

Essential Oils and Neurodegenerative Diseases

Subjects: [Clinical Neurology](#)

Contributor: Aswir RASHED

Despite the improvements in life expectancy, neurodegenerative conditions have arguably become the most dreaded maladies of older people. The neuroprotective and anti-ageing potentials of essential oils (EOs) are widely evaluated around the globe. The entry focuses on analysing the effectiveness of EOs as neuroprotective remedies among the four common age-related neurodegenerative diseases. The literature was extracted from three databases (PubMed, Web of Science and Google Scholar) between the years of 2010 to 2020 using the medical subject heading (MeSH) terms “essential oil”, crossed with “Alzheimer’s disease (AD)”, “Huntington’s disease (HD)”, “Parkinson’s disease (PD)” or “amyotrophic lateral sclerosis (ALS)”. Eighty three percent (83%) of the studies were focused on AD, while another 12% focused on PD. No classifiable study was recorded on HD or ALS. EO from *Salvia officinalis* has been recorded as one of the most effective acetylcholinesterase and butyrylcholinesterase inhibitors. However, only *Cinnamomum* sp. has been assessed for its effectiveness in both AD and PD.

essential oils

neurodegenerative

Alzheimer’s disease

Huntington’s disease

Parkinson’s disease

amyotrophic lateral sclerosis

in vitro

in vivo

1. Introduction

Aromatic plants consist of a wide and diverse array of organic compounds with significant ecological and physiological functions. One of the most vital components synthesised by aromatic plants are essential oils (EOs), along with its secondary metabolites and phenolic compounds ^[1]. EOs can be extracted and obtained from various parts of plants, such as the flower, bark, leaf, root, or peel ^{[2][3][4]}. Generally, monoterpenes and sesquiterpenes are the main constituents of EOs. Phenolic compounds are generated via biochemical synthesis and consist of a chemically heterogeneous group. Phenolic acids, simple phenols, coumarins, flavonoids, stilbenes, lignans, lignins, as well as hydrolysable and condensed tannins are among the well-established phenolic compounds ^{[5][6]}. EOs are volatile and they may play a role in cognitive improvement through olfactory pathways ^[7]. EOs are well-known for various benefits that include its antiviral, antibacterial, antifungal, memory enhancement, medicinal remedy, food preservation, cosmetic preservative, aromatherapy, and many other applications. For example, EO sourced from *Salvia* sp., which is one of the most common medicinal plant species, was reported for its notable remedy in cough, bronchitis, herpes, thrush wounds, as well as in impaired concentration. The EO of this species is also applied in the food industry and cosmetic industry, for ranges of perfume products ^[8].

The brain’s central nervous system (CNS) is composed of diverse neurons responsible for the organisation of neuronal and non-neuronal cells, as well as handling various motor, sensory, regulatory, behavioural, and cognitive functions. The neuronal cells are diverse in their morphology and function, suggesting that each neuronal type may indicate its own genomic profile despite having identical genetic codes. Within the CNS, specific regions were noticed to exhibit different vulnerabilities to ageing and various age-related neurodegenerative diseases ^[9].

Neurodegenerative disorders are often characterised by strong evidence of oxidative stress in their pathogenesis, as a consequence of unregulated synthesis of reactive oxygen species (ROS) ^[10]. Disparity observed in pro-oxidant and antioxidant cellular mechanisms inter-related with mitochondrial dysfunction, lipid peroxidation, neuroinflammatory processes, and endogenous dopamine metabolism are among the contributing factors to deregulation ^{[10][11]}. Many researchers have searched for molecules that activate blocking pathways or minimise the effects of ROS ^{[12][13]}. In an effort to overcome the limitations of current therapeutics available for neurodegenerative disorders, substantial research is being undertaken to explore and identify the availability of other possible natural drugs that are equally effective and without any side effects. As such, natural components composed of various polyphenolic phytochemicals have gained notable insight for this purpose ^[14].

Neurodegenerative disorders are currently incurable, and the available therapies only control symptoms or prolong the disease's growth. EOs have been proposed as an underlying preventive and treatment strategies for anti-ageing and neurodegenerative disorders [15]. Many studies have reported the potential of various EOs and their components to exhibit neuroprotective effects [16][17]. *Cinnamomum* sp. [18][19], *Salvia* sp. [20][21], *Polygonum* sp. [22], *Lavandula* sp. [23][24], *Citrus* sp. [25], *Artemisia* sp. [26][27], and *Zingiber* sp. [28] are among the most widely explored species for evaluating the effectiveness of EOs and its respective components in age-related neurodegenerative disorders. The four most commonly studied age-related neurodegenerative diseases are Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD) and amyotrophic lateral sclerosis (ALS).

AD is the most common cause of dementia in the elderly and is classified as a slow yet progressive neurodegenerative disorder. The highest prevalence rates are reported in North America and Western Europe followed by Latin America, China, and the Western Pacific. In general, it is highlighted that large number of AD cases are noticed among elderly people aged over 75, however early-onset of AD can also develop as early as 30 up to 60 years [29][30]. The direct cost entailed to AD diagnosis covers medical treatment or social services where a caregiver is needed, while a patient's or family members' income loss is referred to as indirect cost [31].

There are several AD profiles, which include deficits in episodic memory, language, semantic knowledge, visuospatial abilities, executive functions in terms of planning and organisation as well as apraxia [32]. Apart from neuronal loss, amyloid plaques and neurofibrillary tangles are inter-related to the presence of reactive astrocytes and activated microglial cells [33][34][35]. A β is the most widely studied component of AD pathogenesis, where it can induce neuronal toxicity and activate microglia leading to the indirect damage of neurons [36]. Proteolytic cleavage from the type I cell-surface protein amyloid precursor protein (APP) was known to yield several forms of A β [37][38]. The pathogenic hypotheses for synaptic and neuronal toxicity in Alzheimer's disease is shown in Figure 1.

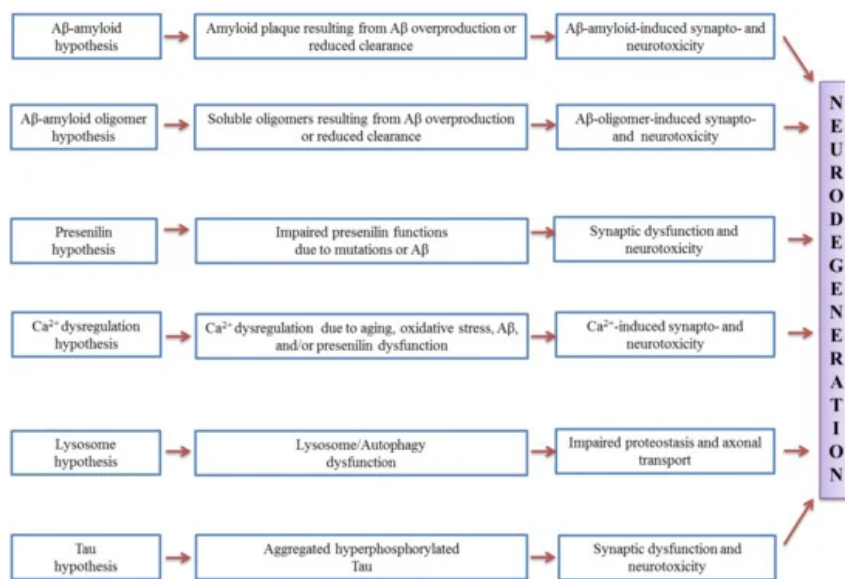


Figure 1. Pathogenic hypotheses for synaptic and neuronal toxicity in Alzheimer's disease (adapted from [39]).

A significant change that correlates to the age-related loss of substantia nigra (SN) dopaminergic cells is the loss of neostriatum dopaminergic innervation. Compelling studies have shown that the involvement of monoamine oxidase (MAO) in AD and neurodegenerative diseases is an important factor in many major pathophysiological pathways [40][41]. MAO-B has been suggested as a biomarker, and its activated form leads to cognitive dysfunction, kills cholinergic neurons, induces cholinergic disorders, and contributes to the development of amyloid plaques. Studies in molecular biology have demonstrated the critical role of A β generation through the modulation of the processing of APP by MAO [42][43][44][45]. The mechanism of A β generation through modulation of APP processing by activated MAO is shown in Figure 2.

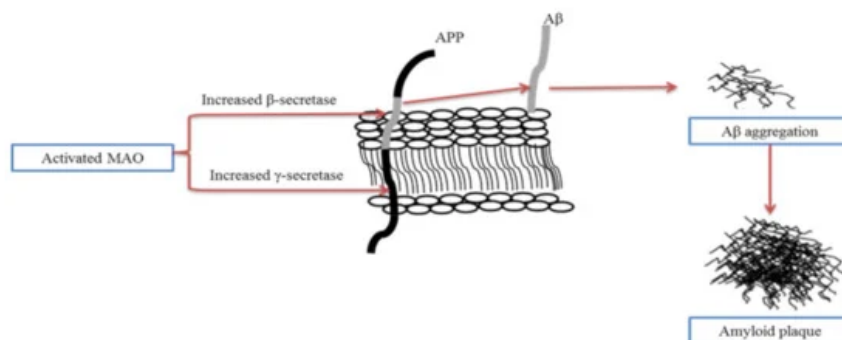


Figure 2. The mechanism of A β generation through modulation of amyloid precursor protein (APP) processing by activated monoamine oxidase (MAO). Activated MAO increases the expression of β -secretase and γ -secretase, improves A β generation, and contributes to the formation of amyloid plaques (adapted from [46]).

PD is the second most prevalent condition after AD, and also develops slowly over time [47]. PD can be identified and clinically characterised via motor impairment, which includes bradykinesia, rigidity, resting tremor and postural instability [48]. PD cases can be divided into sporadic (sPD) and familial (fPD), the latter of which represents approximately 20–25% of all PD cases. A common hallmark of sPD and fPD is the presence of intracellular inclusions, termed Lewy bodies [49][50].

α -Synuclein (α -Syn) has been identified as a major component of Lewy bodies in sporadic and familial cases, and is believed to be the central player in PD aetiology [51]. It is worthwhile to note that research conducted on PD has mainly focused on protein aggregation, neurotoxicity, increased oxidative stress, and mitochondrial dysfunction, as well as defects in the protein degradation machinery [52].

Apart from the role caused by α -Syn, the presence of neurotoxins, in particular 6-hydroxydopamine (6-OHDA) and 1-methyl-4-phenylpyridinium (MPP+), are widely accepted to induce neurotoxicity in PD patients. Both neurotoxins are thought to induce dopaminergic toxicity by intra- and extracellular oxidation, hydrogen peroxide formation, and direct inhibition of the mitochondrial respiratory chain [53].

The next common neurodegenerative disease is HD; caused by the recurrent development of cytosine–adenine–guanine (CAG) in the huntingtin (HTT) gene and involves a network of complex pathogenic mechanisms. HD is a profoundly penetrating, autosomal dominant, progressive neurodegenerative movement and neurobehavioural disorder associated with a variety of motor signs, psychological symptoms and cognitive dysfunction that progress with dementia. Knowledge of HD's causal mutation allows the detection of an ever-expanding number of HD phenotypes and phenocopies. The mean starting age is around 40 years, with a recorded range of 2 to 79+ years [54][55][56][57].

Progressive physical impairment of HD could be contributed to by various movement aspects such as hyperkinetic movements (dystonia, myoclonus, tics) and other motor manifestations (bradykinesia, incoordination, oculomotor function changes, gait impairment) along with chorea as the most distinct involuntary motion. As the disease progresses over time, dystonia becomes more prevalent and replaces chorea.

On the other hand, ALS is categorised as a heterogeneous neurodegenerative condition clinically, genetically, and pathologically [58][59][60]. The degeneration of cortical motor neurons and anterior horn cells of the spinal cord is characterised by ALS, also known as Charcot's or Lou Gehrig's disease. This contributes, usually within 3–5 years of diagnosis, to muscle atrophy, loss of muscle function, and death resulting from respiratory failure. The diverse clinical variability in ALS is believed to be due to differences in upper motor neuron (UMN) and lower motor neuron (LMN) involvement, extra-motor symptoms, onset age, survival, and progression rates. Heterogeneity of the disease prevents biomarker production which hinders the accurate evaluation of candidate drugs in clinical trials [59][60][61]. Various studies have shown that oxidative stress plays a major role in this disease's pathogenesis, identified as an unusual family type that often exhibits superoxide dismutase 1 (SOD1) gene mutations [62][63].

EOs are now commonly used and their advantages in all facets of life are investigated. The attractiveness of EO potential and possible mechanism of actions are ventured on a continual basis around the globe. In this review, we focus on and analyse the effectiveness of EOs as neuroprotective remedies among the four selected age-related neurodegenerative disorders mentioned above. We strongly believe that this review will be beneficial to many researchers and academicians that have great interest in EOs and their extensive applications.

2. The Source of EOs

As mentioned before, EOs can be extracted and obtained from various parts of plants. Clove's EO derived from the *Syzygium aromaticum* tree's aromatic flower buds with origin from Maluku, Indonesia contains the powerful scent used in spiced foods [64][65]. *Eucalyptus globulus* oil is mint-like, with properties such as a decongestant, pain relievers, antimicrobial agent, immunostimulant, flu and cold/cough treatment, as well as for mental clarity in aromatherapy [67][68]. One of the most influential EO is from *Lavandula angustifolia*, which is also known as English lavender. Lavender oil possesses strong antioxidant, anti-inflammatory, antibacterial, and antimicrobial properties, and can be used to treat various skin diseases (e.g., eczema, ringworm, acne), improve digestive system, minimise sore muscle swelling, and additionally, alleviate pain [67][68]. *Citrus limon* EOs are used as antimicrobial and antifungal agents, pain relievers, aids in weight loss, and to reduce extreme nausea as well as for usage as soaps, hair shampoo, furniture polishes, and fresheners [67][68]. Oregano (*Origanum vulgare*) EOs are often used for skin care, menstrual problems, stomach problems, and to control flu and cold infections [67][68]. Rosemary's EO originates from the *Rosmarinus officinalis* evergreen shrub with characteristics of a crisp woody, herbal and balsamic odour, similar to camphor. The usage of rosemary oil ranges from various treatments of skin care, dandruff, and scalp health, as well as to cold prevention and boosting the immune system [67][68]. EO from *Mentha piperita* is called peppermint oil, which is mainly used in the prevention of flu and colds, reduces headache symptoms, and also in relieving muscle and joint pains [67][68].

Apart from the general usage of EOs, their extensive benefits have also been noticed and reported in relevance to age-related neurodegenerative disorders. Based on available studies, EOs have been proposed as an effective preventive and treatment approach for anti-ageing and neurodegenerative disorders. Therefore, we attempted to describe and highlight the various EOs, and the effectiveness of their components with respect to the four common neurodegenerative diseases (AD, PD, HD, and ALS), as mentioned above. The different parameters that are commonly used for the evaluation of each disease are explained, accordingly.

Based on Table 1, a total of sixty-nine types of EOs from different genera of plants were evaluated for their effectiveness against neurodegenerative diseases among studies conducted between 2010 and 2020. In reference to all the compiled literature, we observed that the IC₅₀ results were presented in several formats, in particular to the units of measurement. Among the units that were reported included mg gallic acid equivalents/g [69][70], percentage [71][72], mg/mL or mg/L [26][73][74][75][76], µm [77], and µg/mL [18][78][79][80]. This diversity, however, was considered troublesome because direct comparison among studies with different measurement units is not possible without conversion.

3. Major Component of EOs

The chemical structures of several major components commonly found in EOs that have been reported to have anti-neurodegenerative properties are presented in Figure 3. Based on our review, 1,8-cineole has been identified as one of the major components found from various types of EOs. The compound 1,8-cineole is a saturated monoterpene that can originate from several plant species (e.g., *Eucalyptus*, *Rosmarinus*, and *Salvia*), with *Eucalyptus* leaves recognised as the key source [81]. Sometimes called eucalyptol due to its natural source, 1,8-cineole should not be confused with eucalyptus oil, a combination of many other components [82]. Due to its excellent aroma and taste, 1,8-cineole is mostly used in fruit, fragrances, and cosmetics. Furthermore, pure monoterpene 1,8-cineole is used as an alternative sinusitis remedy for respiratory tract infections, such as common cold or bronchitis [83]. It was indicated as one of the most potent free radical scavengers that may influence anticholinesterase activity based on a study reported by El Euch and colleagues [84]. The antioxidant activity was measured using the free radical 1,1-diphenyl-2-picrylhydrazyl (DPPH) test. Essential oil concentration providing 50% inhibition (IC₅₀) of the initial DPPH concentration was calculated using the linear relationship between the

compound concentration and the percentage of DPPH inhibition. Ascorbic acid was used as a standard. In the study by Abuhamdah et al. [85], EO extracted from the leaves of *Aloysia citrodora Palau* showed neuroprotective activity and a higher presence of 1,8-cineole was reported (23.66%).

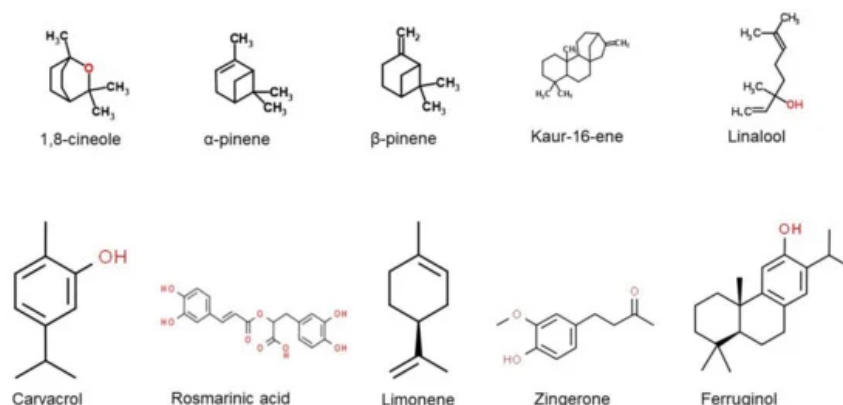


Figure 3. Chemical structures of the major EO components that have been reported to have anti-neurodegenerative properties. (obtained from [86][87][88][89][90][91][92][93][94][95]).

In another study performed by Cutillas and his team [96] on EO from *Thymus mastichina L.*, it was asserted that among all four compounds (α -pinene, β -pinene, limonene and 1,8-cineole), 1,8-cineole was the best AChE inhibitor with an IC_{50} of 35.2 ± 1.5 μ g/mL. They tested the antioxidant activity using five different methods such as the oxygen radical absorbance capacity (ORAC) assay, the 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulphonic acid) (ABTS) antioxidant method, the 2,2-diphenyl-1-picrylhydrazyl (DPPH) method, the thiobarbituric acid reactive substances (TBARS) method, and the chelating power (ChP) method. All the methods recorded that 1,8-cineole was one of the compounds highest in antioxidant capacity. This finding was similar to several other types of research, which suggested the efficacy of 1,8-cineole as the dominant anticholinesterase agent from EOs of different sources [8][21][25][97].

4. Cholinesterase Activity in EOs

The most common criteria used in the determination of AD is related to anti-cholinesterase activity. Cholinesterases (ChEs) are specialised carboxylic ester hydrolases that catalyse the hydrolysis of choline esters. Two types of ChE activity have been identified in mammalian blood and tissues, which are distinguished according to their substrate specificity and sensitivity to selective inhibitors. The first is acetylcholinesterase (AChE), which is systematically known as acetylcholine acetylhydrolase [98]. The second is butyrylcholinesterase (BChE), which is systematically referred to as acetylcholine acyl hydrolase [99][100][101][102]. The preferred substrate for AChE is acetylcholine (ACh), while butyrylcholine (BCh) and propionylcholine (PCh) are ideal for BChE [101][102][103]. AChE activity is known to be inhibited by several compounds, with toxins and drugs as the major inhibitors [104]. AChE activity is used in verifying treatment effects, especially in AD [103].

Both AChE and BChE possess active sites at the bottom of 20 Å-deep gorges with 50% identical amino acid sequence, whereas the gorge entrance locates the peripheral site [105]. The active site for both enzymes comprises a catalytic triad, acyl-binding pocket, and choline binding site [106]. A total of 14 aromatic amino acids are found in the active site of AChE, whereas six of these are substituted by aliphatic amino acids for BChE [107]. Binding and hydrolysis processes of bulky ligands are restricted in AChE due to presence of phenylalanine residues in the acyl binding pocket. In contrast, these residues are substituted with two flexible amino acids that are selective for BChE and allow the binding of bulkier ligands [108]. The different mechanisms involved in relevance to the active gorge site specific for each enzymes have been investigated via molecular modelling, structure-based virtual screening, or even crystallographic studies [107][108][109][110].

Generally, traditional Ellman assay is used with some modifications, applied for the determination of anti-cholinesterase activities [111][112]. This technique is a simple, accurate, and rapid method of measuring ChE activity, that is based on the reaction between thiocholine with the sulfhydryl group of a chromogen such as 5,5'-dithiobis-(2-nitrobenzoic acid) (DTNB or Ellman's reagent). The shift of electrons to sulphur atoms yields a yellow substance called 5-thio-2-nitrobenzoic acid (TNB),

which is measured by monitoring absorbance at 410 nm [113][114][115][116]. DTNB is a water-soluble compound and is useful for its fast reaction with thiocholine and minor side effects at neutral pH [111][113][114][115][116][117]. This technique, however, is also subject to certain limitations; it is restricted for testing antidotes against organophosphorus AChE inhibitors or for measuring AChE activity in samples of such treated individuals [116]. In addition to the Ellman assay, another method that can also be used for measuring ChE activities is the electrometric method of Michel [118]. This technique is applied based on pH changes that arise from H⁺ synthesis via cholinester hydrolysis [101][102][119].

5. Extracellular Plaque Deposits

Extracellular plaque deposits of the A β -peptide and flame-shaped neurofibrillary tangles of the microtubule-binding protein tau are the two hallmark pathologies required for AD patients. Familial early-onset forms of AD are associated with mutations either in the precursor protein for A β (APP) or in presenilin-1 (PS1) or presenilin-2 (PS2). Peptide generation pathways synthesise γ -secretase with either PS1 or PS2 as the catalytic subunit. APP is sequentially cleaved, where β -secretase first cleaves APP to release a large, secreted derivative, sAPP β , followed by γ -secretase that cleaves a fragment of 99 amino acids (CTF β) to generate A β . The process of γ -secretase cleaving can be inaccurate, leading to C-terminal heterogeneity of the resulting peptide population that generates numerous A β species, with A β 1–40 of the highest abundance followed by A β 42. The slightly longer forms of A β , particularly A β 1–42, are the principal species deposited in the brain that are more hydrophobic and fibrillogenic [119]. In view of their vital role in A β synthesis, both β - and γ -secretase are considered as key components in anti-AD pharmaceuticals developments [119][120]. Normal pathology tests refer to the density in the affected brain regions of neuritic amyloid plaques and neurofibrillary tangles of tau protein. AD diagnosis involves the presence of large neuritic plaque portions, consisting of highly insoluble A β in the brain parenchyma. There are also deposits of tau protein, although they occur among less common neurodegenerative disorders, especially in the absence of neuritic plaques. There are some distinctive morphological features of the neurofibrillary tangles in the various diseases, and may exhibit a distinct composition of tau isoforms that vary from AD [121].

It is not only humans which have amyloid beta; non-human primates (NHPs) have the same A β sequences as humans, an almost identical APP sequence, and they overlap with related human biochemical pathways in many aspects, however surprisingly with ageing, they develop relatively few AD-like neuropathologies. Aged canines also develop severe amyloid deposition; canines tend to demonstrate extensive amyloid deposition from about ten years of age, unlike in aged NHPs, where it could take several decades [122]. Amyloid deposition in canines is also interrelated with age-related cognitive dysfunction [123], although little neuronal loss is detected. Due to a poor understanding of AD and the human brain complexity, it has been deduced that there is no natural animal model of the disease [124]. For the past 25 years, pharmacological and genetic AD-models, as well as various animal species (primates, dogs, rodents, etc.), have been used in AD research activities [125][126]. The resurgence of interest in rats as the appropriate animal model of AD led to the usage of various types of rat models. As of current practice, transgenic mice have been extensively used in studies on AD. In any selected models, all need the introduction of some combined familial AD mutation into APP or PS1, or even in both [126].

6. Current EOs on AD

Salvia is the largest genus of plants in the family Lamiaceae, with the number of species estimated to range from 700 to nearly 1000. Fifteen species of *Salvia* (namely, *officinalis* L., *chionantha*, *chrysophylla* Staph, *urmiensis*, *nemorosa* L., *syriaca*, *balsiana*, *cyanescens*, *divaricate*, *hydrangea*, *kronenburgii*, *macrochlamys*, *nydeggeri*, *pachystachys*, *pse* and *rusellii*) were studied for cholinesterase inhibition assay, and happen to be the most widely studied source of EOs. Most of the researchers found that *Salvia* spp. were weak AChE and BChE inhibitors, except for EO from *S. pseudoeuphratica*, which demonstrated the highest inhibitory activity against AChE, at IC₅₀ = 26.00 ± 2.00 μ g/mL compared to *S. cyanescens* and *S. pachystachys*. In contrast, BChE activity did not show 50% inhibition even at the highest concentration, where IC₅₀ was reported to be above 80 μ g/mL for *S. pseudoeuphratica* and *S. hydrangea* [127]. *Salvia officinalis* was studied twice, in 2017 and 2019. Based on GC–MS and GC–FID analysis, the main components of *S. officinalis* were α -thujone, camphor, 1,8-cineole, and β -thujone. *Salvia* has also been used in ex vivo-based research, using the isolated guinea pig ileum method where the major molecule, rosmarinic acid, showed significant contraction responses on an isolated guinea pig

ileum. Docking results of rosmarinic acid also showed a high affinity to the selected target, AChE [128]. The author suggested the potential of rosmarinic acid to become a novel therapeutic candidate for the treatment of AD.

Other than *Salvia* spp., EOs from *Lavandula* spp. have also been studied for the treatment of AD. *L. luisieri* has been found to comprise high contents of oxygen-containing monoterpenes, mainly necrodane derivatives, which are absent from any other oil. This oil was tested on the endogenous beta-site APP-cleaving enzyme 1 (BACE-1) in cultured cells, being responsible for a reduction in A β production, with no significant toxicity. Although the study was conducted in vitro, the low molecular weight and high hydrophobicity of terpenoids are properties that provide a good chance for them to cross cellular membranes and the blood–brain barrier, an essential attribute for BACE-1 inhibition in vivo [129]. However, EO from *L. angustifolia* did not give a required finding because it enhanced the A β aggregation based on the thioflavin T method; this effect was further confirmed by atomic force microscope (AFM) imaging. EO of *L. angustifolia* was also showed to counteract the increase in intracellular reactive oxygen species production and the activation of the pro-apoptotic enzyme caspase-3 induced by A β 1–42 oligomers [23][24]. Meanwhile, EO from *L. pubescens* exhibited strong anti-AChE and anti-BChE effects at IC₅₀ of 0.9 μ L/mL and 6.82 μ L/mL, respectively. Carvacrol (CAR, 2-methyl-5-isopropylphenol) was also found to be higher in *L. pubescens*. Carvacrol was found to be abundant among EOs of the Lamiaceae family, and is known for various benefits including antibacterial, antifungal, antioxidant, antinociceptive, anti-inflammatory, anti-apoptosis, and anti-cancer activities [130]. Several studies on EOs have reported that carvacrol exerts some actions on the neuronal system, including AChE inhibition [131][132], anxiolytic [133], and antidepressant [134] properties. In addition, carvacrol has the ability to modulate central neurotransmitter pathways, such as dopaminergic, serotonergic and γ -aminobutyric acid (GABA)-ergic systems [130].

Only two types of cell lines, SH-SY5Y and PC-12, were reported to have been used in AD research. The in vitro toxic effects of amyloid peptides are usually examined using the human neuroblastoma-derived SH-SY5Y cell line, because differentiated neuron-like SH-SY5Y cells are extra-sensitive to amyloid peptides compared to non-differentiated cells, because the latter lack long neurites [135]. Z-ligustilide (Z-LIG) EOs effectively protect against fibrillar aggregates of A β 25–35- and A β 1–42-induced toxicity in SH-SY5Y and differentiated PC12 cells, possibly through the concurrent activation of the PI3-K/Akt pathway and inhibition of the p38 pathway [136]. A β 25–35 represents a neurotoxic fragment of A β 1–40 or A β 1–42, and retains the toxicity of the full-length peptide [137]. A β 25–35 is often selected as a model for full-length peptides because it retains both its physical and biological properties [138]. In general, declining levels of PI3K subunits as well as blunted Akt kinase phosphorylation have been observed in the AD brain, which is characterised by A β and tau pathologies [139].

There was also a study on the potential therapeutic effect of hybrid EO from Kushui roses. Kushui rose (*R. setate* \times *R. rugosa*) refers to a natural hybrid of cog rose and traditional Chinese rose that has been cultivated for more than 200 years [140]. In this study, transgenic worm strains purchased from the Caenorhabditis Genetics Center (CGC) were used instead of rat or mice models. They found that rose EO (REO) significantly inhibited AD-like symptoms of worm paralysis and hypersensitivity to exogenous serotonin (5-HT) in a dose-dependent manner. Although the GC–MS analysis revealed the presence of 40 components, the major components, β -citronellol and geraniol, were found to act less effectively than the oil itself. Intriguingly, REO significantly suppressed A β deposits and reduced the A β oligomers to alleviate the toxicity induced by A β overexpression [140].

Su He Xiang Wan (SHXW) has also been studied for its neurodegenerative remedy potential. SHXW is a distinct EO, and is a patent medicine comprising borneol, styrax resin, musk, aquilaria, frankincense, piper, benzoin, saussurea, cyperus, sandalwood, clove, terminalia, aristolachia fruit, rhino horn, and cinnabar. This ancient prescription was recorded in the *He Ji Ju Fang* of the Song Dynasty [141]. For this plant, the researchers evaluated the effects of a modified SHXW (KSOP1009 formulation) intake on the AD-like phenotypes of *Drosophila* AD models, which express human A β 1–42 in their developing eyes or neurons. They found that A β 1–42-induced eye degeneration, apoptosis, and locomotive dysfunctions were strongly suppressed. However, A β 1–42 fibril deposits in the A β 1–42 overexpressing model were not affected by treatment with KSOP1009 extract. Conversely, KSOP1009 extract intake significantly suppressed the constitutive active form of hemipterous, a c-Jun N-terminal kinase (JNK) activator, while it induced eye degeneration and JNK activation. In *Drosophila*, flies with mutations that augment JNK signalling accumulate less oxidative damage and live dramatically longer than wild-type flies [142][143].

Cinnamomum zeylanicum consisting of (*E*)-cinnamaldehyde (CAL) (81.39%) and (*E*)-cinnamyl acetate (CAS) (4.20%) as the main compounds showed over 78.0% inhibitory activity in cholinesterase. In MAO-A and MAO-B inhibition assays, EOs from *C. zeylanicum* (96.44%, 95.96%) and CAL (96.32%, 96.29%) demonstrated comparable activity to rasagiline (97.42%, 97.38%, respectively). Research by Murata and co-workers [77] found that kaur-16-ene, nezukol, and ferruginol isolated from plants had anti-AChE (IC₅₀) activity at 640, 300 and 95 μm, respectively. Even though ferruginol activity has already been highlighted before, by Gulacti et al. [144], this study documented the activities of kaur-16-ene and nezukol for the first time.

Meanwhile, *Citrus limon* has been found to significantly lower AChE brain depression in APP/PS1 and wild-type C57BL/6L (WT) mice. PSD95/synaptophysin, the synaptic density index, was substantially improved in histopathological shifts [145]. Based on the previous analysis by other researchers, nobiletin 3',4',5,6,7,8-hexamethoxyflavone was found to be the major component of polymethoxylated flavones in citrus peels, such as *C. depressa*, *C. reticulata*, *C. sinensis*, and *C. limon* [146][147]. Thus, nobiletin may potentially be the compound that substantially alters the development of these diseases. Other than that, *Acori graminei*, which was found to be rich in β-asarone, enhanced cognitive function of AβPP/PS1 mice and decreased neuronal apoptosis in the AβPP/PS1 mouse cortex. In addition, a substantial increase in the expression of CaMKII/CREB/Bcl-2 was observed in the cortex of AβPP/PS1 mice treated with β-asarone.

In a study conducted by Ayuob et al. [148], *Ocimum basilicum* up-regulated the serum corticosterone level, the hippocampal protein glucocorticoid receptor, and the brain-derived neurotrophic factor (BDNF); however, it down-regulated the neurodegenerative and atrophic changes induced in the hippocampus, which decreased after exposure to chronic unpredictable mild stress (CUMS). According to the data collected by Avetisyan and co-workers [149], the major components of *O. basil* includes methyl chavicol and linalool. Interestingly, many linalool-producing plants are commonly used in folk medicine and aromatherapy to alleviate symptoms and treat multiple acute and chronic diseases [150][151]. Linalool is frequently used in the manufacture of fragrances for shampoos, soaps, detergents, and in pharmaceutical formulations [152][153]. Research conducted by Sabogal-Guáqueta et al. [154] found that oral administration of monoterpene linalool to elderly mice (21–24 months old) with a triple transgenic form of AD (3x Tg-AD) at 25 mg/kg for three months at an interval of 48 h resulted in enhanced learning and spatial memory and increased risk assessment activity in the elevated plus maze. Hippocampi and amygdalae from 3x Tg-AD linalool-treated mice also showed a large reduction in extracellular β-amyloidosis, tauopathy, astrogliosis, and microgliosis, as well as pro-inflammatory marker levels of p38 MAPK, NOS2, COX2 and IL-1β. Thus, linalool is suitable as an AD prevention candidate for pre-clinical studies. Based on the articles that we have selected, linalool is a major volatile component of EOs in a number of aromatic plant species, such as *L. angustifolia* Mill., *M. officinalis* L., *R. officinalis* L., and *C. citrate* DC. The presence of linalool can also help to reduce the deposits of Aβ, based on a study conducted by Gradinariu et al. [155] where Aβ1–42-treated rats exhibited the following: a decrease in exploratory activity (crossing number); smaller percentage of time spent and fewer entries in the open arm in the elevated plus-maze test; increase in swim time; and decrease in the immobility time within the forced swimming test.

7. Current EOs on PD

In terms of PD, the current therapy in practice is applied as a combination of gold-standard dopaminergic reposition with 3-(3,4-dihydroxyphenyl)-L-alanine (L-dopa), along with other agents such as MAO-B, catechol *O*-methyltransferase (COMT) inhibitors, dopaminergic agonists, and cholinergic blockers [156]. However, the available treatments are subject to consequences of motor and non-motor side effects, which leads to poor efficacy in advanced stages of PD [157]. These arising phenomena are the main reasons that suggest and emphasise the necessity for the synthesis of anti-PD drugs that could delay the progression of neurodegeneration [157]. As mentioned in the results, PD studies included in this review comprise in vitro and in vivo, as well as combinations of in vitro with in vivo or ex vivo research. *Cinnamomum* sp., *Eryngium* sp., *Myrtus* sp., *Acorus* sp., *Eplingiella* sp., *Foeniculum* sp., *Pulicaria* sp., *Rosa* sp., *Zingiber* sp., and *Lavandula* sp. were among the identified EOs based on the respective included PD studies.

Four of the included studies on PD were based on in vitro approaches, with various EOs. The first study was focused on the evaluation of protective effects of EOs extracted from *Cinnamomum* sp. (*C. verum* and *C. cassia*) and cinnamaldehyde, in comparison to hydroalcoholic extracts using 6-OHDA-induced PC12 cytotoxicity as the representative model of PD [158]. *Cinnamomum* sp., or more commonly known as cinnamon, belongs to the Lauraceae family that is composed of almost

250 species and has been acknowledged for extensive health benefits [159][160]. Among the various species, *C. verum* and *C. cassia* were the two main species that have been widely applied for their medicinal and culinary applications, especially in Iran [161][162][163]. It is important to note that cinnamaldehyde represents one of the key components of both species, and EOs were reported to exhibit strong antioxidant properties [159]. The findings of this study indicated that 6-OHDA led to cell death, cell apoptosis, and suppression of the p44/42 pathway. On the whole, the study concluded that synergistic effects of cinnamaldehyde and EOs as well as other extract components could promote cinnamon's roles as neuroprotective agents, specifically for PD treatment [158].

Eryngium sp. belong to the Apiaceae family, and are recognised for their EOs' potentials in MAO inhibition [164]. MAO is available in two forms (A, B) where MAO-A inhibition is linked to antidepressant effects, while MAO-B is correlated with PD treatment [165][166]. An in vitro study conducted by Klein-Júnior et al. demonstrated the assessment of *Eryngium* sp. (*E. floribundum*: EP, *E. horridum*: EH, *E. pandanifolium*: EP, *E. eriophorum*: EE and *E. nudicaule*: EN) EOs for their MAO inhibitory effect. Intriguingly, the findings of this study indicated that MAO-A activity was not inhibited by any EOs, while EPEO and EHEO resulted in MAO-B inhibition. The literature search has also highlighted that PD patients usually represent elevated levels of MAO-B which arise due to gliosis, and hence contribute towards the collapse of the dopaminergic system [166]. Thus, this study puts forward the claim that *Eryngium* sp. could have potential applications as CNS bioactive secondary metabolites, particularly for neurodegenerative disease, in relevance to characteristics exhibited by EHEO [164].

Another important aspect that is usually associated with studies on PD is on α -Syn fibrillation. It is presumed that protein structural modifications resulting in amyloid fibril formation progresses towards neurodegenerative disorders [167]. However, the exact factors of α -Syn actions of misfolding and aggregation in the brain are still scarce. In addition, it should also be noted that preventive measures against α -Syn fibrillation are yet to be available; hence, the acceleration of the fibrillation process via certain factors should be avoided. Among some of the common factors are metal ions, small molecules, nanoparticles, and, particularly, toxins that could intensify the aggregation process [167][168][169][170][171]. The next two in vitro studies were performed by the same research team, where they highlighted the effects of 15 various Iranian EOs against α -Syn fibrillation [167][172].

Among all the 15 oils tested, it was shown that *M. communis* demonstrated potential benefits because it elevated the fibrillation in a concentration-dependent manner. However, it is necessary to understand that the major components of this oil are not responsible for the observed changes, suggesting complexity of both extract and synergistic effects of the available compounds, regardless of their amount [167]. In the second study, the investigation on *C. cyminum* EO signified the presence of cuminaldehyde as the major active compound that plays its role in the inhibition of α -Syn fibrillation. In addition, cytotoxicity assays on PC12 cells indicated the absence of toxic effects with cuminaldehyde treatment throughout α -Syn fibrillation [172].

Apart from in vitro studies, PD studies are also extensively performed under in vivo conditions. EOs of *Acorus* sp. cover two of the in vivo studies in this review. Both of these studies were conducted by the same research team, and focused on the regulation effect of β -asarone that was isolated from *A. tatarinowii* Schott on 6-OHDA-induced parkinsonian rats via two distinct endoplasmic reticulum (ER) stress pathways [173][174]. ER is known for its role in protein folding, where the build-up of protein unfolding/misfolding could initiate a phenomenon called ER stress that further activates the cellular process of unfolded protein response (UPR) [175]. ER stress has been noticed in a number of PD experimental models, and is also provoked by an increase in wild-type α -Syn [176][177][178]. Three main pathways that are categorised as UPR are inositol requiring enzyme 1 (IRE1), protein kinase RNA (PKR)-like ER kinase (PERK), and activating transcription factor 6 (ATF6) [179].

In general, GRP78 functions in the regulation of ER stress, where it binds the three proteins of the UPR pathway and maintains them as inactive when the cells are not exposed to stress conditions. However, under conditions of accumulated protein unfolding or misfolding, GRP78 will bind to the proteins and release them [180][181][182]. In terms of autophagy, the latest research has claimed that it could be induced due to ER stress [183][184]. Beclin-1 is known for its role in forming autophagosomes and is an essential part of the initial autophagy process. The pro-autophagic role of Beclin-1 could be inhibited via its reaction with Bcl-2, however this interaction is also subject to disruption caused by Bcl-2 phosphorylation that leads to Beclin-1 release and accelerates autophagy [185]. As per the second pathway study, it was proven that β -asarone leads to Beclin-1 downregulation, which highlights that Bcl-2 could possibly be the main linkage between autophagy and ER

stress. The findings of both studies lead us to conclude that diminishing ER stress via β -asarone regulation is proven to be useful in the impairment of PD pathological progression [173][174].

Another study that also applies the use of 6-OHDA-induced PD mouse models focused on the investigation of the effects of zingerone and eugenol on dopamine concentration, behavioural changes, and antioxidant activities upon 6-OHDA administration and treatment of L-dopa [28]. Zingerone is extracted from the ginger root, while eugenol originates from cloves and was reported to be protective against 6-OHDA-induced depletion of striatal dopamine via increases in SOD activity and elevation of reduced glutathione (GSH) and L-Ascorbate (Asc) concentration, respectively [186][187]. Although these groups of researchers previously reported positive findings where pre-treatment with zingerone or eugenol inhibited 6-OHDA-induced dopamine depression by preventing lipid peroxidation, the current study, which involved post-treatment with similar compounds, resulted in contradictory findings, where dopamine decrease was more pronounced [28][187]. Despite the availability of other findings that propose the benefits of consuming these compounds, Kabuto and Yamanushi [28] suggested that intake of these specific substances upon the onset of PD symptoms should be more carefully monitored to prevent further injury aggravation.

In addition to studies among 6-OHDA-induced Parkinson's rat models, the possibility to achieve positive effects of EOs when complexed with β -cyclodextrin (β CD) was evaluated by Filho and colleagues using reserpine-induced progressive models for PD in mice [157]. Cyclodextrins are cyclic oligosaccharides that could form host-guest complexes with hydrophobic molecules and were also reported to protect EOs from heat, evaporation, moisture, oxidation and light effects along with facilitating easy solubility [188][189][190]. Complexation effects of cyclodextrins with EOs were shown to be more prominent in exerting positive effects, especially in the treatment of chronic diseases, as published by several studies [191][192]. In this particular study, the same approach was applied using leaf EO extracted from *Eplingiella fruticosa* (EPL), where one of the key components is 1,8-cineole. *Eplingiella* sp. belongs to the Lamiaceae family, and was reported for its benefits as anti-inflammatory and antioxidant effects [193][194]. This research demonstrated and proved the hypothesis whereby both treated groups of EPL and EPL- β CD deferred reserpine effects on catalepsy time. However, this effect was noticed to be more remarkable with EPL- β CD treated mice groups.

Another study also applied induction with reserpine, with a different PD model of ovariectomized and non-ovariectomized rats [195]. Reserpine is known as an irreversible inhibitor of the vesicular monoamine transporter 2 (VMAT-2). The approach of reserpine injection to rats as a mode of PD model was proposed in response to its action on the depletion of monoamine and locomotor activities [196]. Ovariectomized rats are subjected to oestrogen deficits, similar to surgically menopausal women, where cognitive damage is highly possible [197]. Lower oestrogen levels are correlated with many side effects, such as mental disorders, memory defects, emotional issues, and other cognitive failures [198]. These incidences have led to much attraction towards phytoestrogens for its protective nature against certain diseases. Fennel plant (*Foeniculum vulgare*) is classified as in the Apiaceae family, and is known for its phytoestrogen compounds; it showed promising results in the treatment of cognitive disorders, such as dementia and AD [195][199][200][201]. The evaluation of this study indicated that protective oestrogen effects against neurodegenerative disorders were significantly decreased among reserpine-induced ovariectomized rats. Injection of reserpine resulted in a more remarkable observation on limb movement disorder among ovariectomized rats. Fennel treatment at various doses for both groups gave better results on the motor activity, which stressed the importance of oestrogens and phytoestrogens as a protective measure of dopaminergic neurons and improved PD symptoms [195].

Rotenone administration to rats, as induction of a PD model, is an alternative approach where it induces nigrostriatal dopaminergic neuron degeneration that is associated with α -Syn Lewy bodies [202]. Rotenone is an insecticide with high lipophilic nature, and is known to inhibit mitochondrial complex-1 along with causing oxidative stress [203][204]. This rotenone-induced model was reported in the study by Issa and colleagues, based on the neuroprotective effects of *Pulicaria undulata* EO in male Wistar rats [205]. *P. undulata* belongs to the Asteraceae family, which is commonly distributed in Asia, Europe, and North Africa [206]. From this study, it was shown that EO of *P. undulata* could exert its neuroprotective effects via anti-inflammatory and antioxidant properties. The mechanisms involved in neuroinflammation suppression include downregulation of induced nitric oxide synthase (iNOS) expression, followed by lower gene expression of α -Syn [205].

Compared to individual studies, there are also several approaches that examine combined effects that could incorporate in vitro, in vivo, and also ex vivo applications. One such attractive research is on the combined in vitro/in vivo evaluation of

SHXW EO with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced PD mice and SH-SY5Y cell lines [207]. In this study, SHXW was a Chinese herbal formulation that consisted of 15 crude herbs called KSOP1009, composed of eight medicinal plants of different families (Hamamelidaceae, Myricaceae, Umbelliferae, Santalaceae, Piperaceae, Myrtaceae, Typhaceae, and Lamiaceae). MPTP is known to cause fast degeneration of dopaminergic neurons, and as such, it was believed that the use of this specific model could assist in explaining certain aspects of PD disease mechanisms [207][208]. Positive findings of the study demonstrated that ingestion of KSOP1009 was successful in the protection of MPTP toxicity, where this could be correlated with dopamine reduction that also decreases ROS and restores mitochondrial roles [207].

In terms of in vivo and ex vivo combinatorial approaches, two articles were highlighted in this review, with several authors originating from the same team [209][210]. Both studies explored the approach of L-dopa induction of oxidative toxicity. L-dopa has been recognised as the most effective symptomatic treatment of PD for more than 30 years; however, toxicity issues that were raised via in vitro studies seem to be an unresolved challenge [209]. It was also mentioned that prolonged L-dopa treatment is often associated with side effects that often result in a delay in its administration [209]. Past studies have claimed that L-dopa therapy in combination with antioxidants could lessen the possible side effects. As such, efforts were taken to evaluate the effects; this research team investigated the combined effects of EOs from *Lavandula angustifolia*, *Rosa damascena*, vitamin C and Trolox in the initial study [209], followed by another study in 2019 on the combined effects of pre-treatment with *Rosa damascena* and vitamin C [210].

For the first study, the obtained results indicated that both EOs from herbal plants showed noticeable radical scavenging and antioxidant properties against L-dopa toxicity. Similarly, the second study also put forward equivalent claims with *R. damascena* characteristics where it was in parallel to vitamin C, and exhibited a significant role of rose oil in its interference against the acute oxidative toxicity of L-dopa [209][210]. Based on all the collective studies, it could be observed that PD treatments remain centralised among several parameters that include mainly α -Syn fibrillation, MAO-B, β -asarone regulation of ER stress pathways, toxicity-induced models with 6-OHDA, MPTP, L-dopa, reserpine, and rotenone with common animal models of rats and mice. Although the regulation mechanisms involved in each of the parameters may differ, the main focus remains towards effective and improved treatments for PD patients.

References

1. Osbourn, A.E.; Lanzotti, V. *Plant-Derived Natural Products: Synthesis, Function, and Application*; Springer Science & Business Media: New York, NY, USA, 2009; p. 612.
2. Esposito, E.R.; Bystrek, M.V.; Klein, J.S. An Elective Course in Aromatherapy Science. *Am. J. Pharm. Edu.* 2014, 78, 79.
3. Robins, J.L.W. The Science and Art of Aromatherapy. *J. Holist. Nurs.* 1999, 17, 5–17.
4. Smith, C.A.; Collins, C.T.; Crowther, C.A. Aromatherapy for pain management in labour. *Cochrane Database Syst. Rev.* 2011.
5. Garcia-Salas, P.; Morales-Soto, A.; Segura-Carretero, A.; Fernández-Gutiérrez, A. Phenolic-compound-extraction systems for fruit and vegetable samples. *Molecules* 2010, 15, 8813–8826.
6. Singer, A.C.; Crowley, D.E.; Thompson, I.P. Secondary plant metabolites in phytoremediation and biotransformation. *Trends Biotechnol.* 2003, 21, 123–130.
7. Hsieh, M.T.; Peng, W.H.; Wu, C.R.; Wang, W.H. The ameliorating effects of the cognitive-enhancing Chinese herbs on scopolamine-induced amnesia in rats. *Phytother. Res.* 2000, 14, 375–377.
8. Cioanca, O.; Mircea, C.; Hritcu, L.; Trifan, A.; Marius, M.; Aprotosoiaie, A.C.; Robu, S.; Gille, E.; Hancianu, M. In vitro—In vivo correlation of the antioxidant capacity of *Salviae aetheroleum* essential oil. *Farmacia* 2015, 63, 34–39.
9. Jové, M.; Portero-Otín, M.; Naudí, A.; Ferrer, I.; Pamplona, R. Metabolomics of Human Brain Aging and Age-Related Neurodegenerative Diseases. *J. Neuropath. Exp. Neur.* 2014, 73, 640–657.

10. Yan, M.H.; Wang, X.; Zhu, X. Mitochondrial defects and oxidative stress in Alzheimer disease and Parkinson disease. *Free Radic. Biol. Med.* 2013, 62, 90–101.
11. Sarrafchi, A.; Bahmani, M.; Shirzad, H.; Rafieian-Kopaei, M. Oxidative stress and Parkinson's disease: New hopes in treatment with herbal antioxidants. *Curr. Pharm. Des.* 2016, 22, 238–246.
12. González-Burgos, E.; Gómez-Serranillos, M.P. Terpene compounds in nature: A review of their potential antioxidant activity. *Curr. Med. Chem.* 2012, 19, 5319–5341.
13. Essa, M.; Braidy, N.; Bridge, W.; Subash, S.; Manivasagam, T.; Vijayan, R.; Al-Adawi, S.; Guillemin, G. Review of natural products on Parkinson's disease pathology. *J. Aging Res. Clin. Pract* 2014, 3, 1–8.
14. Karakaya, S.; Koca, M.; Yilmaz, S.V.; Yildirim, K.; Pınar, N.M.; Demirci, B.; Brestic, M.; Sytar, O. Molecular Docking Studies of Coumarins Isolated from Extracts and Essential Oils of *Zosima absinthifolia* Link as Potential Inhibitors for Alzheimer's Disease. *Molecules* 2019, 24, 722.
15. Forrester, L.T.; Maayan, N.; Orrell, M.; Spector, A.E.; Buchan, L.D.; Soares-Weiser, K. Aromatherapy for dementia. *Cochrane Database Syst. Rev.* 2014, CD003150, 1–56.
16. Xu, P.; Wang, K.; Lu, C.; Dong, L.; Gao, L.; Yan, M.; Aibai, S.; Yang, Y.; Liu, X. Protective effects of linalool against amyloid beta-induced cognitive deficits and damages in mice. *Life Sci.* 2017, 174, 21–27.
17. Bansal, A.; Kirschner, M.; Zu, L.; Cai, D.; Zhang, L. Coconut oil decreases expression of amyloid precursor protein (APP) and secretion of amyloid peptides through inhibition of ADP-ribosylation factor 1 (ARF1). *Brain Res.* 2019, 1704, 78–84.
18. Dalai, M.; Bhadra, S.; Chaudhary, S.; Bandyopadhyay, A.; Mukherjee, P. Anti-cholinesterase potential of *Cinnamomum tamala* (Buch.-Ham.) T.Nees & Eberm. leaves. *Indian J. Tradit. Know.* 2014, 13, 691–697.
19. Sihoglu Tepe, A.; Ozaslan, M. Anti-Alzheimer, anti-diabetic, skin-whitening, and antioxidant activities of the essential oil of *Cinnamomum zeylanicum*. *Ind. Crop. Prod.* 2020, 145, 112069.
20. Bahadori, M.B.; Salehi, P.; Sonboli, A. Comparative study of the essential oil composition of *Salvia urmiensis* and its enzyme inhibitory activities linked to diabetes mellitus and Alzheimer's disease. *Int. J. Food Prop.* 2017, 20, 2974–2981.
21. Cutillas, A.B.; Carrasco, A.; Martinez-Gutierrez, R.; Tomas, V.; Tudela, J. *Salvia officinalis* L. Essential Oils from Spain: Determination of Composition, Antioxidant Capacity, Antienzymatic, and Antimicrobial Bioactivities. *Chem. Biodivers.* 2017; 14.
22. Ayaz, M.; Junaid, M.; Ullah, F.; Sadiq, A.; Khan, M.A.; Ahmad, W.; Shah, M.R.; Imran, M.; Ahmad, S. Comparative chemical profiling, cholinesterase inhibitions and anti-radicals properties of essential oils from *Polygonum hydropiper* L: A Preliminary anti- Alzheimer's study. *Lipids Health Dis.* 2015, 14, 141.
23. Ali-Shtayeh, M.S.; Abu-Zaitoun, S.Y.; Dudai, N.; Jamous, R.M. Downy Lavender Oil: A Promising Source of Antimicrobial, Antiobesity, and Anti-Alzheimer's Disease Agents. *Evid-Based Complement. Altern. Med.* 2020, 2020, 5679408.
24. Soheili, M.; Khalaji, F.; Mirhashemi, M.; Salami, M. The Effect of Essential Oil of *Lavandula Angustifolia* on Amyloid Beta Polymerization: An In Vitro Study. *Iran. J. Chem. Chem. Eng.* 2018, 37, 201–207.
25. Chaiyana, W.; Okonogi, S. Inhibition of cholinesterase by essential oil from food plant. *Phytomedicine* 2012, 19, 836–839.
26. Shoaib, M.; Shah, I.; Ali, N.; Ali, W. In vitro acetylcholinesterase and butyrylcholinesterase inhibitory potentials of essential oil of *Artemisia macrocephala*. *Bangladesh J. Pharm.* 2015, 10, 87–91.
27. Ansari, M.; Eslami, H. Preparation and study of the inhibitory effect of nano-niosomes containing essential oil from *artemisia absinthium* on amyloid fibril formation. *Nanomed. J.* 2020, 7, 243–250.
28. Kabuto, H.; Yamanushi, T.T. Effects of zingerone [4-(4-hydroxy-3-methoxyphenyl)-2-butanone] and eugenol [2-methoxy-4-(2-propenyl) phenol] on the pathological progress in the 6-hydroxydopamine-induced

- parkinson's disease mouse model. *Neurochem. Res.* 2011, 36, 2244.
29. Alzheimer's Association. 2019 Alzheimer's disease facts and figures. *Alzheimer's Dement.* 2019, 15, 321–387.
 30. Reitz, C.; Mayeux, R. Alzheimer disease: Epidemiology, diagnostic criteria, risk factors and biomarkers. *Biochem. Pharm.* 2014, 88, 640–651.
 31. Ferretti, C.; Sarti, F.M.; Nitrini, R.; Ferreira, F.F.; Brucki, S.M.D. An assessment of direct and indirect costs of dementia in Brazil. *PLoS ONE* 2018, 13, e0193209.
 32. Weintraub, S.; Wicklund, A.H.; Salmon, D.P. The neuropsychological profile of Alzheimer disease. *Cold Spring Harb. Perspect. Med.* 2012, 2, a006171.
 33. Aliev, G.; Palacios, H.H.; Walrafen, B.; Lipsitt, A.E.; Obrenovich, M.E.; Morales, L. Brain mitochondria as a primary target in the development of treatment strategies for Alzheimer disease. *Int. J. Biochem. Cell Biol.* 2009, 41, 1989–2004.
 34. Serrano-Pozo, A.; Mielke, M.L.; Gómez-Isla, T.; Betensky, R.A.; Growdon, J.H.; Frosch, M.P.; Hyman, B.T. Reactive glia not only associates with plaques but also parallels tangles in Alzheimer's disease. *Am. J. Pathol.* 2011, 179, 1373–1384.
 35. Villa, V.; Thellung, S.; Bajetto, A.; Gatta, E.; Robello, M.; Novelli, F.; Tasso, B.; Tonelli, M.; Florio, T. Novel celecoxib analogues inhibit glial production of prostaglandin E2, nitric oxide, and oxygen radicals reverting the neuroinflammatory responses induced by misfolded prion protein fragment 90–231 or lipopolysaccharide. *Pharm. Res.* 2016, 113, 500–514.
 36. Masters, C.L.; Selkoe, D.J. Biochemistry of amyloid β -protein and amyloid deposits in Alzheimer disease. *Cold Spring Harb. Perspect. Med.* 2012, 2, a006262.
 37. Nhan, H.S.; Chiang, K.; Koo, E.H. The multifaceted nature of amyloid precursor protein and its proteolytic fragments: Friends and foes. *Acta. Neuropathol.* 2015, 129, 1–19.
 38. Campora, M.; Francesconi, V.; Schenone, S.; Tasso, B.; Tonelli, M. Journey on Naphthoquinone and Anthraquinone Derivatives: New Insights in Alzheimer's Disease. *Pharmaceuticals* 2021, 14, 33.
 39. Kocahan, S.; Doğan, Z. Mechanisms of Alzheimer's Disease Pathogenesis and Prevention: The Brain, Neural Pathology, N-methyl-D-aspartate Receptors, Tau Protein and Other Risk Factors. *Clin. Psychopharmacol. Neurosci.* 2017, 15, 1–8.
 40. Zheng, H.; Fridkin, M.; Youdim, M.B. From antioxidant chelators to site-activated multi-target chelators targeting hypoxia inducing factor, beta-amyloid, acetylcholinesterase and monoamine oxidase A/B. *Mini Rev. Med. Chem.* 2012, 12, 364–370.
 41. Huang, L.; Lu, C.; Sun, Y.; Mao, F.; Luo, Z.; Su, T.; Jiang, H.; Shan, W.; Li, X. Multitarget-directed benzylideneindanone derivatives: Anti- β -amyloid ($A\beta$) aggregation, antioxidant, metal chelation, and monoamine oxidase B (MAO-B) inhibition properties against Alzheimer's disease. *J. Med. Chem.* 2012, 55, 8483–8492.
 42. Youdim, M.B.H.; Amit, T.; Bar-Am, O.; Weinreb, O.; Yogev-Falach, M. Implications of co-morbidity for etiology and treatment of neurodegenerative diseases with multifunctional neuroprotective-neurorescue drugs; ladostigil. *Neurotox. Res.* 2006, 10, 181–192.
 43. Bar-Am, O.; Amit, T.; Weinreb, O.; Youdim, M.B.; Mandel, S. Propargylamine containing compounds as modulators of proteolytic cleavage of amyloid-beta protein precursor: Involvement of MAPK and PKC activation. *J. Alzheimers Dis.* 2010, 21, 361–371.
 44. Yogev-Falach, M.; Bar-Am, O.; Amit, T.; Weinreb, O.; Youdim, M.B. A multifunctional, neuroprotective drug, ladostigil (TV3326), regulates holo-APP translation and processing. *FASEB J.* 2006, 20, 2177–2179.

45. Weinreb, O.; Amit, T.; Bar-Am, O.; Sagi, Y.; Mandel, S.; Youdim, M.B.H. Involvement of multiple survival signal transduction pathways in the neuroprotective, neurorescue and APP processing activity of rasagiline and its propargyl moiety. In *Parkinson's Disease and Related Disorders*, 2nd ed.; Springer: Vienna, Austria, 2006; Volume 70, pp. 457–465.
46. Cai, Z. Monoamine oxidase inhibitors: Promising therapeutic agents for Alzheimer's disease (Review). *Mol. Med. Rep.* 2014, 9, 1533–1541.
47. Manouchehrabadi, M.; Farhadi, M.; Azizi, Z.; Torkaman-Boutorabi, A. Carvacrol Protects Against 6-Hydroxydopamine-Induced Neurotoxicity in In Vivo and In Vitro Models of Parkinson's Disease. *Neurotox Res.* 2020, 37, 156–170.
48. Fahn, S. Description of Parkinson's disease as a clinical syndrome. *Ann. N. Y. Acad. Sci.* 2003, 991, 1–14.
49. Goedert, M. Alpha-synuclein and neurodegenerative diseases. *Nat. Rev. Neurosci.* 2001, 2, 492–501.
50. Jellinger, K.A. The pathology of Parkinson's disease. *Adv. Neurol.* 2001, 86, 55–72.
51. Spillantini, M.G.; Crowther, R.A.; Jakes, R.; Hasegawa, M.; Goedert, M. α -Synuclein in filamentous inclusions of Lewy bodies from Parkinson's disease and dementia with Lewy bodies. *Proc. Natl. Acad. Sci. USA* 1998, 95, 6469–6473.
52. Schulz, J.B. Update on the pathogenesis of Parkinson's disease. *J. Neurol.* 2008, 255, 3–7.
53. Blum, D.; Torch, S.; Lambeng, N.; Nissou, M.; Benabid, A.L.; Sadoul, R.; Verna, J.M. Molecular pathways involved in the neurotoxicity of 6-OHDA, dopamine and MPTP: Contribution to the apoptotic theory in Parkinson's disease. *Prog. Neurobiol.* 2001, 65, 135–172.
54. Wexler, N.S. Venezuelan kindreds reveal that genetic and environmental factors modulate Huntington's disease age of onset. *Proc. Natl. Acad. Sci. USA* 2004, 101, 3498–3503.
55. Quarrell, O.W.; Nance, M.A.; Nopoulos, P.; Paulsen, J.S.; Smith, J.A.; Squitieri, F. Managing juvenile Huntington's disease. *Neurodegener. Dis. Manag.* 2013, 3, 267–276.
56. Lee, J.M.; Ramos, E.M.; Lee, J.H.; Gillis, T.; Mysore, J.S.; Hayden, M.R.; Warby, S.C.; Morrison, P.; Nance, M.; Ross, C.A.; et al. CAG repeat expansion in Huntington disease determines age at onset in a fully dominant fashion. *Neurology* 2012, 78, 690–695.
57. Koutsis, G.; Karadima, G.; Kladi, A.; Panas, M. Late-onset Huntington's disease: Diagnostic and prognostic considerations. *Parkinsonism Relat. D.* 2014, 20, 726–730.
58. Miller, R.G.; Mitchell, J.D.; Lyon, M.; Moore, D.H. Riluzole for amyotrophic lateral sclerosis (ALS)/motor neuron disease (MND). *Cochrane Database Syst. Rev.* 2007, 7, 1–26.
59. Katyal, N.; Govindarajan, R. Shortcomings in the current amyotrophic lateral sclerosis trials and potential solutions for improvement. *Front. Neurol.* 2017, 8, 521.
60. Al-Chalabi, A.; Jones, A.; Troakes, C.; King, A.; Al-Sarraj, S.; van den Berg, L.H. The genetics and neuropathology of amyotrophic lateral sclerosis. *Acta. Neuropathol.* 2012, 124, 339–352.
61. Menke, R.A.; Körner, S.; Filippini, N.; Douaud, G.; Knight, S.; Talbot, K.; Turner, M.R. Widespread grey matter pathology dominates the longitudinal cerebral MRI and clinical landscape of amyotrophic lateral sclerosis. *Brain* 2014, 137, 2546–2555.
62. Hadzhieva, M.; Kirches, E.; Wilisch-Neumann, A.; Pachow, D.; Wallesch, M.; Schoenfeld, P.; Paeye, I.; Vielhaber, S.; Petri, S.; Keilhoff, G. Dysregulation of iron protein expression in the G93A model of amyotrophic lateral sclerosis. *Neuroscience* 2013, 230, 94–101.
63. De Assis Lima, I.V.; Bastos, L.F.S.; Limborço-Filho, M.; Fiebich, B.L.; de Oliveira, A.C.P. Role of prostaglandins in neuroinflammatory and neurodegenerative diseases. *Mediat. Inflamm.* 2012, 2012, 1–13.

64. Bassam, A.-S. Antibacterial activities of some plant extracts utilized in popular medicine in Palestine. *Turk. J. Biol.* 2004, 28, 99–102.
65. Rios, J.-L.; Recio, M. Medicinal plants and antimicrobial activity. *J. Ethnopharmacol.* 2005, 100, 80–84.
66. Donelian, A.; Carlson, L.H.C.; Lopes, T.J.; Machado, R.A.F. Comparison of extraction of patchouli (*Pogostemon cablin*) essential oil with supercritical CO₂ and by steam distillation. *J. Supercrit. Fluids* 2009, 48, 15–20.
67. Irshad, M.; Subhani, M.A.; Ali, S.; Hussain, A. Biological Importance of Essential Oils. In *Essential Oils-Oils of Nature*; El-Shemy, H.A., Ed.; IntechOpen: London, UK, 2020.
68. Natural Living Ideas. The 10 Most Popular Essential Oils & 174 Magical Ways To Use Them. Available online: (accessed on 12 December 2020).
69. Movahhedini, N.; Zengin, G.; Bahadori, M.B.; Sarikurkcu, C.; Bahadori, S.; Dinparast, L. *Ajuga chamaecistus* subsp. *scoparia* (Boiss.) Rech.f.: A new source of phytochemicals for antidiabetic, skin-care, and neuroprotective uses. *Ind. Crop. Prod.* 2016, 94, 89–96.
70. Bahadori, S.; Bahadori, M.B.; Zengin, G.; Maggi, F.; Dinparast, L.; Aktumsek, A. Chemical composition profile of the essential oil from *Hymenocrater bituminosus* and its health functionality. *Int. J. Food Prop.* 2017, 20, S972–S980.
71. Karakaya, S.; Eksi, G.; Koca, M.; Demirci, B.; Kaymak, H.C.; Kaplan, M.E.; Aksakal, O. Chemical and morphological characterization of *Allium tuncelianum* (Amaryllidaceae) and its antioxidant and anticholinesterase potentials. *Jard. Bot. Madr.* 2019, 76, 85.
72. Lamamra, M.; Laouer, H.; Amira, S.; Erdogan Orhan, I.; Senol, F.; Demirci, B.; Akkal, S. Chemical Composition and Cholinesterase Inhibitory Activity of Different Parts of *Daucus aristidis* Coss. *Essential Oils from Two Locations in Algeria.* *Rec. Nat. Prod.* 2017, 11, 147–156.
73. Jyotshna; Srivastava, N.; Singh, B.; Chanda, D.; Shanker, K. Chemical composition and acetylcholinesterase inhibitory activity of *Artemisia maderaspatana* essential oil. *Pharm. Biol.* 2015, 53, 1677–1683.
74. Kohoude, M.J.; Gbaguidi, F.; Agbani, P.; Ayedoun, M.A.; Cazaux, S.; Bouajila, J. Chemical composition and biological activities of extracts and essential oil of *Boswellia dalzielii* leaves. *Pharm. Biol.* 2017, 55, 33–42.
75. Ali-Shtayeh, M.S.; Jamous, R.M.; Abu-Zaitoun, S.Y.; Akkawi, R.J.; Kalbouneh, S.R.; Bernstein, N.; Dudai, N. Chemical profile and bioactive properties of the essential oil isolated from *Clinopodium serpyllifolium* (M.Bieb.) Kuntze growing in Palestine. *Ind. Crop. Prod.* 2018, 124, 617–625.
76. Arruda, M.; Viana, H.; Rainha, N.; Neng, N.R.; Rosa, J.S.; Nogueira, J.M.F.; Barreto, M.d.C. Anti-acetylcholinesterase and antioxidant activity of essential oils from *Hedychium gardnerianum* Sheppard ex Ker-Gawl. *Molecules* 2012, 17, 3082–3092.
77. Murata, K.; Tanaka, K.; Akiyama, R.; Noro, I.; Nishio, A.; Nakagawa, S.; Matsumura, S.; Matsuda, H. Anti-cholinesterase Activity of Crude Drugs Selected from the Ingredients of Incense Sticks and Heartwood of *Chamaecyparis obtusa*. *Nat. Prod. Commun.* 2018, 13.
78. Ademosun, A.O.; Oboh, G.; Olupona, A.J.; Oyeleye, S.I.; Adewuni, T.M.; Nwanna, E.E. Comparative Study of Chemical Composition, In Vitro Inhibition of Cholinergic and Monoaminergic Enzymes, and Antioxidant Potentials of Essential Oil from Peels and Seeds of Sweet Orange (*Citrus Sinensis* [L.] Osbeck) Fruits. *J. Food Biochem.* 2016, 40, 53–60.
79. Tundis, R.; Loizzo, M.R.; Bonesi, M.; Menichini, F.; Mastellone, V.; Colica, C.; Menichini, F. Comparative Study on the Antioxidant Capacity and Cholinesterase Inhibitory Activity of *Citrus aurantifolia* Swingle, *C. aurantium* L., and *C. bergamia* Risso and Poit. Peel Essential Oils. *J. Food Sci.* 2012, 77, H40–H46.

80. Loizzo, M.; Ben Jemia, M.; Senatore, F.; Bruno, M.; Menichini, F.; Tundis, R. Chemistry and functional properties in prevention of neurodegenerative disorders of five *Cistus* species essential oils. *Food Chem. Toxicol* 2013, 59.
81. Sadlon, A.E.; Lamson, D.W. Immune-modifying and antimicrobial effects of Eucalyptus oil and simple inhalation devices. *Altern. Med. Rev.* 2010, 15, 33–47.
82. ESCOP. *Alchemillae Herba*, The Scientific Foundation for Herbal Medicinal Products. Available online: (accessed on 12 December 2020).
83. Juergens, U.R. Anti-inflammatory properties of the monoterpene 1,8-cineole: Current evidence for co-medication in inflammatory airway diseases. *Drug Res.* 2014, 64, 638–646.
84. El Euch, S.K.; Hassine, D.B.; Cazaux, S.; Bouzouita, N.; Bouajila, J. *Salvia officinalis* essential oil: Chemical analysis and evaluation of anti-enzymatic and antioxidant bioactivities. *S. Afr. J. Bot.* 2019, 120, 253–260.
85. Abuhamdah, S.; Abuhamdah, R.; Howes, M.J.; Al-Olimat, S.; Ennaceur, A.; Chazot, P.L. Pharmacological and neuroprotective profile of an essential oil derived from leaves of *Aloysia citrodora* Palau. *J. Pharm. Pharm.* 2015, 67, 1306–1315.
86. CSID:2656. 1,8-cineole. Available online: (accessed on 15 January 2021).
87. CSID:6402. α -pinene. Available online: (accessed on 15 January 2021).
88. CSID:14198. β -pinene. Available online: (accessed on 15 January 2021).
89. CSID:28952. Zingerone. Available online: (accessed on 15 January 2021).
90. CSID:388386. Limonene. Available online: (accessed on 15 January 2021).
91. CSID:390582. Ferruginol. Available online: (accessed on 15 January 2021).
92. CSID:454172. Kaur-16-ene. Available online: (accessed on 15 January 2021).
93. CSID:4474888. Rosmarinic acid. Available online: (accessed on 15 January 2021).
94. CSID:13849981. Linalool. Available online: (accessed on 15 January 2021).
95. CSID:21105867. Carvacrol. Available online: (accessed on 15 January 2021).
96. Cutillas, A.B.; Carrasco, A.; Martinez-Gutierrez, R.; Tomas, V.; Tudela, J. *Thymus mastichina* L. essential oils from Murcia (Spain): Composition and antioxidant, antienzymatic and antimicrobial bioactivities. *PLoS ONE* 2018, 13, e0190790.
97. Montalván, M.; Peñafiel, M.A.; Ramírez, J.; Cumbicus, N.; Bec, N.; Larroque, C.; Bicchi, C.; Gilardoni, G. Chemical Composition, Enantiomeric Distribution, and Sensory Evaluation of the Essential Oils Distilled from the Ecuadorian Species *Myrcianthes myrsinoides* (Kunth) Grifo and *Myrcia mollis* (Kunth) DC. (Myrtaceae). *Plants* 2019, 8, 511.
98. Dhanasekaran, S.; Perumal, P.; Palayan, M. In-vitro Screening for acetylcholinesterase enzyme inhibition potential and antioxidant activity of extracts of *Ipomoea aquatica* Forsk: Therapeutic lead for Alzheimer's disease. *J. Appl. Pharm. Sci.* 2015, 5, 012–016.
99. Silver, A. *The Biology of Cholinesterases*, North-Holland Publishing Company: Amsterdam, The Netherlands, 1974; 1–596.
100. Wilson, B.; Philip, W. Cholinesterase inhibition. In *Encyclopedia of Toxicology*; Elsevier: New York, NY, USA, 2005; pp. 588–599.
101. Wilson, B.W. CHAPTER 48-Cholinesterases. In *Handbook of Pesticide Toxicology*, 2nd ed.; Krieger, R.I., Krieger, W.C., Eds.; Academic Press: San Diego, CA, USA, 2001; pp. 967–985.

102. Chatonnet, A.; Lockridge, O. Comparison of butyrylcholinesterase and acetylcholinesterase. *Biochem. J.* 1989, 260, 625–634.
103. Schwarz, M.; Glick, D.; Loewenstein, Y.; Soreq, H. Engineering of human cholinesterases explains and predicts diverse consequences of administration of various drugs and poisons. *Pharm. Ther.* 1995, 67, 283–322.
104. Tecles, F.; Ceron, J. Determination of whole blood cholinesterase in different animal species using specific substrates. *Res. Vet. Sci.* 2001, 70, 233–238.
105. Sramek, J.J.; Cutler, N.R. RBC cholinesterase inhibition: A useful surrogate marker for cholinesterase inhibitor activity in Alzheimer disease therapy? *Alzheimer Dis. Assoc. Disord* 2000, 14, 216–227.
106. Nicolet, Y.; Lockridge, O.; Masson, P.; Fontecilla-Camps, J.C.; Nachon, F. Crystal structure of human butyrylcholinesterase and of its complexes with substrate and products. *J. Biol. Chem.* 2003, 278, 41141–41147.
107. Saxena, A.; Redman, A.M.; Jiang, X.; Lockridge, O.; Doctor, B.P. Differences in active site gorge dimensions of cholinesterases revealed by binding of inhibitors to human butyrylcholinesterase. *Biochemistry* 1997, 36, 14642–14651.
108. Xu, Y.; Colletier, J.-P.; Weik, M.; Jiang, H.; Moul, J.; Silman, I.; Sussman, J.L. Flexibility of aromatic residues in the active-site gorge of acetylcholinesterase: X-ray versus molecular dynamics. *Biophys. J.* 2008, 95, 2500–2511.
109. Dighe, S.N.; Deora, G.S.; de la Mora, E.; Nachon, F.; Chan, S.; Parat, M.-O.; Brazzolotto, X.; Ross, B.P. Discovery and Structure–Activity Relationships of a Highly Selective Butyrylcholinesterase Inhibitor by Structure-Based Virtual Screening. *J. Med. Chem.* 2016, 59, 7683–7689.
110. Zhou, Y.; Lu, X.; Du, C.; Liu, Y.; Wang, Y.; Hong, K.H.; Chen, Y.; Sun, H. Novel BuChE-IDO1 inhibitors from sertaconazole: Virtual screening, chemical optimization and molecular modeling studies. *Bioorg. Med. Chem. Lett.* 2021, 34, 127756.
111. Miles, J.A.; Kapure, J.S.; Deora, G.S.; Courageux, C.; Igert, A.; Dias, J.; McGear, R.P.; Brazzolotto, X.; Ross, B.P. Rapid discovery of a selective butyrylcholinesterase inhibitor using structure-based virtual screening. *Bioorg. Med. Chem. Lett.* 2020, 30, 127609.
112. Ellman, G.L.; Courtney, K.D.; Andres, V., Jr.; Featherstone, R.M. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharm.* 1961, 7, 88–95.
113. Pohanka, M.; Jun, D.; Kuca, K. Improvement of acetylcholinesterase-based assay for organophosphates in way of identification by reactivators. *Talanta* 2008, 77, 451–454.
114. Morizono, M.; Akinaga, Y. Studies on Tissue Cholinesterase in Domestic Animals II. Detection of Tissue Cholinesterase Isoenzymes. *Mem. Fac. Agr. Kagoshima. Univ.* 1981, 17, 219–234.
115. Frasco, M.; Fournier, D.; Carvalho, F.; Guilhermino, L. Do metals inhibit acetylcholinesterase (AChE)? Implementation of assay conditions for the use of AChE activity as a biomarker of metal toxicity. *Biomarkers* 2005, 10, 360–375.
116. Šinko, G.; Čalić, M.; Bosak, A.; Kovarik, Z. Limitation of the Ellman method: Cholinesterase activity measurement in the presence of oximes. *Anal. Biochem.* 2007, 370, 223–227.
117. Marrs, T.C. Organophosphates: History, chemistry, pharmacology. In *Organophosphates and Health*; Imperial College Press: London, UK, 2001; pp. 1–36.
118. Michel, H.O. An electrometric method for the determination of red blood cell and plasma cholinesterase activity. *J. Lab. Clin. Med.* 1949, 34, 1564–1568.
119. Selkoe, D.J. Alzheimer's disease: Genes, proteins, and therapy. *Physiol. Rev.* 2001, 81, 742–760.
120. Rochette, M.; Murphy, M. Gamma-secretase: Substrates and inhibitors. *Mol. Neurobiol.* 2002, 26, 81–95.

121. Lee, V.M.; Goedert, M.; Trojanowski, J.Q. Neurodegenerative tauopathies. *Annu Rev. Neurosci.* 2001, 24, 1121–1159.
122. Satou, T.; Cummings, B.J.; Head, E.; Nielson, K.A.; Hahn, F.F.; Milgram, N.W.; Velazquez, P.; Cribbs, D.H.; Tenner, A.J.; Cotman, C.W. The progression of β -amyloid deposition in the frontal cortex of the aged canine. *Brain Res.* 1997, 774, 35–43.
123. Cummings, B.J.; Head, E.; Afagh, A.J.; Milgram, N.W.; Cotman, C.W. β -amyloid accumulation correlates with cognitive dysfunction in the aged canine. *Neurobiol. Learn. Mem.* 1996, 66, 11–23.
124. De Felice, F.G.; Munoz, D.P. Opportunities and challenges in developing relevant animal models for Alzheimer's disease. *Ageing Res. Rev.* 2016, 26, 112–114.
125. Casadesus, G. *Handbook of Animal Models in Alzheimer's Disease*, 1st ed.; IOS Press: Amsterdam, The Netherlands, 2011; Volume 1, p. 340.
126. Lecanu, L.; Papadopoulos, V. Modeling Alzheimer's disease with non-transgenic rat models. *Alzheimer's Res.* 2013, 5, 17.
127. Temel, H.E.; Demirci, B.; Demirci, F.; Celep, F.; Kahraman, A.; Doğan, M.; Hüsnü Can Başer, K. Chemical characterization and anticholinesterase effects of essential oils derived from *Salvia* species. *J. Essent Oil Res.* 2016, 28, 322–331.
128. Demirezer, L.; Gürbüz, P.; Kelicen Uğur, E.P.; Bodur, M.; Özenver, N.; Uz, A.; Güvenalp, Z. Molecular docking and ex vivo and in vitro anticholinesterase activity studies of *Salvia* sp. and highlighted rosmarinic acid. *Turk. J. Med. Sci.* 2015, 45, 1141–1148.
129. Videira, R.; Castanheira, P.; Grãos, M.; Salgueiro, L.; Faro, C.; Cavaleiro, C. A necrodane monoterpene from *Lavandula luisieri* essential oil as a cell-permeable inhibitor of BACE-1, the β -secretase in Alzheimer's disease. *Flavour Frag J.* 2013, 28, 380–388.
130. Baser, K.H. Biological and pharmacological activities of carvacrol and carvacrol bearing essential oils. *Curr Pharm. Des.* 2008, 14, 3106–3119.
131. Kaufmann, D.; Dogra, A.; Wink, M. Myrtenal inhibits acetylcholinesterase, a known Alzheimer target. *J. Pharm. Pharm.* 2011, 63, 1368–1371.
132. Jukic, M.; Politeo, O.; Maksimovic, M.; Milos, M.; Milos, M. In vitro acetylcholinesterase inhibitory properties of thymol, carvacrol and their derivatives thymoquinone and thymohydroquinone. *Phytother. Res.* 2007, 21, 259–261.
133. Melo, F.H.; Venâncio, E.T.; de Sousa, D.P.; de França Fonteles, M.M.; de Vasconcelos, S.M.; Viana, G.S.; de Sousa, F.C. Anxiolytic-like effect of Carvacrol (5-isopropyl-2-methylphenol) in mice: Involvement with GABAergic transmission. *Fundam. Clin. Pharm.* 2010, 24, 437–443.
134. Melo, F.H.; Moura, B.A.; de Sousa, D.P.; de Vasconcelos, S.M.; Macedo, D.S.; Fonteles, M.M.; Viana, G.S.; de Sousa, F.C. Antidepressant-like effect of carvacrol (5-Isopropyl-2-methylphenol) in mice: Involvement of dopaminergic system. *Fundam. Clin. Pharm.* 2011, 25, 362–367.
135. Krishtal, J.; Bragina, O.; Metsla, K.; Palumaa, P.; Tõugu, V. In situ fibrillizing amyloid-beta 1-42 induces neurite degeneration and apoptosis of differentiated SH-SY5Y cells. *PLoS ONE* 2017, 12, e0186636.
136. Xu, W.; Yang, L.; Li, J. Protection against β -amyloid-induced neurotoxicity by naturally occurring Z-ligustilide through the concurrent regulation of p38 and PI3-K/Akt pathways. *Neurochem. Int.* 2016, 100, 44–51.
137. Pike, C.J.; Walencewicz-Wasserman, A.J.; Kosmoski, J.; Cribbs, D.H.; Glabe, C.G.; Cotman, C.W. Structure-activity analyses of β -amyloid peptides: Contributions of the β 25–35 region to aggregation and neurotoxicity. *J. Neurochem.* 1995, 64, 253–265.

138. Kaminsky, Y.G.; Marlatt, M.W.; Smith, M.A.; Kosenko, E.A. Subcellular and metabolic examination of amyloid- β peptides in Alzheimer disease pathogenesis: Evidence for A β 25–35. *Exp. Neurol.* 2010, 221, 26–37.
139. Gabbouj, S.; Ryhänen, S.; Marttinen, M.; Wittrahm, R.; Takalo, M.; Kemppainen, S.; Martiskainen, H.; Tanila, H.; Haapasalo, A.; Hiltunen, M.; et al. Altered Insulin Signaling in Alzheimer's Disease Brain—Special Emphasis on PI3K-Akt Pathway. *Front. Neurosci.* 2019, 13.
140. Wu, Y.; Han, X.; Yuan, W.; Wang, X.; Meng, D.; Hu, J.; Lv, Z. Salt intervention for the diversities of essential oil composition, aroma and antioxidant activities of Kushui rose (*R. setata* × *R. rugosa*). *Ind. Crop. Prod.* 2020, 150, 112417.
141. Hsu, H.; Hsu, C. Commonly used Chinese herb formulas with illustrations, 2nd ed.; Oriental Healing Arts Institute: Long Beach, CA, USA, 1980; Volume 1, p. 671.
142. Wang, M.C.; Bohmann, D.; Jasper, H. JNK signaling confers tolerance to oxidative stress and extends lifespan in *Drosophila*. *Dev. Cell* 2003, 5, 811–816.
143. Wang, M.C.; Bohmann, D.; Jasper, H. JNK extends life span and limits growth by antagonizing cellular and organism-wide responses to insulin signaling. *Cell* 2005, 121, 115–125.
144. Gulactı, T.; Ufuk, K.; Mehmet, O.; Mehmet, B.; Seda Damla, H.; Fatemeh, B.; Burcu, C.; Tuncay, D. Investigation of Anticholinesterase Activity of a Series of *Salvia* Extracts and the Constituents of *Salvia staminea*. *Nat. Prod. J.* 2013, 3, 3–9.
145. Liu, B.; Kou, J.; Li, F.; Huo, D.; Xu, J.; Zhou, X.; Meng, D.; Ghulam, M.; Artyom, B.; Gao, X.; et al. Lemon essential oil ameliorates age-associated cognitive dysfunction via modulating hippocampal synaptic density and inhibiting acetylcholinesterase. *Aging* 2020, 12, 8622–8639.
146. Chen, J.; Montanari, A.M.; Widmer, W.W. Two New Polymethoxylated Flavones, a Class of Compounds with Potential Anticancer Activity, Isolated from Cold Pressed Dancy Tangerine Peel Oil Solids. *J. Agric. Food Chem.* 1997, 45, 364–368.
147. Nogata, Y.; Sakamoto, K.; Shiratsuchi, H.; Ishii, T.; Yano, M.; Ohta, H. Flavonoid Composition of Fruit Tissues of Citrus Species. *Biosci. Biotechnol. Biochem.* 2006, 70, 178–192.
148. Ayuob, N.N.; El Wahab, M.G.A.; Ali, S.S.; Abdel-Tawab, H.S. *Ocimum basilicum* improve chronic stress-induced neurodegenerative changes in mice hippocampus. *Metab. Brain Dis.* 2018, 33, 795–804.
149. Avetisyan, A.; Markosian, A.; Petrosyan, M.; Sahakyan, N.; Babayan, A.; Aloyan, S.; Trchounian, A. Chemical composition and some biological activities of the essential oils from basil *Ocimum* different cultivars. *BMC Complement. Altern. Med.* 2017, 17, 60.
150. Batista, P.A.; Werner, M.F.; Oliveira, E.C.; Burgos, L.; Pereira, P.; Brum, L.F.; Story, G.M.; Santos, A.R. The antinociceptive effect of (-)-linalool in models of chronic inflammatory and neuropathic hypersensitivity in mice. *J. Pain* 2010, 11, 1222–1229.
151. Elisabetsky, E.; Marschner, J.; Souza, D.O. Effects of Linalool on glutamatergic system in the rat cerebral cortex. *Neurochem. Res.* 1995, 20, 461–465.
152. Letizia, C.S.; Cocchiara, J.; Lalko, J.; Api, A.M. Fragrance material review on linalool. *Food Chem. Toxicol.* 2003, 41, 943–964.
153. Mitić-Culafić, D.; Zegura, B.; Nikolić, B.; Vuković-Gaćić, B.; Knezević-Vukcević, J.; Filipic, M. Protective effect of linalool, myrcene and eucalyptol against t-butyl hydroperoxide induced genotoxicity in bacteria and cultured human cells. *Food Chem. Toxicol.* 2009, 47, 260–266.
154. Sabogal-Guáqueta, A.M.; Osorio, E.; Cardona-Gómez, G.P. Linalool reverses neuropathological and behavioral impairments in old triple transgenic Alzheimer's mice. *Neuropharmacology* 2016, 102, 111–120.

155. Gradinariu, V.; Cioanca, O.; Hritcu, L.; Trifan, A.; Gille, E.; Hancianu, M. Comparative efficacy of *Ocimum sanctum* L. and *Ocimum basilicum* L. essential oils against amyloid beta (1–42)-induced anxiety and depression in laboratory rats. *PhytoChem. Rev.* 2014, 14, 1–9.
156. Jankovic, J.; Aguilar, L.G. Current approaches to the treatment of Parkinson's disease. *Neuropsych. Dis. Treat.* 2008, 4, 743–757.
157. Beserra-Filho, J.I.; de Macêdo, A.M.; Leão, A.H.; Bispo, J.M.M.; Santos, J.R.; de Oliveira-Melo, A.J.; Menezes, P.D.P.; Duarte, M.C.; de Souza Araújo, A.A.; Silva, R.H. Eplingiella fruticosa leaf essential oil complexed with β -cyclodextrin produces a superior neuroprotective and behavioral profile in a mice model of Parkinson's disease. *Food Chem. Toxicol.* 2019, 124, 17–29.
158. Ramazani, E.; YazdFazeli, M.; Emami, S.A.; Mohtashami, L.; Javadi, B.; Asili, J.; Tayarani-Najaran, Z. Protective effects of *Cinnamomum verum*, *Cinnamomum cassia* and cinnamaldehyde against 6-OHDA-induced apoptosis in PC12 cells. *Mol. Biol. Rep.* 2020, 7, 2437–2445.
159. Rao, P.V.; Gan, S.H. Cinnamon: A multifaceted medicinal plant. *Evid-Based Complement. Altern. Med.* 2014, 2014, 1–2.
160. Brahmachari, S.; Jana, A.; Pahan, K. Sodium benzoate, a metabolite of cinnamon and a food additive, reduces microglial and astroglial inflammatory responses. *J. Immunol.* 2009, 183, 5917–5927.
161. Mollazadeh, H.; Hosseinzadeh, H. Cinnamon effects on metabolic syndrome: A review based on its mechanisms. *Iran. J. Basic Med. Sci.* 2016, 19, 1258.
162. Liu, X.C.; Cheng, J.; Zhao, N.N.; Liu, Z.L. Insecticidal activity of essential oil of *Cinnamomum cassia* and its main constituent, trans-cinnamaldehyde, against the booklice, *Liposcelis bostrychophila*. *Trop. J. Pharm. Res.* 2014, 13, 1697–1702.
163. Nabavi, S.F.; di Lorenzo, A.; Izadi, M.; Sobarzo-Sánchez, E.; Daglia, M.; Nabavi, S.M. Antibacterial effects of cinnamon: From farm to food, cosmetic and pharmaceutical industries. *Nutrients* 2015, 7, 7729–7748.
164. Klein-Júnior, L.C.; dos Santos Passos, C.; Tasso de Souza, T.J.; Gobbi de Bitencourt, F.; Salton, J.; de Loreto Bordignon, S.A.; Henriques, A.T. The monoamine oxidase inhibitory activity of essential oils obtained from *Eryngium* species and their chemical composition. *Pharm. Biol.* 2016, 54, 1071–1076.
165. Youdim, M.B.; Bakhle, Y. Monoamine oxidase: Isoforms and inhibitors in Parkinson's disease and depressive illness. *Br. J. Pharm.* 2006, 147, S287–S296.
166. Youdim, M.B.; Edmondson, D.; Tipton, K.F. The therapeutic potential of monoamine oxidase inhibitors. *Nat. Rev. Neurosci.* 2006, 7, 295–309.
167. Morshedi, D.; Nasouti, M. Essential Oils May Lead α -Synuclein towards Toxic Fibrils Formation. *Parkinsons Dis.* 2016, 2016, 6219249.
168. McCormack, A.L.; Mak, S.K.; Shenasa, M.; Langston, W.J.; Forno, L.S.; di Monte, D.A. Pathologic modifications of α -synuclein in 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MPTP)-treated squirrel monkeys. *J. Neuropath Exp. Neur.* 2008, 67, 793–802.
169. Ahmad, A.; Burns, C.S.; Fink, A.L.; Uversky, V.N. Peculiarities of copper binding to α -synuclein. *J. Biomol. Struct. Dyn.* 2012, 29, 825–842.
170. Mohammad-Beigi, H.; Shojaosadati, S.A.; Marvian, A.T.; Pedersen, J.N.; Klausen, L.H.; Christiansen, G.; Pedersen, J.S.; Dong, M.; Morshedi, D.; Otzen, D.E. Strong interactions with polyethylenimine-coated human serum albumin nanoparticles (PEI-HSA NPs) alter α -synuclein conformation and aggregation kinetics. *Nanoscale* 2015, 7, 19627–19640.
171. Silva, B.A.; Breydo, L.; Fink, A.L.; Uversky, V.N. Agrochemicals, α -synuclein, and Parkinson's disease. *Mol. Neurobiol.* 2013, 47, 598–612.

172. Morshedi, D.; Aliakbari, F.; Tayaranian-Marvian, A.; Fassihi, A.; Pan-Montojo, F.; Pérez-Sánchez, H. Cuminaldehyde as the major component of *Cuminum cyminum*, a natural aldehyde with inhibitory effect on alpha-synuclein fibrillation and cytotoxicity. *J. Food Sci.* 2015, 80, H2336–H2345.
173. Ning, B.; Deng, M.; Zhang, Q.; Wang, N.; Fang, Y. β -Asarone inhibits IRE1/XBP1 endoplasmic reticulum stress pathway in 6-OHDA-induced parkinsonian rats. *Neurochem. Res.* 2016, 41, 2097–2101.
174. Ning, B.; Zhang, Q.; Wang, N.; Deng, M.; Fang, Y. β -Asarone regulates ER stress and autophagy via inhibition of the PERK/CHOP/Bcl-2/Beclin-1 Pathway in 6-OHDA-induced parkinsonian rats. *Neurochem. Res.* 2019, 44, 1159–1166.
175. Schröder, M.; Kaufman, R.J. ER stress and the unfolded protein response. *Mutat Res./Fund Mol. M.* 2005, 569, 29–63.
176. Ryu, E.J.; Harding, H.P.; Angelastro, J.M.; Vitolo, O.V.; Ron, D.; Greene, L.A. Endoplasmic reticulum stress and the unfolded protein response in cellular models of Parkinson's disease. *J. Neurosci.* 2002, 22, 10690–10698.
177. Hoozemans, J.; van Haastert, E.; Eikelenboom, P.; de Vos, R.; Rozemuller, J.; Scheper, W. Activation of the unfolded protein response in Parkinson's disease. *Biochem. Biophys. Res. Commun.* 2007, 354, 707–711.
178. Jiang, P.; Gan, M.; Ebrahim, A.S.; Lin, W.-L.; Melrose, H.L.; Yen, S.-H.C. ER stress response plays an important role in aggregation of α -synuclein. *Mol. Neurodegener.* 2010, 5, 1–15.
179. Gardner, B.M.; Pincus, D.; Gotthardt, K.; Gallagher, C.M.; Walter, P. Endoplasmic reticulum stress sensing in the unfolded protein response. *Cold Spring Harb Perspect. Biol.* 2013, 5, a013169.
180. Hotamisligil, G.S. Endoplasmic reticulum stress and atherosclerosis. *Nat. Med.* 2010, 16, 396–399.
181. Hotamisligil, G.S. Endoplasmic reticulum stress and the inflammatory basis of metabolic disease. *Cell* 2010, 140, 900–917.
182. Todd, D.J.; Lee, A.-H.; Glimcher, L.H. The endoplasmic reticulum stress response in immunity and autoimmunity. *Nat. Rev. Immunol.* 2008, 8, 663–674.
183. Senft, D.; Ze'ev, A.R. UPR, autophagy, and mitochondria crosstalk underlies the ER stress response. *Trends Biochem. Sci.* 2015, 40, 141–148.
184. Yorimitsu, T.; Nair, U.; Yang, Z.; Klionsky, D.J. Endoplasmic reticulum stress triggers autophagy. *J. Biol. Chem.* 2006, 281, 30299–30304.
185. Wei, Y.; Pattingre, S.; Sinha, S.; Bassik, M.; Levine, B. JNK1-mediated phosphorylation of Bcl-2 regulates starvation-induced autophagy. *Mol. Cell* 2008, 30, 678–688.
186. Kabuto, H.; Nishizawa, M.; Tada, M.; Higashio, C.; Shishibori, T.; Kohno, M. Zingerone [4-(4-hydroxy-3-methoxyphenyl)-2-butanone] prevents 6-hydroxydopamine-induced dopamine depression in mouse striatum and increases superoxide scavenging activity in serum. *Neurochem. Res.* 2005, 30, 325–332.
187. Kabuto, H.; Tada, M.; Kohno, M. Eugenol [2-methoxy-4-(2-propenyl) phenol] prevents 6-hydroxydopamine-induced dopamine depression and lipid peroxidation inductivity in mouse striatum. *Biol. Pharm. Bull.* 2007, 30, 423–427.
188. Siqueira-Lima, P.S.; Brito, R.G.; Araújo-Filho, H.G.; Santos, P.L.; Lucchesi, A.; Araújo, A.A.; Menezes, P.P.; Scotti, L.; Scotti, M.T.; Menezes, I.R. Anti-hyperalgesic effect of *Lippia grata* leaf essential oil complexed with β -cyclodextrin in a chronic musculoskeletal pain animal model: Complemented with a molecular docking and antioxidant screening. *Biomed. Pharm.* 2017, 91, 739–747.
189. Oliveira, M.A.; Guimarães, A.G.; Araújo, A.A.; Quintans-Júnior, L.J.; Quintans, J.S. New drugs or alternative therapy to blurring the symptoms of fibromyalgia—a patent review. *Expert Opin. Pat.* 2017, 27, 1147–1157.
190. Araújo-Filho, H.G.; Pereira, E.W.; Rezende, M.M.; Menezes, P.P.; Araújo, A.A.; Barreto, R.S.; Martins, A.O.; Albuquerque, T.R.; Silva, B.A.; Alcantara, I.S. D-limonene exhibits superior antihyperalgesic effects in a β -

- cyclodextrin-complexed form in chronic musculoskeletal pain reducing Fos protein expression on spinal cord in mice. *Neuroscience* 2017, 358, 158–169.
191. Santos, P.L.; Brito, R.G.; Quintans, J.S.S.; Araujo, A.A.S.; Menezes, I.R.A.; Brogden, N.K.; Quintans-Junior, L.J. Cyclodextrins as complexation agents to improve the anti-inflammatory drugs profile: A systematic review and meta-analysis. In *Curr. Pharm. Des.*; 2017; Volume 23, pp. 2096–2107.
 192. Brito, R.G.; Araujo, A.A.; Quintans, J.S.; Sluka, K.A.; Quintans-Junior, L.J. Enhanced analgesic activity by cyclodextrins—a systematic review and meta-analysis. *Expert Opin Drug Del.* 2015, 12, 1677–1688.
 193. Andrade, A.M.; Oliveira, J.P.; Santos, A.L.; Franco, C.R.; Antonioli, Â.R.; Estevam, C.S.; Thomazzi, S.M. Preliminary study on the anti-inflammatory and antioxidant activities of the leave extract of *Hyptis fruticosa* Salzm. ex Benth., Lamiaceae. *Rev. Bras. Farm.* 2010, 20, 962–968.
 194. De Lima, A.C.B.; Paixão, M.S.; Melo, M.; de Santana, M.T.; Damascena, N.P.; Dias, A.S.; Porto, Y.C.B.S.; Fernandes, X.A.; Santos, C.C.S.; Lima, C.A.; et al. Orofacial antinociceptive effect and antioxidant properties of the hydroethanol extract of *Hyptis fruticosa* salmz ex Benth. *J. Ethnopharmacol.* 2013, 146, 192–197.
 195. Nemati, M.; Hemmati, A.A.; Najafzadeh, H.; Mansouri, M.T.; Khodayar, M.J. Evaluation of the Effects of *Foeniculum vulgare* Essence on Behavioral-Motor Disorders of Parkinson's Disease induced by Reserpine in Ovariectomized and Non Ovariectomized Rats. *Jundishapur J. Nat. Pharm. Prod.* 2018, 13, e67391.
 196. Tadaiesky, M.T.; Andreatini, R.; Vital, M.A. Different effects of 7-nitroindazole in reserpine-induced hypolocomotion in two strains of mice. *Eur. J. Pharm.* 2006, 535, 199–207.
 197. Ben, J.; Soares, F.M.; Scherer, E.B.; Cechetti, F.; Netto, C.A.; Wyse, A.T. Running exercise effects on spatial and avoidance tasks in ovariectomized rats. *Neurobiol. Learn. Mem.* 2010, 94, 312–317.
 198. Scott, E.L.; Zhang, Q.G.; Han, D.; Desai, B.N.; Brann, D.W. Long-term estrogen deprivation leads to elevation of Dickkopf-1 and dysregulation of Wnt/ β -Catenin signaling in hippocampal CA1 neurons. *Steroids* 2013, 78, 624–632.
 199. Sarkaki, A.; Badavi, M.; Hoseiny, N.; Gharibnaseri, M.K.; Rahim, F. Postmenopausal effects of intrastriatal estrogen on catalepsy and pallidal electroencephalogram in an animal model of Parkinson's disease. *Neuroscience* 2008, 154, 940–945.
 200. Namavar Jahromi, B.; Tartifzadeh, A.; Khabnadideh, S. Comparison of fennel and mefenamic acid for the treatment of primary dysmenorrhea. *Int. J. Gynaecol. Obs.* 2003, 80, 153–157.
 201. Modaress Nejad, V.; Asadipour, M. Comparison of the effectiveness of fennel and mefenamic acid on pain intensity in dysmenorrhoea. *East. Mediterr. Health J.* 2006, 12, 423–427.
 202. Betarbet, R.; Sherer, T.B.; MacKenzie, G.; Garcia-Osuna, M.; Panov, A.V.; Greenamyre, J.T. Chronic systemic pesticide exposure reproduces features of Parkinson's disease. *Nat. Neurosci.* 2000, 3, 1301–1306.
 203. Talpade, D.J.; Greene, J.G.; Higgins, D.S., Jr.; Greenamyre, J.T. In vivo labeling of mitochondrial complex I (NADH:ubiquinone oxidoreductase) in rat brain using [3 H]dihydrorotenone. *J. Neurochem.* 2000, 75, 2611–2621.
 204. Sherer, T.B.; Richardson, J.R.; Testa, C.M.; Seo, B.B.; Panov, A.V.; Yagi, T.; Matsuno-Yagi, A.; Miller, G.W.; Greenamyre, J.T. Mechanism of toxicity of pesticides acting at complex I: Relevance to environmental etiologies of Parkinson's disease. *J. Neurochem.* 2007, 100, 1469–1479.
 205. Issa, M.Y.; Ezzat, M.I.; Sayed, R.H.; Elbaz, E.M.; Omar, F.A.; Mohsen, E. Neuroprotective effects of *Pulicaria undulata* essential oil in rotenone model of parkinson's disease in rats: Insights into its anti-inflammatory and anti-oxidant effects. *S. Afr. J. Bot.* 2020, 132, 289–298.

206. Liu, L.-L.; Yang, J.-L.; Shi, Y.-P. Phytochemicals and biological activities of *Pulicaria* species. *Chem. Biodivers.* 2010, 7, 327–349.
207. Liu, Q.; Sang Heon, K.; Yung-Wei, S.; Sok Cheon, P.; Wonwoong, L.; Jongki, H.; Jaehwan, J.; Kyoung Sang, C.; Songhee, J.; Byung-Soo, K. Neuroprotective effects of Suhexiang Wan on the in vitro and in vivo models of Parkinson's disease. *J. Tradit. Chin. Med.* 2019, 39, 800–808.
208. Lee, K.S.; Lee, J.K.; Kim, H.G.; Kim, H.R. Differential Effects of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine on Motor Behavior and Dopamine Levels at Brain Regions in Three Different Mouse Strains. *Korean J. Physiol. Pharm.* 2013, 17, 89–97.
209. Nikolova, G.; Karamalakova, Y.; Kovacheva, N.; Stanev, S.; Zheleva, A.; Gadjeva, V. Protective effect of two essential oils isolated from *Rosa damascena* Mill. and *Lavandula angustifolia* Mill, and two classic antioxidants against L-dopa oxidative toxicity induced in healthy mice. *Regul. Toxicol. Pharm.* 2016, 81, 1–7.
210. Nikolova, G.; Karamalakova, Y.; Gadjeva, V. Reducing oxidative toxicity of L-dopa in combination with two different antioxidants: An essential oil isolated from *Rosa Damascena* Mill., and vitamin C. *Toxicol. Rep.* 2019, 6, 267–271.
-

Retrieved from <https://encyclopedia.pub/entry/history/show/19745>