QCL) spectrometer **** soda-lime technique.

The diffusion of CH₄ from the atmosphere into the soil strongly affects CH₄ consumption, since the upper-most well-aerated mineral soil located immediately underneath the organic layer exhibits the highest methanotrophic activity [28][46]. The mechanisms associated with the effects of litter atmospheric CH₄ uptake by soils are regulated by moisture [19][27][32], as CH₄ diffusion is 10–4 times slower in water than in air [47].

It is difficult to quantify the effects of litter or litter quality on GHG emissions because of the paucity of reliable data. Two examples (the Figure A1 in Appendix A) indicate that the effects of litter can vary depending on the GHG under consideration, with a positive effect of litter amount on CO₂ fluxes but a negative effect on CH₄ fluxes, assuming that litter thickness reflects the amount of litter present. The correlations are significant but could be influenced by a range of other factors, e.g., variation in soil conditions, tree species, and climate zones.

3. Tree Species-Specific Mechanisms of the Effects of Litter on GHG Fluxes

The contribution of the litter layer to respiration ranged from 5% to 45% of the total CO₂ emissions from temperate forests soils [3]. The emissions of CO₂ were confirmed to be often lower in coniferous forest soils than in deciduous forests [48]. A meta-analysis revealed that natural and doubled litter inputs increased soil respiration in forest ecosystems by 36% and 55%, respectively. The effect of litter inputs on the increase in soil respiration in different types of forests was of the following order: coniferous forests (50.7%) > broad-leaved forests (41.3%) > mixed forests (31.9%) [49]. In coniferous forests, the removal of litter caused a reduction in CO₂ emissions, ranging from 2.61% in a fir forest in Poland to 68% in a *Pinus caribae* plantation in Puerto Rico [50]. After the litter layer removal in a pine forest, CO₂ emissions were reduced by 43%, while CH₄ uptake increased more than twofold under dry and warm soil conditions [51].

Broad leaf forests were found to have a relatively higher mean annual litter fall and a higher litter quality compared to mixed or pine forests [52][53]. The removal of litter reduced CO₂ emissions, to varying extents, depended on the type of deciduous tree species. In hornbeam oak forests, soil respiration decreased only slightly: from 2.88 kg CO₂ m⁻² year⁻¹ (with litter research point) to 2.78 kg CO₂ m⁻² year⁻¹ (litter-free research point), but in the acidophilous beech forest, CO₂ emissions decreased from 2.18 kg CO₂ m⁻² year⁻¹ to 1.32 kg CO₂ m⁻² year⁻¹ after excluding litter. The amount of CO₂ emitted from forest soils also depended on the rate of litter decomposition, which differed in different types of forests [54]. A beech stand was found to have the slowest litter decomposition, and its accumulation was approximately two to three times higher than in mixed stands of deciduous tree species [55]. Similarly, in a hornbeam oak forest litter decomposition processes were faster and CO₂ was released more rapidly than in a beech forest. It was estimated that the average decrease in soil respiration globally after litter removal was 27% for different types of forests. The litter decomposition rate, along with soil respiration decreased after litter removal, in a seasonally flooded forest and an upland forest where litter removal resulted in a 10–20% reduction in soil respiration [56]. Therefore, it can be concluded that the rate of litter decomposition made a significant contribution to differences in soil CO₂ emissions between various ecosystems. Tropical forests were very important in this context [42], and may have contributed about 67% to the total annual global CO₂ efflux [56]. These forests could react differently to litter manipulations, since they differed from temperate forests in terms of

soil age, biotic composition, erosion, and/or uplift rates [57][58]. Tropical and subtropical forests may also have varied significantly in soil abiotic (e.g., soil moisture and temperature) and biotic (e.g., litter quantity and quality, tree species) factors, which also influenced the impact of litter inputs on CO₂ emissions [50][57][59][60].

Similar to CO₂ emissions, forest types differ in soil CH₄ uptake ability. The high consumption of atmospheric CH₄ by forest soils confirms the involvement of high affinity methanotrophs [12] and the process is carried out by different groups of methanotrophs [61]. Among the most abundant methanotrophs, *Methylocystis* spp. and *Methylococcus* more often populate deciduous forest soils than mixed and coniferous forest soils [62]. A number of studies conducted under the same climatic conditions also indicate that tree species affect CH₄ uptake in forest soils, with deciduous forests consuming more CH₄ than coniferous forests [28][40][63][64]. One of the explanations for this is that it is due to vegetation and soil-related differences in the structure and activity of methanotroph communities [62][65]. Of the factors that could be important, litter and soils from coniferous forests have a lower pH than deciduous stands, typically ranging between 3 and 4 in pine-dominated forests [66][67][68][69]. Such conditions are below the optimum typical level for methanotrophs [70] and may result in a lower CH₄ uptake; however, some methanotrophs have adapted to such conditions in forest soils [71]. A study on different temperate tree species shows that soil under beech trees is more acidic and has lower inputs of Ca and Mg via litter in comparison with mixed stands of deciduous tree species [72][55].

The properties of the litter itself are also an important factor. Litter in deciduous forests is characterized by a higher degradability than in coniferous forests, which results in higher soil N turnover rates [73]. Strong interactions between CH₄ oxidation and soil N have also been reported [74][75][76]. In temperate forests, N fertilization is reported to reduce the CH₄ sink [77] due to a salt effect [78] or a higher nitrification rate [79]. In subtropical forests, N deposition can suppress CH₄ uptake by altering methanotroph and methanogen abundance, diversity, and community structure [80].

4. Environmental Controllers of the Impact of Litter on GHG Fluxes

Consideration of the litter effect on soil GHG fluxes should include the role of climatic conditions, mainly temperature and precipitation. Due to the dynamics of these parameters, the litter effect on GHG fluxes may show significant seasonal variability.

In an oak forest rainfall is reported to increase CO₂ emissions, which results in the rapid reactivation of litter-associated microorganisms [38]. On the other hand, in a tropical montane cloud forest in Peru, lower moisture levels do not change the soil respiration after litter removal, which is explained by the fact that the litter and organic matter are decomposed by microorganisms with different moisture sensitivities [39]. Litter respiration also depends largely on moisture content, and the contribution of litter to soil respiration is influenced by the frequency and amount of precipitation [38][40][81][82]. Warm and humid climatic conditions accelerate organic matter decomposition, resulting in increased rates of soil respiration [83]. Changes in the water content of the litter layer are often transient, since litter is directly exposed to wind and solar radiation [84]; nevertheless, they can influence CO₂ emissions, and

therefore the overall forest CO₂ budget [85]. Continuous cycles of wetting and drying of the litter layer led to transient CO₂ emissions [84][86]. An increase in CO₂ emissions followed by an increase in the litter moisture content due to rainfall was also reported in a semi-deciduous old-growth tropical forest (with mostly evergreen species) [41] and in a forest dominated by *Quercus serrata* Thunb. [84]. Studies carried out in coniferous and deciduous forests in different locations in Europe (Italy, the Netherlands, and Finland) show that leaf litter is the main source of CO₂ and the emissions peaked at the higher moisture contents for all types of litter, temperatures, or sites, while the optimum soil CO₂ emissions are achieved at intermediate moisture contents (40–70% WFPS) [87]. Litter removal causes a decrease in soil moisture [15] (compared to soil with litter) and when soil moisture is low, both the transport of nutrients and the metabolism of decomposing microbes are reduced [88][89].

Hydrological conditions are a strong regulator of the CH₄ cycle since saturated (flooded) soil can be a source of methane, while well-aerated soil can be a sink of this gas [10]. After a high rainfall in the temperate zone or during the wet season in a tropical climate, soil may also emit CH₄, although the net exchange between soils and the atmosphere depends on how this impacts the balance between CH₄ production and consumption [19]. The strong dependence of CH₄ oxidation on water content is confirmed as litter addition during the dry season does not significantly affect CH₄ uptake, while it decreases it by 47.1 ± 4.9% during the wet season after doubling the litter level [27]. The litter effect may result from enhanced microbial activity and/or from changes in litter quality and decomposition rate [16]. It is reported that litter may store water during rainfall events. Since water cannot penetrate the mineral soil, a high soil diffusivity is maintained [28]. However, a study on different types of needle leaf and broad leaf litters revealed that the rainfall interception storage capacity of the litter layer varied with physical features and rainfall characteristics [90]. The interception-related storage capacity of needle leaf litters varied significantly with the litter type, while there were no significant differences in water storage across the broad leaf litter types. It was reported that a higher intensity or longer duration of rainfall events could increase the interception storage capacity in all broad leaf and needle leaf litters [90].

The litter layer was an effective insulator, isolating the soil from the effects of variations in irradiance, consequently lowering soil temperature [21]. In a deciduous forest, litter and soil temperatures were responsible for 68% to 81% of the variability in CO₂ emissions, respectively [83]. When there was no litter on the soil surface, the influence of temperature on soil respiration was higher, the activity of soil microbes and their enzymes increased, and the degradation of organic matter was greater. Thus, under these conditions, an increase in soil CO₂ emissions could often be observed [21][91][92][93]. After the removal of litter in a *Quercetum petraeae-cerris* forest in northeastern Hungary, the soil was found to reach higher temperatures in summer and lower temperatures in winter [45]. The greatest reductions in CO₂ emissions after litter exclusion were observed in a *Cinnamomum camphora* forest in China (39.2%) [94], in a beech (*Fagus sylvatica*) forest in Poland (about 39.45%) [54], and in a wet tropical forest dominated by *Tabebuia heterophylla* in Puerto Rico (54%) [50].

References

1. Lal, R. Forest soils and carbon sequestration. *For. Ecol. Manag.* 2005, 220, 242–258.
2. Forest Europe. State of Europe's Forests 2020; Forest Europe: Bonn, Germany, 2020.
3. Fan, J.; Luo, R.; McConkey, B.G.; Ziadi, N. Effects of nitrogen deposition and litter layer management on soil CO₂, N₂O, and CH₄ emissions in a subtropical pine forestland. *Nat. Sci. Rep.* 2020, 10, 8959.
4. Kuzyakov, Y. Sources of CO₂ efflux from soil and review of partitioning methods. *Soil Biol. Biochem.* 2006, 38, 425–448.
5. Baccini, A.; Goetz, S.J.; Walker, W.S.; Laporte, N.T.; Sun, M.; Sulla-Menashe, D.; Hackler, J.; Beck, P.S.A.; Dubayah, R.; Friedl, M.A.; et al. Estimated carbon dioxide emissions from tropical deforestation improved by carbon-density maps. *Nat. Clim.* 2012, 2, 182–185.
6. Carvalho, J.A., Jr.; Amaral, S.S.; Costa, M.A.M.; Soares Neto, T.G.; Veras, C.A.G.; Costa, F.S.; Van Leeuwen, T.T.; Krieger Filho, G.C.; Tourigny, E.; Forti, M.C.; et al. CO₂ and CO emission rates from three forest fire controlled experiments in Western Amazonia. *Atmos. Environ.* 2016, 135, 73–83.
7. Waheed, R.; Chang, D.; Sarwar, S.; Chen, W. Forest, agriculture, renewable energy, and CO₂ emission. *J. Clean. Prod.* 2018, 172, 4231–4238.
8. IPCC. Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change; IPCC: Geneva, Switzerland, 2014.
9. Le Mer, J.; Roger, P. Production, oxidation, emission and consumption of methane by soils: A review. *Eur. J. Soil Biol.* 2001, 37, 25–50.
10. Wnuk, E.; Walkiewicz, A.; Bieganowski, A. Methanogenesis and aerobic methanotrophy in arable soils contaminated with cadmium. *Catena* 2020, 189, 104480.
11. Reay, D.S.; Nadwell, D.B. Methane oxidation in temperate soils: Effects of inorganic N. *Soil Biol. Biochem.* 2004, 36, 2059–2065.
12. Nazaries, L.; Murrell, J.C.; Millard, P.; Baggs, L.; Singh, B.K. Methane, microbes implication for landscape-integrated CH₄ budget. *Glob. Change Biol.* 2013, 23, 966–976.
13. Han, T.; Huang, W.; Liu, J.; Zhou, G.; Xiao, Y. Different soil respiration responses to litter manipulation in three subtropical successional forests. *Sci. Rep.* 2015, 5, 18166.
14. Sayer, E.J. Using experimental manipulation to assess the roles of leaf litter in the function of forest ecosystems. *Biol. Rev.* 2006, 81, 1–31.
15. Xu, S.; Liu, L.L.; Sayer, E.J. Variability of above-ground litter inputs alters soil physicochemical and biological processes: A meta-analysis of litterfall-manipulation. *Biogeosciences* 2013, 10,

7423–7433.

16. Krishna, M.P.; Mohan, M. Litter decomposition in forest ecosystems: A review. *Energ. Ecol. Environ.* 2017, 2, 236–249.
17. Thoms, C.; Gleixner, G. Seasonal differences in tree species' influence on soil microbial communities. *Soil Biol. Biochem.* 2013, 66, 239–248.
18. Cheng, J.; Lee, X.; Zhou, Z.; Wang, B.; Xing, Y.; Cheng, H.; Tang, Y. The effects of litter layer and soil properties on the soil-atmosphere fluxes of greenhouse gases in karst forest, southwest China. *Pol. J. Ecol.* 2013, 61, 79–92.
19. Wang, Y.; Wang, H.; Ma, Z.; Dai, X.; Wen, X.; Liu, Y.; Wang, Z.L. The litter layer acts a moisture-induced bidirectional buffers for atmospheric methane uptake by soil of a subtropical pine plantation. *Soil Biol. Biochem.* 2013, 66, 45–50.
20. Prévost-Bouré, N.C.; Soudani, K.; Damesin, C.; Berveiller, D.; Lata, J.C.; Dufrêne, E. Increase in aboveground fresh litter quantity over-stimulates soil respiration in a temperate deciduous forest. *Appl. Soil Ecol.* 2010, 46, 26–34.
21. Kotroczo, Z.; Veres, Z.; Biró, B.; Tóth, J.A.; Fekete, I. Influence of temperature and organic matter content on soil respiration in a deciduous oak forest. *Eurasian, J. Soil Sci.* 2014, 3, 303–310.
22. Holmes, K.W.; Chadwick, O.A.; Kyriakidis, P.C.; Silva de Filho, E.P.; Soares, J.V.; Roberts, D.A. Large-area spatially explicit estimates of tropical soil carbon stocks and response to land-cover change. *Glob. Biogeochem. Cycles* 2006, 20, GB3004.
23. Crow, S.E.; Lajtha, K.; Bowden, R.D.; Yano, Y.; Brant, J.B.; Caldwell, B.A.; Sulzman, E.W. Increased coniferous needle inputs accelerate decomposition of soil carbon in an old-growth forest. *For. Ecol. Manag.* 2009, 258, 2224–2232.
24. Huang, W.; Spohn, M. Effects of long-term litter manipulation on soil carbon, nitrogen, and phosphorus in a temperate deciduous forest. *Soil Biol. Biochem.* 2015, 83, 12–18.
25. Barrena, I.; Menéndez, S.; Duñabeitia, M.; Merino, P.; Stange, C.F.; Spott, O.; González-Muria, C.; Estavillo, J.M. Greenhouse gas fluxes (CO₂, N₂O and CH₄) from forest soils in the Basque Country: Comparison of different tree species and growth stages. *For. Ecol. Manag.* 2013, 310, 600–611.
26. Mjöfors, K.; Strömgren, M.; Nohrstedt, H.Ö.; Gärdenäs, A.I. Impact of site-preparation on soil-surface CO₂ fluxes and litter decomposition in a clear-cut in Sweden. *Silva Fenn.* 2015, 49, 1–20.
27. Wu, J.; Lu, M.; Feng, J.; Zhang, D.; Chen, Q.; Li, Q.; Long, C.; Zhang, Q.; Cheng, X. Soil net methane uptake rates in response to short-term litter input change in a coniferous forest ecosystem of central China. *Agric. For. Meteorol.* 2019, 271, 307–315.

28. Borken, W.; Beese, F. Methane and nitrous oxide fluxes of soils in pure and mixed stands of European beech and Norway spruce. *Eur. J. Soil Sci.* 2006, 57, 617–625.

29. Eickenscheidt, N.; Brumme, R. Regulation of N₂O and NO_x emission patterns in six acid temperate beech forest soils by soil gas diffusivity, N turnover, and atmospheric NO_x concentrations. *Plant Soil* 2013, 369, 515–529.

30. Epron, D.; Nouvellon, Y.; Roupsard, O.; Mouvondy, W.; Mabiala, A.; Saint-André, L.; Joffre, R.; Jourdan, C.; Bonnefond, J.; Berbigier, P.; et al. Spatial and temporal variations of soil respiration in a Eucalyptus plantation in Congo. *For. Ecol. Manag.* 2004, 202, 149–160.

31. Wang, Y.; Chen, H.; Zhu, Q.; Peng, C.; Wu, N.; Yang, G.; Zhu, D.; Tian, J.; Tian, L.; Kang, X.; et al. Soil methane uptake by grasslands and forests in China. *Soil Biol. Biochem.* 2014, 74, 70–81.

32. Leitner, S.; Sae-Tun, O.; Kranzinger, L.; Zechmeister-Boltenstern, S.; Zimmermann, M. Contribution of litter layer to soil greenhouse gas emissions in a temperate beech forest. *Plant Soil* 2016, 403, 455–469.

33. Mukhopadhyay, S.; Maiti, S.K. Soil CO₂ flux in grasslands, afforested land and reclaimed coalmine overburden dumps: A case study. *Land Degrad. Develop.* 2014, 25, 216–227.

34. Vasconcelos, S.S.; Zarin, D.J.; Capanu, M.; Littell, R.; Davidson, E.A.; Ishida, F.Y.; Santos, E.B.; Araújo, M.M.; Aragão, D.V.; Rangel-Vasconcelos, L.G.T.; et al. Moisture and substrate availability constrain soil trace gas fluxes in an eastern Amazonian regrowth forest. *Glob. Biogeochem. Cycle* 2004, 18, GB2009.

35. Konda, R.; Ohta, S.; Ishizuka, S.; Heriyanto, J.; Wicaksono, A. Seasonal changes in the spatial structures of N₂O, CO₂, and CH₄ fluxes from *Acacia mangium* plantation soils in Indonesia. *Soil Biol. Biochem.* 2010, 42, 1512–1522.

36. Fuentes, J.P.; Bown, H.E.; Perez-Quezada, J.F.; Franck, N. Litter removal in a sclerophyll forest: Short- and medium-term consequences for soil properties. *Soil Sci. Soc. Am. J.* 2014, 78, 634–644.

37. Dou, X.; Zhou, W.; Zhang, Q.; Cheng, X. Greenhouse gas (CO₂, CH₄, N₂O) emissions from soils following afforestation in central China. *Atmos. Environ.* 2016, 126, 98–106.

38. Sun, W.; Maseyk, K.; Lett, C.; Seibt, U. Litter dominates surface fluxes of carbonyl sulfide in a Californian oak woodland. *J. Geophys. Res. Biogeo.* 2016, 121, 438–450.

39. Zimmermann, M.; Meir, P.; Bird, M.; Malhi, Y.; Ccahuana, A. Litter contribution to diurnal and annual soil respiration in a tropical montane cloud forest. *Soil Biol. Biochem.* 2009, 41, 1338–1340.

40. Borken, W.; Davidson, E.A.; Savage, K.; Gaudinski, J.; Trumbore, S.E. Drying and wetting effects on carbon dioxide release from organic horizons. *Soil Sci. Soc. Am. J.* 2003, 67, 1888–1896.

41. Goulden, M.L.; Miller, S.C.; Da Rocha, H.R.; Menton, M.C.; De Freitas, H.C.; Silva Figueira, M.; Dias de Sousa, C.A. Diel and seasonal patterns of tropical forest CO₂ exchange. *Ecol. Appl.* 2004, 14, S42–S54.

42. Sayer, E.J.; Powers, J.S.; Tanner, E.V.J. Increased litterfall in tropical forests Boosts the transfer of soil CO₂ to the atmosphere. *PLoS ONE* 2007, 2, e1299.

43. Sulzman, E.W.; Brant, J.B.; Bowden, R.D.; Lajtha, K. Contribution of aboveground litter, belowground litter, and rhizosphere respiration to total soil CO₂ efflux in an old growth coniferous forest. *Biogeochemistry* 2005, 73, 231–256.

44. Bréchet, L.M.; Lopez-Sangil, L.; George, C.; Birkett, A.J.; Baxendale, C.; Castro Trujillo, B.; Sayer, E.J. Distinc responses of soil respiration to experimental litter manipulation in temperate woodland and tropical forest. *Ecol. Evol.* 2018, 8, 3787–3796.

45. Fekete, I.; Kotroczó, Z.; Varga, C.; Nagy, P.T.; Várbíró, G.; Bowden, R.D.; Tóth, J.A.; Lajtha, K. Alterations in forest detritus inputs influence soil carbon concentration and soil respiration in a Central-European deciduous forest. *Soil Biol. Biochem.* 2014, 74, 106–114.

46. Price, S.J.; Sherlock, R.R.; Kelliher, F.M.; Mcseveny, T.M.; Tate, K.R.; Condron, L.M. Pristine New Zealand forest soil is a strong methane sink. *Glob. Chang. Biol.* 2004, 10, 16–26.

47. Marrero, T.R.; Mason, E.A. Gaseous diffusion coefficients. *J. Phys. Chem. Ref.* 1972, 1, 3–118.

48. Longdoz, B.; Yernaux, M.; Aubinet, M. Soil CO₂ efflux measurements in a mixed forest: Impact of chamber disturbances, spatial variability and seasonal evolution. *Glob. Change Biol.* 2000, 6, 907–917.

49. Zhang, Y.; Zou, J.; Meng, D.; Dang, S.; Zhou, J.; Osborne, B.; Ren, Y.; Liang, T.; Yu, K. Effect of soil microorgnisms and labile C availability on soil respiration in response to litter inputs in forest ecosystems: A meta-analysis. *Ecol. Evol.* 2020, 10, 13602–13612.

50. Li, Y.; Xu, M.; Sun, O.J.; Cui, W. Effects of root and litter exclusion on soil CO₂ efflux and microbial biomass in wet tropical forests. *Soil Biol. Biochem.* 2004, 36, 2111–2114.

51. Peichl, M.; Arain, M.A.; Ullah, S.; Moore, T.R. Carbon dioxide, methane, and nitrous oxide exchanges in an age-sequence of temperate pine forests. *Glob. Change Biol.* 2010, 16, 2198–2212.

52. Zhou, G.; Guan, L.; Wei, X.; Zhang, D.; Zhang, Q.; Yan, J.; Wen, D.; Liu, J.; Liu, S.; Huang, Z.; et al. Litterfall production along successional and altitudinal gradients of subtropical monsoon evergreen broadleaved forests in Guangdong, China. *Plant Ecol.* 2007, 188, 77–89.

53. Chen, H.; Gurmesa, G.A.; Liu, L.; Zhang, T.; Fu, S.; Liu, Z.; Dong, S.; Ma, C.; Mo, J. Effects of litter manipulation on litter decomposition in a successional gradients of tropical forests in southern China. *PLoS ONE* 2014, 9, e99018.

54. Wroński, K.T. Spatial variability of CO₂ fluxes from meadow and forest soils in western part of Wzniesienia Łódzkie (Łódź Hills). *For. Res. Pap.* 2018, **79**, 45–58.

55. Guckland, A.; Flessa, H.; Prenzel, J. Controls of temporal and spatial variability of methane uptake in soils of a temperate deciduous forest with different abundance of European beech (*Fagus sylvatica* L.). *Soil Biol. Biochem.* 2009, **41**, 1659–1667.

56. Borges Pinto, O., Jr.; Vourlitis, G.L.; De Souza Carneiro, E.M.; De França Dias, M.; Hentz, C.; De Souza Nogueira, J. Interactions between vegetation, hydrology, and litter inputs on decomposition and soil CO₂ efflux of tropical forests in the Brazilian Pantanal. *Forests* 2018, **9**, 281.

57. Townsend, A.R.; Asner, G.P.; Cleveland, C.C. The biogeochemical heterogeneity of tropical forests. *Trends Ecol. Evol.* 2008, **23**, 424–431.

58. Leff, J.W.; Wieder, W.R.; Taylor, P.G.; Townsend, A.R.; Nemergut, D.R.; Grandy, A.S.; Cleveland, C.C. Experimental litterfall manipulation drives large and rapid changes in soil carbon cycling in a wet tropical forest. *Glob. Change Biol.* 2012, **18**, 2969–2979.

59. Binkley, D.; Giardina, C. Why do tree species affect soils? The Warp and Woof of tree-soil interactions. *Biogeochemistry* 1998, **42**, 89–106.

60. Hasselquist, N.J.; Allen, M.F.; Santiago, L.S. Water relations of evergreen and drought-deciduous trees along a seasonally dry tropical forest chronosequence. *Oecologia* 2010, **164**, 881–890.

61. Kolb, S.; Knief, C.; Dunfield, P.F.; Conrad, R. Abundance and activity of uncultured methanotrophic bacteria involved in the consumption of atmospheric methane in two forest soils. *Environ. Microbiol.* 2005, **7**, 1150–1161.

62. Kolb, S. The quest for atmospheric methane oxidizers in forest soils. *Environ. Microbiol. Rep.* 2009, **1**, 336–346.

63. Menyailo, O.V.; Hungate, B.A. Interactive effects of tree species and soil moisture on methane consumption. *Soil Biol. Biochem.* 2003, **35**, 625–628.

64. Degelmann, D.M.; Kolb, S.; Borken, W. Methane oxidation kinetics differ in European beech and Norway spruce soils. *Eur. J. Soil Sci.* 2009, **60**, 499–506.

65. Zeng, L.; Tian, J.; Chen, H.; Wu, N.; Yan, Z.; Du, L.; Shen, Y.; Wang, X. Changes in methane oxidation ability and methanotrophic community composition across different climatic zones. *J. Soils Sediments* 2019, **19**, 533–543.

66. Burgess-Conforti, J.R.; Moore, P.A., Jr.; Owens, P.R.; Miller, D.M.; Ashworth, A.J.; Hays, P.D.; Evans-White, M.A.; Anderson, K.R. Are soils beneath coniferous tree stands more acidic than soils beneath deciduous tree stands? *Environ. Sci. Pollut. Res. Int.* 2019, **15**, 14920–14929.

67. Lau, E.; Ahmad, A.; Steudler, P.A.; Cavanaugh, C.M. Molecular characterization of methanotrophic communities in forest soils that consume atmospheric methane. *FEMS Microbiol.*

Ecol. 2007, 60, 490–500.

68. Megonigal, J.P.; Guenther, A.B. Methane emissions from upland forest soils and vegetation. *Tree Physiol.* 2008, 28, 491–498.

69. Machacova, K.; Bäck, J.; Vanhatalo, A.; Halmeenmäki, E.; Kolari, P.; Mammarella, I.; Pumpanen, J.; Acosta, M.; Urban, O.; Pihlatie, M. *Pinus sylvestris* as a missing source of nitrous oxide and methane in boreal forest. *Sci. Rep.* 2016, 6, 23410.

70. Hanson, R.S.; Hanson, T.E. Methanotrophic bacteria. *Microbiol. Rev.* 1996, 60, 439–471.

71. Nguyen, N.L.; Yu, W.J.; Gwak, J.H.; Kim, S.J.; Park, S.J.; Herbold, C.W.; Kim, J.G.; Jung, M.Y.; Rhee, S.K. Genomic insights into the acid adaptation of novel methanotrophs enriched from acidic forest soils. *Front. Microbiol.* 2018, 9, 1982.

72. Guckland, A.; Brauns, M.; Flessa, H.; Thomas, F.; Leuschner, C. Acidity, nutrient stocks and organic matter content in soils of a temperate deciduous forest with different abundance of European beech (*Fagus sylvatica* L.). *J. Plant Nutr. Soil Sci.* 2009, 172, 500–511.

73. Ambus, P.; Zechmeister-Boltenstern, S.; Butterbach-Bahl, K. Sources of nitrous oxide emitted from European forest soils. *Biogeosciences* 2006, 3, 135–145.

74. Bodelier, P.L.E.; Laanbroek, H.J. Nitrogen as regulatory factor of methane oxidation in soils and sediments. *FEMS Microbiol. Ecol.* 2004, 47, 265–277.

75. Bodelier, P.L.E.; Steenbergh, A.K. Interactions between methane and the nitrogen cycle in light of climate change. *Curr. Opin. Env. Sustain.* 2014, 9, 26–36.

76. Mochizuki, Y.; Koba, K.; Yoh, M. Strong inhibitory effect of nitrate on atmospheric methane oxidation in forest soils. *Soil Biol. Biochem.* 2012, 50, 164–166.

77. Steudler, P.A.; Bowden, R.D.; Melillo, J.M.; Aber, J.D. Influence of nitrogen fertilization on methane uptake in temperate forest soils. *Nature* 1989, 341, 314–316.

78. Saari, A.; Rinnan, R.; Martikainen, P.J. Methane oxidation in boreal forest soils: Kinetics and sensitivity to pH and ammonium. *Soil Biol. Biochem.* 2004, 36, 1037–1046.

79. Reay, D.S.; Nedwell, D.B.; McNamara, N.; Ineson, P. Effect of tree species on methane and ammonium oxidation capacity in forest soils. *Soil Biol. Biochem.* 2005, 37, 719–730.

80. Li, Q.; Peng, H.; Zhang, J.; Li, Y.; Song, X. Nitrogen addition decreases methane uptake caused by methanotroph and methanogen imbalances in a Moso bamboo forest. *Sci. Rep.* 2021, 11, 5578.

81. Dannenmann, M.; Gasche, R.; Ledebuhr, A.; Holst, T.; Mayer, H.; Papen, H. The effect of forest management on trace gas exchange at the pedosphere-atmosphere interface in beech (*Fagus sylvatica* L.) forests stocking on calcareous soils. *Eur. J. For. Res.* 2007, 126, 331–346.

82. Hanson, P.J.; O'Neill, E.G.; Chambers, M.L.S.; Riggs, J.S.; Joslin, J.D.; Wolfe, M.H. Soil respiration and litter decomposition. In *North American Temperate Deciduous Forest Response to Changing Precipitation Regimes. Ecological Studies (Analysis and Synthesis)*; Hanson, P.J., Wullschleger, S.D., Eds.; Springer: New York, NY, USA, 2003; Volume 166, pp. 163–189.

83. Vose, J.M.; Bolstad, P.V. Biotic and abiotic factors regulating forest floor CO₂ flux across a range of forest age classes in the southern Appalachians. *Pedobiologia* 2007, 50, 577–587.

84. Ataka, M.; Kominami, Y.; Jomura, M.; Yoshimura, K.; Miyama, T.; Kosugi, Y.; Tani, M. CO₂ efflux from decomposing leaf litter stack is influenced by the vertical distribution of leaf litter water content and its temporal variation. *J. Agric. Meterol.* 2015, 71, 263–270.

85. Wilson, T.B.; Kochendorfer, J.; Meyers, T.P.; Heuer, M.; Sloop, K.; Miller, J. Leaf litter water content and soil surface CO₂ fluxes in a deciduous forest. *Agric. For. Meteorol.* 2014, 192, 42–50.

86. Ataka, M.; Kominami, Y.; Yoshimura, K.; Miyama, T.; Jomura, M.; Tani, M. In situ CO₂ efflux from leaf litter layer showed large temporal variation induced by rapid wetting and drying cycle. *PLoS ONE* 2014, 9, e108404.

87. Gritsch, C.; Egger, F.; Zehetner, F.; Zechmeister-Boltenstern, S. The effect of temperature and moisture on trace gas emissions from deciduous and coniferous leaf litter. *Biogeosciences* 2016, 121, 1339–1351.

88. Sardans, J.; Peñuelas, J. Drought decreases soil enzyme activity in a Mediterranean *Quercus ilex* L. forest. *Soil Biol. Biochem.* 2005, 37, 455–461.

89. Fekete, I.; Kotrczó, Z.; Varga, C.; Hargitai, R.; Townsend, K.; Csányi, G.; Várbiró, G. Variability of organic matter inputs affects soil moisture and soil biological parameters in a European detritus manipulation experiment. *Ecosystems* 2012, 15, 792–803.

90. Li, Q.; Lee, Y.E.; Im, S. Characterizing the interception capacity of floor litter with rainfall simulation experiments. *Water* 2020, 12, 3145.

91. Veres, Z.; Ktroczo, Z.; Magyaros, K.; Tóth, J.A.; Tóthmérész, B. Dehydrogenase activity in a litter manipulation experiment in temperate forest soil. *Acta Silv. Lignaria Hung.* 2013, 9, 25–33.

92. Beni, A.; Soki, E.; Lajtha, K.; Fekete, I. An optimized HPLC method for soil fungal biomass determination and its application to a detritus manipulation study. *J. Microbiol. Methods* 2014, 103, 124–130.

93. Buchmann, N. Biotic and abiotic factors controlling soil respiration rates in *Picea abies* stands. *Soil Biol. Biochem.* 2000, 32, 1625–1635.

94. Yan, W.; Peng, Y.; Zhan, C.; Chen, X. The manipulation of aboveground litter input affects soil CO₂ efflux in a subtropical liquidambar forest in China. *iForest* 2019, 12, 181–186.