KCNJ5 Mutation in Aldosterone Producing Adenoma

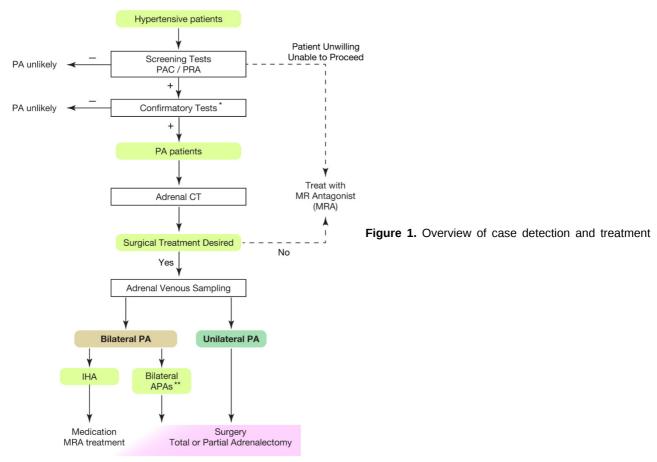
Subjects: Cardiac & Cardiovascular Systems Contributor: Takumi Kitamoto, Tetsuo Nishikawa

The discovery of the *KCNJ5* somatic mutation in aldosteroene producing adenoma (APA) in 2011 and the development of specific CYP11B2 antibodies in 2012 have greatly advanced our understanding of the pathophysiology of primary aldosteronism. PA leads to abnormalities in the renin-angiotensin-aldosterone system and is known to increase the incidence of atrial fibrillation, heart failure, and stroke by 3.52, 2.05, and 2.58 times compared to hypertension due to essential hypertension.

Keywords: primary aldosteronism; KCNJ5; aldosterone producing adenoma

1. Introduction

Hypertension is the leading risk factor for death and disability in the global population [1][2][3][4]. The treatment goal of hypertensive patients is to maintain a healthy life expectancy comparable to that of healthy individuals. To achieve it, researchers need to cure the disease and reduce the risk of cardiovascular complications as much as possible. In this regard, an appropriate classification of hypertension is necessary. Secondary hypertension should be screened for the patients showing following characteristics [5][6][7]: (1) onset of hypertension (<30 years), (2) abrupt onset of hypertension, (3) drug-resistant hypertension, (4) exacerbation of previously controlled hypertension, (5) suspicion of endocrine causes of hypertension or CKD, (6) clinical features suggestive of obstructive sleep apnoea, (7) unprovoked or excessive hypokalemia, (8) disproportionate targeted organ damage for degree of hypertension, (9) onset of diastolic hypertension in older adults (\ge 65 years). Primary aldosteronism (PA) is found in 5–7% of all hypertensive patients $\frac{[8][9][10][11][12][13]}{8}$ and even explains 20% of patients with third-degree hypertension [14]. Despite its frequency, less than 1% of patients with PA undergo screening and progress to treatment throughout their lifetime [15][16][17][18][19][20]. One reason for the underdiagnosis may be that health care providers are not fully aware of the high frequency of PA in hypertensive patients. In addition, there is not sufficient consensus on the targets and methods of screening tests. In most of the institutions, patients to be screened are those with severe hypertension [21][22][23], i.e., in addition to the characteristics indicating secondary hypertension, (1) hypertension and spontaneous or diuretic-induced hypokalemia, (2) hypertension and adrenal incidentaloma, (3) atrial fibrillation in the absence of structural heart disease, (4) a family history of early onset hypertension or stroke at a young age (<40 years), (5) all hypertensive first-degree relatives of patients with PA. Others recommend screening for all hypertensive patients $\frac{[24]}{}$ from a cost-effectiveness perspective based on its high frequency [25][26] and the need for lifelong antihypertensive medication. With suspicions of PA at the first screening test evaluating plasma aldosterone levels and plasma renin activity, researchers should move to definitive diagnosis of PA, performing confirmatory tests (Figure 1).



strategy of primary aldosteronism. Most guidelines recommend to use plasma aldosterone-renin ratio (PAC/PRA) for screening test [21][22][23][24][27]. * Confirmatory tests included captopril challenge test, saline infusion test, and oral sodium loading test. ** Bilateral APAs can be diagnosed by segment selective adrenal venous sampling [28][29]. Partial adrenalectomy to spare normal adrenal tissue should be considered for the dominant side of APA if attempting surgical treatment. This would be helpful when the residual APA may cause MRA-resistant PA in the future. PAC, plasma aldosterone concentration; PRA, plasma renin activity; PA, primary aldosteronism; MRA, mineral corticoid receptor antagonist; CT, computed tomography; IHA, idiopathic hyperaldosteronism; APA, aldosterone producing adenoma.

2. Cardiovascular Outcome of Primary Aldosteronism

PA leads to abnormalities in the renin-angiotensin-aldosterone system and is known to increase the incidence of atrial fibrillation, heart failure, and stroke by 3.52, 2.05, and 2.58 times compared to hypertension due to essential hypertension [30][31][32][33]. Appropriate treatment for PA can greatly improve the prognosis of cardiovascular complications [32], and thus early diagnosis of PA and specific treatment for the condition are important. The etiology of PA is caused by two broad categories: aldosterone-producing adenomas (APA) and idiopathic hyper aldosteronism (IHA). In the former, the pathogenetic localization is clear, and surgical resection can be expected to cure the disease [34]. The frequency of APA and IHA has been reported to vary between primary care and referral centers [11][12][35], but it is difficult to determine the exact frequency. APA should be pathologically diagnosed by morphology and staining for CYP11B2 [36], an aldosteroneproducing rate-limiting enzyme. However, surgical resection is only performed in cases who want to receive surgery and have been pre-surgically diagnosed with APA by adrenal venous sampling (AVS). IHA, on the other hand, is diagnosed by AVS and is usually treated with medication. We, therefore, cannot rule out the possibility of mixed misdiagnosis of APA in most cases. The frequency of APA and IHA diagnoses depends largely on the availability, procedure (e.g., usage of ACTH stimulation) and diagnostic criteria of AVS. Recently, tributary adrenal vein in addition to central adrenal vein sampling (a.k.a., segment selective AVS) has revealed that more than 10% of patients were elucidated as having APA, which could not be diagnosed by classic AVS evaluation of left-right difference [28]. Therefore, it is important to identify the localization of increased aldosterone synthesis within the adrenal gland to accurately diagnose APA, rather than simply looking at the right or left side of the adrenal gland. However, as a practical matter, the number of facilities where AVS is available is still far less than the expected number of patients with PA, and it is difficult to perform AVS on all patients with PA. Furthermore, publications reporting use of segment selective AVS are mostly from Japan $\frac{[28][29][37][38]}{}$. Standardization and cost reduction of the AVS technique would give precise etiology of APA and IHA. The data would help to develop the diagnostic algorithm to predict the cases who require AVS.

3. Great Progress in Our Pathophysiological Understanding of PA

Along with the improvement of diagnostic techniques for PA, there have been remarkable advances in pathophysiological understanding of PA. One is the development of the specific CYP11B2 antibody, and the other is the findings of somatic mutations in APAs.

The corner stone of the specific monoclonal antibody against CYP11B2 was reported in 2014 [39][40]. CYP11B2 is the rate-limiting enzyme for aldosterone synthesis and the antibody was recently used for standardization for the nomenclature and criteria of the pathological diagnosis of surgical specimens of PA patients (HISTALDO) [36]. Classically, the diagnosis of aldosterone-producing adenoma and nodule was made morphologically by HE staining of the resected specimen (classic form). On the other hand, CYP11B2 staining has made it possible to diagnose multiple aldosterone-producing nodules or, less frequently, aldosterone-producing diffuse hyperplasia. In addition, this pathological diagnosis was reproducible among different pathologists (non-classic form). This standardization of the pathological diagnosis was built up by the incorporation of a functional aspect using CYP11B2 into the diagnosis, in parallel with the conventional morphological criteria. A prospective cohort study using these diagnostic concepts showed that non-classical multiple nodules were associated with more postoperative biochemical failures compared to patients diagnosed with a single macro lesion in its classic form represented by APA (33.3 vs. 2.4 (%)) [41]. The possibility that lesions with multiple nodules may be more prone to bilateral disease should be considered, although this is a matter of speculation due to the lack of pathological findings in the residual adrenal gland. Efforts should be made to make the preoperative diagnosis closer to the pathological diagnosis in order to take advantage of the finding that the two forms of classification would have different surgical implications for PA patients.

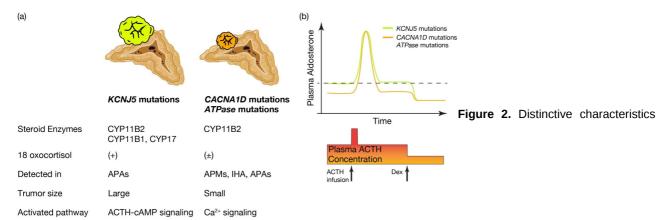
In parallel with these findings, the discovery of the aldosterone driver mutation has greatly advanced the understanding of the pathogenesis of PA patients. The first step in this sequence was the memorable report published in 2011, describing a somatic mutation in the *KCNJ5* gene encoding G protein-activated inward rectifier potassium channel 4 (GIRK-4) [42]. *KCNJ5* mutation was identified in 36% of APA [42](43)(44]. Following this discovery, somatic mutations in *ATP2B3*, *ATP1A1* [45], and *CACNA1D* [46] were identified in 2013; although the frequency of *KCNJ5* mutations in APA is higher in Eastern countries (59.5–76.8%) [47](48)(49)(50) than in Western countries (34–45%) [43](44)(51)(52) [53], *KCNJ5* mutations are similarly the most common mutation across the world. All of these mutations increase intracellular calcium concentrations, followed by activating CYP11B2 enzyme expression and increasing excess aldosterone biosynthesis. However, whether these mutations enhance cell proliferation has not been determined. Particularly, the proliferative effect seen in *KCNJ5* was different due to *KCNJ5* expression levels in the study using HAC15 [54], while another report using APAs did not show any evidence of proliferative effect of *KCNJ5* mutation [55]. However, the evidence from germline *KCNJ5* mutations suggested that *KCNJ5* mutations promote proliferation to form hyperplasia *in vivo* [56](57]. A recent study using CYP11B2 immunohistochemistry-guided high-throughput sequencing rather than Sanger sequencing identified 95% of APAs as showing one of these somatic mutations, and more APAs with *CACNA1D* mutations among *KCNJ5* wild-type APAs [58]. These mutations would play a crucial role in the pathogenesis of PA.

4. Pathological Insights into PA

CYP11B1 and CYCP11B2 play pivotal roles in the synthesis of adrenal corticosteroids, the former responsible for the synthesis of cortisol and corticosterone, and the latter for the synthesis of aldosterone [59]. The advent of CYP11B2 novel monoclonal antibodies has clarified the zonation of these synthesis enzymes in the normal adrenal gland [39][40]. Classically, the adrenal cortex is composed of three main zones, i.e., from the outermost layer, zona glomerulosa (ZG), to the zona fasciculata (ZF) and zona reticularis. Aldosterone and cortisol are produced by ZG and ZF cells, which have been shown by HE staining to be morphologically distinct: ZG cells are smaller, more compact, and have a smaller cytoplasm/nucleus ratio; ZF cells have a larger cytoplasm/nucleus ratio, with more fatty material and transparent appearance. Evaluation of the distribution of each synthase in the normal adrenal gland using specific CYP11B2 and B1 antibodies has shown that the distribution is different in young and elderly individuals [60][61][62]. In younger individuals, CYP11B2-positive cells are present throughout the ZG [61]; CYP11B1 was detected in both the ZF and zona reticularis. An unstained region is identified between the CYP11B2- and CYP11B1-positive cells that is not detected by either antibody. It is not clear whether this region comprises undifferentiated progenitor-like cells [63]. On the other hand, in the elderly, CYP11B2-positive cells are adjacent to the membrane and form a pattern of CYP11B2-immunopositive cell clusters with no CYP11B2 staining around them. Approximately half of the adrenals of normotensive individuals contain these regions, which have been designated as Aldosterone Producing Cell Clusters (APCCs) [60][61][62][64][65]. The nomenclature of APCC was changed to Aldosterone Producing Micronodules (APM) in the HISTALDO study [36]. The question of whether APMs have an autonomous aldosterone secretory capacity has been arisen. This is linked to the fact that the incidence of APMs correlates with the frequency of aldosterone-related hypertension [62][66]. Subsequent studies have shown that these

APMs when found in adult human adrenal tissue with normal adrenal function also contain somatic mutations of the aldosterne driver mutation; mutations were identified in over 30% of APMs, most frequently in the *CACNA1D* mutation [58] [60][65]. This was followed by *ATP2B3*, *ATP1A1*, and few *KCNJ5* mutations. Interestingly, in case reports suggesting APM to APA transition, APM and micro APA were adjacent, with *ATP1A1* and *ATP2B3* mutations from the former and *KCNJ5* mutations from the latter [67][68]. The research efforts have also extended to IHA cases. The study showed somatic mutations in *CACNA1D* are the cause of most IHA [69]. This is consistent with previous reports of image-negative PAs [70]. Interpretation is limited due to the small sample of surgically resected IHAs. However, their findings were much appreciated as supporting the concept of a continuum of pathophysiology from normotensive to hypertensive patients [71]. The different distribution of somatic mutations, with *KCNJ5* mutations found specifically in APA and *CACNA1D* mutations in IHA, may contribute to the difference in clinical features between APA and IHA [35].

What is the pathological characteristics of APA harboring KCNJ5, and are they defined by the expression profile of steroidogenic enzymes? (See Figure 2a.) KCNJ5 mutated APAs are larger than other APAs; 60% are composed of lipidladen clear ZF like cells and 40% are composed of compact ZG like cells [72]. Immunohistological studies on steroid synthase have shown that KCNJ5 mutated APAs are positive for CYP11B2 alone, co-expressing CYP11B2 and CYP17, and in a few cases, cells co-expressing CYP11B2, CYP11B1 and CYP17 [40][72][73]. On the other hand, APA with ATPA2B3 mutations tended to be dominated by compact ZG like cells, while APA with ATP1A1 and CACNA1D mutations showed heterogeneity from tumor to tumor with no clear advantage [49][72]. Very interestingly, the expression of CYP11B2 and CYP17A1, which are involved in aldosterone and cortisol synthesis, correlated positively in KCNJ5 mutant APAs, but negatively in ATP2B3 mutants [72]. The correlation between the two is not clear for ATP1A1 or CACNA1D. Inconsistent with this profile of steroidogenic enzymes, the normally negligible hybrid steroid, 18 oxocortisol (see the section "Development of prediction model for somatic KCNJ5 mutation in PA patinets"), is markedly increased in patients with APAs harboring KCNJ5 mutations. This increase has not been observed in patients with APAs harboring ATPase and CACNA1D mutations [58][74] (Figure 2a). This series of results has suggested that KCNJ5 mutations are associated with the formation of APA and that the pathogenetic process differs from that of ATPase and CACNA1D mutations. APAs harboring KCNJ5 mutations may have a more disorganized process of tumor cell differentiation and formation than APAs with ATPase or CACNA1D mutations. It has not been clear how the KCNJ5 mutation occurred in APAs.



of APA patients harboring KCNJ5 mutations, and CACNA1D or ATPase mutations. Distinctive characteristics of aldosterone producing adenoma (APA) patients harboring KCNJ5 mutation and CACNA1D or ATPase mutations. (a) KCNJ5 mutated APAs showed larger tumor and heterogenous composition of CYP11B2, CYP11B1, and/or CYP17 positive cells, while CACNA1D and ATPase mutated APAs showed smaller tumor and homogeneous composition of CYP11B2 positive cells. 18 oxocortisol is elevated in KCNJ5 mutated APAs, while not in CACNA1D and ATPase mutated APAs. KCNJ5 mutations have been detected mostly in APAs, while CACNA1D and ATPase mutations have been dominantly identified in aldosterone producing micronodules (APMs) and idiopathic hyperaldosteronism (IHA) as well as APAs. Researchers assume distinctive activated pathways of aldosterone synthesis between KCNJ5 mutated and CACNA1D or ATPase mutated APAs, such as the ACTH-cAMP and Ca²⁺ signaling pathways. (b) Conceptual scheme responsiveness of aldosterone secretion ACTH harboring KCNJ5 and CACNA1D or ATPase mutations is shown. Basal aldosterone secretion is higher and its responsiveness to ACTH stimulation is lower in KCNJ5 mutated APAs than CACNA1D and ATPase mutated APAs. ACTH depletion via dexamethasone (Dex) suppression decreased plasma aldosterone levels from KCNJ5 mutated APAs to that from KCNJ5 wild APAs [49][75][76]. Thus researchers assume ACTH-cAMP signaling in KCNJ5 mutated APAs is activated to increase basal aldosterone secretion and to lessen response to extra ACTH stimulation. Figure was created with BioRender.com.

References

- 1. Gilmour, S.; Liao, Y.; Bilano, V.; Shibuya, K. Burden of disease in Japan: Using national and subnational data to inform I ocal health policy. J. Prev. Med. Public Health 2014, 47, 136–143.
- 2. Mills, K.T.; Bundy, J.D.; Kelly, T.N.; Reed, J.E.; Kearney, P.M.; Reynolds, K.; Chen, J.; He, J. Global Disparities of Hyper tension Prevalence and Control: A Systematic Analysis of Population-Based Studies From 90 Countries. Circulation 20 16, 134, 441–450.
- 3. GBD 2017 Risk Factor Collaborators. Global, regional, and national comparative risk assessment of 84 behavioural, en vironmental and occupational, and metabolic risks or clusters of risks for 195 countries and territories, 1990–2017: A sy stematic analysis for the Global Burden of Disease Study 2017. Lancet 2018, 392, 1923–1994.
- 4. Sakaue, S.; Gen, F.; Kanai, M.; Karjalainen, J.; Akiyama, M.; Kurki, M.; Matoba, N.; Takahashi, A.; Hirata, M.; Kubo, M.; et al. Trans-biobank analysis with 676,000 individuals elucidates the association of polygenic risk scores of complex traits with human lifespan. Nat. Med. 2020, 26, 542–548.
- 5. Whelton, P.K.; Carey, R.M.; Aronow, W.S.; Casey, D.E.; Collins, K.J.; Dennison Himmelfarb, C.; DePalma, S.M.; Giddin g, S.; Jamerson, K.A.; Jones, D.W.; et al. 2017 ACC/AHA/AAPA/ABC/ACPM/AGS/APhA/ASH/ASPC/NMA/PCNA Guide line for the Prevention, Detection, Evaluation, and Management of High Blood Pressure in Adults: A Report of the Ameri can College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. Hypertension 2018, 71, e13–e115.
- 6. Williams, B.; Mancia, G.; Spiering, W.; Agabiti Rosei, E.; Azizi, M.; Burnier, M.; Clement, D.L.; Coca, A.; de Simone, G.; Dominiczak, A.; et al. 2018 ESC/ESH Guidelines for the management of arterial hypertension. Eur. Heart J. 2018, 39, 3 021–3104.
- 7. Umemura, S.; Arima, H.; Arima, S.; Asayama, K.; Dohi, Y.; Hirooka, Y.; Horio, T.; Hoshide, S.; Ikeda, S.; Ishimitsu, T.; et al. The Japanese Society of Hypertension Guidelines for the Management of Hypertension (JSH 2019). Hypertens. Re s. 2019, 42, 1235–1481.
- 8. Gordon, R.D.; Ziesak, M.D.; Tunny, T.J.; Stowasser, M.; Klemm, S.A. Evidence that primary aldosteronism may not be uncommon: 12% incidence among antihypertensive drug trial volunteers. Clin. Exp. Pharm. Physiol. 1993, 20, 296–29 8.
- 9. Rossi, G.P.; Sacchetto, A.; Chiesura-Corona, M.; De Toni, R.; Gallina, M.; Feltrin, G.P.; Pessina, A.C. Identification of the etiology of primary aldosteronism with adrenal vein sampling in patients with equivocal computed tomography and magnetic resonance findings: Results in 104 consecutive cases. J. Clin. Endocrinol Metab. 2001, 86, 1083–1090.
- 10. Omura, M.; Saito, J.; Yamaguchi, K.; Kakuta, Y.; Nishikawa, T. Prospective study on the prevalence of secondary hyper tension among hypertensive patients visiting a general outpatient clinic in Japan. Hypertens. Res. 2004, 27, 193–202.
- 11. Rossi, G.P.; Bernini, G.; Caliumi, C.; Desideri, G.; Fabris, B.; Ferri, C.; Ganzaroli, C.; Giacchetti, G.; Letizia, C.; Maccari o, M.; et al. A prospective study of the prevalence of primary aldosteronism in 1,125 hypertensive patients. J. Am. Coll Cardiol. 2006, 48, 2293–2300.
- 12. Monticone, S.; Burrello, J.; Tizzani, D.; Bertello, C.; Viola, A.; Buffolo, F.; Gabetti, L.; Mengozzi, G.; Williams, T.A.; Rabbi a, F.; et al. Prevalence and Clinical Manifestations of Primary Aldosteronism Encountered in Primary Care Practice. J. A m. Coll Cardiol. 2017, 69, 1811–1820.
- 13. Karashima, S.; Kometani, M.; Tsujiguchi, H.; Asakura, H.; Nakano, S.; Usukura, M.; Mori, S.; Ohe, M.; Sawamura, T.; O kuda, R.; et al. Prevalence of primary aldosteronism without hypertension in the general population: Results in Shika st udy. Clin. Exp. Hypertens. 2018, 40, 118–125.
- 14. Brown, J.M.; Siddiqui, M.; Calhoun, D.A.; Carey, R.M.; Hopkins, P.N.; Williams, G.H.; Vaidya, A. The Unrecognized Prevalence of Primary Aldosteronism: A Cross-sectional Study. Ann. Intern. Med. 2020, 173, 10–20.
- 15. Funder, J.W. Primary aldosteronism as a public health issue. Lancet Diabetes Endocrinol. 2016, 4, 972–973.
- 16. Mulatero, P.; Monticone, S.; Burrello, J.; Veglio, F.; Williams, T.A.; Funder, J. Guidelines for primary aldosteronism: Upta ke by primary care physicians in Europe. J. Hypertens. 2016, 34, 2253–2257.
- 17. Jaffe, G.; Gray, Z.; Krishnan, G.; Stedman, M.; Zheng, Y.; Han, J.; Chertow, G.M.; Leppert, J.T.; Bhalla, V. Screening Ra tes for Primary Aldosteronism in Resistant Hypertension. Hypertension 2020, 75, 650–659.
- 18. Cohen, J.B.; Cohen, D.L.; Herman, D.S.; Leppert, J.T.; Byrd, J.B.; Bhalla, V. Testing for primary aldosteronism and min eralocorticoid receptor antagonist use among US veterans: A retrospective cohort study. Ann. Intern. Med. 2021, 174, 2 89–297.
- 19. Gkaniatsa, E.; Ekerstad, E.; Gavric, M.; Muth, A.; Trimpou, P.; Olsson, D.S.; Johannsson, G.; Ragnarsson, O. Increasin g Incidence of Primary Aldosteronism in Western Sweden During 3 Decades-Yet An Underdiagnosed Disorder. J. Clin.

- Endocrinol. Metab. 2021, 106, e3603-e3610.
- 20. Liu, Y.-Y.; King, J.; Kline, G.A.; Padwal, R.S.; Pasieka, J.L.; Chen, G.; So, B.; Harvey, A.; Chin, A.; Leung, A.A. Outcome s of a Specialized Clinic on Rates of Investigation and Treatment of Primary Aldosteronism. JAMA Surg. 2021, 156, 541 –549.
- 21. Funder, J.W.; Carey, R.M.; Mantero, F.; Murad, M.H.; Reincke, M.; Shibata, H.; Stowasser, M.; Young, W.F. The Manag ement of Primary Aldosteronism: Case Detection, Diagnosis, and Treatment: An Endocrine Society Clinical Practice Gu ideline. J. Clin. Endocrinol. Metab. 2016, 101, 1889–1916.
- 22. Mulatero, P.; Monticone, S.; Deinum, J.; Amar, L.; Prejbisz, A.; Zennaro, M.C.; Beuschlein, F.; Rossi, G.P.; Nishikawa, T.; Morganti, A.; et al. Genetics, prevalence, screening and confirmation of primary aldosteronism: A position statement and consensus of the Working Group on Endocrine Hypertension of The European Society of Hypertension. J. Hypertens. 2020, 38, 1919–1928.
- 23. Rossi, G.P.; Bisogni, V.; Bacca, A.V.; Belfiore, A.; Cesari, M.; Concistrè, A.; Del Pinto, R.; Fabris, B.; Fallo, F.; Fava, C.; et al. The 2020 Italian Society of Arterial Hypertension (SIIA) practical guidelines for the management of primary aldost eronism. Int. J. Cardiol. Hypertens. 2020, 5, 100029.
- 24. Naruse, M.; Katabami, T.; Shibata, H.; Sone, M.; Takahashi, K.; Tanabe, A.; Izawa, S.; Ichijo, T.; Otsuki, M.; Omura, M.; et al. Japan Endocrine Society clinical practice guideline for the diagnosis and management of primary aldosteronism 2 021. Endocr. J. 2022, 69, 327–359.
- 25. Lubitz, C.C.; Economopoulos, K.P.; Sy, S.; Johanson, C.; Kunzel, H.E.; Reincke, M.; Gazelle, G.S.; Weinstein, M.C.; G aziano, T.A. Cost-Effectiveness of Screening for Primary Aldosteronism and Subtype Diagnosis in the Resistant Hypert ensive Patients. Circ. Cardiovasc. Qual. Outcomes 2015, 8, 621–630.
- 26. Sato, M.; Morimoto, R.; Seiji, K.; Iwakura, Y.; Ono, Y.; Kudo, M.; Satoh, F.; Ito, S.; Ishibashi, T.; Takase, K. Cost-Effectiv eness Analysis of the Diagnosis and Treatment of Primary Aldosteronism in Japan. Horm. Metab. Res. 2015, 47, 826–8 32
- 27. Wu, V.-C.; Hu, Y.-H.; Er, L.K.; Yen, R.-F.; Chang, C.-H.; Chang, Y.-L.; Lu, C.-C.; Chang, C.-C.; Lin, J.-H.; Lin, Y.-H.; et al. Case detection and diagnosis of primary aldosteronism-The consensus of Taiwan Society of Aldosteronism. J. Formos. Med. Assoc. 2017, 116, 993–1005.
- 28. Kitamoto, T.; Kitamoto, K.K.; Omura, M.; Takiguchi, T.; Tsurutani, Y.; Kubo, H.; Yamazaki, Y.; Sasano, H.; Saito, J.; Nishi kawa, T. Precise Mapping of Intra-Adrenal Aldosterone Activities Provides a Novel Surgical Strategy for Primary Aldoste ronism. Hypertension 2020, 76, 976–984.
- 29. Satoh, F.; Morimoto, R.; Seiji, K.; Satani, N.; Ota, H.; Iwakura, Y.; Ono, Y.; Kudo, M.; Nezu, M.; Omata, K.; et al. Is there a role for segmental adrenal venous sampling and adrenal sparing surgery in patients with primary aldosteronism? Eur J. Endocrinol. 2015, 173, 465–477.
- 30. Milliez, P.; Girerd, X.; Plouin, P.F.; Blacher, J.; Safar, M.E.; Mourad, J.J. Evidence for an increased rate of cardiovascula r events in patients with primary aldosteronism. J. Am. Coll Cardiol. 2005, 45, 1243–1248.
- 31. Catena, C.; Colussi, G.; Nadalini, E.; Chiuch, A.; Baroselli, S.; Lapenna, R.; Sechi, L.A. Cardiovascular outcomes in pat ients with primary aldosteronism after treatment. Arch. Intern. Med. 2008, 168, 80–85.
- 32. Hundemer, G.L.; Curhan, G.C.; Yozamp, N.; Wang, M.; Vaidya, A. Cardiometabolic outcomes and mortality in medically treated primary aldosteronism: A retrospective cohort study. Lancet Diabetes Endocrinol. 2018, 6, 51–59.
- 33. Monticone, S.; D'Ascenzo, F.; Moretti, C.; Williams, T.A.; Veglio, F.; Gaita, F.; Mulatero, P. Cardiovascular events and ta rget organ damage in primary aldosteronism compared with essential hypertension: A systematic review and meta-anal ysis. Lancet Diabetes Endocrinol. 2018, 6, 41–50.
- 34. Williams, T.A.; Lenders, J.W.M.; Mulatero, P.; Burrello, J.; Rottenkolber, M.; Adolf, C.; Satoh, F.; Amar, L.; Quinkler, M.; Deinum, J.; et al. Outcomes after adrenalectomy for unilateral primary aldosteronism: An international consensus on ou tcome measures and analysis of remission rates in an international cohort. Lancet Diabetes Endocrinol. 2017, 5, 689–6
- 35. Omura, M.; Sasano, H.; Saito, J.; Yamaguchi, K.; Kakuta, Y.; Nishikawa, T. Clinical characteristics of aldosterone-producing microadenoma, macroadenoma, and idiopathic hyperaldosteronism in 93 patients with primary aldosteronism. Hypertens. Res. 2006, 29, 883–889.
- 36. Williams, T.A.; Gomez-Sanchez, C.E.; Rainey, W.E.; Giordano, T.J.; Lam, A.K.; Marker, A.; Mete, O.; Yamazaki, Y.; Zerb ini, M.C.N.; Beuschlein, F.; et al. International Histopathology Consensus for Unilateral Primary Aldosteronism. J. Clin. Endocrinol. Metab. 2021, 106, 42–54.
- 37. Satani, N.; Ota, H.; Seiji, K.; Morimoto, R.; Kudo, M.; Iwakura, Y.; Ono, Y.; Nezu, M.; Omata, K.; Ito, S.; et al. Intra-adre nal Aldosterone Secretion: Segmental Adrenal Venous Sampling for Localization. Radiology 2016, 278, 265–274.

- 38. Makita, K.; Nishimoto, K.; Kiriyama-Kitamoto, K.; Karashima, S.; Seki, T.; Yasuda, M.; Matsui, S.; Omura, M.; Nishikaw a, T. A Novel Method: Super-selective Adrenal Venous Sampling. J. Vis. Exp. 2017, 127, e55716.
- 39. Gomez-Sanchez, C.E.; Qi, X.; Velarde-Miranda, C.; Plonczynski, M.W.; Parker, C.R.; Rainey, W.; Satoh, F.; Maekawa, T.; Nakamura, Y.; Sasano, H.; et al. Development of monoclonal antibodies against human CYP11B1 and CYP11B2. M ol. Cell. Endocrinol. 2014, 383, 111–117.
- 40. Nakamura, Y.; Maekawa, T.; Felizola, S.J.; Satoh, F.; Qi, X.; Velarde-Miranda, C.; Plonczynski, M.W.; Ise, K.; Kikuchi, K.; Rainey, W.E.; et al. Adrenal CYP11B1/2 expression in primary aldosteronism: Immunohistochemical analysis using novel monoclonal antibodies. Mol. Cell. Endocrinol. 2014, 392, 73–79.
- 41. Meyer, L.S.; Handgriff, L.; Lim, J.S.; Udager, A.M.; Kinker, I.-S.; Ladurner, R.; Wildgruber, M.; Knösel, T.; Bidlingmaier, M.; Rainey, W.E.; et al. Single-Center Prospective Cohort Study on the Histopathology, Genotype, and Postsurgical Out comes of Patients With Primary Aldosteronism. Hypertension 2021, 78, 738–746.
- 42. Choi, M.; Scholl, U.I.; Yue, P.; Björklund, P.; Zhao, B.; Nelson-Williams, C.; Ji, W.; Cho, Y.; Patel, A.; Men, C.J.; et al. K+ channel mutations in adrenal aldosterone-producing adenomas and hereditary hypertension. Science 2011, 331, 768–772.
- 43. Åkerström, T.; Crona, J.; Verdugo, A.D.; Starker, L.F.; Cupisti, K.; Willenberg, H.S.; Knoefel, W.T.; Saeger, W.; Feller, A.; Ip, J.; et al. Comprehensive re-sequencing of adrenal aldosterone producing lesions reveal three somatic mutations ne ar the KCNJ5 potassium channel selectivity filter. PLoS ONE 2012, 7, e41926.
- 44. Azizan, E.A.; Murthy, M.; Stowasser, M.; Gordon, R.; Kowalski, B.; Xu, S.; Brown, M.J.; O'Shaughnessy, K.M. Somatic mutations affecting the selectivity filter of KCNJ5 are frequent in 2 large unselected collections of adrenal aldosteronom as. Hypertension 2012, 59, 587–591.
- 45. Beuschlein, F.; Boulkroun, S.; Osswald, A.; Wieland, T.; Nielsen, H.N.; Lichtenauer, U.D.; Penton, D.; Schack, V.R.; Am ar, L.; Fischer, E.; et al. Somatic mutations in ATP1A1 and ATP2B3 lead to aldosterone-producing adenomas and secondary hypertension. Nat. Genet. 2013, 45, 440–444, 444e1.1–444e.2.
- 46. Scholl, U.I.; Goh, G.; Stolting, G.; De Oliveira, R.C.; Choi, M.; Overton, J.D.; Fonseca, A.L.; Korah, R.; Starker, L.F.; Ku nstman, J.; et al. Somatic and germline CACNA1D calcium channel mutations in aldosterone-producing adenomas and primary aldosteronism. Nat. Genet. 2013, 45, 1050–1054.
- 47. Wu, V.-C.; Huang, K.-H.; Peng, K.-Y.; Tsai, Y.-C.; Wu, C.-H.; Wang, S.-M.; Yang, S.-Y.; Lin, L.-Y.; Chang, C.-C.; Lin, Y.-H.; et al. Prevalence and clinical correlates of somatic mutation in aldosterone producing adenoma-Taiwanese populati on. Sci. Rep. 2015, 5, 11396.
- 48. Zheng, F.-F.; Zhu, L.-M.; Nie, A.-F.; Li, X.-Y.; Lin, J.-R.; Zhang, K.; Chen, J.; Zhou, W.-L.; Shen, Z.-J.; Zhu, Y.-C.; et al. C linical characteristics of somatic mutations in Chinese patients with aldosterone-producing adenoma. Hypertension 201 5, 65, 622–628.
- 49. Kitamoto, T.; Suematsu, S.; Yamazaki, Y.; Nakamura, Y.; Sasano, H.; Matsuzawa, Y.; Saito, J.; Omura, M.; Nishikawa, T. Clinical and Steroidogenic Characteristics of Aldosterone-Producing Adenomas With ATPase or CACNA1D Gene Mu tations. J. Clin. Endocrinol. Metab. 2016, 101, 494–503.
- 50. Nanba, K.; Yamazaki, Y.; Bick, N.; Onodera, K.; Tezuka, Y.; Omata, K.; Ono, Y.; Blinder, A.R.; Tomlins, S.A.; Rainey, W. E.; et al. Prevalence of Somatic Mutations in Aldosterone-Producing Adenomas in Japanese Patients. J. Clin. Endocrin ol. Metab. 2020, 105, e4066–e4073.
- 51. Boulkroun, S.; Beuschlein, F.; Rossi, G.P.; Golib-Dzib, J.F.; Fischer, E.; Amar, L.; Mulatero, P.; Samson-Couterie, B.; Ha hner, S.; Quinkler, M.; et al. Prevalence, clinical, and molecular correlates of KCNJ5 mutations in primary aldosteronis m. Hypertension 2012, 59, 592–598.
- 52. Williams, T.A.; Monticone, S.; Schack, V.R.; Stindl, J.; Burrello, J.; Buffolo, F.; Annaratone, L.; Castellano, I.; Beuschlei n, F.; Reincke, M.; et al. Somatic ATP1A1, ATP2B3, and KCNJ5 mutations in aldosterone-producing adenomas. Hypert ension 2014, 63, 188–195.
- 53. Lenzini, L.; Rossitto, G.; Maiolino, G.; Letizia, C.; Funder, J.W.; Rossi, G.P. A Meta-Analysis of Somatic KCNJ5 K(+) Ch annel Mutations In 1636 Patients With an Aldosterone-Producing Adenoma. J. Clin. Endocrinol. Metab. 2015, 100, E10 89–E1095.
- 54. Yang, Y.; Gomez-Sanchez, C.E.; Jaquin, D.; Aristizabal Prada, E.T.; Meyer, L.S.; Knösel, T.; Schneider, H.; Beuschlein, F.; Reincke, M.; Williams, T.A. Primary Aldosteronism: KCNJ5 Mutations and Adrenocortical Cell Growth. Hypertension 2019, 74, 809–816.
- 55. Lerario, A.M.; Nanba, K.; Blinder, A.R.; Suematsu, S.; Omura, M.; Nishikawa, T.; Giordano, T.J.; Rainey, W.E.; Else, T. Genetics of aldosterone-producing adenomas with pathogenic KCNJ5 variants. Endocr. Relat. Cancer 2019, 26, 463.

- 56. Tamura, A.; Nishimoto, K.; Seki, T.; Matsuzawa, Y.; Saito, J.; Omura, M.; Gomez-Sanchez, C.E.; Makita, K.; Matsui, S.; Moriya, N.; et al. Somatic KCNJ5 mutation occurring early in adrenal development may cause a novel form of juvenile p rimary aldosteronism. Mol. Cell. Endocrinol. 2017, 441, 134–139.
- 57. Maria, A.G.; Suzuki, M.; Berthon, A.; Kamilaris, C.; Demidowich, A.; Lack, J.; Zilbermint, M.; Hannah-Shmouni, F.; Fauc z, F.R.; A Stratakis, C. Mosaicism for KCNJ5 Causing Early-Onset Primary Aldosteronism due to Bilateral Adrenocortica I Hyperplasia. Am. J. Hypertens. 2020, 33, 124–130.
- 58. De Sousa, K.; Boulkroun, S.; Baron, S.; Nanba, K.; Wack, M.; Rainey, W.E.; Rocha, A.; Giscos-Douriez, I.; Meatchi, T.; Amar, L.; et al. Genetic, Cellular, and Molecular Heterogeneity in Adrenals with Aldosterone-Producing Adenoma. Hype rtension 2020, 75, 1034–1044.
- 59. Spat, A.; Hunyady, L. Control of aldosterone secretion: A model for convergence in cellular signaling pathways. Physiol. Rev. 2004, 84, 489–539.
- 60. Nishimoto, K.; Tomlins, S.A.; Kuick, R.; Cani, A.K.; Giordano, T.J.; Hovelson, D.H.; Liu, C.-J.; Sanjanwala, A.R.; Edward s, M.A.; Gomez-Sanchez, C.E.; et al. Aldosterone-stimulating somatic gene mutations are common in normal adrenal gl ands. Proc. Natl. Acad. Sci. USA 2015, 112, E4591–E4599.
- 61. Nishimoto, K.; Seki, T.; Hayashi, Y.; Mikami, S.; Al-Eyd, G.; Nakagawa, K.; Morita, S.; Kosaka, T.; Oya, M.; Mitani, F.; et al. Human Adrenocortical Remodeling Leading to Aldosterone-Producing Cell Cluster Generation. Int. J. Endocrinol. 20 16, 2016, 1–6.
- 62. Nanba, K.; Vaidya, A.; Williams, G.H.; Zheng, I.; Else, T.; Rainey, W.E. Age-Related Autonomous Aldosteronism. Circula tion 2017, 136, 347–355.
- 63. Gomez-Sanchez, C.E.; Gomez-Sanchez, E.P.; Nishimoto, K. Immunohistochemistry of the Human Adrenal CYP11B2 in Normal Individuals and in Patients with Primary Aldosteronism. Horm. Metab. Res. 2020, 52, 421–426.
- 64. Nishimoto, K.; Nakagawa, K.; Li, D.; Kosaka, T.; Oya, M.; Mikami, S.; Shibata, H.; Itoh, H.; Mitani, F.; Yamazaki, T.; et a I. Adrenocortical zonation in humans under normal and pathological conditions. J. Clin. Endocrinol. Metab. 2010, 95, 22 96–2305.
- 65. Omata, K.; Anand, S.K.; Hovelson, D.H.; Liu, C.-J.; Yamazaki, Y.; Nakamura, Y.; Ito, S.; Satoh, F.; Sasano, H.; Rainey, W.E.; et al. Aldosterone-Producing Cell Clusters Frequently Harbor Somatic Mutations and Accumulate with Age in Nor mal Adrenals. J. Endocr. Soc. 2017, 1, 787–799.
- 66. Nanba, K.; Vaidya, A.; Rainey, W.E. Aging and Adrenal Aldosterone Production. Hypertension 2018, 71, 218-223.
- 67. Nishimoto, K.; Seki, T.; Kurihara, I.; Yokota, K.; Omura, M.; Nishikawa, T.; Shibata, H.; Kosaka, T.; Oya, M.; Suematsu, M.; et al. Case Report: Nodule Development From Subcapsular Aldosterone-Producing Cell Clusters Causes Hyperald osteronism. J. Clin. Endocrinol. Metab. 2016, 101, 6–9.
- 68. Zennaro, M.C.; Boulkroun, S.; Fernandes-Rosa, F. Genetic Causes of Functional Adrenocortical Adenomas. Endocr. R ev. 2017, 38, 516–537.
- 69. Omata, K.; Satoh, F.; Morimoto, R.; Ito, S.; Yamazaki, Y.; Nakamura, Y.; Anand, S.K.; Guo, Z.; Stowasser, M.; Sasano, H.; et al. Cellular and Genetic Causes of Idiopathic Hyperaldosteronism. Hypertension 2018, 72, 874–880.
- 70. Yamazaki, Y.; Nakamura, Y.; Omata, K.; Ise, K.; Tezuka, Y.; Ono, Y.; Morimoto, R.; Nozawa, Y.; Gomez-Sanchez, C.E.; Tomlins, S.A.; et al. Histopathological Classification of Cross-Sectional Image-Negative Hyperaldosteronism. J. Clin. En docrinol. Metab. 2017, 102, 1182–1192.
- 71. Brown, J.M.; Robinson-Cohen, C.; Luque-Fernandez, M.A.; Allison, M.A.; Baudrand, R.; Ix, J.H.; Kestenbaum, B.; De B oer, I.H.; Vaidya, A. The Spectrum of Subclinical Primary Aldosteronism and Incident Hypertension: A Cohort Study. An n. Intern. Med. 2017, 167, 630–641.
- 72. Ono, Y.; Yamazaki, Y.; Omata, K.; Else, T.; A Tomlins, S.; Rhayem, Y.; Williams, T.A.; Reincke, M.; Carling, T.; Monticon e, S.; et al. Histological Characterization of Aldosterone-producing Adrenocortical Adenomas with Different Somatic Mut ations. J. Clin. Endocrinol. Metab. 2019, 105, e282–e289.
- 73. Nakamura, Y.; Kitada, M.; Satoh, F.; Maekawa, T.; Morimoto, R.; Yamazaki, Y.; Ise, K.; Gomez-Sanchez, C.E.; Ito, S.; Ar ai, Y.; et al. Intratumoral heterogeneity of steroidogenesis in aldosterone-producing adenoma revealed by intensive dou ble-and triple-immunostaining for CYP11B2/B1 and CYP17. Mol. Cell. Endocrinol. 2016, 422, 57–63.
- 74. Tezuka, Y.; Yamazaki, Y.; Kitada, M.; Morimoto, R.; Kudo, M.; Seiji, K.; Takase, K.; Kawasaki, Y.; Mitsuzuka, K.; Ito, A.; e t al. 18-Oxocortisol Synthesis in Aldosterone-Producing Adrenocortical Adenoma and Significance of KCNJ5 Mutation S tatus. Hypertension 2019, 73, 1283–1290.
- 75. Wannachalee, T.; Zhao, L.; Nanba, K.; Nanba, A.T.; Shields, J.J.; E Rainey, W.; Auchus, R.J.; Turcu, A.F. Three Discret e Patterns of Primary Aldosteronism Lateralization in Response to Cosyntropin During Adrenal Vein Sampling. J. Clin. Endocrinol. Metab. 2019, 104, 5867–5876.

76. Inoue, K.; Yamazaki, Y.; Kitamoto, T.; Hirose, R.; Saito, J.; Omura, M.; Sasano, H.; Nishikawa, T. Aldosterone Suppressi on by Dexamethasone in Patients with KCNJ5-Mutated Aldosterone-Producing Adenoma. J. Clin. Endocrinol. Metab. 2 018, 103, 3477–3485.

Retrieved from https://encyclopedia.pub/entry/history/show/64130