

Value-Added Polyhydroxyalkanoates

Subjects: Biotechnology & Applied Microbiology | Biochemistry & Molecular Biology

Contributor: Brian Johnston, Grazyna Adamus, Anabel Itohowo Ekere, Marek Kowalczyk, Fideline Tchuenbou-Magaia, Iza Radecka

The synthesis of polyhydroxyalkanoates (PHAs), a bioplastic that can be used to replace traditional (petrol-based) plastics, is an important focus in today's politically and environmentally conscious society.

Keywords: polyethylene (PE) ; polypropylene (PP) ; polyhydroxyalkanoate (PHA) ; PHAS ; bioplastic ; biotechnology

1. Introduction

The synthesis of polyhydroxyalkanoates (PHAs), a bioplastic that can be used to replace traditional (petrol-based) plastics, is an important focus in today's politically and environmentally conscious society. PHAs are part of a group of organic polymers containing 3-, 4-, 5-, and 6-hydroxyalkanoic acids that are biocompatible, 100% biodegradable, and nontoxic to the environment [1][2]. PHAs can be considered a greener alternative to synthetic plastic compounds, and they can be produced by plants and various strains of bacteria (as documented later). In addition, these bioplastics can be generated through microbial fermentation using waste materials which could offer more sustainability in a closed-cycle system of carbon materials [2]. Some of the waste materials that have been used to make PHAs have included used synthetic plastics, such as PE, PS, and PP [2][3][4]. From 2022, by changing the co-monomer type and distribution in PHAs, the properties can be considered to be comparable with seven of the most profitable crude-oil-based plastics, which is estimated to be 230 million tons of plastic per year [5]. There have also been global policies put in place and capacity expansions for the next 5 years for over 1.4 million tons, so there is a lot of encouragement for the industry to adopt biomaterials [5][6]. It is also predicted that 12,000 million tons of plastic waste will be added to landfills or the natural environment by 2050 [7]. **Figure 1** shows a timeline of milestones related to bioplastic development over the last hundred years.

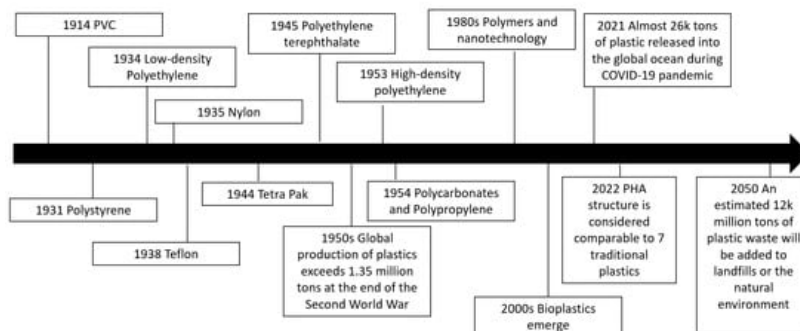


Figure 1. Milestones over the last century summarizing how bioplastics fit into the plastics industry.

Research projects around the world have focused on using microbes to break down some the most persistent types of plastics found in the environment. Currently, the most common way plastics are disposed of is by incineration, mechanical and chemical recycling, and the relatively cheap method of landfill sites [8]. However, all of these methods have their disadvantages; landfill occupies too much land space, and incineration creates secondary pollutants, such as dioxins and carbon monoxide. Even though mechanical recycling has become the main technique for refuge plastic, the chemical properties are usually compromised via processing, which results in reduced commercial value [9].

Chemical recycling is known to be able to recover monomers from plastic waste, but its success depends heavily on the efficiency of catalysts [9]. With up to 79% of waste plastic being discarded into landfills (or the surrounding environment), there is a huge requirement for novel recycling methodologies, and bioconversion is one possible answer [4][10][11]. The transfer of current feedstocks could be smoother if the true economical value (including the carbon footprint of products and practical benefits) were considered in detail. The efficiency of biotechnological methods can also be further improved using metabolic engineering, which could help achieve the aims of the internationally agreed Paris Agreement, a treaty on climate change [10]. Moreover, the recently estimated impact of the COVID-19 pandemic on plastic discharge indicates

that around 8.4 million tons of pandemic-associated plastic waste was generated from 193 countries, as of 23 August 2021, and over 25.9 thousand tons were released into the global ocean [12]. With these issues in mind, the diagram in **Figure 2** displays a possible system for generating biomaterials, such as PHAs from waste plastics via fermentation.

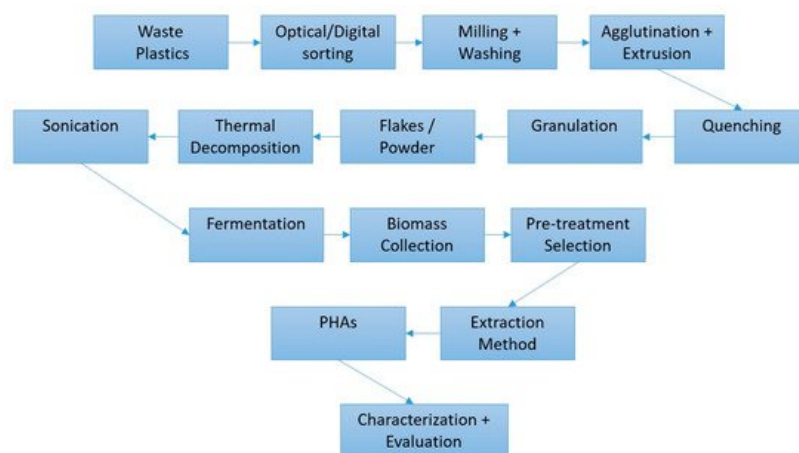


Figure 2. Potential operation stages for waste plastic material for the bio-generation of PHA bioplastics, adapted from [3]. The ‘Pre-treatment Selection’ of ‘Biomass’ process could include methods such as ionic liquid soaking, sonication, glass sphere mixing, and blade/pressure homogenization. Every procedure such as this in today’s economy should end with an evaluation step to ascertain any shortcomings and potential investigation avenues.

Elements of this kind of system exist in parts, such as the optical scanning and separation and sorting of plastics; the innovation would be in having all of these sections interlinked. In some cases, processes such as milling, agglutination, or sonication would have to be selected on the basis of the target material’s properties. After thermal pre-treatments of PS or PE, where oxygenated groups were incorporated into the unsaturated carbon backbone, sonication was found to greatly increase the mixing of plastics with the growth media for fermentation [2][3][4]. Due to the nature of microbial cultivation, requiring time (normally 48hrs or more) and optimal growth conditions (usually ranging from 30 to 37 °C at 50 to 150 rpm), a lot of energy is needed [4][10]. This is perhaps the major bottleneck for such a system, which is why the choice of micro-organisms used is so important. Additionally, any biological PHA generation has the often-unreported issue of difficulty in controlling the precise purity and biopolymer composition. Moreover, there is evidence showing that the extraction processes can alter the PHA-polymer structure when conventional chemicals (such as chloroform) and Soxhlet extraction are used [13]. The extraction yields and the PHA properties can also be affected. Data revealed the two different extraction methods alter the crystallization degree and the chemical composition. When pure bioplastic is required, pre-treatments such as homogenization provided a 15% more extractive yield than the others, especially at high pressures, which also improved the visual appearance (transparency and clearness), thermal stability, and mechanical performances, which is ideal for medical grade PHAs [13]. For packaging (the major application of PHAs), these polymers have already been proposed to effectuate a significant shift in the industry, which currently utilizes almost 40% of plastics created [14].

2. Value-Added Bioplastic Synthesis

The issues of many of the current ways of managing waste plastics (burning and landfill) can often lead to the creation of secondary pollution events. The techniques featured in **Figure 2** employ large amounts of energy, which are generally not environmentally friendly or financially viable. Due to poor recycling strategies globally and the durable properties of plastic materials, serious environmental issues, such as oceanic and soil pollution are happening [15]. For these reasons certain plastics have gained attention as carbon sources, especially those that need milder temperatures and less energy consumption for their pre-treatments.

PHAs have attracted a lot of attention mainly due to their similarities to petrochemical polymers, such as those mentioned previously, which makes them a sustainable alternative for a wide range of uses. They can be dissolved in chlorinated solvents and PHAs show a range of properties, from brittle thermoplastics to gummy elastomers, depending on the nature of the fermentation conditions and the carbon-source metabolized by the PHA producer organism [16][17][18]. The structure and the composition of the biopolymers dictate the degradation rate in the environment, and the microbes that generate PHAs cover a broad range, including both Gram positive and Gram negative bacterial strains, as shown in **Table 1**.

Table 1. Notable research of bioplastic production from the last 15 years on Gram-positive (+) and Gram-negative (–) bacterial strains, with their respective carbon sources and biopolymers produced. * Bio-PU a novel bio-based poly (amide urethane).

Strain	Carbon Source	Polymer Synthesised	References
<i>Bacillus megaterium</i> (+)	Glucose salt medium	PHB	[19]
<i>Bacillus</i> spp. (+)	Soy molasses, nutrient broth, glucose, butyrate, valerate, hexanoate, octanoate, decanoate, 4-hydroxybutanoate, ϵ -caprolactone	PHB, PHBV, copolymers	[19]
<i>Burkholderia cepacia</i> (-)	Palm olein, palm stearin, crude palm oil, palm kernel oil, oleic acid, xylose, levulinic acid, sugar beet molasses, sugar maple hemicellulosic hydrolysate	PHB, PHBV	[20]
<i>Caryophanon latum</i> (+)	Nutrient broth	PHA	[21]
<i>Cupriavidus necator</i> (-)	Glucose, soybean oil, waste PE, PP, PS, plastics, biodiesel by-product substrates	PHB, PHBV, PHBH, PHBHx, copolymers	[2][3][4][22][23][24][25][26]
<i>Caldimonas taiwanensis</i> (-)	Potatoe and wheat starch	PHBV	[27]
<i>Bacillus odysseyi</i> SUK3 (+)	PS plastic	PHB	[28]
<i>Haloferax mediterranei</i> (-)	Molasses and wastewater	PHBV	[29]
<i>Pseudomonas umsongensis</i> GO16 (-)	Ethylene glycol	PHA, * Bio-PU	[30]
<i>Zoogloea</i> spp. (-)	Nutrient broth (activated sludge/wastewater)	PHA	[31]

Gram-negative *Cupriavidus necator* has been shown to have an accumulation yield of up to 90% cell dry weight, and, for this reason, it is the most studied microbe for PHA production [32]. The metabolic pathways used by *C. necator* are well-documented, both for aerobic and anaerobic conditions. In the cases of alternative carbon substrates (such as pretreated plastics), biochemical pathways I, II, and III can be selected by the organism. **Figure 3** displays these pathways and how they intersect. These pathways are similar to those utilized by *Bacillus megaterium*, which can accumulate up to 62% cell dry weight, a different archaea species from the family *Halobacteriaceae* that has also been found to produce PHA biopolymers [33][34][35][36][37].

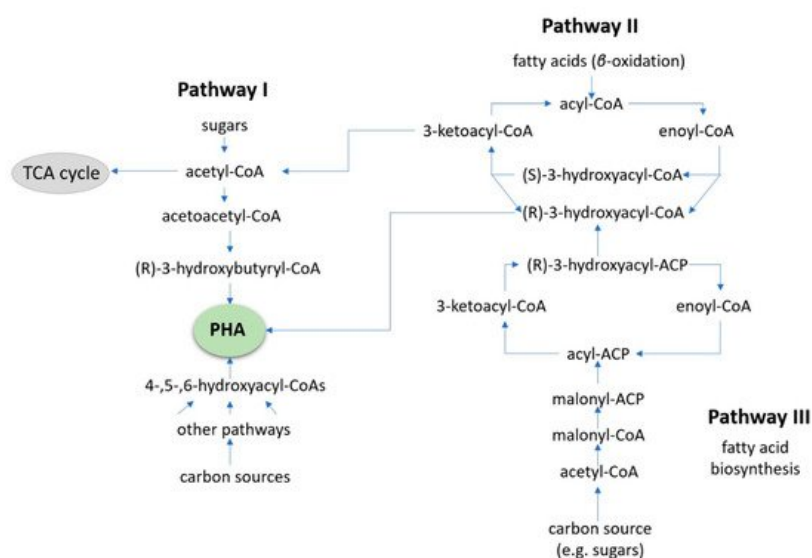


Figure 3. The combined pathways I, II, and III used by *C. necator* for PHA bioconversion. Similar biochemical pathways would be expected for other bacterial species capable of PHA synthesis where oxidized PS, PE, or PP waste plastic is treated as a fatty acid.

Glucose and fructose are normally processed in pathway I, generating PHB homopolymers. Fatty acids or sugars are metabolized via pathways II, III, or potentially other routes where copolymers can be produced [38][39][40]. Oxidized PE, PS, and PP particles are thought to enter the β -oxidation pathway forming acetyl-CoA that is then metabolized along pathway I, creating PHA polymers [36][37]. Both pure and mixed cultures (such as a cascade set-up with biosurfactant or hydrolase synthesizers) have the ability to make use of waste materials as feedstock to produce value-added PHAs. This approach, combined with the use of and using locally sourced refuse, could contribute to vastly reducing bioplastic expenses. PHAs

are a good alternative to traditional plastics, but they have a long way to go before they can surpass them, due to their high production costs, lack of specific policies, and the downstream processing [37][40]. With that said, the next evolution in bioconversion to produce PHAs is likely to be focus towards extremophile strains. The reason for this is that they combine pure culture advantages (easier optimization of conditions), plus time can be saved by working in non-sterile conditions, simplifying the extraction and reducing the running costs.

References

1. Somleva, M.N.; Peoples, O.P.; Snell, K.D. PHA Bioplastics, Biochemicals, and Energy from Crops. *Plant Biotechnol. J.* 2013, 11, 233–252.
2. Johnston, B.; Jiang, G.; Hill, D.; Adamus, G.; Kwiecien, I.; Zieba, M.; Sikorska, W.; Green, M.; Kowalczyk, M.; Radecka, I. The Molecular Level Characterization of Biodegradable Polymers Originated from Polyethylene Using Non-Oxygenated Polyethylene Wax as a Carbon Source for Polyhydroxyalkanoate Production. *Bioengineering* 2017, 4, 73.
3. Johnston, B.; Radecka, I.; Hill, D.; Chiellini, E.; Ilieva, V.I.; Sikorska, W.; Musioł, M.; Zieba, M.; Marek, A.A.; Keddie, D.; et al. The Microbial Production of Polyhydroxyalkanoates from Waste Polystyrene Fragments Attained Using Oxidative Degradation. *Polymers* 2018, 10, 957.
4. Johnston, B.; Radecka, I.; Hill, D.; Chiellini, E.; Sikorska, W.; Musioł, M.; Zieba, M.; Marek, A.A.; Mendrek, B.; Ekere, I.; et al. Mass Spectrometry Reveals Molecular Structure of Polyhydroxyalkanoates Derived from Waste Polypropylene Attained Using Oxidative Degradation. *Polymers* 2019, 11, 1580.
5. Koller, M.; Mukherjee, A. A new wave of industrialization of PHA biopolyesters. *Bioengineering* 2022, 9, 74.
6. Da Cruz Pradella, J.G. Economics and Industrial Aspects of PHA Production. In *The Handbook of Polyhydroxyalkanoates*; Koller, M., Ed.; CRC Press: Boca Raton, FL, USA, 2020; Volume 3, pp. 389–404.
7. Geyer, R.; Jambeck, J.; Law, K.L. Production, use, and fate of all plastics ever made. *Sci. Adv.* 2017, 3, e1700782.
8. Peng, R.T.; Xia, M.L.; Ru, J.K.; Huo, Y.X.; Yang, Y. Microbial degradation of polyurethane plastics. *Chin. J. Biotechnol.* 2018, 34, 1398–1409.
9. Ru, J.; Huo, Y.; Yang, Y. Microbial Degradation and Valorization of Plastic Wastes. *Front. Microbiol.* 2020, 11, 442.
10. Blank, L.M.; Narancic, T.; Mampel, J.; Tiso, T.; O'Connor, K.E. Biotechnological upcycling of plastic waste and other non-conventional feedstocks in a circular economy. *Curr. Opin. Biotechnol.* 2020, 62, 212–219.
11. Garcia, J.M.; Robertson, M.L. The future of plastics recycling. *Science* 2017, 358, 870–872.
12. Peng, Y.; Wu, P.; Schartup, A.T.; Zhang, Y. Plastic waste release caused by COVID-19 and its fate in the global ocean. *Proc. Natl. Acad. Sci. USA* 2021, 118, 47.
13. Palmieri, S.; Tittarelli, F.; Sabbatini, S.; Cespi, M.; Bonacucina, G.; Eusebi, A.L.; Fatone, F.; Stipa, P. Effects of different pre-treatments on the properties of polyhydroxyalkanoates extracted from sidestreams of a municipal wastewater treatment plant. *Sci. Total Environ.* 2021, 801, 149633.
14. Rabnawaz, M.; Wyman, I.; Auras, R.; Cheng, S. A roadmap towards green packaging: The current status and future outlook for polyesters in the packaging industry. *Green Chem.* 2017, 19, 4737–4753.
15. Harshvardhan, K.; Jha, B. Biodegradation of low-density polyethylene by marine bacteria from pelagic waters, Arabian Sea, India. *Mar. Pollut. Bull.* 2013, 77, 100–106.
16. Raza, Z.A.; Abid, S.; Banat, I.M. Polyhydroxyalkanoates: Characteristics, production, recent developments and applications. *Int. Biodeterior. Biodegrad* 2018, 126, 45–56.
17. Dietrich, K.; Dumont, M.-J.; Del Rio, L.F.; Orsat, V. Producing PHAs in the bioeconomy—Towards a sustainable bioplastic. *Sustain. Prod. Consum.* 2017, 9, 5870.
18. Rodríguez-Contreras, A.; Koller, M.; Brauneegg, G.; Marqués-Calvo, M.S. Poly production under different salinity conditions by a novel *Bacillus megaterium* strain. *New Biotechnol.* 2016, 33, 73–77.
19. Valappil, S.; Boccaccini, A.R.; Bucke, C.; Roy, I. Polyhydroxyalkanoates in Gram-positive bacteria: Insights from the genera *Bacillus* and *Streptomyces*. *Antonie Leeuwenhoek* 2007, 91, 1–17.
20. Pan, W.; Perrotta, J.A.; Stipanovic, A.J.; Nomura, C.T.; Nakas, J.P. Production of polyhydroxyalkanoates by *Burkholderia cepacia* ATCC 17759 using a detoxified sugar maple hemicellulosic hydrolysate. *J. Ind. Microbiol. Biotechnol.* 2012, 39, 459–469.
21. Jendrossek, D.; Selchow, O.; Hoppert, M. Poly(3-Hydroxybutyrate) Granules at the Early Stages of Formation Are Localized Close to the Cytoplasmic Membrane in *Caryophanon latum*. *Appl. Environ. Microbiol.* 2007, 73, 586–593.

22. Radecka, I.; Irorere, V.; Jiang, G.; Hill, D.; Williams, C.; Adamus, G.; Kwiecień, M.; Marek, A.A.; Zawadiak, J.; Johnston, B.; et al. Oxidized Polyethylene Wax as a Potential Carbon Source for PHA Production. *Materials* 2016, 9, 367.
23. Ghatge, S.; Yang, Y.; Ahn, J.H.; Hur, H.G. Biodegradation of polyethylene: A brief review. *Appl. Biol. Chem.* 2020, 63, 27.
24. Sudesh, K.; Abe, H.; Doi, Y. Synthesis, structure and properties of polyhydroxyalkanoates: Biological polyesters. *Prog. Polym. Sci.* 2000, 25, 1503–1555.
25. Flores-Sánchez, A.; López-Cuellar, M.R.; Pérez-Guevara, F.; López, U.F.; Martín-Bufájer, J.M.; Vergara-Porras, B. Synthesis of Poly-(R-hydroxyalkanoates) by *Cupriavidus necator* ATCC 17699 Using Mexican Avocado (*Persea americana*) Oil as a Carbon Source. *Int. J. Polym. Sci.* 2017, 2017, 6942950.
26. Sharma, P.K.; Fu, J.; Spicer, V.; Krokhin, O.V.; Cicek, N.; Sparling, R.; Levin, D.B. Global changes in the proteome of *Cupriavidus necator* H16 during poly-(3-hydroxybutyrate) synthesis from various biodiesel by-product substrates. *AMB Express* 2016, 6, 36.
27. Sheu, D.S.; Chen, W.M.; Yang, J.Y.; Chang, R.C. Thermophilic bacterium *Caldimonas taiwanensis* produces poly(3-hydroxybutyrate-co-3-hydroxyvalerate) from starch and valerate as carbon sources. *Enzyme Microb. Technol.* 2009, 44, 289–294.
28. Tan, G.Y.A.; Chen, C.L.; Ge, L.; Li, L.; Tan, S.N.; Wang, J.Y. Bioconversion of styrene to poly(hydroxyalkanoate) (PHA) by the new bacterial strain *Pseudomonas putida* NBUS12. *Microbes Environ.* 2015, 30, 76–85.
29. Cui, Y.W.; Gong, X.Y.; Shia, Y.P.; Wang, Z. Salinity effect on production of PHA and EPS by *Haloferax mediterranei*. *RSC Adv.* 2017, 7, 53587–53595.
30. Tiso, T.; Narancic, T.; Wei, R.; Pollet, E.; Beagan, N.; Schröder, K.; Honak, A.; Jiang, M.; Kenny, S.T.; Wierckx, N.; et al. Towards bio-upcycling of polyethylene terephthalate. *Metab. Eng.* 2021, 66, 167–178.
31. Muller, E.E.L.; Narayanasamy, S.; Zeimes, M.; Laczny, C.C.; Lebrun, L.A.; Herold, M.; Hicks, N.D.; Gillece, J.D.; Schupp, J.M.; Keim, P.; et al. First draft genome sequence of a strain belonging to the *Zoogloea* genus and its gene expression in situ. *Stand. Genom. Sci.* 2017, 12, 64.
32. Emadian, S.M.; Onay, T.T.; Demirel, B. Biodegradation of bioplastics in natural environments. *Waste Manag.* 2017, 59, 526–536.
33. Passanha, P.; Esteves, S.R.; Kedia, G.; Dinsdale, R.M.; Guwy, A.J. Increasing polyhydroxyalkanoate (PHA) yields from *Cupriavidus necator* by using filtered digestate liquors. *Bioresour. Technol.* 2013, 147, 345–352.
34. Faccin, D.J.L.; Rech, R.; Secchi, A.R.; Cardozo, N.S.M.; Ayub, M.A.Z. Influence of oxygen transfer rate on the accumulation of poly(3-hydroxybutyrate) by *Bacillus megaterium*. *Process. Biochem.* 2013, 48, 420–425.
35. Hermann-Krauss, C.; Koller, M.; Muhr, A.; Fasl, H.; Stelzer, F.; Braunegg, G. Archaeal production of polyhydroxyalkanoate (PHA) Co- and terpolyesters from biodiesel industry-derived by-products. *Archaea* 2013, 2013, 129268.
36. Tsuge, T. Metabolic Improvements and Use of Inexpensive Carbon Sources in Microbial Production of Polyhydroxyalkanoates. *J. Biosci. Bioeng.* 2002, 94, 579–584.
37. Palmeiro-Sánchez, T.; O'Flaherty, V.; Lens, P.N. Polyhydroxyalkanoate bio-production and its rise as biomaterial of the future. *J. Biotechnol.* 2022, 348, 10–25.
38. Aldor, I.S.; Keasling, J.D. Process design for microbial plastic factories: Metabolic engineering of polyhydroxyalkanoates. *Curr. Opin. Biotechnol.* 2003, 14, 475–483.
39. Steinbüchel, A.; Lütke-Eversloh, T. Metabolic engineering and pathway construction for biotechnological production of relevant polyhydroxyalkanoates in microorganisms. *Biochem. Eng. J.* 2003, 16, 81–96.
40. Rosenboom, J.G.; Langer, R.; Traverso, G. Bioplastics for a circular economy. *Nat. Rev. Mater.* 2022, 7, 117–137.