

Gut Microbiome in Schizophrenia

Subjects: **Nutrition & Dietetics**

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The gut microbiome (GMB) plays an important role in developmental processes and has been implicated in the etiology of psychiatric disorders.

gut microbiome

gut–brain axis

schizophrenia

antipsychotics

prebiotics

probiotics

psychobiotics

1. Introduction

A relationship by which the gut microbiota (GMB) may interact with the central nervous system (CNS) through the gut–brain axis has long been construed. The gut–brain axis is a complex bilateral communication network that ensures proper maintenance of the gastrointestinal (GI) homeostasis. The fundamental mechanisms of the gut–brain axis communications involve neuro-immunoendocrine mediators through the CNS, the autonomic nervous system, the enteric nervous system and the hypothalamic pituitary adrenal axis ^[1]. In recent decades, studies have reported causal effects of the GMB on human behaviour and on the structure and function of the brain along with elucidating the underlying molecular mechanisms ^{[2][3][4]}.

Current evidence indicates that communication from the GMB to the brain primarily occurs through neuroimmune and neuroendocrine mechanisms, often involving the vagus nerve ^[5]. This bottom-up communication is mediated by several microbial-derived molecules, with the best-elucidated examples including short-chain fatty acids, secondary bile acids and tryptophan metabolites ^{[6][7][8]}. While some of these microbial-derived molecules interact directly with enteroendocrine cells, enterochromaffin cells and the mucosal immune system, others may penetrate the intestinal barrier to enter systemic circulation. Although some of the molecules are able to cross the blood–brain barrier, it remains unclear whether they interact with brain sites directly or only through neural signaling via spinal and/or vagal afferents ^[5]. Additionally, the GMB can independently produce or contribute to the production of several neuroactive molecules such as serotonin (5HT), norepinephrine, dopamine and γ -aminobutyric acid ^{[9][10][11]}. However, it is unknown if they reach significant concentrations or relevant receptors to elicit a response. Through evolution, the human immune system has maintained a symbiotic relationship with the microbiota. The GMB was shown to play an important role in regulating various immune functions in mice such as autoimmunity, defense and inflammation. The GMB has been implicated in the development and function of the CNS immune cells, specifically microglia ^[12]. Relative to controls, GF mice have diminished microglia maturation leading to higher risk of pathogen exposure. Adult specific pathogen-free mice treated with antibiotics led to microglia re-entering immature status which can be rescued through recolonization with a complex microbiota. Treatment with

antibiotics reverts microglia back to an immature state, suggesting continuous signaling of the GMB is needed for microglial function ^[12]. Disruption of the dynamic interaction between the two can result in profound effects on human health.

2. The Gut Microbiome in Behavior and Psychiatric Diseases

Several experimental approaches in animal models have been used to study the influence of the GMB on the gut–brain axis, including fecal microbial transplantation ^{[13][14]}, colonization with human ^{[14][15]} or synthetic ^[16] microbiota, manipulation of the GMB with antibiotics ^{[13][17][18]} and probiotics ^{[19][20][21]} and germ-free (GF) animal models ^{[13][22][23][24]}. It was demonstrated that the absence of normal GMB in GF mice has significant effects on behaviour such as abnormal memory, sociability, anxiety-like behaviours and motor performance ^[23]. These behavioural abnormalities could be rescued by colonization at the neonatal stage. In addition to anxiety-like behaviours ^[19], the GMB is also implicated in relation to stress-responsiveness ^{[19][22]} and depression-like behaviours ^{[25][26]}.

3. Antipsychotics and the Potential Role of the Gut Microbiome

3.1. Antipsychotic-Induced Metabolic Side Effects

Psychiatric patients have a higher relative risk of metabolic syndrome, a combination of cardiovascular risk factors such as dyslipidemia, hypertension, stroke and obesity, than the general population ^[27]. Moreover, the prevalence of metabolic side effects, type 2 diabetes and antipsychotic-induced weight gain (AIWG), is increased with the use of antipsychotics, specifically with olanzapine and clozapine ^{[27][28][29]}. Although the exact mechanisms by which metabolic dysfunction occurs remain unclear, several specific mechanisms of AIWG are supported by research evidence. One mechanism proposed is the antagonism of serotonergic and histaminergic receptors, resulting in increased appetite ^[30]. Additionally, genetic factors such as associations with 5HTR2C gene promoter polymorphisms or MC4R gene variants have been found to be associated with AIWG ^{[30][31][32]}.

Evidence also suggests that peptide hormones might mediate AIWG as studies have found correlations between antipsychotics, weight gain and expression of peptide hormones such as leptin and C-peptide ^{[33][34]}. Antipsychotics can disrupt mitochondria function, related enzyme activity and ATP levels through downregulation in genes encoding subunits of the electron transport chain ^[35]. Our groups initiated a pilot study examining the effects of antipsychotics in patients with early psychosis, as well as in patients starting clozapine and in patients being chronically treated with clozapine in order to better understand the potential causal roles of antipsychotic medications and induced metabolic abnormalities ^[36].

The GMB has been proposed as a potential target in relation to AIWG and other metabolic dysfunction due to its ability to regulate metabolism, homeostasis and energy balance ^{[37][38][39][40]}. Morgan et al. (2014) demonstrated that the presence of a gut microbiome was necessary and sufficient for olanzapine-induced weight gain in GF

mice. They proposed olanzapine's effect on AIWG to be through its in vitro antimicrobial activity against resident enteric bacteria [41]. Sex may play a role as females have been observed to be more susceptible to metabolic side effects such as weight gain than males in both animal and human models, with some studies reporting a decrease in body weight in male rats [29][39][41][42][43]. Evidence suggests a lipogenic effect when administering olanzapine in rodents; however, significant weight gain is only seen in female rats [42]. Furthermore, co-treatment of minocycline with olanzapine prevents AIWG in mice. However, coadministration of tetracycline, a closely related antibiotic showed no effects on AIWG, suggesting the mechanism behind minocycline is distinct from its antibiotic properties [44]. Olanzapine was also observed to alter the GMB profile in both male and female-treated rats, specifically increased levels of *Firmicutes* and decreased levels of *Bacteroidetes* [37]. Furthermore, treatment with prebiotics or antibiotics attenuated the olanzapine-induced metabolic dysfunction, including AIWG in female rats [38][45]. However, Kao et al. (2018) observed no change in GMB composition after olanzapine treatment, which may have been due to the short duration and variable dose of olanzapine treatment. The prebiotic mixture alone was reported to have some notable effects such as increasing *Bifidobacteria* spp. and reducing species within the *Firmicutes* (*Coprococcus*, *Oscillibacter*, *C. coccoides*, *Roseburia intestinalis* cluster, *Clostridium XVIII* cluster) and *Proteobacteria* (*Escherichia/Shigella* spp.) phyla [45]. Furthermore, olanzapine was observed to attenuate this effect of the prebiotic mixture, which suggests the antipsychotic does influence the GMB.

3.2. Effects of Antipsychotics on the Microbiome

Variety of antipsychotics used in various research studies may play a role in this inconsistency as different antipsychotics have been shown to have different effects on the GMB [38][39][43]. A study examining the effects of APs on the GMB in patients with bipolar disorder found a significant decrease in species diversity in females [46]. Recent in vitro work extensively examined over 1000 marketed drugs and found that 24% of the drugs with human targets inhibited the growth of at least one strain [47]. Among these drugs, APs were overrepresented with 26 out of 37 having antibacterial activity and displayed a similar pattern when targeting specific species. The authors hypothesized that direct bacterial inhibition may not only be a side effect of the APs, but also be part of their mechanism of action [47]. However, clozapine, risperidone and olanzapine were reported to lack any antibacterial activity.

When investigating the GMB in first-episode SCZ patients, chronic SCZ patients and healthy controls, Ma et al. (2020) found that chronically antipsychotic-treated SCZ patients had increased abundance in family *Enterococcaceae* and *Lactobacillaceae* and in genus *Enterococcus*, *Escherichia*, *Lactobacillus*, *Shigella*, *Streptococcus* and *Veillonella* relative to first-episode SCZ patients [48]. The same microbiota levels were similar or increased in abundance in first-episode SCZ patients compared to healthy controls, suggesting that antipsychotics may have negative effects on microbiota levels. In a similar fashion, relative abundances of family *Peptostreptococcaceae* and *Veillonellaceae* and genus *Fusobacterium* and *Megasphaera* were increased in chronic SCZ patients compared to first-episode SCZ patients but remained unchanged between first-episode SCZ patients and healthy controls, suggesting a normalizing effect of antipsychotics on these microbiota levels [48].

Bahr et al. (2015) reported that chronic treatment of risperidone in male children ($n = 18$) was associated with a lower *Bacteroidetes:Firmicutes* ratio and an increase in BMI. Furthermore, a gradual decrease in the *Bacteroidetes:Firmicutes* ratio was seen over the course of risperidone treatment [39].

Yuan et al. (2018) looked at the effects of risperidone in drug-naïve, normal weight, first-episode SCZ patients. After a 24-week treatment plan, there were significant increases in relative abundance of fecal *Bifidobacterium* spp. and *Escherichia coli* and significant decreases in the abundance of fecal *Clostridium coccoides* group and *Lactobacillus* spp. Increases in body weight, BMI, blood-glucose, triglycerides and C-reactive protein were observed as well. Antipsychotic-naïve patients with first-episode SCZ were found to have reduced levels of *Bifidobacterium* spp., *Escherichia coli* and *Lactobacillus* spp. and increased levels of *Clostridium coccoides* group compared to matched healthy controls. However, levels of *Lactobacillus* spp. and *Bifidobacterium* spp. were elevated in first-episode psychosis patients compared to controls following antipsychotic treatment [49][50]. Weight gain was found to be correlated with GMB composition of first-episode SCZ patients treated with risperidone. Specifically, increase in the relative abundance of *Bifidobacterium* spp. strongly correlated with weight gain.

However, another study conducted by Pelka-Wysiecka et al. (2019) showed differing results. The study analyzed the effect of olanzapine on the GMB in 20 SCZ patients [50]. Patients underwent a 7-day washout of all psychiatric medication, received standard hospital diet and were administered olanzapine treatment. Stool samples were taken at baseline after the washout and after 6 weeks of treatment. Similar to previous studies, only women experienced a significant increase in BMI. No significant changes were observed in alpha diversity and GMB composition [51]. However, the absence of significant changes may be due to the short intervention period.

Zhu et al. (2020) looked at the effects of APs on the GMB by following up with 38 medication-free patients with SCZ at baseline and 3 months after treatment. 26 microbial species were identified to differ between medication-free SCZ patients and HCs at baseline [52]. However, 20 species remained altered after 3 months of AP treatment compared with controls. Throughout AP treatment, authors reported 28 differentially abundant bacterial species, five of which were included in the 26 operational taxonomic unit SCZ classifiers [52]. The authors suggest that the GMB is influenced by APs but is not completely restored from SCZ-associated alterations.

Current clinical studies provide little evidence on the relationship between the APs, GMB and metabolic changes. Given the inconsistent findings highlighted in this review, future studies should ideally address the following: larger patient samples, with standardized medications, use of homogeneous populations (e.g., age, sex, BMI, symptom severity), comparable study designs and accounting for geographical diversity.

4. Evidence for Psychobiotic Intervention in Schizophrenia

A GMB is essential for optimal function of the immune system and immune dysfunction has been implicated in SCZ. Consequently, alterations in the GMB may play a key role in the etiology and treatment response of SCZ through bacterial infections and contributes to a chronic inflammatory state. The studies by Shen et al. (2018) and Zhang et al. (2020) found reduced levels of the genera: *Roseburia* and *Faecalibacterium* [53][54]. Notably, both

genera play an important role in maintaining the intestinal barrier through the production of butyrate [55][56]. Braniste et al. (2014) concluded that not only the presence of a GMB is essential for normal development of the blood–brain barrier, but studies also indicated that the GMB can also regulate its permeability [57]. Therefore, through such mechanisms, disturbances to the GMB can lead to CNS infection and inflammation.

Although many findings were inconsistent, a few taxonomic groups were repeatedly reported to be altered in SCZ patients (Table 1). Multiple studies reported increased species within the *Fusobacterium*, *Lactobacillus*, *Megasphaera* and *Prevotella* genera, most of which are gram-negative bacteria [58][59][52][53][54][60][48][61]. While gram-negative bacteria are common in normal gut flora, increased permeability of the gut wall may result in systemic circulation of enteric inflammatory molecules such as lipopolysaccharides (LPS). An in vitro study examined the effects of LPS on intestinal epithelial cells and found that acute administration of LPS resulted in altered and reduced distribution of tight junctions [62]. LPS has been shown to be an effective neurodevelopmental model of SCZ in rodents [63][64][65]. In addition, to LPS, SCZ patients have been shown to have increased inflammatory cytokines which may contribute to the change in gut permeability and development of a “leaky gut” [66].

Table 1. List of taxonomic groups that were significantly increased/decreased in multiple studies.

Taxa	Increased in Schizophrenia	Decreased in Schizophrenia
Phylum: <i>Firmicutes</i>	Castro-Nallar et al., 2015 (Oral) [59]	Shen et al., 2018 [53]
Phylum: <i>Proteobacteria</i>	Shen et al., 2018 [53]	
	Ma et al., 2020 (vs HC only) [48]	Nguyen et al., 2019 [54]
	Zhang et al., 2020 (FSCZ) [67]	
Class: <i>Deltaproteobacteria</i>	Xu et al., 2020 [61]	
	Zhang et al., 2020 (FSCZ) [67]	
Order: <i>Actinomycetales</i>	Xu et al., 2020 [61]	
	Zhang et al., 2020 (FSCZ) [67]	
Order: <i>Clostridiales</i>	He et al., 2018 (High-risk for SCZ) [58]	Shen et al., 2018 [53]

Taxa	Increased in Schizophrenia	Decreased in Schizophrenia	
Family: <i>Enterobacteriaceae</i>	Shen et al., 2018 [53]	Zheng et al., 2019 [60]	
	Ma et al., 2020 (vs. HC only) [48]		
Family: <i>Enterococcaceae</i>	Ma et al., 2020 [48]	Xu et al., 2020 [61]	
Family: <i>Lactobacillaceae</i>	Shen et al., 2018 [53]	Shen et al., 2018 [53] Zheng et al., 2019 [60] Zhang et al., 2020 (FSCZ) [67]	
	Ma et al., 2020 [48]		
Family: <i>Lachnospiraceae</i>			
Family: <i>Prevotellaceae</i>	Shen et al., 2018 [53]		
	Zheng et al., 2019 [60]		
Family: <i>Rikenellaceae</i>	Zheng et al., 2019	Xu et al., 2020 [61]	
Family: <i>Veillonellaceae</i>	Shen et al., 2018 [53]		
	Zheng et al., 2019 [60]		
	Ma et al., 2020 (vs. FSCZ only) [48]		
Genus: <i>Acidaminococcus</i>	Shen et al., 2018 [53]		
	Zhu et al., 2020 (FSCZ) [52]		
Genus: <i>Akkermansia</i>	Zheng et al., 2019 [60]		

Taxa	Increased in Schizophrenia	Decreased in Schizophrenia
Genus: <i>Anaerotruncus</i>	Zhu et al., 2020 (FSCZ) [52]	
	Zhang et al., 2020 (FSCZ) [67]	
	Zhu et al., 2020 (FSCZ) [52]	
Genus: <i>Bifidobacterium</i>	Castro-Nallar et al., 2015 (Oral) [59]	Yuan et al., 2018 (FSCZ) [49]
	Zhu et al., 2020 (FSCZ) [52]	
Genus: <i>Blautia</i>	Nguyen et al., 2019 [54]	Shen et al., 2018 [53]
	Zhang et al., 2020 (FSCZ) [67]	Zheng et al., 2019 [60]
Genus: <i>Citrobacter</i>	Shen et al., 2018 [53]	
	Ma et al., 2020 (vs. FSCZ only) [48]	Zheng et al., 2019 [60]
	Zhu et al., 2020 (FSCZ) [52]	
Genus: <i>Clostridium</i>	Shen et al., 2018 [53]	
	Ma et al., 2020 (vs. FSCZ only) [48]	
Genus: <i>Coprobacillus</i>	Ma et al., 2020 (vs. HC only) [48]	
	Zhu et al., 2020 (FSCZ) [52]	
Genus: <i>Coprococcus</i>		Shen et al., 2018 [53]
	Zhang et al., 2020 (FSCZ) [67]	Zheng et al., 2019 [60]
Genus: <i>Eggerthella</i>	Xu et al., 2020 [61]	

Taxa	Increased in Schizophrenia	Decreased in Schizophrenia
	Zhang et al., 2020 (FSCZ) [67]	
Genus: <i>Enterococcus</i>	Ma et al., 2020 [48] Zhu et al., 2020 (FSCZ) [52]	Xu et al., 2020 [61]
Genus: <i>Fusobacterium</i>	Shen et al., 2018 [53] Zheng et al., 2019 [60] Ma et al., 2020 (vs. FSCZ only) [48]	
Genus: <i>Lactobacillus</i>	Castro-Nallar et al., 2015 (Oral) [59] He et al., 2018 (High-risk for SCZ) [58] Shen et al., 2018 [53] Ma et al., 2020 [48] Zhu et al., 2020 (FSCZ) [52]	Yuan et al., 2018 (FSCZ) [49]
Genus: <i>Megasphaera</i>	Shen et al., 2018 [53] Nguyen et al., 2019 [54] Zheng et al., 2019 [60] Ma et al., 2020 (vs. FSCZ only) [48] Xu et al., 2020 [61]	
Genus: <i>Prevotella</i>	He et al., 2018 (High-risk for SCZ) [58] Shen et al., 2018 [53]	Yolken et al., 2020 (Oral) [68]

Taxa	Increased in Schizophrenia	Decreased in Schizophrenia
	Zheng et al., 2019 [60] Zhang et al., 2020 (FSCZ) [67]	
Genus: <i>Ruminococcus</i>	Nguyen et al., 2019 [54] Ma et al., 2020 (vs. FSCZ only) [48]	Zhang et al., 2020 (FSCZ) [67]
Genus: <i>Streptococcus</i>	Ma et al., 2020 [48] Yolken et al., 2020 (Oral) [68]	Shen et al., 2018 [53]
Genus: <i>Veillonella</i>	Ma et al., 2020 [48] Zhu et al., 2020 (FSCZ) [52]	
Species: <i>Akkermansia muciniphila</i>	Xu et al., 2020 [61] Zhu et al., 2020 (vs. HC only) [52]	
Species: <i>Bacteroides eggerthii</i>		Shen et al., 2018 [53] Zheng et al., 2019 [60]
Species: <i>Bifidobacterium adolescentis</i>	Shen et al., 2018 [53] Xu et al., 2020 [61] Zhu et al., 2020 (vs. HC only) [52]	
Species: <i>Escherichia coli</i>	Zhu et al., 2020 (vs. FSCZ only) [52]	Yuan et al., 2018 (FSCZ) [49]
Species: <i>Eubacterium hallii</i>	Castro-Nallar et al., 2015 (Oral) [59]	Zhu et al., 2020 (vs FSCZ only) [52]

Notably, SCZ patients were reported to have lower levels of the family *Lachnospiraceae*, which has been reported to be beneficial in health such as butyrate and other short-chain fatty acid production. As previously mentioned, the

Taxa	Increased in Schizophrenia	Decreased in Schizophrenia [55] [56]
Species: <i>Lactobacillus gasseri</i>	[69] Castro-Nallar et al., 2015 (Oral) [59]	
	[19] [22] [70] [71] [72] [73] Xu et al., 2020 [61]	
Species: <i>Lactobacillus ruminis</i>	[74] [75]	
	He et al., 2019 (High-risk for SCZ) [58]	[76]
	Zhu et al., 2020 (vs. HC only) [52]	Zhu et al., 2020 (vs FSCZ) [52] [26] [76] [77] [78] [79] .
Species: <i>Lactobacillus salivarius</i>	Castro-Nallar et al., 2015 (Oral) [59]	
	Zhu et al., 2020 (vs. HC only) [52] _{TM}	
Species: <i>Haemophilus parainfluenzae</i>		[45]
		Nguyen et al., 2019 [54]
		Zheng et al., 2019 [60]
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rhamnosus and *Bifidobacterium animalis*. Probiotic supplementation led to a reduction of the acute phase reactant von Willebrand factor in SCZ patients, suggested to be the secondary effect of probiotic-induced improvement of intestinal epithelium integrity. Patients treated with probiotics also were less likely to develop severe bowel difficulties [\[81\]](#). Bowel difficulty was reported to be positively correlated with seropositivity of *Candida albicans* [\[82\]](#). Positive symptoms of SCZ were significantly improved in males who were seronegative for *Candida albicans* compared to those who were seropositive within the 13-week timepoint. However, no significant effects on the PANSS scores were seen for the full duration of the study.

In a more recent study, probiotic *Bifidobacterium breve* A-1 was given to SCZ patients for 4 weeks and improvements were seen in anxiety, depression and PANSS scores [\[83\]](#). Elevated levels of various interleukins were also measured, including IL-22 and tumour necrosis factor-related activation induced cytokine (TRANCE). The authors suggested that the reported symptom improvements could be due to the critical roles IL-22 and TRANCE play in the function of the gut epithelial barrier.

Ghaderi et al. (2019) investigated probiotic supplement treatment in SCZ patients by administering a combination of vitamin D and probiotic mixture containing *Bifidobacterium bifidum*, *Lactobacillus acidophilus*, *Lactobacillus fermentum* and *Lactobacillus reuteri* for 12 weeks [\[84\]](#). A significant decrease in metabolic abnormalities and circulating C-reactive protein were noted, indicating reduced inflammation, alongside improvements in general and total PANSS scores and plasma total antioxidant capacity. However, it is uncertain whether vitamin D, probiotic supplement, or a combination of both are responsible for the observed improvements.

References

1. Carabotti, M.; Scirocco, A.; Maselli, M.A.; Severi, C. The gut-brain axis: Interactions between enteric microbiota, central and enteric nervous systems. *Ann. Gastroenterol.* 2015, 28, 203–209.
2. Rea, K.; Dinan, T.G.; Cryan, J.F. The microbiome: A key regulatory of stress and neuroinflammation. *Neurobiol. Stress* 2016, 4, 23–33.
3. Dinan, T.G.; Cryan, J.F. Gut-brain axis in 2016: Brain-gut-microbiota axis mood, metabolism and behavior. *Nat. Rev. Gastroenterol. Hepatol.* 2017, 14, 69–70.
4. Ma, Q.; Xing, C.; Long, W.; Wang, H.Y.; Liu, Q.; Wang, R.-F. Impact of microbiota on central nervous system and neurological diseases: The gut-brain axis. *J. NeuroInflamm.* 2019, 16, 1–14.
5. Tolhurst, G.; Heffron, H.; Lam, Y.S.; Parker, H.E.; Habib, A.M.; Diakogiannaki, E.; Cameron, J.; Grosse, J.; Reimann, F.; Gribble, F.M. Short-Chain Fatty Acids Stimulate Glucagon-Like Peptide-1 Secretion via the G-Protein-Coupled Receptor FFAR2. *Diabetes* 2011, 61, 364–371.
6. Shishov, V.A.; Kirovskaia, T.A.; Kudrin, V.S.; Oleskin, A.V. Amine neuromediators, their precursors, and oxidation products in the culture of *Escherichia coli* K-12. *Prikl. Biokhimiia i Mikrobiol.* 2009, 45, 550–554.
7. Wikoff, W.R.; Anfora, A.T.; Liu, J.; Schultz, P.G.; Lesley, S.A.; Peters, E.C.; Siuzdak, G. Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. *Proc. Natl. Acad. Sci. USA* 2009, 106, 3698–3703.
8. Yano, J.M.; Yu, K.; Donaldson, G.P.; Shastri, G.G.; Ann, P.; Ma, L.; Nagler, C.R.; Ismagilov, R.F.; Mazmanian, S.K.; Hsiao, E.Y. Indigenous Bacteria from the Gut Microbiota Regulate Host Serotonin Biosynthesis. *Cell* 2015, 161, 264–276.
9. Russell, W.R.; Gratz, S.W.; Duncan, S.H.; Holtrop, G.; Ince, J.; Scobbie, L.; Duncan, G.; Johnstone, A.M.; E Lobley, G.; Wallace, R.J.; et al. High-protein, reduced-carbohydrate weight-loss diets promote metabolite profiles likely to be detrimental to colonic health. *Am. J. Clin. Nutr.* 2011, 93, 1062–1072.
10. Asano, Y.; Hiramoto, T.; Nishino, R.; Aiba, Y.; Kimura, T.; Yoshihara, K.; Koga, Y.; Sudo, N. Critical role of gut microbiota in the production of biologically active, free catecholamines in the gut lumen of mice. *Am. J. Physiol. Liver Physiol. Gastrointest. Liver Physiol.* 2012, 303, G1288–G1295.
11. Barrett, E.; Ross, R.P.; O'Toole, P.W.; Fitzgerald, G.F.; Stanton, C. γ -Aminobutyric acid production by culturable bacteria from the human intestine. *J. Appl. Microbiol.* 2012, 113, 411–417.
12. Erny, D.; De Angelis, A.L.H.; Jaitin, D.; Wieghofer, P.; Staszewski, O.; David, E.; Keren-Shaul, H.; Mhlahoi, T.; Jakobshagen, K.; Buch, T.; et al. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat. Neurosci.* 2015, 18, 965–977.

13. Bercik, P.; Denou, E.; Collins, J.; Jackson, W.; Lu, J.; Jury, J.; Deng, Y.; Blennerhassett, P.; Macri, J.; McCoy, K.D.; et al. The Intestinal Microbiota Affect Central Levels of Brain-Derived Neurotropic Factor and Behavior in Mice. *Gastroenterology* 2011, 141, 599–609.e3.
14. De Palma, G.; Lynch, M.D.J.; Lu, J.; Dang, V.T.; Deng, Y.; Jury, J.; Umeh, G.; Miranda, P.M.; Pastor, M.P.; Sidani, S.; et al. Transplantation of fecal microbiota from patients with irritable bowel syndrome alters gut function and behavior in recipient mice. *Sci. Transl. Med.* 2017, 9, eaaf6397.
15. Wrzosek, L.; Ciocan, D.; Borentain, P.; Spatz, M.; Puchois, V.; Hugot, C.; Ferrere, G.; Mayeur, C.; Perlemuter, G.; Cassard, A.-M. Transplantation of human microbiota into conventional mice durably reshapes the gut microbiota. *Sci. Rep.* 2018, 8, 1–9.
16. Desai, M.S.; Seekatz, A.M.; Koropatkin, N.M.; Kamada, N.; Hickey, C.A.; Wolter, M.; Pudlo, N.A.; Kitamoto, S.; Terrapon, N.; Muller, A.; et al. A Dietary Fiber-Deprived Gut Microbiota Degrades the Colonic Mucus Barrier and Enhances Pathogen Susceptibility. *Cell* 2016, 167, 1339–1353.e21.
17. Lee, K.; Vuong, H.E.; Nusbaum, D.J.; Hsiao, E.Y.; Evans, C.J.; Taylor, A.M.W. The gut microbiota mediates reward and sensory responses associated with regimen-selective morphine dependence. *Neuropsychopharmacology* 2018, 43, 2606–2614.
18. Lowe, P.P.; Gyongyosi, B.; Satishchandran, A.; Iracheta-Vellve, A.; Cho, Y.; Ambade, A.; Szabo, G. Reduced gut microbiome protects from alcohol-induced neuroinflammation and alters intestinal and brain inflammasome expression. *J. NeuroInflamm.* 2018, 15, 1–12.
19. Bravo, J.A.; Forsythe, P.; Chew, M.V.; Escaravage, E.; Savignac, H.M.; Dinan, T.G.; Bienenstock, J.; Cryan, J.F. Ingestion of *Lactobacillus* strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc. Natl. Acad. Sci. USA* 2011, 108, 16050–16055.
20. Celiberto, L.S.; Pinto, R.A.; Rossi, E.A.; Vallance, B.A.; Cavallini, D.C.U. Isolation and Characterization of Potentially Probiotic Bacterial Strains from Mice: Proof of Concept for Personalized Probiotics. *Nutrients* 2018, 10, 1684.
21. Lee, H.-J.; Lee, K.-E.; Kim, J.-K.; Kim, D.-H. Suppression of gut dysbiosis by *Bifidobacterium longum* alleviates cognitive decline in 5XFAD transgenic and aged mice. *Sci. Rep.* 2019, 9, 1–12.
22. Sudo, N.; Chida, Y.; Aiba, Y.; Sonoda, J.; Oyama, N.; Yu, X.-N.; Kubo, C.; Koga, Y. Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. *J. Physiol.* 2004, 558, 263–275.
23. Luk, B.; Veeraragavan, S.; Engevik, M.; Balderas, M.; Major, A.; Runge, J.; Luna, R.A.; Versalovic, J. Postnatal colonization with human "infant-type" *Bifidobacterium* species alters behavior of adult gnotobiotic mice. *PLoS ONE* 2018, 13, e0196510.
24. Luo, Y.; Zeng, B.; Zeng, L.; Du, X.; Li, B.; Huo, R.; Liu, L.; Wang, H.; Dong, M.; Pan, J.; et al. Gut microbiota regulates mouse behaviors through glucocorticoid receptor pathway genes in the

- hippocampus. *Transl. Psychiatry* 2018, 8, 1–10.
25. Kelly, J.R.; Borre, Y.; Brien, C.O.; Patterson, E.; El Aidy, S.; Deane, J.; Kennedy, P.J.; Beers, S.; Scott, K.; Moloney, G.; et al. Transferring the blues: Depression-associated gut microbiota induces neurobehavioural changes in the rat. *J. Psychiatr. Res.* 2016, 82, 109–118.
 26. Zheng, P.; Zeng, B.; Zhou, C.; Liu, M.; Fang, Z.; Xu, X.; Zeng, L.; Chen, J.; Fan, S.; Du, X.; et al. Gut microbiome remodeling induces depressive-like behaviors through a pathway mediated by the host's metabolism. *Mol. Psychiatry* 2016, 21, 786–796.
 27. Penninx, B.W.J.H.; Lange, S.M.M. Metabolic syndrome in psychiatric patients: Overview, mechanisms, and implications. *Dialog. Clin. Neurosci.* 2018, 20, 63–73.
 28. Gressier, F.; Porcelli, S.; Calati, R.; Serretti, A. Pharmacogenetics of clozapine response and induced weight gain: A comprehensive review and meta-analysis. *Eur. Neuropsychopharmacol.* 2016, 26, 163–185.
 29. Vancampfort, D.; Correll, C.U.; Galling, B.; Probst, M.; De Hert, M.; Ward, P.B.; Rosenbaum, S.; Gaughran, F.P.; Lally, J.A.; Stubbs, B. Diabetes mellitus in people with schizophrenia, bipolar disorder and major depressive disorder: A systematic review and large scale meta-analysis. *World Psychiatry* 2016, 15, 166–174.
 30. Aringhieri, S.; Carli, M.; Kolachalam, S.; Verdesca, V.; Cini, E.; Rossi, M.; McCormick, P.J.; Corsini, G.U.; Maggio, R.; Scarselli, M. Molecular targets of atypical antipsychotics: From mechanism of action to clinical differences. *Pharmacol. Ther.* 2018, 192, 20–41.
 31. Sicard, M.N.; Zai, C.C.; Tiwari, A.K.; Souza, R.P.; Meltzer, H.Y.; A Lieberman, J.; Kennedy, J.L.; Müller, D.J. Polymorphisms of the HTR2C gene and antipsychotic-induced weight gain: An update and meta-analysis. *Pharmacogenomics* 2010, 11, 1561–1571.
 32. Chowdhury, N.I.; Souza, R.P.; Tiwari, A.K.; Brandl, E.J.; Sicard, M.; Meltzer, H.Y.; Lieberman, J.A.; Kennedy, J.L.; Muller, D.J. Investigation of melanocortin system gene variants in antipsychotic-induced weight gain. *World J. Biol. Psychiatry* 2014, 15, 251–258.
 33. Ragguett, R.-M.; Hahn, M.; Messina, G.; Chieffi, S.; Monda, M.; De Luca, V. Association between antipsychotic treatment and leptin levels across multiple psychiatric populations: An updated meta-analysis. *Hum. Psychopharmacol. Clin. Exp.* 2017, 32, e2631.
 34. Pisano, S.; Coppola, G.; Catone, G.; Carotenuto, M.; Iuliano, R.; D'Esposito, V.; Cabaro, S.; Del Giudice, E.M.; Bravaccio, C.; Formisano, P. Differences in Metabolic Factors Between Antipsychotic-Induced Weight Gain and Non-pharmacological Obesity in Youths. *Clin. Drug Investig.* 2018, 38, 457–462.
 35. Scaini, G.; Quevedo, J.; Velligan, D.; Roberts, D.L.; Raventos, H.; Walss-Bass, C. Second generation antipsychotic-induced mitochondrial alterations: Implications for increased risk of

- metabolic syndrome in patients with schizophrenia. *Eur. Neuropsychopharmacol.* 2018, 28, 369–380.
36. Gorbovskaya, I.; Kanji, S.; Liu, J.C.; MacKenzie, N.E.; Agarwal, S.M.; Marshe, V.S.; Srirenakumar, V.; Verdu, E.F.; Bercik, P.; De Palma, G.; et al. Investigation of the Gut Microbiome in Patients with Schizophrenia and Clozapine-Induced Weight Gain: Protocol and Clinical Characteristics of First Patient Cohorts. *Neuropsychobiology* 2020, 79, 5–12.
 37. Davey, K.J.; O'Mahony, S.M.; Schellekens, H.; O'Sullivan, O.; Bienenstock, J.; Cotter, P.D.; Dinan, T.G.; Cryan, J.F. Gender-dependent consequences of chronic olanzapine in the rat: Effects on body weight, inflammatory, metabolic and microbiota parameters. *Psychopharmacology* 2012, 221, 155–169.
 38. Davey, K.J.; Cotter, P.D.; Osullivan, O.; Crispie, F.; Dinan, T.G.; Cryan, J.F.; O'Mahony, S.M. Antipsychotics and the gut microbiome: Olanzapine-induced metabolic dysfunction is attenuated by antibiotic administration in the rat. *Transl. Psychiatry* 2013, 3, e309.
 39. Bahr, S.M.; Tyler, B.; Wooldridge, N.; Butcher, B.; Burns, T.L.; Teesch, L.M.; Oltman, C.L.; Azcarate-Peril, M.; Kirby, J.R.; A Calarge, C. Use of the second-generation antipsychotic, risperidone, and secondary weight gain are associated with an altered gut microbiota in children. *Transl. Psychiatry* 2015, 5, e652.
 40. Torres-Fuentes, C.; Schellekens, H.; Dinan, T.G.; Cryan, J.F. The microbiota–gut–brain axis in obesity. *Lancet Gastroenterol. Hepatol.* 2017, 2, 747–756.
 41. Morgan, A.P.; Crowley, J.J.; Nonneman, R.J.; Quackenbush, C.R.; Miller, C.N.; Ryan, A.K.; Bogue, M.A.; Paredes, S.H.; Yourstone, S.; Carroll, I.M.; et al. The Antipsychotic Olanzapine Interacts with the Gut Microbiome to Cause Weight Gain in Mouse. *PLoS ONE* 2014, 9, e115225.
 42. Fernø, J.; Ersland, K.; Duus, I.; González-García, I.; Fossan, K.; Berge, R.; Steen, V.; Skrede, S. Olanzapine depot exposure in male rats: Dose-dependent lipogenic effects without concomitant weight gain. *Eur. Neuropsychopharmacol.* 2015, 25, 923–932.
 43. Kraal, A.Z.; Ward, K.M.; Ellingrod, V.L. Sex Differences in Antipsychotic Related Metabolic Functioning in Schizophrenia Spectrum Disorders. *Psychopharmacol. Bull.* 2017, 47, 8–21.
 44. Perez-Gomez, A.; Carretero, M.; Weber, N.; Peterka, V.; To, A.; Titova, V.; Solis, G.; Osborn, O.; Petrascheck, M. A phenotypic *Caenorhabditis elegans* screen identifies a selective suppressor of antipsychotic-induced hyperphagia. *Nat. Commun.* 2018, 9, 1–12.
 45. Kao, A.C.-C.; Spitzer, S.; Anthony, D.C.; Lennox, B.; Burnet, P.W.J. Prebiotic attenuation of olanzapine-induced weight gain in rats: Analysis of central and peripheral biomarkers and gut microbiota. *Transl. Psychiatry* 2018, 8, 1–12.
 46. Flowers, S.A.; Evans, S.J.; Ward, K.M.; McInnis, M.G.; Ellingrod, V.L. Interaction between Atypical Antipsychotics and the Gut Microbiome in a Bipolar Disease Cohort. *Pharmacother. J. Hum.*

Pharmacol. Drug Ther. 2017, 37, 261–267.

47. Maier, L.; Pruteanu, M.; Kuhn, M.; Zeller, G.; Telzerow, A.; Anderson, E.E.; Brochado, A.R.; Fernandez, K.C.; Dose, H.; Mori, H.; et al. Extensive impact of non-antibiotic drugs on human gut bacteria. *Nat. Cell Biol.* 2018, 555, 623–628.
48. Ma, X.; Asif, H.; Dai, L.; He, Y.; Zheng, W.; Wang, D.; Ren, H.; Tang, J.; Li, C.; Jin, K.; et al. Alteration of the gut microbiome in first-episode drug-naïve and chronic medicated schizophrenia correlate with regional brain volumes. *J. Psychiatr. Res.* 2020, 123, 136–144.
49. Yuan, X.; Zhang, P.; Wang, Y.; Liu, Y.; Li, X.; Kumar, B.U.; Hei, G.; Lv, L.; Huang, X.-F.; Fan, X.; et al. Changes in metabolism and microbiota after 24-week risperidone treatment in drug naïve, normal weight patients with first episode schizophrenia. *Schizophr. Res.* 2018, 201, 299–306.
50. Schwarz, E.; Maukonen, J.; Hyytiäinen, T.; Kieseppä, T.; Orešič, M.; Sabunciyan, S.; Mantere, O.; Saarela, M.; Yolken, R.; Suvisaari, J. Analysis of microbiota in first episode psychosis identifies preliminary associations with symptom severity and treatment response. *Schizophr. Res.* 2018, 192, 398–403.
51. Pełka-Wysiecka, J.; Kaczmarczyk, M.; Bąba-Kubiś, A.; Liśkiewicz, P.; Wroński, M.; Skonieczna-Żydecka, K.; Marlicz, W.; Misiak, B.; Starzyńska, T.; Kucharska-Mazur, J.; et al. Analysis of Gut Microbiota and Their Metabolic Potential in Patients with Schizophrenia Treated with Olanzapine: Results from a Six-Week Observational Prospective Cohort Study. *J. Clin. Med.* 2019, 8, 1605.
52. Zhu, F.; Ju, Y.; Wang, W.; Wang, Q.; Guo, R.; Ma, Q.; Sun, Q.; Fan, Y.; Xie, Y.; Yang, Z.; et al. Metagenome-wide association of gut microbiome features for schizophrenia. *Nat. Commun.* 2020, 11, 1612.
53. Shen, Y.; Xu, J.; Li, Z.; Huang, Y.; Yuan, Y.; Wang, J.; Zhang, M.; Hu, S.; Liang, Y. Analysis of gut microbiota diversity and auxiliary diagnosis as a biomarker in patients with schizophrenia: A cross-sectional study. *Schizophr. Res.* 2018, 197, 470–477.
54. Nguyen, T.T.; Kosciolk, T.; Maldonado, Y.; Daly, R.E.; Martin, A.S.; McDonald, D.; Knight, R.; Jeste, D.V. Differences in gut microbiome composition between persons with chronic schizophrenia and healthy comparison subjects. *Schizophr. Res.* 2019, 204, 23–29.
55. Peng, L.; Li, Z.-R.; Green, R.S.; Holzman, I.R.; Lin, J. Butyrate Enhances the Intestinal Barrier by Facilitating Tight Junction Assembly via Activation of AMP-Activated Protein Kinase in Caco-2 Cell Monolayers. *J. Nutr.* 2009, 139, 1619–1625.
56. Machiels, K.; Joossens, M.; Sabino, J.; De Preter, V.; Arijs, I.; Eeckhaut, V.; Ballet, V.; Claes, K.; Van Immerseel, F.; Verbeke, K.; et al. A decrease of the butyrate-producing species *Roseburia hominis* and *Faecalibacterium prausnitzii* defines dysbiosis in patients with ulcerative colitis. *Gut* 2013, 63, 1275–1283.

57. Braniste, V.; Al-Asmakh, M.; Kowal, C.; Anuar, F.; Abbaspour, A.; Tóth, M.; Korecka, A.; Bakocevic, N.; Ng, L.G.; Kundu, P.; et al. The gut microbiota influences blood-brain barrier permeability in mice. *Sci. Transl. Med.* 2014, 6, 263ra158.
58. He, Y.; Kosciolk, T.; Tang, J.; Zhou, Y.; Li, Z.; Ma, X.; Zhu, Q.; Yuan, N.; Yuan, L.; Li, C.; et al. Gut microbiome and magnetic resonance spectroscopy study of subjects at ultra-high risk for psychosis may support the membrane hypothesis. *Eur. Psychiatry* 2018, 53, 37–45.
59. Castro-Nallar, E.; Bendall, M.L.; Pérez-Losada, M.; Sabuncyan, S.; Severance, E.G.; Dickerson, F.B.; Schroeder, J.R.; Yolken, R.H.; Crandall, K.A. Composition, taxonomy and functional diversity of the oropharynx microbiome in individuals with schizophrenia and controls. *PeerJ* 2015, 3, e1140.
60. Zheng, P.; Zeng, B.; Liu, M.; Chen, J.; Pan, J.; Han, Y.; Liu, Y.; Cheng, K.; Zhou, C.; Wang, H.; et al. The gut microbiome from patients with schizophrenia modulates the glutamate-glutamine-GABA cycle and schizophrenia-relevant behaviors in mice. *Sci. Adv.* 2019, 5, eaau8317.
61. Xu, R.; Wu, B.; Liang, J.; He, F.; Gu, W.; Li, K.; Luo, Y.; Chen, J.; Gao, Y.; Wu, Z.; et al. Altered gut microbiota and mucosal immunity in patients with schizophrenia. *Brain Behav. Immun.* 2020, 85, 120–127.
62. Gorecki, A.M.; Preskey, L.; Bakeberg, M.C.; Kenna, J.E.; Gildenhuis, C.; MacDougall, G.; Dunlop, S.A.; Mastaglia, F.L.; Akkari, P.A.; Koengten, F.; et al. Altered gut microbiome in Parkinson's disease and the influence of lipopolysaccharide in a human α -synuclein over-expressing mouse model. *Front. Neurosci.* 2019, 13, 839.
63. Basta-Kaim, A.; Szczęśny, E.; Leśkiewicz, M.; Głombik, K.; Ślusarczyk, J.; Budziszewska, B.; Regulski, M.; Kubera, M.; Nowak, W.; Wędzony, K.; et al. Maternal immune activation leads to age-related behavioral and immunological changes in male rat offspring—The effect of antipsychotic drugs. *Pharmacol. Rep.* 2012, 64, 1400–1410.
64. Zhu, F.; Zhang, L.; Ding, Y.-Q.; Zhao, J.; Zheng, Y. Neonatal intrahippocampal injection of lipopolysaccharide induces deficits in social behavior and prepulse inhibition and microglial activation in rats: Implication for a new schizophrenia animal model. *Brain Behav. Immun.* 2014, 38, 166–174.
65. Waterhouse, U.; Roper, V.E.; Brennan, K.A.; Ellenbroek, B.A. Nicotine ameliorates schizophrenia-like cognitive deficits induced by maternal LPS exposure: A study in rats. *Dis. Models Mech.* 2016, 9, 1159–1167.
66. Prestwood, T.R.; Asgariroozbehani, R.; Wu, S.; Agarwal, S.M.; Logan, R.W.; Ballon, J.S.; Hahn, M.K.; Freyberg, Z. Roles of inflammation in intrinsic pathophysiology and antipsychotic drug-induced metabolic disturbances of schizophrenia. *Behav. Brain Res.* 2021, 402, 113101.

67. Zhang, X.; Pan, L.-Y.; Zhang, Z.; Zhou, Y.-Y.; Jiang, H.-Y.; Ruan, B. Analysis of gut mycobiota in first-episode, drug-naïve Chinese patients with schizophrenia: A pilot study. *Behav. Brain Res.* 2020, 379, 112374.
68. Yolken, R.; Prandovszky, E.; Severance, E.G.; Hatfield, G.; Dickerson, F. The oropharyngeal microbiome is altered in individuals with schizophrenia and mania. *Schizophr. Res.* 2020, in press.
69. Dinan, T.G.; Stanton, C.; Cryan, J.F. Psychobiotics: A Novel Class of Psychotropic. *Biol. Psychiatry* 2013, 74, 720–726.
70. Desbonnet, L.; Garrett, L.; Clarke, G.; Kiely, B.; Cryan, J.F.; Dinan, T.G. Effects of the probiotic *Bifidobacterium infantis* in the maternal separation model of depression. *Neuroscience* 2010, 170, 1179–1188.
71. Smith, C.J.; Emge, J.R.; Berzins, K.; Lung, L.; Khamishon, R.; Shah, P.; Rodrigues, D.M.; Sousa, A.J.; Reardon, C.; Sherman, P.M.; et al. Probiotics normalize the gut-brain-microbiota axis in immunodeficient mice. *Am. J. Physiol. Liver Physiol.* 2014, 307, G793–G802.
72. Bruce-Keller, A.J.; Salbaum, J.M.; Berthoud, H.-R. Harnessing Gut Microbes for Mental Health: Getting from Here to There. *Biol. Psychiatry* 2018, 83, 214–223.
73. Nishida, K.; Sawada, D.; Kuwano, Y.; Tanaka, H.; Rokutan, K. Health Benefits of *Lactobacillus gasseri* CP2305 Tablets in Young Adults Exposed to Chronic Stress: A Randomized, Double-Blind, Placebo-Controlled Study. *Nutrients* 2019, 11, 1859.
74. Gardsjord, E.S.; Romm, K.L.; Friis, S.; Barder, H.E.; Evensen, J.; Haahr, U.; Hegelstad, W.T.V.; Joa, I.; Johannessen, J.O.; Langeveld, J.; et al. Subjective quality of life in first-episode psychosis. A ten year follow-up study. *Schizophr. Res.* 2016, 172, 23–28.
75. Uptegrove, R.; Marwaha, S.; Birchwood, M. Depression and Schizophrenia: Cause, Consequence, or Trans-diagnostic Issue? *Schizophr. Bull.* 2016, 43, 240–244.
76. Burokas, A.; Arboleya, S.; Moloney, R.D.; Peterson, V.L.; Murphy, K.; Clarke, G.; Stanton, C.; Dinan, T.G.; Cryan, J.F. Targeting the Microbiota-Gut-Brain Axis: Prebiotics Have Anxiolytic and Antidepressant-like Effects and Reverse the Impact of Chronic Stress in Mice. *Biol. Psychiatry* 2017, 82, 472–487.
77. Lin, P.; Ding, B.; Feng, C.; Yin, S.; Zhang, T.; Qi, X.; Lv, H.; Guo, X.; Dong, K.; Zhu, Y.; et al. *Prevotella* and *Klebsiella* proportions in fecal microbial communities are potential characteristic parameters for patients with major depressive disorder. *J. Affect. Disord.* 2017, 207, 300–304.
78. Chen, Z.; Li, J.; Gui, S.; Zhou, C.; Chen, J.; Yang, C.; Hu, Z.; Wang, H.; Zhong, X.; Zeng, L.; et al. Comparative metaproteomics analysis shows altered fecal microbiota signatures in patients with major depressive disorder. *NeuroReport* 2018, 29, 417–425.

79. Tillmann, S.; Abildgaard, A.; Winther, G.; Wegener, G. Altered fecal microbiota composition in the Flinders sensitive line rat model of depression. *Psychopharmacology* 2019, 236, 1445–1457.
80. Tomasik, J.; Yolken, R.H.; Bahn, S.; Dickerson, F.B. Immunomodulatory Effects of Probiotic Supplementation in Schizophrenia Patients: A Randomized, Placebo-Controlled Trial. *Biomark. Insights* 2015, 10, 47–54.
81. Dickerson, F.B.; Stallings, C.; Origoni, A.; Katsafanas, E.; Savage, C.L.G.; Schweinfurth, L.A.B.; Goga, J.; Khushalani, S.; Yolken, R.H. Effect of Probiotic Supplementation on Schizophrenia Symptoms and Association with Gastrointestinal Functioning: A Randomized, Placebo-Controlled Trial. *Prim. Care Companion CNS Disord.* 2014, 16, PCC.13m01579.
82. Severance, E.G.; Gressitt, K.L.; Stallings, C.R.; Katsafanas, E.; Schweinfurth, L.A.; Savage, C.L.; Adamos, M.B.; Sweeney, K.M.; Origoni, A.E.; Khushalani, S.; et al. Probiotic normalization of *Candida albicans* in schizophrenia: A randomized, placebo-controlled, longitudinal pilot study. *Brain Behav. Immun.* 2017, 62, 41–45.
83. Okubo, R.; Koga, M.; Katsumata, N.; Odamaki, T.; Matsuyama, S.; Oka, M.; Narita, H.; Hashimoto, N.; Kusumi, I.; Xiao, J.; et al. Effect of bifidobacterium breve A-1 on anxiety and depressive symptoms in schizophrenia: A proof-of-concept study. *J. Affect. Disord.* 2019, 245, 377–385.
84. Ghaderi, A.; Banafshe, H.R.; Mirhosseini, N.; Moradi, M.; Karimi, M.-A.; Mehrzad, F.; Bahmani, F.; Asemi, Z. Clinical and metabolic response to vitamin D plus probiotic in schizophrenia patients. *BMC Psychiatry* 2019, 19, 77.

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