

Dendrimers Integration in Cancer Imaging and Theranostics

Subjects: **Oncology**

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Cancer is a result of abnormal cell proliferation. This pathology is a serious health problem since it is a leading cause of death worldwide. Anti-cancer therapies rely on surgery, radiation, and chemotherapy. However, these treatments still present major associated problems, namely the absence of specificity. Nanoparticles, particularly dendrimers, have been paving their way to the front line of cancer treatment, mostly for drug and gene delivery, diagnosis, and disease monitoring. This is mainly derived from their high versatility, which results from their ability to undergo distinct surface functionalization, leading to improved performance.

cancer

nanocarriers

anticancer dendrimers

intracellular targeting

theranostics

1. Introduction

All types of human cells may suffer an abnormal proliferation that can lead to cancer cells. Cancer classification/identification is performed according to the tissue and cell type from which the cancer cells arise. Therefore, there are multiple distinct types of cancer, which can vary significantly in their behavior and response to treatments ^[1]. This disease is a major public health problem and a leading cause of death worldwide in countries of all income levels.

According to the World Health Organization (WHO), the social and economic impact caused by cancer is increasing. In a 2014 report, an annual cost of EUR 1.04 trillion was estimated for global cancer expenses. The report also declared that it is important to continue investing in care and control, which will prevent a considerable number of deaths for years to come. ^[2] Extended lifespan associated with environmental factors (e.g., exposure to pollution, carcinogenic agents, radiation, viruses and bacteria) and the low efficacy of available treatments, closely associated with an increased drug resistance, also contributes to cancer development ^[3].

Currently, the standard cancer treatments used in clinical settings are radiotherapy, surgery, and conventional chemotherapy ^[4]. However, these therapies present several drawbacks, such as high toxicity, due to insufficient selectivity and unspecific targeting of cancer cells, which leads to increased resistance to anticancer drugs.

It is therefore relevant to find new anticancer agents able to control tumor growth with minimal side effects. In recent years, the use of nanotechnology in cancer treatment has offered some exciting possibilities, including improvement of detection and elimination of cancer cells before tumor development. This includes the use of dendrimer-based nanotherapeutics as a novel strategy for diagnosis and therapy (theranostics) ^{[5][6][7]}. Dendrimers

are synthetic 3D polymers with well-defined layered architecture [8]. Owing to their high functionality and loading capacity, as well as their precisely controlled chemical composition and molecular weight, dendrimer-based anticancer therapies offer great advantages over conventional formulations. These include the use of dendrimers as advanced contrast agents (diagnosis) [6], nanocarriers (treatment) [5][9], and theranostic agents (diagnosis, treatment and disease monitoring) [7]. In the quest for new anticancer drugs using dendrimer-based therapies, Shao et al. demonstrated that dendrimers may display innate anticancer activity and anti-metastatic properties without the loading of any therapeutic agent [10].

2. Dendrimer Nanoparticles

Multifunctional nanoparticles display great potential for drug and gene delivery, especially for cancer therapy [11][12]. Dendrimers are a class of hyperbranched synthetic polymers with a very low polydispersity, also known as “cascade polymers” [13]. Biologically, dendrimers are highly biocompatible, with a predictable biodistribution and cell-membrane-interacting features mostly determined by their size and surface charge [14]. They were first synthesized in the late 1970s by Tomalia et al., with the desire to mimic a common pattern in nature with vast potential applications [15]. Due to their hyperbranched structure, dendrimers are extremely versatile macromolecules. Their structure can be defined by three main elements: the inner core, repetitive branching units (dendrons), and terminal groups that provide surface tuning (Figure 1) [16].

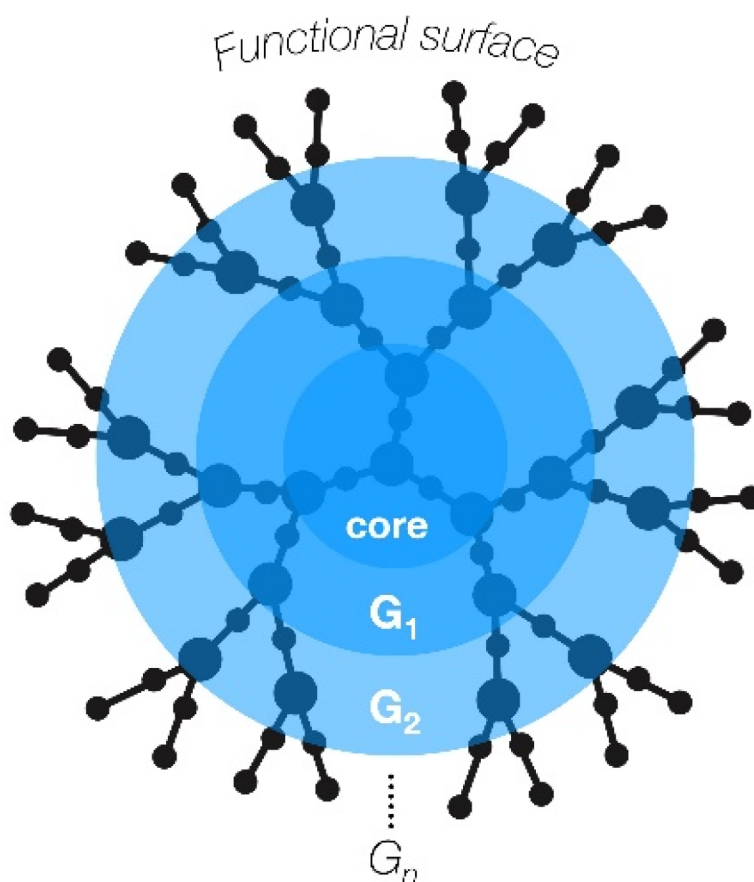


Figure 1. Schematic representation of a dendrimer nanoparticle, showing the core, repetitive branching units (dendrons) that constitute the growing layers (generations, G) and terminal groups (functional surface).

Dendrimers are classically obtained by two main approaches: a divergent or a convergent synthesis. In the first methodology, the dendrimer structure is constructed starting from a core molecule. The core reacts with monomers containing one reactive group and two dormant groups originating the first generation (G1) dendrimer. Then, the periphery of the G1 dendrimer may be activated for reaction with more monomers forming the second-generation dendrimer (G2), and so on. The divergent approach typically originates lower dendrimer generations of open structures and asymmetric shape [16]. In the convergent approach, the individual branches (dendrons) are first synthesized and then attached to a functional core molecule [17]. This methodology minimizes the occurrence of structural problems and facilitates the purification of the product [16]. Later, with the development of “click chemistry” [18], the preparation of dendronized systems became more efficient, requiring minimal purification [19]. She et al. [20], for instance, synthesized G2-poly-L-lysine dendrons that were connected at the core to a heparin sulphate moiety via “click chemistry”, and doxorubicin (DOX) was conjugated to the terminal ends using acyl hydrazine. As a consequence of the dendrimer architecture, the number of peripheral groups increases exponentially with generations, which results in nanosized particles suitable for drug loading and release [21]. However, when a critical branched state is reached, dendrimers cannot grow further because of the steric restriction imposed by the increasing branch density. This phenomenon is known as the “starburst effect” and is usually observed in high generations [16].

Since the discovery of these polymers [15], a variety of dendrimers have been developed, polyamidoamine (PAMAM) being the most studied [22]. PAMAM dendrimers are synthesized by the divergent method, mostly using an ethylenediamine core, and are hydrophilic, biocompatible, and non-immunogenic. Polypropylene imine (PPI) dendrimers, along with PAMAM, have been also widely investigated. PPIs are based on a 1,4-diaminobutane core or similar molecules and grow via double Michael addition reactions [13]. Poly-L-lysine (PLL) dendrimers are amino-acid-based polymers [23] that differ from PAMAM and PPI dendrimers in shape, since they are mostly asymmetrical. They have lysines as branching units and amines as terminal groups [22]. Phosphorous-based dendrimers are another interesting and well-studied class of dendrimers [24]. The potential of phosphorous dendrimers has been largely demonstrated, especially as cancer therapeutics. Similarly, carbosilane dendrimers have been explored as antimetastatic agents when complexed with ruthenium derivatives [25].

The different classes of dendrimers and their chemical structures are summarized in **Table 1**.

Table 1. Summary of different classes of dendrimers depicted in above section.

Classes of Dendrimers	Chemical Structure
PAMAM	Ethylenediamine-based core and terminal groups with primary amines

Classes of Dendrimers	Chemical Structure
PPL	Amino acid lysine-base core and branching units
PPI	1,4-Diaminobutane-based core and terminal groups with primary amines
Phosphorous dendrimers	P-Cl-based core, azabisphosphonates are possible terminal groups
Carbosilane dendrimers	Si-based dendrimer

Dendrimers can be used in a vast number of theranostic applications [26][27][28][29]. It is, therefore, important to be aware of the properties they have to display in order to be employed as biomedical devices. Biocompatibility is crucial to preventing undesirable responses from the host, a property that can only be defined depending on specific applications [29]. In order to prevent bioaccumulation and consequent toxicity, biodegradability is a must. Another important aspect for the development of biomedical devices is their pharmacokinetics, namely their fate in the body after administration [29]. Additionally, the water solubility of dendrimer–drug conjugates enhance the bioavailability of poorly soluble drugs [30]. Lastly, polyvalence, i.e., the ability to support versatile surface functionalization and multiple interactions with biological receptors, is a key property for highly versatile platforms [27].

Dendrimers, as shown in several studies, have a high potential to be used as nanocarriers for both diagnostic and therapeutic approaches [31][32]. Dendrimer–drug interactions might occur in many ways and are dependent on multiple factors such as size, charge, or the chemical nature of the dendrimer/drug. The chemistry behind nanocarriers is the same used in diagnostic or therapeutic schemes, with the agent selected for conjugation being the key player. In general, dendrimers could be used as nanocarriers via two major approaches: loading or conjugation at the surface of the drug and/or target molecule. Encapsulation solves solubility problems indicated by many chemotherapeutics and drugs in general. When a drug is entrapped into the dendrimer's cavity, the polymer works as a dendritic box [33][34]; in this case, the dendrimers can cargo the drug of interest by forming structures that are stabilized via non-covalent interactions. In a different context, dendrimers could also be used as gene vectors, especially cationic dendrimers. When the strategy is dendrimer–drug conjugation, systemic effects can be reduced, increasing the efficacy of cellular targeting. This strategy also improves the half-life of the drug. The conjugate linker is also key to understanding release mechanisms. In many cases, ester and amide conjugate linkers are used that allow enzymatic or hydrolytic cleavage [34][35], easier for esters than amides [36]. Dendrimer–drug conjugation may influence the efficacy of drug itself. Importantly, these nano-polymers can cross cellular barriers by transcellular or paracellular pathways.

To use dendrimers as an alternative diagnostic tool or improve the properties of a contrast agent, it is important to guarantee some criteria. Dendrimers offer many advantages to improve the free delivery of contrast agents or drugs, including high solubility and low polydispersity, which are properties of all dendrimer classes (**Table 2**).

Table 2. Suitable properties of dendrimers to be used as delivery systems in different strategies.

Properties	Observations	References
Low polydispersity index	Common to all classes of dendrimers	[37][38]
EPR effect	Size/generation/ M_w dependent	[39]
Permeability towards BBB	Already observed for PAMAM dendrimers	[40][41][42][43]
Highly solubility	Common to the majority of dendrimer classes	[44][45]
Multifunctional platform	Common to all classes of dendrimers	[46]
Highly loading capacity	Size/generation/ M_w dependent	[47]
Stability	Common to all classes of dendrimers	[39]
Low toxicity and immunogenicity	Size/generation/ M_w /charge dependent	[48][49][50]

EPR: enhanced permeability and retention, BBB: blood–brain barrier.

References

4. Dendrimers Cellular Uptake and Mechanism of Action at Cell Organelle Level

1. Hassanipour, S.H., Dehghani, M. Review of cancer from perspective of molecular. J. Cancer Res. Pract. 2017, 4, 127–129.

Dendrimers and other nanosized particles naturally accumulate in tumor sites by a possible enhanced permeability and retention (EPR) effect. This happens because the tumor microenvironment is rich in blood vessels due to increased angiogenesis. Using the blood vessels surrounding tumors, the nanoparticles can be driven into the

3. Parsa, N. Environmental factors inducing human cancers. Iran J. Public Health 2012, 41, 1–9.

tumors, crossing cellular barriers by transcellular or paracellular pathways [31][32]. The main question at this point is: how does the intracellular transportation of these macromolecules occur? In the case of dendrimers, their entry into

4. Pérez-Hernández, E.; Fernández-Medrano, S.; Apáiz, A. Advanced targeted therapies (Figure 2). Drug delivery and drug delivery systems: the future of chemotherapy. *Eur. J. Pharm. Biopharm.* 2015, 93, 527–79.

5. Quintana, A.; Raczka, E.; Piehler, L.; Lee, I.; Myc, A.; Majoros, I.; Patri, A.K.; Thomas, T.; Mule, J.; Baker, J.R., Jr. Design and function of a dendrimer-based therapeutic nanodevice targeted to tumor cells through the folate receptor. *Pharm. Res.* 2002, 19, 1310–1316.

6. Wiener, F.C.; Brechbiel, M.W.; Brothers, H.; Magin, R.L.; Gansow, O.A.; Tomalia, D.A.; Lauterbur, P.C. Dendrimer-based metal chelates: A new class of magnetic resonance imaging contrast agents. *Magn. Reson. Med.* 1994, 31, 1–8.

7. Alibolandi, M.; Hoseini, F.; Mohammadi, M.; Ramdani, P.; Einafshar, E.; Taghdisi, S.M.; Ramezani, M.; Abnous, K. Curcumin-entrapped MUC-1 aptamer targeted dendrimer-gold hybrid nanostructure as a theranostic system for colon adenocarcinoma. *Int. J. Pharm.* 2018, 549, 67–75.

8. Restani, R.B.; Morgado, P.I.; Ribeiro, M.P.; Correia, I.J.; Aguiar-Ricardo, A.; Bonifácio, V.D. Biocompatible polyurea dendrimers with pH-dependent fluorescence. *Angew. Chem. Int. Ed. Engl.* 2012, 51, 5162–5165.

9. Wu, Y.; Sefah, K.; Liu, H.; Wang, R.; Tan, V. DNA aptamer-micelle as an efficient detection/delivery vehicle toward cancer cells. *Proc. Natl. Acad. Sci. USA* 2010, 107, 5–10.

10. Shao, S.; Zhou, Q.; Si, J.; Tang, J.; Liu, X.; Wang, M.; Gao, J.; Wang, K.; Xu, R.; Shen, Y. A non-cytotoxic dendrimer with innate and potent anticancer and anti-metastatic activities. *Nat. Biomed. Eng.* 2017, 1, 745–757.

11. Sanvicens, N.; Marco, M.P. Multifunctional nanoparticles--properties and prospects for their use in human medicine. *Trends Biotechnol.* 2008, 26, 425–433.

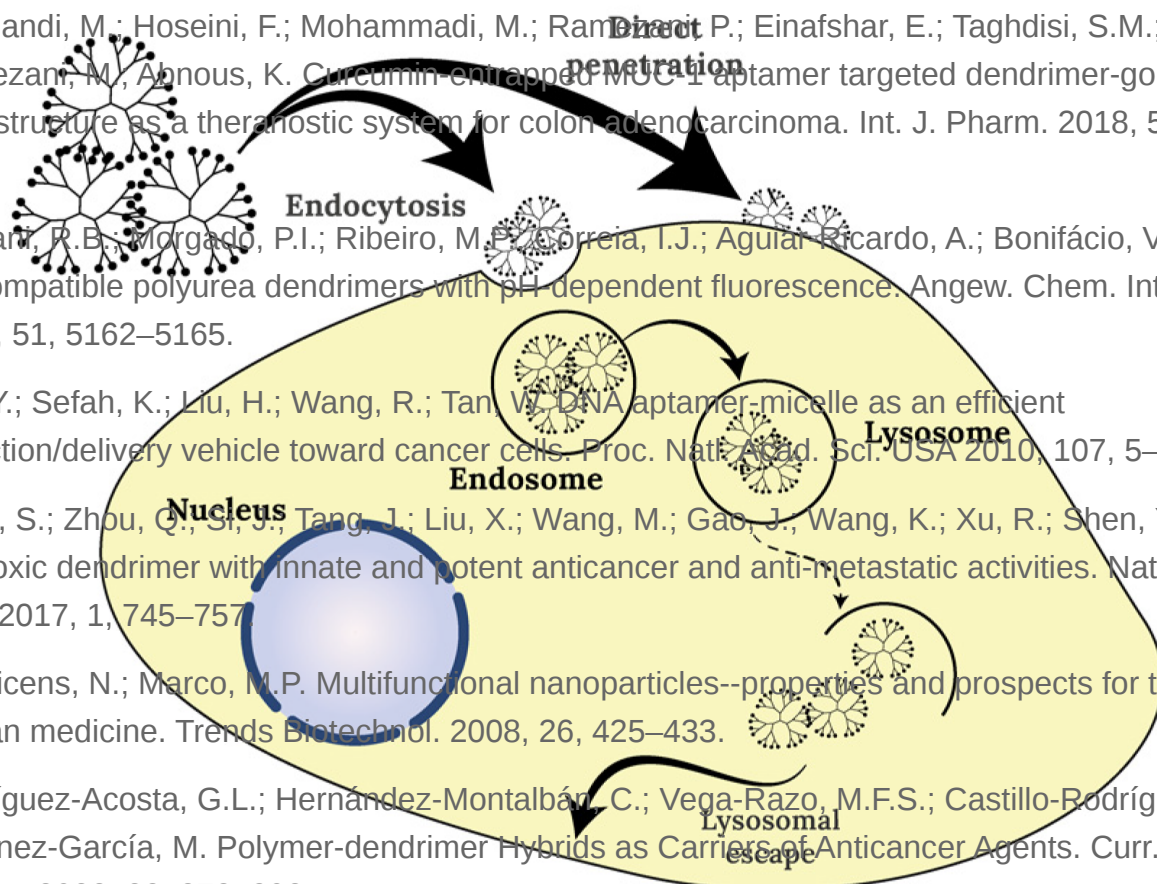
12. Rodríguez-Acosta, G.L.; Hernández-Montalbán, C.; Vega-Razo, M.F.S.; Castillo-Rodríguez, I.O.; Martínez-García, M. Polymer-dendrimer Hybrids as Carriers of Anticancer Agents. *Curr. Drug Targets* 2022, 23, 373–392.

13. Buhleier, E.; Wehner, W.; Vögtle, F. "Cascade"- and "Nonskid-Chain-like" Syntheses of Molecular Cavity Topologies. *Synthesis* 1978, 1978, 155–158.

14. Freygaard, P.W.; Boas, U.; Sørensen, N.S. Dendrimers for vaccine and immunostimulatory uses. A review. *Bioconjug. Chem.* 2010, 21, 405–418.

15. Tomalia, D.A.; Baker, H.; Dewald, J.; Hall, M.; Kallos, G.; Martin, S.; Roeck, J.; Ryder, J.; Smith, P. A New Class of Polymers: Starburst-Dendritic Macromolecules. *Polym. J.* 1985, 17, 117–132.

16. Wu, L.P.; Ficker, M.; Christensen, J.B.; Frohnapoles, P.N.; Maghni, S.M. Dendrimers in medicine: Therapeutic Concepts and Pharmaceutical Challenges. *Bioconjug. Chem.* 2019, 26, 1199–1211.



17. Hawker, C.J.; Frechet, J.M.J. The Preparation of Polymers with Controlled Molecular Architecture: A that
 18. Kolb, H.C.; Finn, M.G.; Sharpless, K.B. Click Chemistry: Diverse Chemical Function from a Few
 Good Reactions. *Angew. Chem. Int. Ed. Engl.* 2001, 40, 2004–2021.
19. Decker, L.; Daniel, M.C. Dendronized Systems for the Delivery of Chemotherapeutics. *Adv. Anionic*
PAMAM dendrimers 2018, 130, 85–120.
 the authors observed that PAMAM_{G2}-NH₂ and PAMAM_{G1.5}-CO₂H dendrimers co-localized at the lysosome level at
 20. She, W.; Li, N.; Luo, K.; Guo, C.; Wang, G.; Geng, Y.; Gu, Z. Dendronized heparin-doxorubicin
 conjugate based nanoparticle as pH-responsive drug delivery system for cancer therapy.
Biomaterials 2013, 34, 2252–2264.
 In HeLa cells (cervical cancer cells), PAMAM dendrimers were shown to be internalized by two major mechanisms:
 21. Lo, S.T.; Kumar, A.; Hsieh, J.T.; Sun, X. Dendrimer nanoscaffolds for potential theranostics of
 prostate cancer with a focus on radiochemistry. *Mol. Pharm.* 2013, 10, 793–812.
- In addition to endocytosis, the uptake of polycationic polymers by cancer cells can follow a direct penetration
 22. Palmerston Mendes, L.; Pan, J.; Torchilin, V.P. Dendrimers as Nanocarriers for Nucleic Acid and
 Drug Delivery in Cancer Therapy. *Molecules* 2017, 22, 1401.
23. Rahimi, A.; Amjad-iranagh, S.; Modarress, H. Molecular dynamics simulation of coarse grained
 poly(L-lysine) dendrimers. *J. Mol. Model.* 2016, 22, 59.
24. Camilade, A.M. Phosphorus Dendrimers as Nanotools against Cancers. *Molecules* 2020, 25,
 3333.
 formation can be reversible [58]. The pore-forming ability of cationic molecules such as dendrimers and peptides
 can be used to develop formulations for anticancer therapy, as this mechanism of action can potentially lead to cell
 25. Maroto-Díaz, M.; Elie, B.T.; Gómez-Sal, P.; Pérez-Serrano, J.; Gómez, R.; Contel, M.; Javier de la
 Mata, F. Synthesis and anticancer activity of carbosilane metallodendrimers based on arene
 ruthenium(II) complexes. *Dalton Trans.* 2016, 45, 7049–7066.
- Importantly, positively charged surface groups produce a localized charge density that is known to influence the
 interaction (and, consequently, toxicity) of dendrimers with cell membranes that possess a relevant content of
 26. Lee, C.C.; Mackay, J.A.; Frechet, J.M.J.; Szoka, F.C. Designing dendrimers for biological
 applications. *Nat. Biotechnol.* 2005, 23, 1517–1526.
27. Abbasi, F.; Aval, S.F.; Akbarzadeh, A.; Milani, M.; Nasrabadi, H.T.; Joo, S.W.; Hanifehpour, Y.;
 Nejati-Koshki, K.; Pashaei-Asl, R. Dendrimers: Synthesis, applications, and properties. *Nanoscale*
Res. Lett. 2014, 9, 247.
- Dendrimers can then be designed to target a specific organelle or tissue (with a distinct mechanism of action) and
 induce, for instance, less side effects to normal/healthy cells. Chemical modifications in the dendrimer core or
 28. Noriega-Luna, B.; Godínez, L.A.; Rodríguez, F.J.; Rodríguez, A.; Zaldivar-Leo de Lanrea, G.;
 Sosa-Pereyra, C.F.; Mercado-Cunel, R.F.; Manríquez, J.; Bustos, E. Applications of Dendrimers
 in Drug Delivery Agents, Diagnosis, Therapy, and Detection. *J. Nanomater.* 2014, 2014, 507273.
29. Duncan, R.; Izzo, L. Dendrimer biocompatibility and toxicity. *Adv. Drug Deliv. Rev.* 2005, 57,
 2215–2237.
30. Choudhary, S.; Gupta, L.; Rani, S.; Dave, K.; Gupta, U. Impact of Dendrimers on Solubility of
 Hydrophobic Drug Molecules. *Front. Pharm.* 2017, 8, 261.

31. Alven, S.; Aderibigbe, B.A. The Therapeutic Efficacy of Dendrimer and Micelle Formulations for Breast Cancer Treatment. *Pharmaceutics* 2020, 12, 1212.
32. Trembleau, L.; Simpson, M.; Cheyne, R.W.; Escofet, I.; Appleyard, M.V.C.A.L.; Murray, K.; Sharp, S.; Thompson, A.M.; Smith, T.A.D. Development of ^{18}F -fluorinatable dendrons and their application to cancer cell targeting. *New J. Chem.* 2011, 35, 2496–2502.
33. Liu, M.; Fréchet, J.M. Designing dendrimers for drug delivery. *Pharm. Sci. Technol. Today* 1999, 2, 393–401.
34. Menjoge, A.R.; Kannan, R.M.; Tomalia, D.A. Dendrimer-based drug and imaging conjugates: Design considerations for nanomedical applications. *Drug Discov. Today* 2010, 15, 171–185.
35. Wang, J.; Li, B.; Qiu, L.; Qiao, X.; Yang, H. Dendrimer-based drug delivery systems: History, challenges, and latest developments. *J. Biol. Eng.* 2022, 16, 18.
36. Najlah, M.; Freeman, S.; Attwood, D.; D'Emanuele, A. In vitro evaluation of dendrimer prodrugs for oral drug delivery. *Int. J. Pharm.* 2004, 275, 115–124.
37. Kolhe, P.; Khandare, S.; Shal, O.; Kannan, S.; Lieh-Lai, M.; Kannan, R. Dendrimer-branched polymer-drug conjugates with high drug payload for enhanced cellular delivery. *Pharm. Res.* 2004, 21, 2185–2191.
38. Patel, S.; Rajani, C.; Paul, D.; Borica, P.; Rajpoot, K.; Youngren-Ortiz, S.R.; Tekade, R.K. Chapter 8—Dendrimers as novel drug-delivery system and its applications. In *Drug Delivery Systems*; Tekade, R.K., Ed.; Academic Press: Cambridge, MA, USA, 2020; pp. 333–392.
39. Gai, D.B.; Gupta, N.; Pooja, D.; Kulhari, H. Dendrimers for diagnostic applications. In *Pharmaceutical Applications of Dendrimers*; Chaudhan, A., Kulhari, H., Eds.; Elsevier: Amsterdam, The Netherlands, 2020; pp. 291–324.

- Figure 3.** Schematic representation of potential intracellular cellular organelles (including plasma membrane, lipid droplets, cell nucleus, and mitochondria) targeted by dendrimers used in anticancer strategies. Cationic surface of conjugated dendrimer nanoparticles for glioblastoma multimodal delivery. *J. Colloid. Interface Sci.* 2015, 450, 396–403.
40. Zhao, J.; Zhang, B.; Shen, S.; Chen, J.; Zhang, Q.; Jiang, X.; Pang, Z. CREKA peptide-conjugated dendrimer nanoparticles for glioblastoma multimodal delivery. *J. Colloid. Interface Sci.* 2015, 450, 396–403.
41. Sharma, A.; Porterfield, J.E.; Smith, E.; Sharma, R.; Kannan, S.; Kannan, R.M. Effect of mannose targeting of hydroxyl PAMAM dendrimers on cellular and organ bio-distribution in a neonatal brain injury model. *J. Control. Release* 2018, 282, 175–189.
42. Srinageshwar, B.; Peruzzaro, S.; Andrews, M.; Johnson, K.; Hletpas, A.; Clark, B.; McGuire, C.; Petersen, E.; Kippe, J.; Stewart, A.; et al. PAMAM Dendrimers Cross the Blood-Brain Barrier When Administered through the Carotid Artery in C57BL/6J Mice. *Int. J. Mol. Sci.* 2017, 18, 628.
43. Srinageshwar, B.; Dils, A.; Sturgis, J.; Wedster, A.; Kathirvelu, B.; Baivasi, S.; Swanson, D.; Sharma, A.; Dunbar, G.L.; Rossignol, J. Surface-Modified G4 PAMAM Dendrimers Cross the

- The Blood-Brain Barrier Following Multiple Tail Vein Injections in MCF-7/ADR and SKOV-3 Cells. *Neurosci. Biomed. Res.* 2019, 10, 4145–4150.
44. Yiyun, C.; Tongwen, X. Dendrimers as potential drug carriers. Part I. Solubilization of non-steroidal anti-inflammatory drugs in the presence of polyamidoamine dendrimers. *Eur. J. Med. Chem.* 2005, 40, 1188–1192.
45. Hawker, C.J.D.; Wooley, K.L.; Fréchet, J.M.J. With unimolecular micelles and globular amphiphiles as model macromolecular scaffolds, novel biodegradable solubilization agents. *J. Chem. Soc. Perkin Trans. 1* 1993, 1, 1267–1267.
46. Vaidya, A.; Jain, S.; Pathak, K.; Pathak, D. Dendrimers: Nanosized Multifunctional Platform for Drug Delivery. *Drug Deliv. Lett.* 2018, 8, 3–19.
47. Singh, J.; Jain, K.; Mehra, N.K.; Jain, N.K. Dendrimers in anticancer drug delivery: Mechanism of interaction of drug and dendrimers. *Artif. Cells Nanomed. Biotechnol.* 2016, 44, 1626–1634.
48. Jam, K.; Kesharwani, P.; Gupta, U.; Jain, N.K. Dendrimer toxicity: Let's meet the challenge. *Int. J. Pharm.* 2010, 394, 122–142.
49. Pinto, S.N.; Mil-Homens, D.; Pires, R.F.; Alves, M.M.; Serafim, G.; Martinho, N.; Melo, M.; Fialho, A.M.; Bonifácio, V.D.B. Core-shell polycationic polyurea pharma dendrimers: New-generation of sustainable broad-spectrum antibiotics and antifungals. *Biomater. Sci.* 2022, 10, 5197–5207.
50. Pryor, J.B.; Harper, B.J.; Harper, S.L. Comparative toxicological assessment of PAMAM and thiophosphoryl dendrimers using embryonic zebrafish. *Int. J. Nanomed.* 2014, 9, 1947–1956.
51. Singh, V.; Sahebkar, A.; Kesharwani, P. Poly (propylene imine) dendrimer as an emerging polymeric nanocarrier for anticancer drug and gene delivery. *Eur. Polym. J.* 2021, 158, 110683.
52. Zhang, J.; Liu, D.; Zhang, M.; Sun, Y.; Zhang, X.; Guan, G.; Zhao, X.; Qiao, M.; Chen, D.; Hu, H. The cellular uptake mechanism, intracellular transportation, and exocytosis of polyamidoamine dendrimers in multidrug-resistant breast cancer cells. *Int. J. Nanomed.* 2016, 11, 3677–3690.
53. Shinde Patil, V.R.; Campbell, C.J.; Yun, Y.H.; Slack, S.M.; Goetz, D.J. Particle diameter influences adhesion under flow. *Biophys. J.* 2001, 80, 1733–1743.
54. Bamrungsap, S.; Zhao, Z.; Chen, T.; Wang, L.; Li, C.; Fu, T.; Tan, W. Nanotechnology in therapeutics: A focus on nanoparticles as a drug delivery system. *Nanomedicine* 2012, 7, 1253–1271.
55. Kitchens, K.M.; Foraker, A.B.; Kolhatkar, R.B.; Swaan, P.W.; Ghandehari, H. Endocytosis and interaction of poly (amidoamine) dendrimers with Caco-2 cells. *Pharm. Res.* 2007, 24, 2138–2145.
56. Kitchens, K.M.; Kolhatkar, R.B.; Swaan, P.W.; Ghandehari, H. Endocytosis inhibitors prevent poly(amidoamine) dendrimer internalization and permeability across Caco-2 cells. *Mol. Pharm.*

- 2008, 5, 364–369.
57. Albertazzi, L.; Serresi, M.; Albanese, A.; Beltram, F. Dendrimer internalization and intracellular trafficking in living cells. *Mol. Pharm.* 2010, 7, 680–688.
58. Hong, S.; Bielinska, A.U.; Mecke, A.; Keszler, B.; Beals, J.L.; Shi, X.; Balogh, L.; Orr, B.G.; Baker, J.R., Jr.; Banaszak Holl, M.M. Interaction of Poly(amidoamine) Dendrimers with Supported Lipid Bilayers and Cells: Hole Formation and the Relation to Transport. *Bioconjugate Chem.* 2004, 15, 774–782.
59. Jahanafrooz, Z.; Mokhtarzadeh, A. Pore-forming Peptides: A New Treatment Option for Cancer. *Curr. Med. Chem.* 2022, 29, 4078–4096.
60. Chernyshova, D.N.; Tyulin, A.A.; Ostroumova, O.S.; Efimova, S.S. Discovery of the Potentiator of the Pore-Forming Ability of Lantibiotic Nisin: Perspectives for Anticancer Therapy. *Membranes* 2022, 12, 1166.
61. Tajarobi, F.; El-Sayed, M.; Rege, B.D.; Polli, J.E.; Ghandehari, H. Transport of poly amidoamine dendrimers across Madin-Darby canine kidney cells. *Int. J. Pharm.* 2001, 215, 263–267.
62. Szwed, A.; Miłowska, K.; Michlewska, S.; Moreno, S.; Shcharbin, D.; Gomez-Ramirez, R.; de la Mata, F.J.; Majoral, J.P.; Bryszewska, M.; Gabryelak, T. Generation Dependent Effects and Entrance to Mitochondria of Hybrid Dendrimers on Normal and Cancer Neuronal Cells In Vitro. *Biomolecules* 2020, 10, 427.
63. Albertazzi, L.; Storti, B.; Marchetti, L.; Beltram, F. Delivery and subcellular targeting of dendrimer-based fluorescent pH sensors in living cells. *J. Am. Chem. Soc.* 2010, 132, 18158–18167.
64. Mukherjee, S.P.; Lyng, F.M.; Garcia, A.; Davoren, M.; Byrne, H.J. Mechanistic studies of in vitro cytotoxicity of poly(amidoamine) dendrimers in mammalian cells. *Toxicol. Appl. Pharm.* 2010, 248, 259–268.
65. Czarnomysy, R.; Muszyńska, A.; Rok, J.; Rzepka, Z.; Bielawski, K. Mechanism of Anticancer Action of Novel Imidazole Platinum(II) Complex Conjugated with G2 PAMAM-OH Dendrimer in Breast Cancer Cells. *Int. J. Mol. Sci.* 2021, 22, 5581.
66. Zhang, X.; Zhang, Z.; Xu, X.; Li, Y.; Li, Y.; Jian, Y.; Gu, Z. Bioinspired therapeutic dendrimers as efficient peptide drugs based on supramolecular interactions for tumor inhibition. *Angew. Chem. Int. Ed. Engl.* 2015, 54, 4289–4294.
67. Antunes, P.; Cruz, A.; Barbosa, J.; Bonifácio, V.D.B.; Pinto, S.N. Lipid Droplets in Cancer: From Composition and Role to Imaging and Therapeutics. *Molecules* 2022, 27, 991.
68. Sharma, A.; Khatchadourian, A.; Khanna, K.; Sharma, R.; Kakkar, A.; Maysinger, D. Multivalent niacin nanoconjugates for delivery to cytoplasmic lipid droplets. *Biomaterials* 2011, 32, 1419–1429.

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