

# Immunomodulatory Effects of Histone Deacetylase 6 Inhibitors

Subjects: **Cell Biology**

Contributor: Husvinee Sundaramurthi , Zoltán Giricz , Breandán N. Kennedy

Patients diagnosed with metastatic uveal melanoma (MUM) have a poor survival prognosis. Unfortunately for this rare disease, there is no known cure and suitable therapeutic options are limited. Histone deacetylase 6 inhibitors (HDAC6i) are currently in clinical trials for other cancers and show potential beneficial effects against tumor cell survival in vitro and in vivo. In MUM cells, HDAC6i show an anti-proliferative effect in vitro and in preclinical zebrafish xenograft models.

primary and metastatic uveal melanoma

HDAC6 inhibitors

HDAC isozymes

combinatorial therapies

## 1. Introduction

Uveal melanoma (UM) is a poor prognosis cancer with no cure and limited treatment options [1][2][3]. Originating in the eye, UM is the most common form of adult ocular cancer [4]. Metastatic UM (MUM) is difficult to treat. A recent meta-analysis estimated a 60% relative survival rate of 15 to 20 years for patients after initial UM diagnosis [5]. Approximately 50% of UM patients progress to MUM, with only 15% of patients reported to survive beyond one-year post diagnosis [1][4][6]. In a meta-analysis (1 January 1980 to 29 March 2017) from a combined 2494 MUM patients, the median overall survival probability was 1.07 years across all forms of treatment [7]. Common treatment strategies for MUM include chemotherapeutic agents, immunotherapy, site-directed (e.g., chemoembolization, immunoembolization) therapy and surgical tumor resection [1][2][8][9]. Additional targeted and combinatorial pharmacological-based treatment approaches such as receptor tyrosine kinases, MAPK pathway inhibitors, checkpoint inhibitors and histone deacetylase inhibitors have entered clinical trials [9][10][11][12].

A milestone in UM was achieved in January 2022 with FDA approval of Tebentafusp-tebn (KIMMTRAK<sup>®</sup>), an immunotherapy, for treatment of HLA-A\*02:01-positive adult patients with unresectable or metastatic UM [13]. Very recently, Tebentafusp-tebn received EMA approval for use in the EU [14][15]. A phase III clinical trial (ClinicalTrials.gov Identifier: NCT03070392/EudraCT registration number: 2015-003153-18) reported that MUM patients (252) treated with Tebentafusp had improved one-year overall survival (OS) and six-months progression-free survival (PFS; 73% and 31%, respectively), on average, compared to patients (126) treated with control treatment (59% and 19%, respectively) options [13][16]. Overall it was reported that the patient cohort treated with Tebentafusp achieved 21.7 months median OS compared to 16 months (hazard ratio (HR) for death = 0.51; 95% CI 0.37 to 0.71;  $p < 0.001$ ) in the control cohort; median PFS was 3.3 months in Tebentafusp treatment group

compared to 2.9 months (HR for disease progression or death = 0.73; 95% CI, 0.58 to 0.94;  $p = 0.01$ ) in the control group [13]. However, at present this drug is only approved for use in a sub-cohort of patients, making it essential that more therapies and therapeutic targets are discovered for the treatment of a wider cohort of patients. Additionally, in an article published by Olivier and Prasad (2022), the authors put forth three key concerns regarding this study; (1) limited treatment options offered to control patient cohort in trial, (2) only 43% of the trial patients received treatment for disease progression post trial, and (3) interestingly, the observed median OS is “*far larger than, and disproportionate to the PFS benefit in terms of hazard ratios*” and that mathematically, Tebentafusp “*is an outlier across trials in melanoma*”, suggesting that further investigation is required for Tebentafusp [17].

In recent decades, histone deacetylase inhibitors (HDACi) were investigated as anti-cancer agents, with some receiving FDA/EMA approval for T-cell lymphoma and multiple myeloma [18][19][20]. However, pan-HDACi are often skeptically considered due to histone deacetylase (HDAC) function in epigenetics, suggesting off-target effects and undesirable side-effects associated with the pleiotropic inhibition of HDAC isozymes. The true therapeutic potential of pan-HDACi in oncology remains to be determined. Several reviews have been published on HDACs and HDACi in cancer, and Moschos et al. eloquently summarized the potential of broad HDACi as anti-UM agents [21].

Selective histone deacetylase 6 inhibitors (HDAC6i) are small molecule drugs with higher affinity for HDAC6 inhibition than other HDAC isozymes or other targets. HDAC6i are currently in clinical trials for various maladies, including cancer [22][23]. Histone deacetylase 6 (HDAC6), classified as a Class IIb enzyme is different to other HDACs. HDAC6 primarily resides in the cytoplasm and deacetylates cytosolic proteins, although it has the ability to shuttle between the nucleus and cytoplasm [24][25]. HDAC6 is a  $Zn^{2+}$ -dependent deacetylase composed of two deacetylases catalytic domains, dynein motor binding domain and ubiquitin-binding zinc-finger functional domains [26]. HDAC6 functions in cancer-related processes such as tumorigenesis, angiogenesis, metastasis, the aggresome–autophagy pathway and inflammation as detailed in previous review articles [22][23][27][28].

## 2. Selective HDAC6i as a Therapeutic Option for Uveal Melanoma

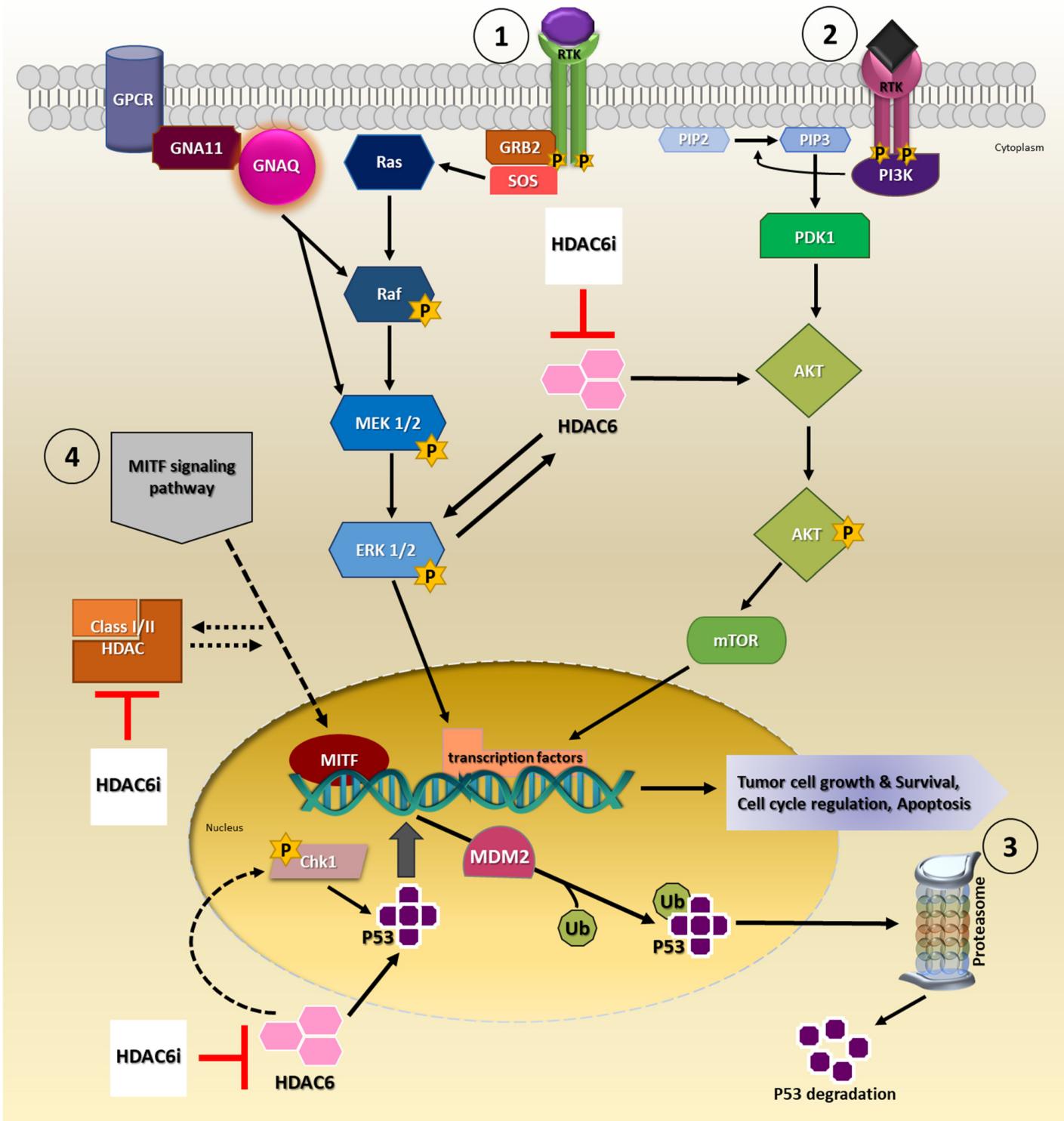
Interestingly, there are presently no registered clinical trials involving selective HDAC6i in UM/MUM, even though there are clinical trials ongoing for relapsed or refractory multiple myeloma, lymphoma, non-small cell lung cancer, metastatic breast cancer and solid tumor (Table 1) [23][29][30][31]. Likewise, to date, few studies have assessed the efficacy of selective HDAC6i in UM or MUM cells in vitro or in vivo. However, evidence in vitro and in pre-clinical UM models suggests inhibiting HDAC6 may offer therapeutic benefit. Nencetti et al. reported that novel compound VS13, with increased HDAC6 selectivity, significantly reduced 92.1 and Mel270 (primary) UM cell viability (up to a 100% reduction) in a dose-dependent manner [32]. Additionally, ACY-1215, a selective HDAC6i, induced a dose-dependent, significant reduction in UM and MUM cell survival by up to 99.99% in vitro and significantly decreased UM cell fluorescence by 65% in zebrafish xenografts with MUM (OMM2.5) cells in vivo [33].

**Table 1.** HDAC6 inhibitors in clinical trials for various cancers.

Cancer Type	Drug Combination	Clinical Trial Identifier(s)	Phase	No. of Participants	Status	Outcome	Reference (PMID)
Biliary Tract Cancer (advanced)	KA2507	NCT04186156 EUCTR2019-001459-38-GB	Ib/II	N/A	Withdrawn	N/A	
Relapsed/Refractory Multiple Myeloma	ACY-1215 + Bortezomib/Dexamethasone	NCT01323751	I/II	120	Completed	N/A	
Lymphoid Malignancies, Lymphoma	ACY-1215	NCT02091063	I/II	24	Completed	Under review	
Non-Small Cell Lung Cancer	ACY-241 + Nivolumab	NCT02635061	Ib	17	Active, not recruiting	Ongoing	34552864
Solid Tumor, Adult	KA2507	NCT03008018	I	20	Completed	Published	33947698
Metastatic Breast Cancer, Breast Carcinoma	ACY-1215 + Nab-paclitaxel	NCT02632071	Ib	17	Completed	N/A	
Unresectable/Metastatic Cholangiocarcinoma	ACY-1215 + Gemcitabine/Cisplatin	NCT02856568	Ib	N/A	Withdrawn	N/A	
Relapsed/Refractory Multiple Myeloma	ACY-1215 + Pomalidomide/Dexamethasone	EUCTR2014-002338-29-IT EUCTR2014-002338-29-GR NCT01997840	Ib/II	103	Active, not recruiting	Ongoing	

Cancer Type	Drug Combination	Clinical Trial Identifier(s)	Phase	No. of Participants	Status	Outcome	Reference (PMID)
Gynecological Cancer	ACY-1215 + Paclitaxel/Bevacizumab	NCT02661815	Ib	6	Terminated	Available	ival and d kinase [23][34][35]
Malignant Melanoma [36][37][38][39]	ACY-241 + Nivolumab + Ipilimumab	NCT02935790	Ib	1	Completed	N/A	tance as signaling
Solid Tumor (advanced)	JBI-802	NCT05268666	I/II	126	Recruiting	N/A	$\beta$ -binding GNA11),

upstream of this signaling cascade [41][42]. Uveal melanoma cells treated with a MEK and PI3K inhibitor combination inhibited cell proliferation and induced apoptosis [43]. Moreover, several other studies explored MEK and PI3K/AKT pathway inhibitors as a therapeutic option for UM; however, the therapeutic efficacy has been contentious [40][44][45][46][47].



**Figure 1.** Involvement of HDAC6 in cancer signaling pathways. Proposed model for HDAC6i mechanism of action by targeting either ① MAPK/ERK, ② PI3K/AKT, ③ P53 and/or ④ MITF signaling pathway(s). Consequently, targeting these pathways inhibits biological processes that promotes cell survival and proliferation in the cancer cell.

Studies show modulation of HDAC6 in HEK293T cells, either by siRNA mediated knock-down or ACY-1215 pharmacological inhibition, led to increased phospho-ERK levels and ERK1/2 activation, respectively, hence

regulating ERK signaling [48][49]. In LNCaP prostate cancer cells, treatment with Panobinostat inhibited HDAC6 activity and triggered ERK activation, which consequently resulted in the arrest of cell cycle [48]. In colorectal cancer cells, HCT116 and HT29, knock-down of HDAC6, blocked cell proliferation, migration and invasion in part via the MAPK/ERK signaling pathway [50]. A reduction in the expression levels of phospho-MEK, phospho-ERK and phospho-AKT was demonstrated in these cells. Peng et al. (2017), reported that inhibition of cell proliferation and survival with ACY-1215 and/or Vemurafenib in A375 melanoma cells was partly mediated through the inhibition of ERK activation [51]. Similarly, it was observed that ACY-1215 treatment of esophageal squamous cell carcinoma cells, EC109 and TE-1, led to a decrease in phospho-ERK1/2, phospho-AKT levels and inhibited cell proliferation and survival [52].

The PI3K/AKT signaling pathway is constitutively activated in UM which facilitates tumorigenesis processes such as cell survival, inhibition of apoptosis and angiogenesis [53]. In ACY-1215 treated cholangiocarcinoma cells, cell proliferation was blocked, and apoptosis was triggered via the PI3K/AKT pathway [54]. Kaliszczak et al. (2016) demonstrated that cotreatment of HCT116 with HDAC6i and pan-AKT inhibitor/dual PI3K/mTOR inhibitor enhanced anti-tumor effects both in vitro and in vivo [55]. Other studies reveal that dual inhibition with HDACi and/or PI3K/AKT/mTOR inhibitor effectively reduced tumor cell proliferation in cancer cells, e.g., prostate cancer, multiple myeloma, relapsed or refractory diffuse large B-cell lymphoma, neuroblastoma, hematologic tumor(s), hepatocarcinoma and breast cancer cells [56][57][58][59][60].

Another significant pathway involved in UM disease pathogenesis may be attributed to the p53 signaling pathway. Mutation(s) in *TP53* is rare in UM tumors; however, its activity is commonly disrupted as a consequence of mutations and dysregulation of other key factors in this pathway [61][62]. Cao et al. demonstrated in vitro and in vivo, that ACY-1215 increased the transcriptional activity of p53, which consequently led to cell cycle arrest and apoptosis in triple-negative breast cancer cells [63]. In colorectal cancer cells, HDAC6 inhibition upregulated p53 levels and increased acetylated p53 levels, and consequently increasing apoptosis [64]. In another study, the authors proposed that in ovarian cancer cells, HDAC6 inhibition in combination with Paclitaxel activated p53 and induced apoptosis [37]. Recently, it was reported that ACY-1215 inhibited cell proliferation through the promotion of apoptosis through mitotic catastrophe in a p53-dependent manner by repressing p-Chk1 activity, in head and neck carcinoma cells [65]. Promisingly, a Class III specific HDAC inhibitor, Tenovin-6, prevented tumor cell growth and induced apoptosis by increasing p53 expression in uveal melanoma tumor and cancer stem cells [66].

Interestingly, an additional pathway receiving renewed attention is the microphthalmia-associated transcription factor (MITF) signaling pathway in UM. MITF signaling is of particular interest in the context of UM as mutations in this transcription factor and its interacting partners are associated with oncogenic functions and disease pathogenesis in cutaneous melanoma [67][68][69][70]. MITF is widely known as the master regulator of melanogenesis and melanocyte differentiation, in addition to regulating various cellular processes [71]. Comparably, in UM, MITF seems to have an intricate role balancing between UM tumor growth and tumor suppression activities [72]. Most recently, Phelps et al. presented that MITF acts as a bona fide tumor suppressor in UM [73]. The authors report using zebrafish models that loss of *mitfa* resulted in growth of UM tumors. Yokoyama et al. (2008) has also shown

that pan-HDACi repressed M-MITF expression in clear cell sarcoma and melanoma cells [74]. Regardless, the exact relationship between HDACi/HDAC6i and MITF still remains to be determined.

As such these pathways offer additional novel putative targets that needs to be thoroughly investigated. Taken together, there is a strong possibility that using HDAC6i may offer therapeutic benefits mediated through the regulation of these highlighted pathways either as a monotherapy or as combinatorial therapy for UM, advanced UM and/or other solid tumors, which needs to be explored extensively.

## 4. Immunomodulatory Effects of HDAC6 Inhibitors

The tumor microenvironment (TME) supports processes involved in tumorigenesis [75][76]. Composed of tumor cells, immune cells, stromal cells, signaling molecules, extracellular matrix and blood vessels, the microenvironment can ensure tumor cells are able to grow, survive, spread and even to gain resistance to therapy [76]. An increasing number of studies are focusing on understanding and identifying novel therapeutic targets within the tumor microenvironment for cancer treatment. In UM, tumors with chromosomal defects such as loss of one copy of chromosome 3 or gain of chromosome 8q, present with increased levels of inflammatory mediators and immune cells leading to a tumor-promoting inflammatory TME [77]. Differing from other cancers, increased amounts of tumor infiltrating lymphocytes (TILs) and tumor associated macrophages (TAMs) are correlated to poor prognosis and high metastasis risk in UM [78][79]. A delicate balance is required to ensure the prevention of UM cells from evading immune surveillance.

HDACi are capable of immunomodulation in cancer [35][80][81][82][83]. Inhibition of Class I HDACs by HDACi or in combination with the immunomodulatory drug Lenalidomide, resulted in the downregulation of cellular Myc proto-oncogene protein (c-MYC) and increased cytotoxicity in multiple myeloma cells [84]. Additionally, ACY-1215 treatment, alone or combined with Lenalidomide, significantly reduced c-MYC, IKAROS family zinc finger (IKZF)1/IKZF3 and interferon regulatory factor 4 (IRF4) expression levels triggering immune system activation, which was postulated to be involved in the anti-tumor cell survival effects. The HDAC6 inhibitor A452 in combination with Lenalidomide or Pomalidomide, displayed significantly increased synergistic anti-proliferative effects which was attributed to the augmented reduction of c-MYC, IKZF1/IKZF3 and IRF4 expression in multiple myeloma cells [85]. In a preclinical mouse model of non-small cell lung cancer (NSCLC), ACY-1215 treatment increased the expression of MHC Class II molecules, CD86 and CD96 co-stimulatory molecules, suggesting that ACY-1215 may play a role in T-cell activation and antigen presentation, consequently promoting anti-tumor immunity [86]. Another study described that the HDAC6 inhibitor ACY-241 alone, and when combined with Oxaliplatin (chemotherapy drug), promoted T cell functions, thereby increasing the immunogenicity of tumor cells, in an NSCLC mouse model [87]. Inhibition of HDAC6 in melanoma cells resulted in the increased expression of MHC class I and presentation of tumor-related antigens [88]. Knox et al. demonstrated that treatment of murine melanoma model with the HDAC6i Nextrastat A, combined with the anti-PD1 antibody, increased tumor infiltration of CD8<sup>+</sup> and natural killer cells and reduced the level of pro-tumorigenic M2 macrophages [89]. Combining HDAC6i with immunomodulatory agents can improve therapeutic efficacy [90].

## 5. Conclusion

A combined therapeutic approach involving HDAC6i with chemotherapeutic agents, may be more beneficial and should be explored in detail as treatment options for UM/MUM. Promising results from Entinostat pembrolizumab gives us hope that it is worthwhile to pursue combinatorial treatment strategy in UM/MUM.

## References

1. Carvajal, R.D.; Schwartz, G.K.; Tezel, T.; Marr, B.; Francis, J.H.; Nathan, P.D. Metastatic disease from uveal melanoma: Treatment options and future prospects. *Br. J. Ophthalmol.* 2017, **101**, 38–44.
2. Branisteanu, D.C.; Bogdanici, C.M.; Branisteanu, D.E.; Maranduca, M.A.; Zemba, M.; Balta, F.; Branisteanu, C.I.; Moraru, A.D. Uveal melanoma diagnosis and current treatment options (Review). *Exp. Med.* 2021, **22**, 1428.
3. Beasley, A.B.; Chen, F.K.; Isaacs, T.W.; Gray, E.S. Future perspectives of uveal melanoma blood based biomarkers. *Br. J. Cancer* 2022, **126**, 1511–1528.
4. Kaliki, S.; Shields, C.L. Uveal melanoma: Relatively rare but deadly cancer. *Eye* 2017, **31**, 241–257.
5. Stalhammar, G.; Gill, V.T. The long-term prognosis of patients with untreated primary uveal melanoma: A systematic review and meta-analysis. *Crit. Rev. Oncol. Hematol.* 2022, **172**, 103652.
6. Smit, K.N.; Jager, M.J.; de Klein, A.; Kili, E. Uveal melanoma: Towards a molecular understanding. *Prog. Retin. Eye Res.* 2020, **75**, 100800.
7. Rantala, E.S.; Hernberg, M.; Kivela, T.T. Overall survival after treatment for metastatic uveal melanoma: A systematic review and meta-analysis. *Melanoma Res.* 2019, **29**, 561–568.
8. Rodriguez-Vidal, C.; Fernandez-Diaz, D.; Fernandez-Marta, B.; Lago-Baameiro, N.; Pardo, M.; Silva, P.; Paniagua, L.; Blanco-Teijeiro, M.J.; Piñeiro, A.; Bande, M. Treatment of Metastatic Uveal Melanoma: Systematic Review. *Cancers* 2020, **12**, 2557.
9. Mallone, F.; Sacchetti, M.; Lambiase, A.; Moramarco, A. Molecular Insights and Emerging Strategies for Treatment of Metastatic Uveal Melanoma. *Cancers* 2020, **12**, 2761.
10. Schank, T.E.; Hassel, J.C. Immunotherapies for the Treatment of Uveal Melanoma-History and Future. *Cancers* 2019, **11**, 1048.
11. Wang, J.Z.; Lin, V.; Toumi, E.; Wang, K.; Zhu, H.; Conway, R.M.; Madigan, M.C.; Murray, M.; Cherepanoff, S.; Zhou, F.; et al. Development of new therapeutic options for the treatment of

uveal melanoma. *FEBS J.* 2021, **288**, 6226–6249.

12. Schefler, A.C.; Kim, R.S. Recent advancements in the management of retinoblastoma and uveal melanoma. *Fac. Rev.* 2021, **10**, 51.

13. Nathan, P.; Hassel, J.C.; Rutkowski, P.; Baurain, J.F.; Butler, M.O.; Schlaak, M.; Sullivan, R.J.; Ochsenreither, S.; Dummer, R.; Kirkwood, J.M.; et al. Overall Survival Benefit with Tebentafusp in Metastatic Uveal Melanoma. *N. Engl. J. Med.* 2021, **385**, 1196–1206.

14. Dhillon, S. Tebentafusp: First Approval. *Drugs* 2022, **82**, 703–710.

15. Agency, E.M. Kimmtrak. Available online: <https://www.ema.europa.eu/en/medicines/human/EPAR/kimmtrak> (accessed on 18 June 2022).

16. Killock, D. Tebentafusp for uveal melanoma. *Nat. Rev. Clin. Oncol.* 2021, **18**, 747.

17. Olivier, T.; Prasad, V. Tebentafusp in first-line melanoma trials: An outperforming outlier. *Transl. Oncol.* 2022, **20**, 101408.

18. Eckschlager, T.; Plch, J.; Stiborova, M.; Hrabeta, J. Histone Deacetylase Inhibitors as Anticancer Drugs. *Int. J. Mol. Sci.* 2017, **18**, 1414.

19. McClure, J.J.; Li, X.; Chou, C.J. Advances and Challenges of HDAC Inhibitors in Cancer Therapeutics. *Adv. Cancer Res.* 2018, **138**, 183–211.

20. Jenke, R.; Ressing, N.; Hansen, F.K.; Aigner, A.; Buch, T. Anticancer Therapy with HDAC Inhibitors: Mechanism-Based Combination Strategies and Future Perspectives. *Cancers* 2021, **13**, 634.

21. Moschos, M.M.; Dettoraki, M.; Androudi, S.; Kalogeropoulos, D.; Lavaris, A.; Garmpis, N.; Damaskos, C.; Garmpi, A.; Tsatsos, M. The Role of Histone Deacetylase Inhibitors in Uveal Melanoma: Current Evidence. *Anticancer Res.* 2018, **38**, 3817–3824.

22. Aldana-Masangkay, G.I.; Sakamoto, K.M. The role of HDAC6 in cancer. *J. Biomed. Biotechnol.* 2011, **2011**, 875824.

23. Li, T.; Zhang, C.; Hassan, S.; Liu, X.; Song, F.; Chen, K.; Zhang, W.; Yang, J. Histone deacetylase 6 in cancer. *J. Hematol. Oncol.* 2018, **11**, 111.

24. Boyault, C.; Sadoul, K.; Pabion, M.; Khochbin, S. HDAC6, at the crossroads between cytoskeleton and cell signaling by acetylation and ubiquitination. *Oncogene* 2007, **26**, 5468–5476.

25. Li, Y.; Shin, D.; Kwon, S.H. Histone deacetylase 6 plays a role as a distinct regulator of diverse cellular processes. *FEBS J.* 2013, **280**, 775–793.

26. Batchu, S.N.; Brijmohan, A.S.; Advani, A. The therapeutic hope for HDAC6 inhibitors in malignancy and chronic disease. *Clin. Sci.* 2016, **130**, 987–1003.

27. Li, G.; Tian, Y.; Zhu, W.G. The Roles of Histone Deacetylases and Their Inhibitors in Cancer Therapy. *Front. Cell Dev. Biol.* 2020, 8, 576946.

28. Hontecillas-Prieto, L.; Flores-Campos, R.; Silver, A.; de Alava, E.; Hajji, N.; Garcia-Dominguez, D.J. Synergistic Enhancement of Cancer Therapy Using HDAC Inhibitors: Opportunity for Clinical Trials. *Front. Genet.* 2020, 11, 578011.

29. Vogl, D.T.; Raje, N.; Jagannath, S.; Richardson, P.; Hari, P.; Orlowski, R.; Supko, J.G.; Tamang, D.; Yang, M.; Jones, S.S.; et al. Ricolinostat, the First Selective Histone Deacetylase 6 Inhibitor, in Combination with Bortezomib and Dexamethasone for Relapsed or Refractory Multiple Myeloma. *Clin. Cancer Res.* 2017, 23, 3307–3315.

30. Amengual, J.E.; Lue, J.K.; Ma, H.; Lichtenstein, R.; Shah, B.; Cremers, S.; Jones, S.; Sawas, A. First-in-Class Selective HDAC6 Inhibitor (ACY-1215) Has a Highly Favorable Safety Profile in Patients with Relapsed and Refractory Lymphoma. *Oncologist* 2021, 26, 184.e366.

31. Awad, M.M.; Le Bruchec, Y.; Lu, B.; Ye, J.; Miller, J.; Lizotte, P.H.; Cavanaugh, M.E.; Rode, A.J.; Dumitru, C.D.; Spira, A. Selective Histone Deacetylase Inhibitor ACY-241 (Citarinostat) Plus Nivolumab in Advanced Non-Small Cell Lung Cancer: Results from a Phase Ib Study. *Front. Oncol.* 2021, 11, 696512.

32. Nencetti, S.; Cuffaro, D.; Nuti, E.; Ciccone, L.; Rossello, A.; Fabbi, M.; Ballante, F.; Ortore, G.; Carbotti, G.; Campelli, F.; et al. Identification of histone deacetylase inhibitors with (arylidene)aminoxy scaffold active in uveal melanoma cell lines. *J. Enzym. Inhib. Med. Chem.* 2021, 36, 34–47.

33. Sundaramurthi, H.; Garcia-Mulero, S.; Tonelotto, V.; Slater, K.; Marcone, S.; Piulats, J.M.; Watson, R.W.; Tobin, D.J.; Jensen, L.D.; Kennedy, B.N. Uveal Melanoma Cell Line Proliferation Is Inhibited by Ricolinostat, a Histone Deacetylase Inhibitor. *Cancers* 2022, 14, 782.

34. Zuidervaart, W.; van Nieuwpoort, F.; Stark, M.; Dijkman, R.; Packer, L.; Borgstein, A.M.; Pavey, S.; van der Velden, P.; Out, C.; Jager, M.J.; et al. Activation of the MAPK pathway is a common event in uveal melanomas although it rarely occurs through mutation of BRAF or RAS. *Br. J. Cancer* 2005, 92, 2032–2038.

35. Cosenza, M.; Pozzi, S. The Therapeutic Strategy of HDAC6 Inhibitors in Lymphoproliferative Disease. *Int. J. Mol. Sci.* 2018, 19, 2337.

36. Li, J.; Yu, M.; Fu, S.; Liu, D.; Tan, Y. Role of Selective Histone Deacetylase 6 Inhibitor ACY-1215 in Cancer and Other Human Diseases. *Front. Pharm.* 2022, 13, 907981.

37. Yoo, J.; Jeon, Y.H.; Lee, D.H.; Kim, G.W.; Lee, S.W.; Kim, S.Y.; Park, J.; Kwon, S.H. HDAC6-selective inhibitors enhance anticancer effects of paclitaxel in ovarian cancer cells. *Oncol. Lett.* 2021, 21, 201.

38. Huang, Z.; Xia, Y.; Hu, K.; Zeng, S.; Wu, L.; Liu, S.; Zhi, C.; Lai, M.; Chen, D.; Xie, L.; et al. Histone deacetylase 6 promotes growth of glioblastoma through the MKK7/JNK/c-Jun signaling pathway. *J. Neurochem.* 2020, 152, 221–234.

39. Shoushtari, A.N.; Carvajal, R.D. GNAQ and GNA11 mutations in uveal melanoma. *Melanoma Res.* 2014, 24, 525–534.

40. Babchia, N.; Calipel, A.; Mouriaux, F.; Faussat, A.M.; Mascarelli, F. The PI3K/Akt and mTOR/P70S6K signaling pathways in human uveal melanoma cells: Interaction with B-Raf/ERK. *Investig. Ophthalmol. Vis. Sci.* 2010, 51, 421–429.

41. Coupland, S.E.; Lake, S.L.; Zeschnigk, M.; Damato, B.E. Molecular pathology of uveal melanoma. *Eye* 2013, 27, 230–242.

42. Li, Y.; Shi, J.; Yang, J.; Ge, S.; Zhang, J.; Jia, R.; Fan, X. Uveal melanoma: Progress in molecular biology and therapeutics. *Adv. Med. Oncol.* 2020, 12, 1758835920965852.

43. Khalili, J.S.; Yu, X.; Wang, J.; Hayes, B.C.; Davies, M.A.; Lizée, G.; Esmaeli, B.; Woodman, S.E. Combination small molecule MEK and PI3K inhibition enhances uveal melanoma cell death in a mutant GNAQ- and GNA11-dependent manner. *Clin. Cancer Res.* 2012, 18, 4345–4355.

44. Faiao-Flores, F.; Emmons, M.F.; Durante, M.A.; Kinose, F.; Saha, B.; Fang, B.; Koomen, J.M.; Chellappan, S.P.; Maria-Engler, S.S.; Rix, U.; et al. HDAC Inhibition Enhances the In Vivo Efficacy of MEK Inhibitor Therapy in Uveal Melanoma. *Clin. Cancer Res.* 2019, 25, 5686–5701.

45. Steeb, T.; Wessely, A.; Ruzicka, T.; Heppt, M.V.; Berking, C. How to MEK the best of uveal melanoma: A systematic review on the efficacy and safety of MEK inhibitors in metastatic or unresectable uveal melanoma. *Eur. J. Cancer* 2018, 103, 41–51.

46. Farhan, M.; Silva, M.; Xingan, X.; Zhou, Z.; Zheng, W. Artemisinin Inhibits the Migration and Invasion in Uveal Melanoma via Inhibition of the PI3K/AKT/mTOR Signaling Pathway. *Oxid. Med. Cell Longev.* 2021, 2021, 9911537.

47. Musi, E.; Ambrosini, G.; de Stanchina, E.; Schwartz, G.K. The phosphoinositide 3-kinase alpha selective inhibitor BYL719 enhances the effect of the protein kinase C inhibitor AEB071 in GNAQ/GNA11-mutant uveal melanoma cells. *Mol. Cancer* 2014, 13, 1044–1053.

48. Chuang, M.J.; Wu, S.T.; Tang, S.H.; Lai, X.M.; Lai, H.C.; Hsu, K.H.; Sun, K.H.; Sun, G.H.; Chang, S.Y.; Yu, D.S.; et al. The HDAC inhibitor LBH589 induces ERK-dependent prometaphase arrest in prostate cancer via HDAC6 inactivation and down-regulation. *PLoS ONE* 2013, 8, e73401.

49. Wu, J.Y.; Xiang, S.; Zhang, M.; Fang, B.; Huang, H.; Kwon, O.K.; Zhao, Y.; Yang, Z.; Bai, W.; Bepler, G.; et al. Histone deacetylase 6 (HDAC6) deacetylates extracellular signal-regulated kinase 1 (ERK1) and thereby stimulates ERK1 activity. *J. Biol. Chem.* 2018, 293, 1976–1993.

50. Zhang, S.Z.H.; Zhou, B.; Chu, Y.; Huo, J.; Tan, Y.; Liu, D. Histone deacetylase 6 is overexpressed and promotes tumor growth of colon cancer through regulation of the MAPK/ERK signal pathway. *Onco. Targets Ther.* 2019, 12, 2409–2419.

51. Peng, U.; Wang, Z.; Pei, S.; Ou, Y.; Hu, P.; Liu, W.; Song, J. ACY-1215 accelerates vemurafenib induced cell death of BRAF-mutant melanoma cells via induction of ER stress and inhibition of ERK activation. *Oncol. Rep.* 2017, 37, 1270–1276.

52. Cao, J.; Lv, W.; Wang, L.; Xu, J.; Yuan, P.; Huang, S.; He, Z.; Hu, J. Ricolinostat (ACY-1215) suppresses proliferation and promotes apoptosis in esophageal squamous cell carcinoma via miR-30d/PI3K/AKT/mTOR and ERK pathways. *Cell Death Dis.* 2018, 9, 817.

53. Katopodis, P.; Khalifa, M.S.; Anikin, V. Molecular characteristics of uveal melanoma and intraocular tumors. *Oncol. Lett.* 2021, 21, 9.

54. Kim, C.; Lee, S.; Kim, D.; Lee, D.S.; Lee, E.; Yoo, C.; Kim, K.P. Blockade of GRP78 Translocation to the Cell Surface by HDAC6 Inhibition Suppresses Proliferation of Cholangiocarcinoma Cells. *Anticancer Res.* 2022, 42, 471–482.

55. Kaliszczak, M.; Trousil, S.; Ali, T.; Aboagye, E.O. AKT activation controls cell survival in response to HDAC6 inhibition. *Cell Death Dis.* 2016, 7, e2286.

56. Ellis, L.; Ku, S.Y.; Ramakrishnan, S.; Lasorsa, E.; Azabdaftari, G.; Godoy, A.; Pili, R. Combinatorial antitumor effect of HDAC and the PI3K-Akt-mTOR pathway inhibition in a Pten defecient model of prostate cancer. *Oncotarget* 2013, 4, 2225–2236.

57. Yan, Z.; Zhang, K.; Ji, M.; Xu, H.; Chen, X. A Dual PI3K/HDAC Inhibitor Downregulates Oncogenic Pathways in Hematologic Tumors In Vitro and In Vivo. *Front. Pharm.* 2021, 12, 741697.

58. Chilamakuri, R.; Agarwal, S. Dual Targeting of PI3K and HDAC by CUDC-907 Inhibits Pediatric Neuroblastoma Growth. *Cancers* 2022, 14, 1067.

59. Ranganna, K.; Selvam, C.; Shivachar, A.; Yousefipour, Z. Histone Deacetylase Inhibitors as Multitarget-Directed Epi-Drugs in Blocking PI3K Oncogenic Signaling: A Polypharmacology Approach. *Int. J. Mol. Sci.* 2020, 21, 8198.

60. Liao, W.; Yang, W.; Xu, J.; Yan, Z.; Pan, M.; Xu, X.; Zhou, S.; Zhu, Y.; Lan, J.; Zeng, M.; et al. Therapeutic Potential of CUDC-907 (Fimepinostat) for Hepatocarcinoma Treatment Revealed by Tumor Spheroids-Based Drug Screening. *Front. Pharm.* 2021, 12, 658197.

61. Brantley, M.A., Jr.; Harbour, J.W. Deregulation of the Rb and p53 pathways in uveal melanoma. *Am. J. Pathol.* 2000, 157, 1795–1801.

62. Helgadottir, H.; Hoiom, V. The genetics of uveal melanoma: Current insights. *Appl. Clin. Genet.* 2016, 9, 147–155.

63. Cao, W.; Shen, R.; Richard, S.; Liu, Y.; Jalalirad, M.; Cleary, M.P.; D'Assoro, A.B.; Gradilone, S.A.; Yang, D.Q. Inhibition of triplenegative breast cancer proliferation and motility by reactivating p53 and inhibiting overactivated Akt. *Oncol. Rep.* 2022, 47, 1–8.

64. Ryu, H.W.; Shin, D.H.; Lee, D.H.; Choi, J.; Han, G.; Lee, K.Y.; Kwon, S.H. HDAC6 deacetylates p53 at lysines 381/382 and differentially coordinates p53-induced apoptosis. *Cancer Lett.* 2017, 391, 162–171.

65. Miyake, K.; Takano, N.; Kazama, H.; Kikuchi, H.; Hiramoto, M.; Tsukahara, K.; Miyazawa, K. Ricolinostat enhances adavosertibinduced mitotic catastrophe in TP53mutated head and neck squamous cell carcinoma cells. *Int. J. Oncol.* 2022, 60, 1–12.

66. Dai, W.; Zhou, J.; Jin, B.; Pan, J. Class III-specific HDAC inhibitor Tenovin-6 induces apoptosis, suppresses migration and eliminates cancer stem cells in uveal melanoma. *Sci. Rep.* 2016, 6, 22622.

67. Garraway, L.A.; Widlund, H.R.; Rubin, M.A.; Getz, G.; Berger, A.J.; Ramaswamy, S.; Beroukhim, R.; Milner, D.A.; Granter, S.R.; Du, J.; et al. Integrative genomic analyses identify MITF as a lineage survival oncogene amplified in malignant melanoma. *Nature* 2005, 436, 117–122.

68. Kawakami, A.; Fisher, D.E. The master role of microphthalmia-associated transcription factor in melanocyte and melanoma biology. *Lab. Investig.* 2017, 97, 649–656.

69. Levy, C.; Khaled, M.; Fisher, D.E. MITF: Master regulator of melanocyte development and melanoma oncogene. *Trends Mol. Med.* 2006, 12, 406–414.

70. Yajima, I.; Kumasaka, M.Y.; Thang, N.D.; Goto, Y.; Takeda, K.; Iida, M.; Ohgami, N.; Tamura, H.; Yamanoshita, O.; Kawamoto, Y.; et al. Molecular Network Associated with MITF in Skin Melanoma Development and Progression. *J. Ski. Cancer* 2011, 2011, 730170.

71. Goding, C.R.; Arnheiter, H. MITF—the first 25 years. *Genes Dev.* 2019, 33, 983–1007.

72. Gelmi, M.C.; Houtzagers, L.E.; Strub, T.; Krossa, I.; Jager, M.J. MITF in Normal Melanocytes, Cutaneous and Uveal Melanoma: A Delicate Balance. *Int. J. Mol. Sci.* 2022, 23, 6001.

73. Phelps, G.B.; Hagen, H.R.; Amsterdam, A.; Lees, J.A. MITF deficiency accelerates GNAQ-driven uveal melanoma. *Proc. Natl. Acad. Sci. USA* 2022, 119, e2107006119.

74. Yokoyama, S.; Feige, E.; Poling, L.L.; Levy, C.; Widlund, H.R.; Khaled, M.; Kung, A.L.; Fisher, D.E. Pharmacologic suppression of MITF expression via HDAC inhibitors in the melanocyte lineage. *Pigment. Cell Melanoma Res.* 2008, 21, 457–463.

75. Anderson, N.M.; Simon, M.C. The tumor microenvironment. *Curr. Biol.* 2020, 30, R921–R925.

76. Baghban, R.; Roshangar, L.; Jahanban-Esfahlan, R.; Seidi, K.; Ebrahimi-Kalan, A.; Jaymand, M.; Kolahian, S.; Javaheri, T.; Zare, P. Tumor microenvironment complexity and therapeutic implications at a glance. *Cell Commun. Signal.* 2020, 18, 59.

77. Bronkhorst, I.H.; Jager, M.J. Uveal melanoma: The inflammatory microenvironment. *J. Innate Immun.* 2012, 4, 454–462.

78. Basile, M.S.; Mazzon, E.; Fagone, P.; Longo, A.; Russo, A.; Fallico, M.; Bonfiglio, V.; Nicoletti, F.; Avitabile, T.; Reibaldi, M. Immunobiology of Uveal Melanoma: State of the Art and Therapeutic Targets. *Front. Oncol.* 2019, 9, 1145.

79. Tosi, A.; Cappellessi, R.; Dei Tos, A.P.; Rossi, V.; Aliberti, C.; Pigozzo, J.; Fabozzi, A.; Sbaraglia, M.; Blandamura, S.; Del Bianco, P.; et al. The immune cell landscape of metastatic uveal melanoma correlates with overall survival. *J. Exp. Clin. Cancer Res.* 2021, 40, 154.

80. Khan, A.N.; Gregorie, C.J.; Tomasi, T.B. Histone deacetylase inhibitors induce TAP, LMP, Tapasin genes and MHC class I antigen presentation by melanoma cells. *Cancer Immunol. Immunother.* 2008, 57, 647–654.

81. Shen, L.; Orillion, A.; Pili, R. Histone deacetylase inhibitors as immunomodulators in cancer therapeutics. *Epigenomics* 2016, 8, 415–428.

82. Shanmugam, G.; Rakshit, S.; Sarkar, K. HDAC inhibitors: Targets for tumor therapy, immune modulation and lung diseases. *Transl. Oncol.* 2022, 16, 101312.

83. Blaszczak, W.; Liu, G.; Zhu, H.; Barczak, W.; Shrestha, A.; Albayrak, G.; Zheng, S.; Kerr, D.; Samsonova, A.; La Thangue, N.B. Immune modulation underpins the anti-cancer activity of HDAC inhibitors. *Mol. Oncol.* 2021, 15, 3280–3298.

84. Hideshima, T.; Cottini, F.; Ohguchi, H.; Jakubikova, J.; Gorgun, G.; Mimura, N.; Tai, Y.T.; Munshi, N.C.; Richardson, P.G.; Anderson, K.C. Rational combination treatment with histone deacetylase inhibitors and immunomodulatory drugs in multiple myeloma. *Blood Cancer J.* 2015, 5, e312.

85. Won, H.R.; Lee, D.H.; Yeon, S.K.; Ryu, H.W.; Kim, G.W.; Kwon, S.H. HDAC6selective inhibitor synergistically enhances the anticancer activity of immunomodulatory drugs in multiple myeloma. *Int. J. Oncol.* 2019, 55, 499–512.

86. Adeegbe, D.O.; Liu, Y.; Lizotte, P.H.; Kamihara, Y.; Aref, A.R.; Almonte, C.; Dries, R.; Li, Y.; Liu, S.; Wang, X.; et al. Synergistic Immunostimulatory Effects and Therapeutic Benefit of Combined Histone Deacetylase and Bromodomain Inhibition in Non-Small Cell Lung Cancer. *Cancer Discov.* 2017, 7, 852–867.

87. Bag, A.; Schultz, A.; Bhimani, S.; Stringfield, O.; Dominguez, W.; Mo, Q.; Cen, L.; Adeegbe, D. Coupling the immunomodulatory properties of the HDAC6 inhibitor ACY241 with Oxaliplatin promotes robust anti-tumor response in non-small cell lung cancer. *Oncoimmunology* 2022, 11, 2042065.

88. Woan, K.V.; Lienlaf, M.; Perez-Villaroel, P.; Lee, C.; Cheng, F.; Knox, T.; Woods, D.M.; Barrios, K.; Powers, J.; Sahakian, E.; et al. Targeting histone deacetylase 6 mediates a dual anti-melanoma

effect: Enhanced antitumor immunity and impaired cell proliferation. *Mol. Oncol.* 2015, 9, 1447–1457.

89. Knox, T.; Sahakian, E.; Banik, D.; Hadley, M.; Palmer, E.; Noonepalle, S.; Kim, J.; Powers, J.; Gracia-Hernandez, M.; Oliveira, V.; et al. Selective HDAC6 inhibitors improve anti-PD-1 immune checkpoint blockade therapy by decreasing the anti-inflammatory phenotype of macrophages and down-regulation of immunosuppressive proteins in tumor cells. *Sci. Rep.* 2019, 9, 6136.

90. Yussuf Khamis, M.; Wu, H.P.; Ma, Q.; Li, Y.H.; Ma, L.Y.; Zhang, X.H.; Liu, H.M. Overcome the tumor immunotherapy resistance by combination of the HDAC6 inhibitors with antitumor immunomodulatory agents. *Bioorg. Chem.* 2021, 109, 104754.

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