B-Cell Activating Factor in IBD

Subjects: Immunology Contributor: Josko Bozic

B-cell activating factor (BAFF), a member of the tumor necrosis factor (TNF) superfamily. BAFF is predominantly produced by myeloid cells (monocytes, macrophages, dendritic cells and neutrophils), and its main role is regulation of mature B cell survival and differentiation into antibody-producing plasma cells. Overproduction of BAFF has been observed in various autoimmune diseases, most notably in systemic lupus erythematosus (SLE), where BAFF-inhibitor belimumab was approved for treatment.

B-cell activating factor inflammatory bowel disease

1. Role of B-Cells in IBD

Despite myriad of distinct biological pathways have been implicated in its pathophysiology, it is generally believed that IBD is a result of a maladaptive immune response to gut-resident commensal bacteria in a genetically susceptible host ^[1]. Inflammation in CD seems to be mainly driven by T_h1 responses, whereas T_h2 responses dominate the pathobiology of UC ^[2]. Nevertheless, additional lymphocytes, such as innate lymphoid cells and T_h17 cells have also arisen as key players in the pathogenesis of IBD ^[3]. Specifically, abnormal pro-inflammatory CD4⁺ T-cell responses mediated by effector T_h1 , T_h2 , or T_h17 cells disrupt homeostasis and causes IBD by outweighing anti-inflammatory CD4⁺ T-cell responses orchestrated by T regulatory (T_{reg}) cells ^[4]. Even though T-cell system is predominant in studies concerning the IBD pathogenesis and therapeutic approach, emerging data suggest a role of B-cell lineage in IBD as well. Firstly, humoral homeostasis seems to be impaired in IBD. For instance, it has been shown that production of functional, dimeric immunoglobulin A (IgA) is impaired in patients with IBD ^[5]. As IgA exerts local anti-inflammatory effects by coating commensal bacteria after undergoing transepithelial translocation in gut, its depletion favors gut inflammation ^[6]. Moreover, B-cell expression of the pro-inflammatory cytokine IL-8, as well as production of mucosal IgG in gut are upregulated in IBD, thereby further promoting inflammation ^[7]. Accordingly, expression of both IL-8 and TLR-2 in IBD patients positively correlated with CD activity ^[8]

In murine IBD model, poorly regulated B-cells have been shown to exacerbate inflammation by blocking T_{reg} cell function ^{[11][12]}. Moreover, B-cells promote ileitis in UC by producing epithelial cell-specific autoantibodies ^{[11][13]}. Mucosal IgG in IBD can be directed against microbial elements, such as anti-saccharomyces-cerevisiae antibodies (ASCA) and anti-flagellin antibodies, or autoantigens, such as anti-neutrophil cytoplasmic antibodies (ANCA) and anti-epithelial antibodies ^{[14][15]}. Notably, the role of anti-granulocyte macrophage colony-stimulating factor (anti-GM-CSF) in IBD is fairly complex, yet the presence of anti-GM-CSF in the setting of IBD is associated with ileal phenotype and intricated behavior of the disease ^[16]. The latter observation is in line with preclinical data, as NOD2

KO mice treated with anti-GM-CSF antibodies develop transmural ileitis subsequent to NSAID exposure [16]. Alterations of the B-cell lineage in IBD are less obvious, and more complex for that matter, than those associated with derangement of T-cell system [17]. Acknowledgement of poor understanding of the complexity of B-cell responses in IBD allows us to evaluate recently failed therapeutic attempts in UC involving the CD20-targeting agent rituximab more critically ^[18]. Namely, negative outcomes of therapeutic approaches including rituximab should not discourage from considering B-cells as potential therapeutic target in IBD, especially since unresponsiveness to rituximab can be also observed in certain cases of B-cell-related autoimmune disorders such as rheumatoid arthritis (RA) and vasculitis [19][20]. For example, in certain group of patients, paradoxical proinflammatory manifestations can occur subsequent to rituximab administration [21][22]. While dysfunctional B-cell lineage can promote autoimmunity via autoreactive, long-lived plasma cells, regulatory B-cells can attenuate inflammation too. For instance, anti-CD20-treated mice deficient in peripheral B-cells failed to undergo spontaneous recovery and even developed chronic disease in a model of murine autoimmune encephalomyelitis [23][24]. In addition, B-cells may produce anti-inflammatory IL-10 but may also promote the anti-inflammatory effect of T_{reg} cells ^[25]. Even more perplexing is the fact that tissue resident plasma cells do not express CD20 and thus cannot be targeted by rituximab ^[26]. In the setting of IBD, in vitro experiment showed that plasma cells subset expanded in the mucosa of IBD patients and was resistant to rituximab-induced apoptosis [27]. Nevertheless, evidence of rituximab in IBD is conflicting, as some studies showed that rituximab could improve colonic inflammation, whereas case reports showed that rituximab could trigger colitis [28][29][30][31]. In fact, a retrospective cohort study showed that patients on rituximab have a sixfold increased risk of developing IBD compared to the general population ^[32]. Finally, a phase II randomized controlled trial, in which effects of rituximab on UC patients was assessed, showed no significant effect on inducing remission in moderately active UC not responding to oral steroids with possible short-term response that was not sustained $\frac{[28]}{2}$.

2. Pathophysiological Background of BAFF in IBD

Even though BAFF is widely considered as a cytokine affecting B-cells primarily, and to a lesser extent T-cells, BAFF has been also shown to affect innate immunity by multiple effector arms ^[33]. Namely, it has been well documented that BAFF improves human monocyte survival, upregulates proinflammatory cytokine secretion, and positively regulates secretion of multiple costimulatory molecules ^[34]. These findings may be important for explaining the role of BAFF in IBD, as genome-wide association studies have highlighted the importance of host innate immune responses to microbes in the pathogenesis of IBD ^[31]. Single nucleotide polymorphisms associated with increased risk of developing IBD were identified in genes encoding microbial sensing and clearance, as well as integrating antimicrobial adaptive immune responses ^[35]. Accordingly, evidence suggests that macrophages and dendritic cells residing in gastrointestinal system have important interactions with the microbial environment, resolution of mucosal inflammation, proinflammatory tissue injury, and induction of adaptive immune responses ^[36]. ^[37]BB. Despite the presence of BAFF is most commonly associated with the induction of autoimmune inflammatory injury has also been explored ^[39]. It was recently demonstrated that mice with DSS-colitis exhibit a persistent decrease in colonic CD5(+) regulatory B-cells (B_{rea}), suggesting that persistent altered mucosal B-cell population

caused by chronic gut inflammation may be involved in the pathogenesis of IBD ^[40]. These conclusions were corroborated in a small-sample clinical study, as UC patients had significantly reduced frequencies of CD5(+) B_{reg} in peripheral blood and intestinal tissues, accompanied by lower serum IL-10 levels ^[41]. In the same study, Mayo clinic scores, CRP, and ESR in UC patients negatively correlated with the frequency of B_{reg} and the IL-10 concentration. Nevertheless, as we further discuss, BAFF serum levels positively correlate with clinical disease activity and inflammatory biomarkers, thus indicating that the role of BAFF in IBD is primarily proinflammatory ^[42]. Finally, the strongest evidence indicating contribution of BAFF to IBD pathogenesis is data from colonic biopsies from UC and CD patients ^[42]. In colonic bioptates of both UC and CD patients, mRNA and BAFF protein expression were higher than in the control group. Furthermore, in inflamed regions of UC mucosa, upregulation of BAFF was predominant in mononuclear cells residing in lamina propria ^[42].

Finally, an important link connecting BAFF with IBD is the Nuclear Factor kappa-light-chain-enhancer of activated B-cells (NFκB), protein complex involved in control of transcription of DNA in almost all mammalian cells ^[43]. In response to various pro-inflammatory stimuli, NFκB activates and leads to in the increased expression of adhesion molecules and chemokines by endothelial cells and in the tissue, thus favoring the recruitment and activation of effector immune cells ^[44]. Multiple line of evidence suggests important role of NFκB activation in IBD pathogenesis ^{[45][46]}. Activation of NFκB is detected in both epithelial cells and macrophages from IBD patients and relates to the intensity of inflammation ^[47]. Furthermore, administration of antisense phosphorothioate oligonucleotides to the p65 subunit of NFκB has been shown to suppress colitis in mice model of TNBS-induced colitis ^[48]. Finally, effectiveness of corticosteroid treatment in IBD flares is in part owing to steroid-induced decrease of NFκB activation ^[49]. As BAFF is capable of activating NFκB in lymphoid and myeloid cells via both canonical and non-canonical pathway, thus promoting intestinal inflammation, it is plausible that NFκB is the missing link between BAFF and IBD ^[50].

3. Clinical Implications of BAFF in IBD

In light of the prominent role of BAFF in B-cells and autoimmunity, Zhang et al. sought to explore the putative role of BAFF in management of IBD ^[42]. The principal aim of that pivotal study was to determine the value of BAFF to discriminate patients with IBD from healthy controls and patients with IBS by measuring BAFF serum and fecal levels, as well as its mucosal expression. An additional aim was to establish whether there is a correlation between BAFF and disease activity in IBD patients. It was demonstrated that BAFF expression is increased in serum, feces and colonic mucosa of patients with IBD when compared to controls. Moreover, in comparison to IBS patients, significantly higher fecal BAFF concentrations were observed in patients with IBD, regardless of disease activity, with fecal BAFF concentrations in IBS patients being identical to those of healthy controls. In fact, for BAFF fecal levels above cut-off of 325 pg/mL, respectively. Even higher sensitivity (90%) was observed for discrimination between BAFF and disease activity, TNF- α and IL-1 β in patients with UC. The discriminative power of serum BAFF had comparable specificity (93%), but markedly lower sensitivity (55%) than fecal BAFF. Nevertheless, it appears that, following the reduction in disease activity, BAFF serum levels return to values similar to that of healthy subjects

rapidly, unlike BAFF levels in feces which persist much longer. Thus, it is plausible that serum BAFF may be utilized in monitoring the disease activity.

Clinical distinction between IBD and IBS is of utmost importance, as these two conditions can present similar symptoms, but have very different underlying pathophysiology and, more importantly, severity of consequences ^[51]. In the absence of additional symptoms indicating on IBD, such as rectal bleeding and systemic illness, it is very challenging to distinguish between the two. On the other hand, although it is a common practice, it is not cost-effective to use colonoscopy as a part of diagnostic algorithm in the workup of patients suspected of having IBS ^[52]. Moreover, colonoscopy is associated with serious complications such as bleeding and perforations, adverse events related to the anesthesia, and increased discomfort of patients ^[54]. Hence, it would be beneficial to use sensitive and specific biomarker for IBD/IBS differentiation. So far, calprotectin has been used for this purpose with relative success owing to sufficient sensitivity, yet what calprotectin lacks is adequate specificity ^[55].

In this sense, Fu et al. compared the efficacy of fecal BAFF, calprotectin and fecal occult blood test (FOBT) to find the "best non-invasive marker" $\frac{56}{56}$. The study showed that for discriminating IBD from IBS, fecal BAFF ≥ 227.3 pg/mL yielded 84% sensitivity and 100% specificity, calprotectin \geq 50 µg/g yielded 76% sensitivity and 93% specificity whereas FOBT yielded 65% sensitivity and 93% specificity. Moreover, combination of BAFF with calprotectin yielded 94% sensitivity and 93% specificity, thus increasing the accuracy of differential diagnosis. Notably, fecal BAFF concentration exhibited stronger correlation with endoscopic inflammatory score in comparison to calprotectin in both UC (r = 0.69, p < 0.0001 vs. r = 0.58, p < 0.0001) and CD (r = 0.58, p < 0.0001 vs. r = 0.580.52, p = 0.0003). Accordingly, a separate study confirmed that fecal BAFF is more sensitive and specific in predicting UC activity and severity than fecal calprotectin ^[57]. On a separate note, neither fecal BAFF nor calprotectin showed significant correlation with Crohn's Disease Activity Index (CDAI) in CD patients, yet they both showed correlation with Mayo score in patients with UC (r = 0.415 and 0.365, respectively). Furthermore, Xie et al. explored whether BAFF can discriminate patients with IBD and malignancy from other gastrointestinal diseases among population of patients presenting with abdominal discomfort [58]. It was demonstrated that fecal BAFF was able to accurately distinguish patients with either IBD or tumor from patients without any of these, giving a sensitivity of 85% and specificity of 91%. On the other hand, BAFF was also able to discriminate IBD from patients without it with sensitivity of 89% and specificity of 77%. An important finding of this study is that BAFF was found to be temperature-stable for 7 days and equally distributed in the feces, thus implying that for BAFF measurement no specific storage conditions are required and only a small number of samples are needed to accurately measure it. Altogether, the above-noted results imply that BAFF could be used as a complementary biomarker in diagnostic workup of patients with suspected IBD and that it may also be utilized as a sensitive surrogate for assessment of endoscopic inflammation in IBD. Nevertheless, further research is warranted to elucidate in deep the values of fecal BAFF in IBD clinical scenery, such as whether fecal BAFF can be used to predict relapse in IBD patients in remission, as shown in calprotectin [59].

Usefulness of BAFF in the setting of IBD has been explored in pediatric population as well. The problem of IBD diagnosis and follow-up represents an even bigger challenge children then among the adults. Firstly, colonoscopy is not easily accepted by children nor caregivers compared to adults ^[60]. Moreover, currently no consensus was

established with regard to the definition of remission in pediatric IBD. The term "clinical remission" is the most widely accepted term defined by composite scores based on clinical parameters (abdominal pain, rectal bleeding, stool frequency and consistency, number of stools per 24 h, nocturnal stools, activity level, weight). Notably, laboratory findings (ESR and albumin) are used in the wPCDAI score exclusively. As multiple studies showed that there is a discrepancy between the clinical remission and the laboratory, endoscopic and histologic findings, there is a need for a comprehensive approach, including clinical, imaging, histologic and laboratory parameters ^{[61][62][63]}. In a recent prospective study, Fodor et al. found no differences in serum BAFF between IBD, IBS, and healthy group, yet they found that fecal BAFF was higher in the IBD group in comparison to IBS and healthy group ^[64]. In comparison of different types of IBD, it was shown that BAFF is higher in pediatric patients with UC compared to CD patients. ROC curve analysis for fecal BAFF showed that with cut-off of 16 pg/mL, sensitivity and specificity for discrimination between IBD and IBS in pediatric population is 51% and 93%, respectively. The observed lack of sensitivity could be owing to the fact that only mild cases of IBD were included and limited number of participants. Finally, as fecal BAFF was the highest in patients with increased calprotectin levels, the authors proposed that fecal BAFF may be a promising marker in the evaluation of the remission status in pediatric IBD.

References

- 1. Zhang, Y.Z.; Li, Y.Y. Inflammatory bowel disease: Pathogenesis. World J. Gastroenterol. 2014, 20, 91–99.
- 2. Schirbel, A.; Fiocchi, C. Inflammatory bowel disease: Established and evolving considerations on its etiopathogenesis and therapy. J. Dig. Dis. 2010, 11, 266–276.
- Zhao, J.; Lu, Q.; Liu, Y.; Shi, Z.; Hu, L.; Zeng, Z.; Tu, Y.; Xiao, Z.; Xu, Q. Th17 Cells in Inflammatory Bowel Disease: Cytokines, Plasticity, and Therapies. J. Immunol. Res. 2021, 2021, 8816041.
- 4. Yan, J.B.; Luo, M.M.; Chen, Z.Y.; He, B.H. The Function and Role of the Th17/Treg Cell Balance in Inflammatory Bowel Disease. J. Immunol. Res. 2020, 2020, 8813558.
- 5. Ware, C.F. Decoy receptors thwart B cells. Nature 2000, 404, 949–950.
- Okai, S.; Usui, F.; Ohta, M.; Mori, H.; Kurokawa, K.; Matsumoto, S.; Kato, T.; Miyauchi, E.; Ohno, H.; Shinkura, R. Intestinal IgA as a modulator of the gut microbiota. Gut Microbes. 2017, 8, 486– 492.
- Yu, G.; Boone, T.; Delaney, J.; Hawkins, N.; Kelley, M.; Ramakrishnan, M.; McCabe, S.; Qiu, W.R.; Kornuc, M.; Xia, X.Z.; et al. APRIL and TALL-I and receptors BCMA and TACI: System for regulating humoral immunity. Nat. Immunol. 2000, 1, 252–256.
- 8. Do, R.K.; Chen-Kiang, S. Mechanism of BLyS action in B cell immunity. Cytokine Growth Factor Rev. 2002, 13, 19–25.

- 9. Morimoto, M.; Watanabe, T.; Yamori, M.; Takebe, M.; Wakatsuki, Y. Isoflavones regulate innate immunity and inhibit experimental colitis. J. Gastroenterol. Hepatol. 2009, 24, 1123–1129.
- McDonnell, M.; Liang, Y.; Noronha, A.; Coukos, J.; Kasper, D.L.; Farraye, F.A.; Ganley-Leal, L.M. Systemic Toll-like receptor ligands modify B-cell responses in human inflammatory bowel disease. Inflamm. Bowel Dis. 2011, 17, 298–307.
- 11. Goetz, M.; Atreya, R.; Ghalibafian, M.; Galle, P.R.; Neurath, M.F. Exacerbation of ulcerative colitis after rituximab salvage therapy. Inflamm. Bowel Dis. 2007, 13, 1365–1368.
- 12. Olson, T.S.; Bamias, G.; Naganuma, M.; Rivera-Nieves, J.; Burcin, T.L.; Ross, W.; Morris, M.A.; Pizarro, T.T.; Ernst, P.B.; Cominelli, F.; et al. Expanded B cell population blocks regulatory T cells and exacerbates ileitis in a murine model of Crohn disease. J. Clin. Investig. 2004, 114, 389–398.
- 13. Takahasi, F.; Shah, H.S.; Wise, L.S.; Das, K.M. Circulating antibodies against human colonic extract enriched with a 40 kDa protein in patients with ulcerative colitis. Gut 1990, 31, 1016–1020.
- Chao, L.P.; Steele, J.; Rodrigues, C.; Lennard-Jones, J.; Stanford, J.L.; Spiliadis, C.; Rook, G.A. Specificity of antibodies secreted by hybridomas generated from activated B cells in the mesenteric lymph nodes of patients with inflammatory bowel disease. Gut 1988, 29, 35–40.
- Russell, M.W.; Reinholdt, J.; Kilian, M. Anti-inflammatory activity of human IgA antibodies and their Faba fragments: Inhibition of IgG-mediated complement activation. Eur. J. Immunol. 1989, 19, 2243–2249.
- Han, X.; Uchida, K.; Jurickova, I.; Koch, D.; Willson, T.; Samson, C.; Bonkowski, E.; Trauernicht, A.; Kim, M.O.; Tomer, G.; et al. Granulocyte-macrophage colony-stimulating factor autoantibodies in murine ileitis and progressive ileal Crohn's disease. Gastroenterology 2009, 136, 1261–1271.
- 17. Larabi, A.; Barnich, N.; Nguyen, H.T.T. New insights into the interplay between autophagy, gut microbiota and inflammatory responses in IBD. Autophagy 2020, 16, 38–51.
- 18. Uzzan, M.; Colombel, J.F.; Cerutti, A.; Treton, X.; Mehandru, S. B Cell-Activating Factor (BAFF)-Targeted B Cell Therapies in Inflammatory Bowel Diseases. Dig. Dis. Sci. 2016, 61, 3407–3424.
- 19. Selmi, C.; Generali, E.; Massarotti, M.; Bianchi, G.; Sciré, C.A. New treatments for inflammatory rheumatic disease. Immunol. Res. 2014, 60, 277–288.
- 20. Dumoitier, N.; Terrier, B.; London, J.; Lofek, S.; Mouthon, L. Implication of B lymphocytes in the pathogenesis of ANCA-associated vasculitides. Autoimmun. Rev. 2015, 14, 996–1004.
- 21. Jayasekera, P.; Parslew, R.; Al-Sharqi, A. A case of tumour necrosis factor-a inhibitor- and rituximab-induced plantar pustular psoriasis that completely resolved with tocilizumab. Br. J. Dermatol. 2014, 171, 1546–1549.
- 22. Fiorillo, L.; Wang, C.; Hemmati, I. Rituximab induced psoriasis in an infant. Pediatr. Dermatol. 2014, 31, e149–e151.

- Wolf, S.D.; Dittel, B.N.; Hardardottir, F.; Janeway, C.A. Experimental autoimmune encephalomyelitis induction in genetically B cell-deficient mice. J. Exp. Med. 1996, 184, 2271– 2278.
- 24. Ray, A.; Mann, M.K.; Basu, S.; Dittel, B.N. A case for regulatory B cells in controlling the severity of autoimmune-mediated inflammation in experimental autoimmune encephalomyelitis and multiple sclerosis. J. Neuroimmunol. 2011, 230, 1–9.
- Ray, A.; Basu, S.; Williams, C.B.; Salzman, N.H.; Dittel, B.N. A novel IL-10-independent regulatory role for B cells in suppressing autoimmunity by maintenance of regulatory T cells via GITR ligand. J. Immunol. 2012, 188, 3188–3198.
- He, Y.; Shimoda, M.; Ono, Y.; Villalobos, I.B.; Mitra, A.; Konia, T.; Grando, S.A.; Zone, J.J.; Maverakis, E. Persistence of autoreactive IgA-secreting B cells despite multiple immunosuppressive medications including Rituximab. JAMA Dermatol. 2015, 151, 646–650.
- Cupi, M.L.; Sarra, M.; Marafini, I.; Monteleone, I.; Franzè, E.; Ortenzi, A.; Colantoni, A.; Sica, G.; Sileri, P.; Rosado, M.M.; et al. Plasma cells in the mucosa of patients with inflammatory bowel disease produce granzyme B and possess cytotoxic activities. J. Immunol. 2014, 192, 6083– 6091.
- Leiper, K.; Martin, K.; Ellis, A.; Subramanian, S.; Watson, A.J.; Christmas, S.E.; Howarth, D.; Campbell, F.; Rhodes, J.M. Randomised placebo-controlled trial of rituximab (anti-CD20) in active ulcerative colitis. Gut 2011, 60, 1520–1526.
- 29. Swaminath, A.; Magro, C.M.; Dwyer, E. Refractory urticarial vasculitis as a complication of ulcerative colitis successfully treated with rituximab. J. Clin. Rheumatol. 2011, 17, 281–283.
- 30. Ardelean, D.S.; Gonska, T.; Wires, S.; Cutz, E.; Griffiths, A.; Harvey, E.; Tse, S.M.; Benseler, S.M. Severe ulcerative colitis after rituximab therapy. Pediatrics 2010, 126, e243–e246.
- 31. Cho, J.H.; Brant, S.R. Recent insights into the genetics of inflammatory bowel disease. Gastroenterology 2011, 140, 1704–1712.
- Kristjánsson, V.B.; Lund, S.H.; Gröndal, G.; Sveinsdóttir, S.V.; Agnarsson, H.R.; Jónasson, J.G.; Björnsson, E.S. Increased risk of inflammatory bowel disease among patients treated with rituximab in Iceland from 2001 to 2018. Scand. J. Gastroenterol. 2021, 56, 46–52.
- 33. Vincent, F.B.; Morand, E.F.; Mackay, F. BAFF and innate immunity: New therapeutic targets for systemic lupus erythematosus. Immunol. Cell Biol. 2012, 90, 293–303.
- 34. Chang, S.K.; Arendt, B.K.; Darce, J.R.; Wu, X.; Jelinek, D.F. A role for BLyS in the activation of innate immune cells. Blood 2006, 108, 2687–2694.
- 35. Jostins, L.; Ripke, S.; Weersma, R.K.; Duerr, R.H.; McGovern, D.P.; Hui, K.Y.; Lee, J.C.; Schumm, L.P.; Sharma, Y.; Anderson, C.A.; et al. Host-microbe interactions have shaped the genetic

architecture of inflammatory bowel disease. Nature 2012, 491, 119-124.

- 36. Steinbach, E.C.; Plevy, S.E. The role of macrophages and dendritic cells in the initiation of inflammation in IBD. Inflamm. Bowel Dis. 2014, 20, 166–175.
- Thosani, N.; Singh, H.; Kapadia, A.; Ochi, N.; Lee, J.H.; Ajani, J.; Swisher, S.G.; Hofstetter, W.L.; Guha, S.; Bhutani, M.S. Diagnostic accuracy of EUS in differentiating mucosal versus submucosal invasion of superficial esophageal cancers: A systematic review and meta-analysis. Gastrointest. Endosc. 2012, 75, 242–253.
- 38. Mowat, A.M.; Bain, C.C. Mucosal macrophages in intestinal homeostasis and inflammation. J. Innate Immun. 2011, 3, 550–564.
- 39. Liu, Z.; Davidson, A. BAFF and selection of autoreactive B cells. Trends Immunol. 2011, 32, 388– 394.
- Mishima, Y.; Ishihara, S.; Amano, Y.; Oshima, N.; Kadota, C.; Otani, A.; Moriyama, I.; Li, Y.Y.; Aziz, M.M.; Kinoshita, Y. Alterations of peripheral blood CD5+ B cells in inflammatory bowel disease. Scand. J. Gastroenterol. 2009, 44, 172–179.
- 41. Wang, X.; Zhu, Y.; Zhang, M.; Wang, H.; Jiang, Y.; Gao, P. Ulcerative Colitis Is Characterized by a Decrease in Regulatory B Cells. J. Crohn's Colitis 2016, 10, 1212–1223.
- 42. Zhang, P.; Liu, X.; Guo, A.; Xiong, J.; Fu, Y.; Zou, K. B cell-activating factor as a new potential marker in inflammatory bowel disease. Dig. Dis. Sci. 2016, 61, 2608–2618.
- 43. Mitchell, S.; Vargas, J.; Hoffmann, A. Signaling via the NFκB system. Wiley Interdiscip. Rev. Syst. Biol. Med. 2016, 8, 227–241.
- 44. Hayden, M.S.; West, A.P.; Ghosh, S. NF-kappaB and the immune response. Oncogene 2006, 25, 6758–6780.
- 45. Ordaś, I.; Eckmann, L.; Talamini, M.; Baumgart, D.C.; Sandborn, W.J. Ulcerative colitis. Lancet 2012, 380, 1606–1619.
- 46. Baumgart, D.C.; Sandborn, W.J. Crohn's disease. Lancet 2012, 380, 1590–1605.
- Rogler, G.; Brand, K.; Vogl, D.; Page, S.; Hofmeister, R.; Andus, T.; Knuechel, R.; Baeuerle, P.A.; Schölmerich, J.; Gross, V. Nuclear factor kappaB is activated in macrophages and epithelial cells of inflamed intestinal mucosa. Gastroenterology 1998, 115, 357–369.
- 48. Neurath, M.F.; Pettersson, S.; Zum Büschenfelde, K.H.M.; Strober, W. Local administration of antisense phosphorothioate oligonucleotides to the p65 subunit of NF-kappa B abrogates established experimental colitis in mice. Nat. Med. 1996, 2, 998–1004.
- 49. Schreiber, S.; Nikolaus, S.; Hampe, J. Activation of nuclear factor kappa B inflammatory bowel disease. Gut 1998, 42, 477–484.

- 50. Demchenko, Y.N.; Kuehl, W.M. A critical role for the NFkB pathway in multiple myeloma. Oncotarget 2010, 1, 59–68.
- 51. Kane, S. Fecal lactoferrin is a sensitive and specific marker in identifying intestinal inflammation. Am. J. Gastroenterol. 2003, 98, 1309–1314.
- 52. Suleiman, S.; Sonnenberg, A. Cost-effectiveness of endoscopy in irritable bowel syndrome. Arch. Intern. Med. 2001, 161, 369–375.
- Lacy, B.E.; Pimentel, M.; Brenner, D.M.; Chey, W.D.; Keefer, L.A.; Long, M.D.; Moshiree, B. ACG Clinical Guideline: Management of Irritable Bowel Syndrome. Am. J. Gastroenterol. 2021, 116, 17–44.
- 54. Thakkar, K.; El-Serag, H.B.; Mattek, N.; Gilger, M. Complications of pediatric colonoscopy: A fiveyear multicenter experience. Clin. Gastroenterol. Hepatol. 2008, 6, 515–520.
- 55. Ricciuto, A.; Griffiths, A.M. Clinical value of fecal calprotectin. Crit. Rev. Clin. Lab. Sci. 2019, 56, 307–320.
- 56. Fu, Y.; Wang, L.; Xie, C.; Zou, K.; Tu, L.; Yan, W.; Hou, X. Comparison of non-invasive biomarkers faecal BAFF, calprotectin and FOBT in discriminating IBS from IBD and evaluation of intestinal inflammation. Sci. Rep. 2017, 7, 2669.
- 57. Hussein, H.A.; Mohamed, R.S. Fecal B-cell-activating factor as a new noninvasive marker in the evaluation of ulcerative colitis Egyptian patients: A comparative cross-sectional study. Egypt J. Intern. Med. 2019, 31, 563–572.
- 58. Xie, C.; Quan, R.; Wang, L.; Chen, C.; Yan, W.; Fu, Y. Diagnostic value of fecal B cell activating factor in patients with abdominal discomfort. Clin. Exp. Immunol. 2019, 198, 131–140.
- Gisbert, J.P.; Bermejo, F.; Pérez-Calle, J.L.; Taxonera, C.; Vera, I.; McNicholl, A.G.; Algaba, A.; López, P.; López-Palacios, N.; Calvo, M.; et al. Fecal calprotectin and lactoferrin for the prediction of inflammatory bowel disease relapse. Inflamm. Bowel Dis. 2009, 15, 1190–1198.
- 60. Singh, H.K.; Ee, L.C. Recurrent Abdominal Pain in Children: Is Colonoscopy Indicated? J. Pediatr. Gastroenterol. Nutr. 2019, 68, 214–217.
- Aggarwal, V.; Day, A.S.; Connor, S.; Leach, S.T.; Brown, G.; Singh, R.; Friedman, A.; Zekry, A.; Craig, P.I. Role of capsule endoscopy and fecal biomarkers in small-bowel Crohn's disease to assess remission and predict relapse. Gastrointest. Endosc. 2017, 86, 1070–1078.
- Kopylov, U.; Yablecovitch, D.; Lahat, A.; Neuman, S.; Levhar, N.; Greener, T.; Klang, E.; Rozendorn, N.; Amitai, M.M.; Ben-Horin, S.; et al. Detection of small bowel mucosal healing and deep remission in patients with known small bowel Crohn's disease using biomarkers, capsule endoscopy, and imaging. Am. J. Gastroenterol. 2015, 110, 1316–1323.

- 63. El-Matary, W.; Abej, E.; Deora, V.; Singh, H.; Bernstein, C.N. Impact of fecal calprotectin measurement on decision-making in children with inflammatory bowel disease. Front. Pediatr. 2017, 5, 7.
- 64. Fodor, I.; Serban, O.; Serban, D.E.; Farcau, D.; Man, S.C.; Dumitrascu, D.L. B cell-activating factor (BAFF) in children with inflammatory bowel disease. Pediatr. Res. 2021, 89, 1798–1803.

Retrieved from https://encyclopedia.pub/entry/history/show/44649