

Food-Borne Chemical Carcinogens

Subjects: Toxicology

Contributor: Tetyana Kobets, Benjamin P. C. Smith, Gary M. Williams

Food-borne carcinogens span a range of chemical classes and can arise from natural or anthropogenic sources, as well as form endogenously.

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1. Introduction

Foods and beverages are essentially complex mixtures of chemicals consumed for either sustenance or pleasure. The diversity of chemicals found in food is vast, as are their varying properties. It has long been known that chemicals with carcinogenic activity in rodent models can be found in many commonly consumed foods ^{[1][2][3][4][5]} from a variety of sources including plants, microorganisms, contaminations, additive uses and reactions which occur during storage, processing and cooking ^[2] (**Table 1**). In addition, carcinogens can be formed endogenously, from food materials ^{[6][7][8]}.

Table 1. Sources of detectable carcinogens in food.

Source	Examples ^a	
1. Naturally occurring		
Plant:	alkenylbenzene derivatives aristolochic acid cycasin ptaquiloside d-limonene	psoralen pyrrolizidine alkaloids pulegone β-myrcene
Microbial/Fungal:	various mycotoxins	
2. Contaminants		
Introduced before processing:	daminozide dioxins	DDT flumequine
Introduced during processing:	trichloroethylene	methylene chloride
Food contact materials:	plastics (polyolefins, polyesters, polystyrene, polyamides, etc.) polymeric coatings	monomers (vinyl chloride, styrene, acrylonitrile)
3. Additives		
Anthropogenic:	α,β-aldehydes butylated hydroxyanisole and butylated hydroxytoluene	hexenal saccharin
4. Formed from food components		
During processing:	acrylamide chloropropanols ethyl carbamate (urethane)	furan various nitrosamines alkylated imidazoles
During packaging:	bisphenol A furan	phthalates
During storage:	benzene	
During cooking:	acrylamide benzo[a]pyrene	various heterocyclic amines

Source	Examples ^a	
In the body:	nitrosamines and nitrosamides α,β -aldehydes	ethylene oxide

^a Many of the agents listed are detectable only at minute levels by highly sensitive analytical techniques.

2. Mechanisms of Carcinogenicity of DNA-Reactive Carcinogens

DNA-reactive carcinogens have structures that permit formation of electrophilic reactants that covalently bind (adduct) to nucleophilic sites in nuclear DNA, as well as in other macromolecules, including RNA and proteins, in the target tissue(s) of carcinogenicity [9][10][11]. In target tissue(s), a single DNA reactant can form different DNA adducts on various nucleophilic sites either on a single base or on different bases. Each adduct can undergo different rates of repair depending upon its location in the genome. For example, adducts in transcriptionally active regions are repaired by a transcription-coupled repair system whereas adducts in transcriptionally silent regions are repaired by a global repair system [12]. The levels of DNA adducts resulting from exposures are a function of several metrics including dose levels, the frequency of exposure, and rates of DNA repair for specific adducts. Each adduct has a characteristic efficiency with which it gives rise to mutations, with those at sites of base pairing being more mutagenic.

Pro-mutagenic DNA alterations are converted to mutations during cell replication [13][14][15]. Mutations in critical growth control genes lead to neoplastic conversion, and subsequent neoplastic development [14][16]. DNA-reactive carcinogens can also exert other cellular effects, such as cytotoxicity, leading to enhanced cell proliferation, which can contribute to their carcinogenic activity [17][18]. DNA-reactive carcinogens can have additive effects with one another in their target organ(s).

Some DNA adducts evidently do not lead to carcinogenicity, since some adducts can be found in tissues where no tumors are induced following administration of a carcinogen [19][20][21][22]. For example, acrylamide, which is discussed below, forms adducts in target and non-target tissues [23]. It could also be the case that epigenetic effects are required to enable neoplastic conversion resulting from some adducts [24][25].

As a result of DNA interactions, DNA-reactive carcinogens are typically genotoxic in assay systems in which appropriate bioactivation is represented [10][26][27][28][29]. Moreover, DNA-reactive carcinogens often produce tumors at multiple sites and with a short duration of exposure, even after administration of a single dose for some. This property underlies their activity in limited short-term bioassays [27].

Some DNA-reactive carcinogens have been demonstrated to exhibit no-observed-adverse-effect-levels (NOAELs) for carcinogenic effects in animal models [11][17][30][31][32][33][34], although conflicting data have been reported. Based on the steps for tumorigenesis, it is evident that biological thresholds that may influence the likelihood of cancer progression for genotoxic carcinogens exist. Nevertheless, currently, thresholds are not generally accepted for DNA-reactive carcinogens from a risk assessment and management perspective [35]. It is acknowledged that the derivation of NOAELs can be dependent on the study design, and more research is needed in this space. It is outside the scope of this research to discuss thresholds for carcinogens in detail; however, this topic is reviewed elsewhere [11][17][30][31][32][33][34].

3. Mechanisms of Carcinogenicity of Epigenetic Carcinogens

Epigenetic carcinogens do not chemically react with DNA [26][36][37][38][39][40][41]. In the target tissue(s) of carcinogenicity, MoAs of these types of carcinogens involve molecular or cellular effects, which through secondary mechanisms, can either indirectly result in modification of DNA function or cell behavior [26][37]. For example, epigenetic carcinogens can induce oxidative stress, resulting in oxidative DNA damage [42][43][44], leading to either neoplastic conversion or stimulation of cell proliferation, thereby facilitating neoplastic development, often from cryptogenic pre-neoplastic cells. Epigenetic carcinogens can also affect gene expression [45][46], leading to neoplastic conversion. Such effects are often specific for rodents (e.g., d-limonene). Epigenetic carcinogens can enhance carcinogenicity of DNA-reactive carcinogens through interactive effects such as neoplasm promotion (e.g., butylated hydroxyanisole).

Due to their lack of direct DNA reactivity, epigenetic carcinogens, in contrast to DNA-reactive agents, are typically negative in genotoxicity assays, even in the presence of bioactivation, unless some artifact, such as extreme cytotoxicity, mediates mutagenicity. To exert their carcinogenicity, epigenetic agents often require prolonged high-level exposures. Their MoA underlies the fact that in limited bioassays they are negative for initiating activity, but may be positive for promoting activity [27].

Epigenetic carcinogens are well established to exhibit NOAELs for the cellular effect underlying their carcinogenicity in animal models [26][47], as discussed for several of the food-borne carcinogens reviewed herein. Accordingly, thresholds are generally accepted for DNA-reactive carcinogens from a risk assessment perspective [35].

4. Risk Assessment of Food-Derived Carcinogens

4.1. Application of Carcinogenicity Data to Human Risk

Two types of carcinogenicity data are used in the assessment of risk: human epidemiologic data and tumor data obtained in testing in rodent models [48]. The former is considered more relevant for a variety of reasons [49][50][51][52], although such data are often limited in human exposure information and can be poorly controlled [53].

Animal data are usually more robust, but frequently involve findings whose relevance to humans is uncertain [27][54][55], because the tumorigenic effect involves MoAs operational only in rodents. In addition, rodent studies do not mimic real life human exposures with respect to both the concentration and frequency of exposure. The human diet is also composed of mixture of components, which can both enhance and inhibit carcinogenicity.

Thus, in assessing human risk, two considerations are critical, i.e., the MoA of carcinogenicity and human exposure dose [11][56].

Once a chemical has been identified in a food product and its structure determined, it is possible to undertake an in silico analysis to determine, based on structure-activity relationships, the potential for DNA reactivity [57]. While this works well for relatively simple compounds, with the complexity of many natural products, the subtleties of metabolic activation become increasingly difficult to predict. If sufficient material is available, direct testing for DNA reactivity is the preferred approach [27].

This research focuses primarily on chemicals present in food that have sufficient evidence of carcinogenicity in either humans or experimental animals and which were classified by the International Agency for Research on Cancer (IARC) as either carcinogenic to humans (Group 1), probably (Group 2A) or possibly (Group 2B) carcinogenic to humans [48][58]. IARC also recognizes a third group of substances (Group 3) which lack sufficient evidence to be classified as carcinogenic to humans but nonetheless can have the potential to cause carcinogenicity in animals. Moreover, a variety of chemicals has not yet been characterized as to their carcinogenic risk to humans. Where available, evaluations by other expert groups are cited. Data on classification of carcinogens by government agencies and their carcinogenic potencies (TD₅₀) calculated based on the tumorigenicity findings in rodents are provided in **Table 2**.

Table 2. Classifications and characteristics of food-borne carcinogens.

Chemical Name	CAS Registry Number	Classification		Carcinogenic Potency (TD ₅₀ , mg/kg/d) ^c	MoA
		IARC ^a	NTP ^b		
1. Human carcinogens					
Aflatoxins		1	1	0.343 (mouse) 0.0032 (rat)	GTX
Aristolochic acid I	313-67-7	1	1	N/A	GTX
Benzene	71-43-2	1	1	77.5 (mouse) 169 (rat)	GTX
Benzo[a]pyrene	50-32-8	1	2	3.47 (mouse) 0.956 (rat)	GTX
Dioxin (TCDD)	1746-01-6	1	1	0.000156 (mouse) 0.0000235 (rat)	EPI
Dioxin-like compounds (PBCs)		1	N/L	N/A	EPI
Ethylene oxide	75-21-8	1	1	63.7 (mouse) 21.3 (rat)	GTX
Methoxsalen with UV A radiation	298-81-7	1	1	32.4 (rat)	GTX
Processed meat		1	N/L	N/A	GTX
Salted fish		1	N/L	N/A	GTX

Chemical Name	CAS Registry Number	Classification		Carcinogenic Potency (TD ₅₀ , mg/kg/d) ^c	MoA
		IARC ^a	NTP ^b		
2. Likely to be human carcinogens					
Acrylamide	79-06-1	2A	2	3.75 (rat)	GTX
2-Amino-3-methylimidazo[4,5- <i>f</i>]quinoline	76180-96-6	2A	2	19.6 (mouse) 0.812 (rat)	GTX
<i>p,p'</i> -Dichlorodiphenyl-trichloroethane (DDT)	50-29-3	2A	2	12.8 (mouse) 84.7 (rat)	EPI
Ethyl carbamate (urethane)	51-79-6	2A	2	16.9 (mouse) 41.3 (rat)	GTX
5-Methoxypsoralen	484-20-8	2A	N/L	N/A	GTX
<i>N</i> -nitrosodiethylamine	55-18-5	2A	2	0.0265 (rat)	GTX
Red meat		2A	N/L	N/A	GTX
2-Amino-3,4-dimethylimidazo[4,5- <i>f</i>]quinoline	77094-11-2	2B	2	15.5 (mouse)	GTX
2-Amino-3,8-dimethylimidazo[4,5- <i>f</i>]quinoline	77500-04-0	2B	2	24.3 (mouse) 1.66 (rat)	GTX
2-Amino-1-methyl-6-phenylimidazo[4,5- <i>b</i>]pyridine	105650-23-5	2B	2	33.2 (mouse) 1.78 (rat)	GTX
Benzophenone	119-61-9	2B	N/L	152 (rat) 379 (mouse)	EPI
Bracken fern		2B	N/L	N/A	GTX
Butylated hydroxyanisole	25013-16-5	2B	2	5530 (mouse) 405 (rat)	EPI
3-Chloro-1,2-propanediol	96-24-2	2B	N/L	117 (rat)	Uncertain
Crotonaldehyde	4170-30-3	2B	N/L	4.2 (rat)	GTX
Cycasin	14901-08-7	2B	N/L	N/A	GTX
1,3-Dichloro-2-propanol	96-23-1	2B	N/L	46.4 (rat)	GTX
Di(2-ethylhexyl) phthalate	117-81-7	2B	2	476 (rat) 484 (mouse)	EPI
1,4-Dioxane	123-91-1	2B	2	204 (mouse) 267 (rat)	Uncertain/EPI
Fumonisin B ₁	116355-83-0	2B	N/L	6.79 (mouse) 5.75 (rat)	Uncertain/EPI
Fusarin C	79748-81-5	2B	N/L	N/A	Uncertain/EPI
Furan	110-00-9	2B	2	2.72 (mouse) 0.396 (rat)	EPI
Lasiocarpine	303-34-4	2B	N/L	0.389 (rat)	GTX
Methyl eugenol	93-15-2	2B	2	19.3 (mouse) 19.7 (rat)	GTX
Methylazoxymethanol	592-62-1	2B	N/L	N/A	GTX
2-Methylimidazole	693-98-1	2B	N/L	782 (mouse) 868 (rat)	EPI
4-Methylimidazole	822-36-6	2B	N/L	387 (mouse) 317 (rat)	EPI
Methyl isobutyl ketone	108-10-1	2B	N/L	612 (rat)	EPI
Monocrotaline	315-22-0	2B	N/L	0.94 (rat)	GTX
β-Myrcene	123-35-3	2B	N/L	15,400 (rat)	EPI

Chemical Name	CAS Registry Number	Classification		Carcinogenic Potency (TD ₅₀ , mg/kg/d) ^c	MoA
		IARC ^a	NTP ^b		
<i>N</i> -nitrosodiethanolamine	1116-54-7	2B	2	3.17 (rat)	GTX
Ochratoxin A	303-47-9	2B	2	6.41 (mouse) 0.136 (rat)	GTX/EPI
Pickled vegetables		2B	N/L	N/A	GTX
Pulegone	89-82-7	2B	N/L	232 (mouse) 156 (rat)	EPI
Riddelliine	23246-96-0	2B	2	1.97 (mouse) 0.119 (rat)	GTX
Safrole	94-59-7	2B	2	51.3 (mouse) 441 (rat)	GTX
<i>trans,trans</i> -2,4-Hexadienal	142-83-6	2B	N/L	176 (mouse) 62.2 (rat)	GTX
3. Unknown carcinogenic potential					
Agaritrine ^d	2757-90-6	3	N/L	N/A	GTX
Butylated hydroxytoluene	128-37-0	3	N/L	653 (mouse)	EPI
Carrageenan (native) ^d	9000-07-1	3	N/L	N/A	
Chlorate (sodium salt) ^d	7775-09-9	3	N/L	69.1 (mouse) 0.865 (rat)	EPI
Eugenol ^d	97-53-0	3	N/L	N/A	
Furfural ^d	98-01-1	3	N/L	197 (mouse) 683 (rat)	Uncertain
Hydroquinone	123-31-9	3	N/L	225 (mouse) 82.8 (rat)	EPI
Isatidine ^d	15503-86-3	3	N/L	0.716 (rat)	GTX
d-Limonene ^d	5989-27-5	3	N/L	204 (rat)	EPI
Malondialdehyde	24382-04-5	3	N/L	14.1 (mouse) 122 (rat)	GTX
Patulin ^d	149-29-1	3	N/L	N/A	Uncertain
Ptaquiloside	87625-62-5	3	N/L	N/A	GTX
Quercetin ^d	117-39-5	3	N/L	10.1 (rat)	EPI
Retrorsine ^d	480-54-6	3	N/L	0.862 (rat)	GTX
Senkirkine ^d	2318-18-5	3	N/L	1.7 (rat)	GTX
Sodium saccharin ^d	128-44-9	3	N/L	2140 (rat)	EPI
Symphytine ^d	22571-95-5	3	N/L	1.91	GTX
Zearalenone ^d	17924-92-4	3	N/L	39 (mouse)	EPI
4. Not classified by IARC/NTP					
Daminozide ^d	1596-84-5	N/L	N/L	1030 (mouse) 2500 (rat)	EPI
Estragole	140-67-0	N/L	N/L	51.8 (mouse)	GTX
Genistein ^d	446-72-0	N/L	N/L	27.1 (rat)	EPI
<i>N</i> -methyl- <i>N</i> -formylhydrazine ^d	758-17-8	N/L	N/L	1.37 (mouse)	GTX

^a IARC group 1—carcinogenic to humans; group 2A—probably carcinogenic to humans; group 2B—possibly carcinogenic to humans; group 3—not classifiable as to its carcinogenicity to humans; group 4—probably not carcinogenic to humans.

Source—Agents Classified by the IARC Monographs, Volumes 1–131 [58] . ^b 1—known to be a human carcinogen; 2—reasonably anticipated to be a human carcinogen. Source—NTP Report on Carcinogens, 15th Edition [59] . ^c Only rodent data was included for comparison; Source—Lhasa Carcinogenicity Database, <https://carcdb.lhasalimited.org/> (accessed on 9 July 2022). ^d Not discussed in this research. EPI, epigenetic modifications; GTX, genotoxicity; N/A, not available N/L, not listed.

In this research, the evidence for human cancer risk from intake of food borne carcinogens of both the DNA-reactive and epigenetic types is discussed. In the assessment of risk from experimental studies, the greatest weight is given to studies with oral administration since that route of intake is most relevant to human consumption. The demonstration of human carcinogenicity is made in epidemiologic studies, although, the absence of an effect can be due to inadequacy of the studies.

4.2. Risk Assessment of DNA-Reactive Rodent Carcinogens

In order to evaluate possible safety concerns arising from presence of carcinogens with DNA-reactive MoA in the diet, many regulatory and advisory agencies, including the European Food Safety Authority Panel on Contaminants in the Food Chain (EFSA CONTAM) and the Joint Food and Agriculture Organization of the United Nations (FAO)/World Health Organization (WHO) the Expert Committee on Food Additives (JECFA) use a margin of exposure (MoE) approach [60][61]. MoE is calculated as a ratio between an appropriate Point of Departure for a tumor response, such as NOAELs obtained from animal studies, and a predicted or estimated human exposure level. A number of considerations should be taken into account when a MoE is derived, including the biological relevance of carcinogenic MoAs to humans [55].

Among DNA-reactive rodent carcinogens, only aflatoxins, aristolochic acid I, benzene, benzo[a]pyrene and ethylene oxide, have been found to be associated with cancer causation in humans (Table 2). Nevertheless, all materials in this class are genotoxic, indicating an MoA that represents human risk [62].

4.3. Risk Assessment of Epigenetic Carcinogens

The contribution and relevance of epigenetic mechanisms produced by dietary factors leading to the development of cancer in humans is uncertain [63], and the best approach to risk assessment of such carcinogens remains a topic of a debate [64]. Nevertheless, at low intermittent exposures (less than 1 mg/day) epigenetic carcinogens are not considered to pose cancer risks to humans [56]. This may reflect the absence in humans of the processes involved in the MoAs in rodents, e.g., d-limonene alpha 2μ(α_{2μ})-globulin nephropathy in male rats leading to kidney tumors [65], or the much lower exposures of humans, e.g., forestomach irritation in rats caused by butylated hydroxyanisole leading to squamous cell carcinoma [66]. Additionally, the fact that epigenetic changes can be reversible could contribute to lack of human risk. Hence, for epigenetic carcinogens NOAELs are used to derive safety values, such as tolerable daily intake (TDI) [56].

References

1. National Research Council Committee, on Comparative Toxicity of Naturally Occurring Carcinogens. Carcinogens and Anticarcinogens in the Human Diet: A Comparison of Naturally Occurring and Synthetic Substances; The National Academies Collection: Reports Funded by National Institutes of Health; National Academies Press (US): Washington, DC, USA, 1996.
2. Williams, G.M. Food-borne carcinogens. *Prog. Clin. Biol. Res.* 1986, 206, 73–81.
3. Abnet, C.C. Carcinogenic food contaminants. *Cancer Investig.* 2007, 25, 189–196.
4. Sugimura, T. Nutrition and dietary carcinogens. *Carcinogenesis* 2000, 21, 387–395.
5. Jackson, L.S. Chemical food safety issues in the United States: Past, present, and future. *J. Agric. Food Chem.* 2009, 57, 8161–8170.
6. Rietjens, I.M.C.M.; Michael, A.; Bolt, H.M.; Siméon, B.; Andrea, H.; Nils, H.; Christine, K.; Angela, M.; Gloria, P.; Daniel, R.; et al. The role of endogenous versus exogenous sources in the exposome of putative genotoxins and consequences for risk assessment. *Arch. Toxicol.* 2022, 96, 1297–1352.
7. Hecht, S.S.; Hoffmann, D. N-nitroso compounds and man: Sources of exposure, endogenous formation and occurrence in body fluids. *Eur. J. Cancer Prev.* 1998, 7, 165–166.
8. Tricker, A.R.; Preussmann, R. Carcinogenic N-nitrosamines in the diet: Occurrence, formation, mechanisms and carcinogenic potential. *Mutat. Res./Genet. Toxicol.* 1991, 259, 277–289.

9. Miller, E.C.; Miller, J.A. Biochemical mechanisms of chemical carcinogenesis. *Mol. Biol. Cancer* 1974, 377–402.
10. Preston, R.J.; Williams, G.M. DNA-reactive carcinogens: Mode of action and human cancer hazard. *Crit. Rev. Toxicol.* 2005, 35, 673–683.
11. Hartwig, A.; Arand, M.; Epe, B.; Guth, S.; Jahnke, G.; Lampen, A.; Martus, H.-J.; Monien, B.; Rietjens, I.M.C.M.; Schmitz-Spanke, S.; et al. Mode of action-based risk assessment of genotoxic carcinogens. *Arch. Toxicol.* 2020, 94, 1787–1877.
12. Hanawalt, P. Functional characterization of global genomic DNA repair and its implications for cancer. *Mutat. Res./Rev. Mutat. Res.* 2003, 544, 107–114.
13. Tong, C.; Fazio, M.; Williams, G.M. Cell cycle-specific mutagenesis at the hypoxanthine phosphoribosyltransferase locus in adult rat liver epithelial cells. *Proc. Natl. Acad. Sci. USA* 1980, 77, 7377–7379.
14. Kaufmann, W.K.; Kaufman, D.G. Cell cycle control, DNA repair and initiation of carcinogenesis. *FASEB J.* 1993, 7, 1188–1191.
15. Pagès, V.; Fuchs, R.P. How DNA lesions are turned into mutations within cells? *Oncogene* 2002, 21, 8957–8966.
16. Vogelstein, B.; Papadopoulos, N.; Velculescu, V.E.; Zhou, S.; Diaz, L.A., Jr.; Kinzler, K.W. Cancer genome landscapes. *Science* 2013, 339, 1546–1558.
17. Williams, G.M.; Iatropoulos, M.J.; Jeffrey, A.M. Mechanistic basis for nonlinearities and thresholds in rat liver carcinogenesis by the DNA-reactive carcinogens 2-acetylaminofluorene and diethylnitrosamine. *Toxicol. Pathol.* 2000, 28, 388–395.
18. Cohen, S.M.; Arnold, L.L. Chemical carcinogenesis. *Toxicol. Sci.* 2011, 120 (Suppl. S1), S76–S92.
19. Poirier, M.C. Chemical-induced DNA damage and human cancer risk. *Discov. Med.* 2012, 14, 283–288.
20. Paini, A.; Scholz, G.; Marin-Kuan, M.; Schilter, B.; O'Brien, J.; van Bladeren, P.J.; Rietjens, I.M.C.M. Quantitative comparison between in vivo DNA adduct formation from exposure to selected DNA-reactive carcinogens, natural background levels of DNA adduct formation and tumour incidence in rodent bioassays. *Mutagenesis* 2011, 26, 605–618.
21. Hwa Yun, B.; Guo, J.; Bellamri, M.; Turesky, R.J. DNA adducts: Formation, biological effects, and new biospecimens for mass spectrometric measurements in humans. *Mass Spectrom. Rev.* 2020, 39, 55–82.
22. Poirier, M.C.; Beland, F.A. DNA adduct measurements and tumor incidence during chronic carcinogen exposure in rodents. *Environ. Health Perspect.* 1994, 102 (Suppl. S6), 161–165.
23. Doerge, D.R.; Gamboa da Costa, G.; McDaniel, L.P.; Churchwell, M.I.; Twaddle, N.C.; Beland, F.A. DNA adducts derived from administration of acrylamide and glycidamide to mice and rats. *Mutat. Res.* 2005, 580, 131–141.
24. Lafferty, J.S.; Kamendulis, L.M.; Kaster, J.; Jiang, J.; Klaunig, J.E. Subchronic acrylamide treatment induces a tissue-specific increase in DNA synthesis in the rat. *Toxicol. Lett.* 2004, 154, 95–103.
25. Pavanello, S.; Bollati, V.; Pesatori, A.C.; Kapka, L.; Bolognesi, C.; Bertazzi, P.A.; Baccarelli, A. Global and gene-specific promoter methylation changes are related to anti-BPDE-DNA adduct levels and influence micronuclei levels in polycyclic aromatic hydrocarbon-exposed individuals. *Int. J. Cancer* 2009, 125, 1692–1697.
26. Kobets, T.; Iatropoulos, M.J.; Williams, G.M. Mechanisms of DNA-reactive and epigenetic chemical carcinogens: Applications to carcinogenicity testing and risk assessment. *Toxicol. Res.* 2019, 8, 123–145.
27. Williams, G.M.; Iatropoulos, M.J.; Enzmann, H.G.; Deschl, U. Carcinogenicity of chemicals: Assessment and human extrapolation. In *Hayes' Principles and Methods of Toxicology*, 6th ed.; Hayes, A., Kruger, C.L., Eds.; Taylor and Francis: Philadelphia, PA, USA, 2014; pp. 1251–1303.
28. Williams, G.M.; Iatropoulos, M.J.; Weisburger, J.H. Chemical carcinogen mechanisms of action and implications for testing methodology. *Exp. Toxicol. Pathol.* 1996, 48, 101–111.
29. Phillips, D.H.; Arlt, V.M. Genotoxicity: Damage to DNA and its consequences. *EXS* 2009, 99, 87–110.
30. Neumann, H.-G. Risk assessment of chemical carcinogens and thresholds. *Crit. Rev. Toxicol.* 2009, 39, 449–461.
31. Kobets, T.; Williams, G.M. Thresholds for hepatocarcinogenicity of DNA-reactive compounds. In *Thresholds of Genotoxic Carcinogens*; Academic Press: Cambridge, MA, USA, 2016; pp. 19–36.
32. Nohmi, T. Thresholds of genotoxic and non-genotoxic carcinogens. *Toxicol. Res.* 2018, 34, 281–290.
33. Kobets, T.; Williams, G.M. Review of the evidence for thresholds for DNA-reactive and epigenetic experimental chemical carcinogens. *Chem. Biol. Interact.* 2019, 301, 88–111.

34. Williams, G.M.; Iatropoulos, M.J.; Jeffrey, A.M. Dose-effect relationships for DNA-reactive liver carcinogens. In *Cellular Response to the Genotoxic Insult: The Question of Threshold for Genotoxic Carcinogens*; Oxford Academic: Oxford, UK, 2012; pp. 33–51.
35. EFSA CONTAM Panel, European Food Safety Authority, Panel on Contaminants in the Food Chain. Scientific Opinion of the Panel on Contaminants in the Food Chain on a Request from the European Commission on Polycyclic Aromatic Hydrocarbons in Food; The EFSA Journal Series, No. 724; European Food Safety Authority: Parma, Italy, 2008; 114p.
36. Weisburger, J.H.; Williams, G.M. The distinction between genotoxic and epigenetic carcinogens and implication for cancer risk. *Toxicol. Sci.* 2000, 57, 4–5.
37. Williams, G.M. DNA reactive and epigenetic carcinogens. *Exp. Toxicol. Pathol.* 1992, 44, 457–463.
38. Klaunig, J.E.; Kamendulis, L.M.; Xu, Y. Epigenetic mechanisms of chemical carcinogenesis. *Hum. Exp. Toxicol.* 2000, 19, 543–555.
39. Sawan, C.; Vaissière, T.; Murr, R.; Herceg, Z. Epigenetic drivers and genetic passengers on the road to cancer. *Mutat. Res./Fundam. Mol. Mech. Mutagen.* 2008, 642, 1–13.
40. Pogribny, I.P.; Rusyn, I.; Beland, F.A. Epigenetic aspects of genotoxic and non-genotoxic hepatocarcinogenesis: Studies in rodents. *Environ. Mol. Mutagen.* 2008, 49, 9–15.
41. Pogribny, I.P.; Rusyn, I. Environmental toxicants, epigenetics, and cancer. *Adv. Exp. Med. Biol.* 2013, 754, 215–232.
42. Williams, G.M.; Jeffrey, A.M. Oxidative DNA damage: Endogenous and chemically induced. *Regul. Toxicol. Pharmacol.* 2000, 32, 283–292.
43. Cooke, M.S.; Evans, M.D.; Dizdaroglu, M.; Lunec, J. Oxidative DNA damage: Mechanisms, mutation, and disease. *FASEB J.* 2003, 17, 1195–1214.
44. Klaunig, J.E.; Kamendulis, L.M. The role of oxidative stress in carcinogenesis. *Annu. Rev. Pharmacol. Toxicol.* 2004, 44, 239–267.
45. Jones, P.A.; Baylin, S.B. The epigenomics of cancer. *Cell* 2007, 128, 683–692.
46. Baylin, S.B.; Jones, P.A. Epigenetic determinants of cancer. *Cold Spring Harb. Perspect. Biol.* 2016, 8, a019505.
47. Williams, G.M. Mechanisms of chemical carcinogenesis and application to human cancer risk assessment. *Toxicology* 2001, 166, 3–10.
48. IARC, International Agency for Research on Cancer. Preamble to the IARC Monographs (Amended January 2019); International Agency for Research on Cancer: Lyon, France, 2019.
49. Wiltse, J.; Dellarco, V.L. U.S. Environmental Protection Agency guidelines for carcinogen risk assessment: Past and future. *Mutat. Res./Rev. Genet. Toxicol.* 1996, 365, 3–15.
50. Jeffrey, A.M.; Williams, G.M. Risk assessment of DNA-reactive carcinogens in food. *Toxicol. Appl. Pharm.* 2005, 207, 628–635.
51. Felter, S.P.; Bhat, V.S.; Botham, P.A.; Bussard, D.A.; Casey, W.; Hayes, A.W.; Hilton, G.M.; Magurany, K.A.; Sauer, U.G.; Ohanian, E.V. Assessing chemical carcinogenicity: Hazard identification, classification, and risk assessment. Insight from a Toxicology Forum state-of-the-science workshop. *Crit. Rev. Toxicol.* 2021; 51, 653–694.
52. Barlow, S.; Schlatter, J. Risk assessment of carcinogens in food. *Toxicol. Appl. Pharm.* 2010, 243, 180–190.
53. Raffaele, K.; Vulimiri, S.; Bateson, T. Benefits and barriers to using epidemiology data in environmental risk assessment. *Open Epidemiol. J.* 2011, 411, 99–105.
54. Kobets, T.; Williams, G.M. Chemicals with carcinogenic activity primarily in rodent liver. In *Comprehensive Toxicology*; Elsevier: Amsterdam, The Netherlands, 2018; pp. 409–442.
55. Edler, L.; Hart, A.; Greaves, P.; Carthew, P.; Coulet, M.; Boobis, A.; Williams, G.M.; Smith, B. Selection of appropriate tumour data sets for Benchmark Dose Modelling (BMD) and derivation of a Margin of Exposure (MoE) for substances that are genotoxic and carcinogenic: Considerations of biological relevance of tumour type, data quality and uncertainty assessment. *Food Chem. Toxicol.* 2014, 70, 264–289.
56. Williams, G.M. Application of mode-of-action considerations in human cancer risk assessment. *Toxicol. Lett.* 2008, 180, 75–80.
57. Rosenkranz, H.S. SAR modeling of genotoxic phenomena: The consequence on predictive performance of deviation from a unity ratio of genotoxicants/non-genotoxicants. *Mutat. Res./Genet. Toxicol. Environ. Mutagen.* 2004, 559, 67–71.
58. IARC, International Agency for Research on Cancer. Agents Classified by the IARC Monographs; International Agency for Research on Cancer: Lyon, France, 2022; Volume 1–131.

59. NTP, National Toxicology Program. Report on Carcinogens, (RoC), 15th ed.; National Toxicology Program: Research Triangle, NC, USA, 2021.
60. O'Brien, J.; Renwick, A.G.; Constable, A.; Dybing, E.; Müller, D.J.G.; Schlatter, J.; Slob, W.; Tueting, W.; van Benthem, J.; Williams, G.M.; et al. Approaches to the risk assessment of genotoxic carcinogens in food: A critical appraisal. *Food Chem. Toxicol.* 2006, 44, 1613–1635.
61. Benford, D.; Bolger, P.M.; Carthew, P.; Coulet, M.; DiNovi, M.; Leblanc, J.-C.; Renwick, A.G.; Setzer, W.; Schlatter, J.; Smith, B.; et al. Application of the Margin of Exposure (MOE) approach to substances in food that are genotoxic and carcinogenic. *Food Chem. Toxicol.* 2010, 48, S2–S24.
62. Williams, G.M. Chemicals with carcinogenic activity in the rodent liver; mechanistic evaluation of human risk. *Cancer Lett.* 1997, 117, 175–188.
63. Herceg, Z. Epigenetics and cancer: Towards an evaluation of the impact of environmental and dietary factors. *Mutagenesis* 2007, 22, 91–103.
64. Braakhuis, H.M.; Slob, W.; Olthof, E.D.; Wolterink, G.; Zwart, E.P.; Gremmer, E.R.; Rorije, E.; van Benthem, J.; Woutersen, R.; van der Laan, J.W.; et al. Is current risk assessment of non-genotoxic carcinogens protective? *Crit. Rev. Toxicol.* 2018, 48, 500–511.
65. Swenberg, J.A.; Lehman-McKeeman, L.D. alpha 2-Urinary globulin-associated nephropathy as a mechanism of renal tubule cell carcinogenesis in male rats. *IARC Sci. Publ.* 1999, 147, 95–118.
66. Williams, G.M.; Whysner, J. Epigenetic carcinogens: Evaluation and risk assessment. *Exp. Toxicol. Pathol.* 1996, 48, 189–195.

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