### γδ T Cells in Hepatocellular Carcinoma Immunotherapy

#### Subjects: Gastroenterology & Hepatology

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Hepatocellular carcinoma (HCC) remains a global health challenge with limited treatment options and a poor prognosis for advanced-stage patients. Recent advancements in cancer immunotherapy have generated significant interest in exploring novel approaches to combat HCC. One such approach involves the unique and versatile subset of T cells known as  $\gamma\delta$  T cells.  $\gamma\delta$  T cells represent a distinct subset of T lymphocytes that differ from conventional  $\alpha\beta$  T cells in terms of antigen recognition and effector functions. They play a crucial role in immunosurveillance against various malignancies, including HCC.

Keywords: yδ T cells ; hepatocellular carcinoma (HCC) ; immunotherapy

#### 1. Introduction

#### 1.1. Hepatocellular Carcinoma: Understanding, Challenges, and Therapeutic Horizons

In 2020, approximately 906,000 individuals worldwide received a diagnosis of liver cancer, mainly hepatocellular carcinoma (HCC), ranking it as the third leading cause of cancer-related deaths <sup>[1][2]</sup>, accompanied by a 5-year survival rate of merely 18% [3]. HCC mostly affects individuals aged 60-70 years and is more prevalent in men. The causative factors show regional and ethnic variations, mainly attributed to distinct risk factors. Chronic liver diseases arising from hepatitis B (HBV) or C (HCV), alcohol abuse, fatty liver diseases, obesity, diabetes, and certain rare conditions contribute significantly to its development <sup>[4]</sup>. Predominantly, 80% of cases occur in sub-Saharan Africa and eastern Asia, where chronic hepatitis B and exposure to aflatoxin B1 are primary risk factors. For hepatitis B patients, the incidence correlates with viral load, infection duration, and liver disease severity. Occult hepatitis B virus infection also heightens the risk due to DNA damage induced by virus integration <sup>[5]</sup>. In the USA, Europe, and Japan, hepatitis C, coupled with excessive alcohol intake, stands as the primary risk factor. The epidemiology of hepatocellular carcinoma exhibits dynamic temporal trends, with Japan and Europe experiencing plateauing or declining incidence, contrasting with the ongoing increase in the USA, predicted to stabilize by 2020 <sup>[5]</sup>. Concurrently, there is a noticeable rise in the incidence of metabolic dysfunctionassociated fatty liver disease (MAFLD)/metabolic dysfunction-associated steatohepatitis (MASH)-related liver cancers, now standing as the most prevalent chronic liver diseases globally, impacting approximately 25% of the population <sup>[6]</sup>. Lifestyle factors, such as obesity, diabetes, alcohol consumption, and genetic variants (PNPLA3, TM6SF2, HSD17B13), are associated with an increased risk of HCC, even in the absence of cirrhosis <sup>[I]</sup>. Notably, enhancing glycemic control and maintaining a healthy weight are independently linked to a reduced HCC risk <sup>[B]</sup>. Interestingly, while alcohol-related liver disease is more common in men, women face a higher relative risk of developing HCC <sup>[9]</sup>. Epigenetic mechanisms significantly influence the progression of HCC [10].

#### 1.2. Decoding yo T Cell Diversity: Bridging Innate and Adaptive Immunity

Lymphocytes in the vertebrate adaptive immune system utilize somatic recombination to generate diverse antigen receptor repertoires, enabling recognition of a wide range of pathogens. B cells,  $\alpha\beta$  T cells, and  $\gamma\delta$  T cells, evolving over approximately 500 million years, are crucial components of this system <sup>[11]</sup>. While research on T cells has traditionally focused on the  $\alpha\beta$  compartment, groundbreaking studies revealed the existence of a separate lineage,  $\gamma\delta$  T lymphocytes, distinguished by a distinct somatically recombined  $\gamma\delta$  T cell receptor (TCR) and non-MHC restriction <sup>[12]</sup>. Despite proposing diverse roles in immunity, such as protection against pathogens, tumor immunosurveillance, and the maintenance of epithelial surfaces, the functions and antigen recognition requirements of  $\gamma\delta$  T cells remain unclear <sup>[13]</sup>.

Regarding human peripheral  $\gamma\delta$  T cells, two major subsets, innate-like V $\gamma$ 9V $\delta$ 2 T cells and adaptive-like subsets, exhibit distinct immunobiologies <sup>[12]</sup>. Innate-like V $\gamma$ 9V $\delta$ 2 T cells, predominant in peripheral blood (PB), undergo a phenotypic transition from rare and naïve in cord blood to mature effector cells with CD45RO expression in early childhood <sup>[14]</sup>. They respond universally to pathogen-derived phosphoantigens (pAgs), including microbial-derived 4-hydroxy-3-methyl-but-2-

enyl pyrophosphate (HMBPP), in a TCR-dependent manner <sup>[15]</sup>. This subset displays a semi-invariant TCR repertoire, resembling innate lymphocytes, such as i-NKT and mucosal-associated invariant T (MAIT) cells.

On the other hand, adaptive-like subsets, such as  $V\delta 2^{neg} \gamma \delta T$  cells and  $V\gamma 9^{neg}V\delta 2 T$  cells, constitute a minority in PB <sup>[11]</sup>.  $V\delta 2^{neg} T$  cells, enriched in solid tissues, respond to viral infections and display clonal expansions, exhibiting an adaptive-like immunobiology <sup>[12]</sup>. These cells undergo clonal expansions in response to various infections, including CMV, suggesting an adaptive response <sup>[16]</sup>.  $V\gamma 9^{neg}V\delta 2 T$  cells, unable to respond to pAgs, adopt a T<sub>naive</sub> phenotype, but in rare instances, they can transition to a T<sub>effector</sub> phenotype with clonal expansion upon stimulation <sup>[11]</sup>.

In summary, recent studies reveal a complex landscape of  $\gamma\delta$  T cell subsets with distinct immunobiologies, combining features of both innate and adaptive immunity <sup>[127]</sup>. The transition from naïve to effector status in these subsets appears linked to environmental exposures, particularly infections, resembling conventional adaptive immune responses <sup>[11]</sup>.  $\gamma\delta$  T cells possess unique characteristics that make them well suited for recognizing and responding to transformed cells, including HCC.  $\gamma\delta$  T cells exhibit distinct advantages over conventional  $\alpha\beta$  T cells. Unlike  $\alpha\beta$  T cells that mainly recognize peptide antigens presented by major histocompatibility complexes (MHC),  $\gamma\delta$  T cells recognize a diverse array of nonpeptide antigens, including stress-induced molecules and phosphorylated metabolites.  $\gamma\delta$  T cells provide a rapid response to antigens, acting as a frontline defense in epithelial tissues and mucosa. Their innate-like immune functions enable immediate responses without prior sensitization, contributing to immune surveillance against infections and tumors. Importantly,  $\gamma\delta$  T cells can recognize antigens independently of MHC presentation, broadening their capacity to detect various stress-related molecules. In contrast,  $\alpha\beta$  T cells primarily function as adaptive immune cells, requiring specific antigen recognition and prior activation <sup>[18]</sup>.

#### 1.3. yo T Cells: Dynamic Modulators of Inflammation in Chronic Liver Diseases

 $y\delta$  T cells play a crucial role in the establishment and regulation of inflammatory processes, with their involvement varying based on disease etiology and specific subsets <sup>[19]</sup>. The involvement of  $y\delta$  T cells in various inflammatory processes is underscored by several studies, with their role dependent on disease etiology and the specific subset engaged. Notably, Vδ1 and Vy9Vδ2 T cells can adopt a CD103<sup>+</sup> liver tissue-resident memory (TRM) phenotype, exhibiting higher levels of C-X-C chemokine receptor (CXCR) 6 and CXCR3 compared to circulating  $y\delta$  T cells in HCC patients <sup>[18]</sup>. In acute viral HBV,  $y\delta$  T cells are depleted in blood but accumulate in the liver, potentially driven by CXCR3 and C-C chemokine receptor (CCR)5 <sup>[20]</sup>. In chronic HCV infection, V $\delta$ 1 T cells are enriched in the liver, correlating with increased liver inflammation <sup>[21]</sup>. In models of acute liver injury,  $y\delta$  T cells contribute to pro-fibrotic processes through IL-17A production <sup>[22]</sup>. However, in humans,  $y\delta$  T cells are more likely to exhibit type 1 effector functions, producing interferon (IFN)-y and engaging in tumor-killing activities <sup>[23]</sup>.

In liver transplantation for chronic liver disease,  $Vy9^+V\delta2^+$  T cells decrease in circulation and liver sinusoids, particularly in acute-on-chronic liver failure [24]. Chronically activated, these cells exhibit reduced antibacterial effector functions in response to microbial metabolites, possibly contributing to bacterial infections.

In decompensated cirrhosis, bacterial translocation in the peritoneal cavity leads to a comparable fraction of  $\gamma\delta$  T cells as in the blood, but with a TRM phenotype <sup>[25][26]</sup>. This phenotype is associated with enhanced local inflammation in response to a bacterial infection. The detection of both migratory and resident  $\gamma\delta$  T cell populations suggest a possible redistribution between the gut, liver, circulation, and peritoneum <sup>[26]</sup>.

# 2. Exploring Epigenetic Regulation of $y\delta$ T Cells: Insights into Development, Function, and Therapeutic Strategies in Cancer Immunotherapy

Epigenetic regulation plays a crucial role in the process of carcinogenesis  $^{[27][28][29][30][31]}$ . γδ T cells, positioned at the interface of innate and adaptive immunity, exhibit a unique repertoire that necessitates in-depth investigation. As our comprehension of the epigenetic landscape evolves, the analysis of the molecular pathways regulating the behavior of γδ T cells becomes invaluable for the comprehension of immune regulation  $^{[32]}$ . For example, the utilization of single-cell RNA-sequencing technology and immune functional assays provided evidence that bacille Calmette –Guerin (BCG) induces changes in the epigenetic transcriptional programs of γδ T cells at the chromatin level  $^{[33]}$  and at histone H3 acetylation at lysine 27 (H3K27ac)  $^{[34]}$ , enhancing their responsiveness to heterologous bacterial and fungal stimuli, such as lipopolysaccharides (LPS) and Candida albicans  $^{[35]}$ , characterized by a higher production of TNF and IFN-γ several weeks after the vaccination  $^{[35]}$ . Taking a step further, distinct and temporally restricted epigenetic mechanisms regulate the development of TCRαβ and TCRγδ T cells  $^{[36]}$ . Chromatin accessibility dynamics exhibit stage specificity, with T cell lineage commitment marked by the GATA binding protein 3 (GATA3)- and B-cell lymphoma/leukemia 11B (BCL11B)-

dependent closing of PU.1 sites. Notably, β-selection is characterized by a temporary increase in histone H3 lysine 27 (H3K27me3) without open chromatin modifications. Emerging  $\gamma\delta$  T cells, originating from common precursors of  $\beta$ selected cells, exhibit significant chromatin accessibility changes due to strong TCR signaling [36]. Chromatin modifiers, particularly histone-modifying factors, play a significant role in the differentiation of  $y\delta$  T cells [37]. Specifically, an upregulation of chromatin modifiers is evident during the differentiation trajectories of yo T cell subsets (yo T17 and IFN-yproducing T cells). The late activated molecules during effector differentiation, particularly in the γδ T17 lineage, showed an increased expression of histone-modifying factors, such as histone-lysine N-methyltransferase (Kmt)2a, Kmt2c, lysine demethylase (Kdm)5a, Kdm5b, and the histone deacetylase (Hdac)7 [37]. These modifications are involved in shaping the functional characteristics and phenotypic diversity of yo T cell subsets. A growing body of evidence suggests that distinct epigenetic patterns at certain gene loci differentiate the IL-17-producing CD27<sup>-</sup> γδ T cells from the IFN-y-expressing CD27<sup>+</sup> yo T cells [38]. In CD27<sup>-</sup> yo T cells, active H3K4me2 marks were found to accumulate at the II17a, IL17f, and II22 loci, indicating permissive chromatin configurations. However, in γδ CD27<sup>+</sup> T cells, these marks were notably absent at these loci. Conversely, the Ifng locus displayed H3K4me2 marks in both γδ cell subsets, suggesting a shared epigenetic regulation for Ifng in both subsets. An additional investigation into histone modifications revealed that acetylation of histone H3 (H3ac) was exclusive to the *ll17a* locus in  $\gamma\delta$  CD27<sup>-</sup> cells, while both CD27<sup>+</sup> and  $\gamma\delta$  CD27<sup>-</sup> T cells exhibited H3ac marks at the Ifng locus. These epigenetic modifications were associated with differential cytokine gene expression levels. II17a was highly expressed in yo CD27<sup>-</sup> T cells, while Ifng showed higher expression in CD27<sup>+</sup> yo T cells, consistent with the epigenetic patterns observed [38].

The combination of immune checkpoint blockade (ICB) and epigenetic modifiers significantly enhances  $\gamma\delta$  T cell functions, providing a promising strategy for improving clinical outcomes in immunotherapy <sup>[39]</sup>. Using 3D melanoma models, it is observed that  $\gamma\delta$  T cells infiltrated rapidly but exhibited exhausted phenotypes, limiting tumor killing. ICB enhances  $\gamma\delta$  T cell killing, and epigenetic modifiers (Entinostat, Vorinostat) improve functions by regulating MHC class I chain-related protein A (MICA)/ MHC class I chain-related protein B (MICB) and programmed death-ligand 1 (PD-L1).  $\gamma\delta$  T cells, serving as innate immune sentinels, infiltrate melanoma spheroids effectively, with early IFN- $\gamma$  production and enhanced effector functions. However, they display exhausted phenotypes. Combining  $\gamma\delta$  T cells with anti-programmed cell death protein 1 (PD1)/cytotoxic T lymphocyte associated protein 4 (CTLA-4) monoclonal antibodies (mAbs) enhanced functions and reduced spheroid size. Epigenetic modifiers such as Tubastatin-A, Ricolinostat, and Vorinostat inhibit HDAC6/7 pathways, improving  $\gamma\delta$  T cell antitumor functions. These modifiers increase MICA/MICB expression and decrease NKG2A expression in tumor cells.

## 3. Insights into the Utilization of $\gamma\delta$ T Cells in HCC Immunotherapy Clinical Evidence

HCC treatment necessitates innovative immunotherapeutic strategies. The HCC tumor microenvironment (TME) is acknowledged for its immunosuppressive characteristics, attributed to various regulatory factors, such as a low pH, hypoxia, a nutritional deficiency, metabolic remodeling, and inflammation <sup>[40]</sup>.  $\gamma\delta$  T cells have emerged as potential assets in this landscape, and their role is currently under intense investigation in patients with chronic liver disease <sup>[25]</sup>.

HCC transcriptomic data reveal a significant upregulation of inhibitory checkpoint molecules in HCC tissues compared to normal tissues [41]. Notably, these molecules, including CTLA4, hepatitis A virus cellular receptor 2 (HAVCR2), lymphocyte-activation gene 3 (LAG3), programmed cell death protein 1 (PDCD1), programmed cell death 1 ligand 2 (PDCD1LG2), T cell immunoreceptor with immunoglobulin (Ig) and immunoreceptor tyrosine-based inhibitory motif (ITIM) domains (TIGIT), and sialic acid-binding Ig-like lectin 15 (SIGLEC15), show consistent expression across different patient pTNM stages, suggesting a uniform responsiveness to ICB therapy. Hu et al. uncovered a distinct immunosuppressive profile in the HCC TME. This is characterized by increased infiltration of regulatory T cells (Tregs), activated mast cells, and M0 macrophages, coupled with the suppression of  $v\delta$  T cell infiltration and hindrance in the differentiation of macrophages from M0 to M2. Strikingly, these immune cell infiltrations exhibited no statistical differences among different pTNM stages, affirmin g the highly immunosuppressive nature of the HCC TME across various disease stages. The authors further extended their investigation to the phenotypic profiles of circulating  $\gamma\delta$  T cells in HCC and healthy populations [41]. The analysis reveals a dominance of the V $\delta 2^+$  subset in the PB of healthy individuals, while under immune-suppressed conditions in HCC, the  $V\delta 2^+$  subset is significantly depleted. The proposed  $V\delta 1^+/V\delta 2^+$  ratio emerges as a potential indicator for clinical prognosis, with higher ratios associating with more suppressed immunity and poorer outcomes. The study also identified reduced NKG2D expression in the V $\delta$ 1<sup>+</sup> population, suggesting depressed cell activation and enhanced survival ability, while a higher pPD1 expression in the  $V\delta 2^+$  population indicated suppressed cytotoxicity [41].

Employing the CIBERSORT algorithm and weighted gene co-expression network analysis (WGCNA), they identified  $\gamma\delta$  T cell-specific genes associated with immunological activity and notably T cell activation. A risk signature comprising 11 hub genes demonstrated robust prognostic capabilities. This signature exhibited correlations with clinical features and tumor grade and stage, providing valuable prognostic insights. High-risk HCC samples exhibited an immune-activated phenotype and increased susceptibility to ICB pathways. The risk signature demonstrated associations with chemotherapy drug sensitivity, suggesting its potential utility in guiding treatment strategies. They also investigated the role of the Rieske Fe-S domain-containing (*RFESD*) gene in HCC, identifying it as a poor prognosis predictor and correlating it with immune cell infiltration and HCC cell line proliferation. Collectively, they underscored the significance of immune infiltration of  $\gamma\delta$  T cells in the context of HCC, while the constructed risk signature may serve as a valuable tool for risk stratification and treatment decision making in HCC <sup>[42]</sup>.

Sun et al. explored the immune characteristics of hepatitis B virus-related hepatocellular carcinoma (HBV-HCC) and focused on the changes in peripheral immunity in patients with HBV-HCC <sup>[43]</sup>. They documented that the expression of NKp46, a marker associated with cytotoxicity, is up-regulated in the PB of HBV-HCC patients. Additionally, they investigated  $\gamma\delta$  T cells, highlighting an increase in PD-1 expression on Vd1 cells in HBV-HCC patients. They suggested a correlation between T cell Ig and mucin domain 3 (TIM3) expression on  $\gamma\delta$  T cells and disease progression, with increased TIM3<sup>+</sup>  $\gamma\delta$  T cells in the PB of advanced HBV-HCC patients. This highlights the need for further exploration of the liver immune microenvironment in HBV-HCC patients, while single-cell RNA sequencing (scRNA-seq) could serve towards a more comprehensive understanding of the pathogenesis and treatment strategies of HCC <sup>[43]</sup>.

#### 4. The Role of $y\delta$ T Cell in the Immune Landscape of HCC

#### 4.1. y $\delta$ T Cell Features and Interactions in the HCC TME

Research has highlighted unique characteristics of  $\gamma\delta$  T cells in the HCC TME. These features determine the interactions between  $\gamma\delta$  T cells and neoplastic cell and the distribution of different  $\gamma\delta$  T cells between the PB and the liver tissue, thus affecting antitumor immunity.

Toutirais investigated the interaction between Vy9Vδ2 T cells and HCC cells, focusing on the role of DNAM-1 and CD96, two activating NK receptors expressed by Vy9Vδ2 T cells <sup>[44]</sup>. The ligands for these receptors, Nectin-like-5, Nectin-2, and CD96, were found on all analyzed HCC cell lines. Through mAb-mediated masking experiments, they demonstrated that DNAM-1 plays a crucial role in the cytotoxicity against HCC cells and IFN-y production in Vy9Vδ2 T cells. Specifically, Nectin-like-5, but not Nectin-2, was identified as the ligand responsible for DNAM-1-dependent functions in Vy9Vδ2 T cells, CD96, however, did not appear to contribute to the killing of HCC cells. Moreover, they revealed that DNAX accessory molecule-1 (DNAM-1) and NKG2D could cooperate in the cell lysis of HCC, providing insights into potential therapeutic targets for antitumor immunotherapy involving  $\gamma\delta$  T cells <sup>[44]</sup>. Given the liver's abundance of tissue-resident  $\gamma\delta$ T cells and the established efficacy of allogeneic  $V\delta 2^+ \gamma \delta$  T cell adoptive transfer in liver cancer control, Hu et al. investigated the mechanisms underlying  $\gamma\delta$  T cell repression in the HCC TME <sup>[41]</sup>. Analyzing circulating  $\gamma\delta$  T cells in healthy individuals and HCC patients, they observed a significant imbalance in subset proportions. Specifically, the  $V\delta 1^+$ subset exhibited a substantial increase, while the  $V\delta 2^+$  subset showed a marked reduction in the HCC population, resulting in a noteworthy elevation of the  $V\delta 1^+/V\delta 2^+$  ratio. Furthermore, they revealed a significant decrease in NKG2D expression within the Vd1<sup>+</sup> subset, suggesting a potential impairment in antitumor immunity. In contrast, the V $\delta$ 2<sup>+</sup> subset in the HCC group displayed a notable increase in PD1 expression, indicative of compromised cytotoxicity in V $\delta 2^+$  y $\delta T$ cells.

#### 4.2. y $\delta$ T Cell Reshaping the HCC Tumor Microenvironment

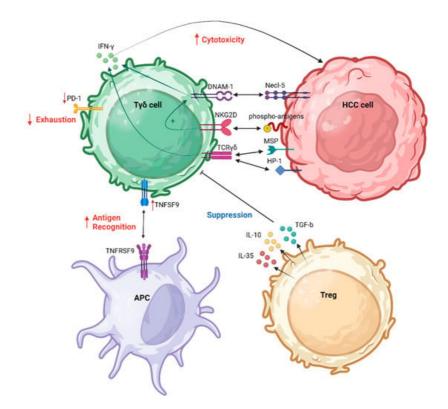
The immune TME plays a critical role in the dynamics of HCC, influencing its progression and response to treatment <sup>[45]</sup>. Comprising a complex interplay of immune cells, stromal elements, and signaling molecules, the TME significantly impacts the trajectory of HCC <sup>[46][47]</sup>. In this context, the immune TME often takes on an immunosuppressive profile, fostering conditions conducive to tumor growth and immune evasion <sup>[48][49]</sup>.

Several T cell populations comprise the immune HCC TME <sup>[50]</sup>. Within the tumor, an increased presence of cells expressing CD137, and inducible T cell co-stimulator (ICOS) suggested recent antigenic activation. Notably, distinct T cell populations were identified, such as functionally impaired proliferating CD4<sup>+</sup> cells co-expressing ICOS and TIGIT, functionally active CD8<sup>+</sup> cells co-expressing CD38 and PD1, and CD4-CD8 double-negative T cell receptor  $\alpha\beta$  and  $\gamma\delta$  cells [non-major histocompatibility complex (MHC)-restricted T cells)]. Histologic classification correlated with the accumulation of activated T cells, indicating an immune-inflamed HCC phenotype <sup>[50]</sup>. Recent findings indicate a synergistic interaction between  $\gamma\delta$  T cells and NK cells within the HCC TME <sup>[44][51][52]</sup>.  $\gamma\delta$  T cells exhibit a capacity to

recognize human tumor cells in a non-restricted MHC manner. The mechanisms for malignant cell recognition by  $\gamma\delta$  T cells involve both TCR-dependent pathways and activation through NK receptors <sup>[44]</sup>. Toutirais et al. explored the role of two NK receptors, DNAM-1 and CD96, in the lytic function of  $\gamma\delta$  T cells, as was analyzed previously. These specific interactions between DNAM-1 and Necl-5 on HCC cells was highlighted as a novel regulatory mechanism for  $\gamma\delta$  T cell cytotoxicity, enhancing IFN- $\gamma$  production. Moreover, the authors suggested a cooperative role of DNAM-1 and *NKG2D* receptors in triggering the cytotoxic effector function of  $\gamma\delta$  T cells <sup>[44]</sup>.

Numerous molecules play pivotal roles in shaping the immune TME of HCC, impacting the abundance and functionality of  $\gamma\delta$  T cells. Emerging evidence suggests that transforming growth factor (TGF)-b1 is associated with a poor prognosis in HCC <sup>[53]</sup>. Specifically, seven members of the TGF-b family, including bone morphogenetic protein (BMP)2, BMP6, growth differentiation factor (GDF)6, GDF7, GDF10, left-right determination factor 2 (LEFTY2), and TGF-b1, were identified as potential independent prognostic factors for HCC.

**Figure 1** summarizes the interactions between  $\gamma\delta$  T cells and HCC cells as well as with other immune cells responsible for the regulation of  $\gamma\delta$  T cell functions in the HCC TME.



**Figure 1.** γδ T cell interaction in HCC TME. APC, antigen-presenting cell; DNAM-1, DNAX accessory molecule-1; HCC, hepatocellular carcinoma; MSP, macrophage-stimulating protein; *NKG2D*, natural killer group 2D; PD-1, programmed cell death protein 1; TCR, T cell receptor; TNFSF9, tumor necrosis factor (TNF) superfamily member 9; Treg, regulatory T cell. Created with <u>BioRender.com</u> (accessed on 4 December 2023).

#### 5. γδ T Cells: An Immunotherapeutic Odyssey for Hepatocellular Carcinoma

#### 5.1. Strategies to Enhance yo T Cell Antitumor Efficacy

#### 5.1.1. y $\delta$ T Cells in CAR T Therapies: Targeting HCC Breakthroughs

The development of allogeneic V $\delta$ 1 T cells expressing a glypican-3 (GPC-3)-targeted chimeric antigen receptor (CAR) <sup>[54]</sup> and secreting IL-15 is explored as a potential cell therapy for HCC and other GPC-3-expressing tumors <sup>[55]</sup>. Autologous CAR T cell therapy has shown success in hematologic malignancies, but challenges exist in treating solid tumors due to issues such as poor T cell homing, CAR target antigen variability, and the immunosuppressive TME. GPC-3 is an attractive target for immunotherapy, being highly expressed in HCC <sup>[18][56]</sup>. Autologous GPC-3.CAR  $\alpha\beta$  T cells have shown promise, but logistical challenges and concerns about immune fitness have hindered widespread adoption. Allogeneic cell therapies, derived from healthy donors, present a potential solution by overcoming manufacturing challenges. In fact, recent evidence exploring the safety and efficacy of combining locoregional therapy with the adoptive transfer of allogeneic  $\gamma\delta$  T cells for advanced HCC and intrahepatic cholangiocarcinoma (ICC) demonstrated that adverse events

were manageable, indicating the safety of this novel combination therapy  $^{[57]}$ , as opposed to allogeneic  $\alpha\beta$  T cells that may pose a risk of graft-versus-host disease (GvVHD), necessitating complex gene editing  $^{[58]}$ .

#### 5.1.2. Zoledronic Acid/Artesunate and y $\delta$ T Cells Reshaping HCC Therapeutics

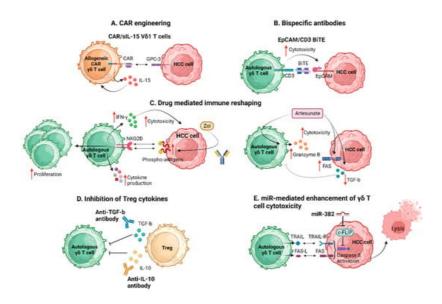
Ongoing research in the field of HCC investigates the interplay between zoledronic acid (Zol) and  $\gamma\delta$  T cells <sup>[19][59][60][61]</sup>. Zol has demonstrated immunomodulatory properties, notably activating  $\gamma\delta$  T cells, which show antitumor capabilities. The exploration of combining Zol with  $\gamma\delta$  T cell therapy emerges as a prospective approach to enhance the immune response against HCC.

Zakeri et al. investigated the potential of Zol in enhancing the antitumor activity of Vy9Vo2 T cells in the context of HCC with a focus on the TRM subset of  $V_{y}9V\delta^2$  T cells found in the human liver <sup>[19]</sup>. They began characterizing the liverresident Vy9Vo2 TRM, highlighting their tissue-homing capabilities through the expression of chemokine receptors CXCR6 and CXCR3. These TRMs displayed a reduced expression of the endothelial homing receptor CX3CR1 and lacked the expression of the lymph node homing receptor CD62L. Functionally, the intrahepatic Vy9Vo2 TRMs exhibited an activated profile with higher expression of the T cell activation marker HLA-DR. However, they showed reduced cytotoxic potential, marked by a lower expression of the serine protease granzyme B and a significant capacity for rapid production of the pro-survival cytokine IL-2 and IFN-y. The authors also explored the antitumor potential of Vy9Vδ2 TRM against Zol-sensitized HCC cell lines. While fresh intrahepatic Vy9Vδ2 T cells exhibited minimal effector function in response to co-culture with HCC cell lines, Zol pre-treatment of the tumor cells significantly enhanced the effector function of CD69<sup>+</sup>CD49a<sup>+</sup> Vy9Vδ2 TRM. This response was further observed with tumor-infiltrating lymphocytes (TILs) isolated from HCC tumors, suggesting the potential for Zol to boost the antitumor function of intratumoral Vy9V $\delta$ 2 T cells <sup>[19]</sup>. The above are further supported by Sugai et al., who demonstrated that Vy9Vδ2 T cells, activated by Zol-induced phosphoantigens, exhibited significant antitumor activity against various HCC cell lines (HepG2, HLE, HLF, HuH-1, JHH5, JHH7, and Li-7). Zol treatment not only increased HCC cell susceptibility to T cell killing but also triggered T cell proliferation and induced cytokine production (IL-4, IL-5, IL-13, IFN-y, GM-CSF, TNF-α) and grandzyme B <sup>[59]</sup>. Towards the same direction, Tian et al. utilized Zol in combination with IL-2 in vitro, concluding that, in conjunction, they could expand circulating yo T cells in HCC patients, enabling them to lyse HCC cells without raising immunosuppressive factors during amplification [61]. Importantly, they investigated immunosuppressive factors, demonstrating that Tregs, yo T17 cells, and IL-17A do not increase during  $y\delta$  T cell amplification, indicating the safety of this approach for HCC immunotherapy [61].

#### 5.1.3. Therapeutic Manipulation of $y\delta$ T Cells in HCC beyond Known Pathways

Yi et al. exploring the immune dynamics within the HCC TME revealed a significant attenuation in the infiltration and cytotoxic function of  $\gamma\delta$  T cells within HCC tissue <sup>[62]</sup>. Importantly, Treg cells within the tumor were found to directly suppress the cytotoxic ability and IFN- $\gamma$  secretion of  $\gamma\delta$  T cells, with this effect being dependent on TGF- $\beta$  and IL-10. They emphasized the novel insight into the distribution, cytotoxic function, and Treg-mediated suppression of  $\gamma\delta$  T cells in the TME. They documented a negative correlation between Treg cells and  $\gamma\delta$  T cells in HCC tissues. The in vitro analysis revealed a partial reversal of Treg cell-mediated suppression by anti-TGF $\beta$  or anti-IL-10 antibodies, suggesting a soluble factor-dependent mechanism in the Treg cell-mediated suppression of  $\gamma\delta$  T cells [62]. In conclusion, they emphasized the importance of enhancing the antitumor effect of  $\gamma\delta$  T cells and eliminating suppressive factors, particularly Treg cells, within the TME. The combination of these approaches is suggested as a potential strategy to improve outcomes for patients with HCC.

Figure 2 summarizes the proposed strategies applied to enhance  $\gamma\delta$  T cell antitumor efficacy in HCC.



**Figure 2.** Strategies to enhance γδ T cell antitumor efficacy. BiTE, bispecific T cell engager; CAR, chimeric antigen receptor; c-FLIP, cellular FADD-like interleukin-1b-converting enzyme-inhibitory protein; EpCAM, epithelial cell adhesion molecule; FAS-L, FAS ligand; GPC-3, glypican-3; HCC, hepatocellular carcinoma; IFN-γ, interferon-γ; IL, interleukin; miR, micro RNA; *NKG2D*, natural killer group 2D; sIL-15, soluble interleukin 15; TGF-b, transforming growth factor b; TRAIL, TNF-related apoptosis-inducing ligand; Zol, zolendronic acid. Created with <u>BioRender.com</u> (accessed on 4 December 2023).

#### 5.2. T Cell Exhaustion in y $\delta$ T Cells within HCC

In HCC, T cell exhaustion is a complex process influenced by various factors. The liver is an immunologically unique organ, and HCC often develops in the setting of chronic liver inflammation, commonly caused by viral hepatitis or cirrhosis <sup>[63][64]</sup>. The HCC TME is widely acknowledged for its immunosuppressive nature, influenced by various regulatory factors, such as low pH, hypoxia, nutritional deficiencies, and metabolic pathway alterations <sup>[65][66]</sup>. The prolonged exposure to antigens in the TME, along with the immunosuppressive signals produced by both the tumor and the liver environment, can lead to T cell exhaustion <sup>[67]</sup>. Transcriptomic analysis of HCC tissue indicates significantly elevated expression of genes encoding for inhibitory checkpoint molecules, including *CTLA4*, *HAVCR2*, *LAG3*, *PDCD1*, *PDCD1LG2*, *TIGIT*, and *SIGLEC15*, irrespective of patient pTNM stages <sup>[41]</sup>.

More recently, increased evidence highlights the role of the PD-1/PD-L1 axis in the context of  $\gamma\delta$  T cell function in HCC pathogenesis <sup>[68][69][70][71]</sup>. Jiang et al. documented that circulating  $\gamma\delta$  T cells from HCC patients exhibit impaired cytotoxicity compared to healthy controls, a finding attributed to a lower frequency of V $\delta$ 2 T cells and reduced IL-2 and IL-21 expression <sup>[69]</sup>. In vitro expansion with Zol enhanced cytotoxicity but did not fully restore it. PD-1 expression was elevated in HCC patients, and a co-culture with  $\gamma\delta$  T cells increased PD-L1 in HCC cell lines. Blocking PD-1 during the expansion improved cytotoxicity against all HCC lines, while blocking during assays enhanced it specifically against HepG2 and SNU-398, indicating a connection between reduced cytotoxicity and altered IL-2, IL-21, and PD-1 expression in circulating  $\gamma\delta$  T cells of HCC patients <sup>[69]</sup>.

LAG-3 stands out as one of the primarily studied indicators of T cell exhaustion  $^{[T1][T2]]}$ . He et al. investigated the functional state of  $\gamma\delta$  T cells infiltrating HCC and explored the potential of allogenic  $V\delta2^+ \gamma\delta$  T cells in HCC immunotherapy. scRNA-seq on  $\gamma\delta$  T cells from HCC tumors and healthy donor livers revealed that  $\gamma\delta$  T cells in the HCC TME exhibit G2/M cell cycle arrest, express cytotoxic molecules, and show functional exhaustion with elevated LAG3 expression. The LAG3<sup>+</sup>V $\delta1^+$  population dominated, while  $V\delta2^+ \gamma\delta$  T cells were depleted in the HCC TME. Glutamine metabolism was upregulated in  $\gamma\delta$  T cells due to glutamine deficiency in the TME, resulting in increased LAG3 expression. Ex vivo-expanded  $V\delta2^+ \gamma\delta$  T cells from healthy donors complement the loss of TCRclonality and effector functions in HCC-derived  $\gamma\delta$  T cells  $^{[T2]}$ . They provided insight into the dysfunctional signatures of HCC-infiltrating  $\gamma\delta$  T cells and supported the potential use of allogenic  $V\delta2^+ \gamma\delta$  T cells in HCC cellular therapy

Conclusively, several mechanisms contributing to exhaustion in  $\gamma\delta$  T cells have been identified. These multiple mechanisms collectively contribute to the reduced cytotoxic capacity and functional exhaustion of  $\gamma\delta$  T cells in HCC. The reversal of the identified mechanisms contributing to  $\gamma\delta$  T cell exhaustion in HCC presents a unique and promising therapeutic target.

#### References

- Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J. Clin. 2021, 71, 209–249.
- Vogel, A.; Meyer, T.; Sapisochin, G.; Salem, R.; Saborowski, A. Hepatocellular Carcinoma. Lancet 2022, 400, 1345– 1362.
- 3. Allaire, M.; Bruix, J.; Korenjak, M.; Manes, S.; Maravic, Z.; Reeves, H.; Salem, R.; Sangro, B.; Sherman, M. What to Do about Hepatocellular Carcinoma: Recommendations for Health Authorities from the International Liver Cancer Association. JHEP Rep. 2022, 4, 100578.
- 4. Forner, A.; Reig, M.; Bruix, J. Hepatocellular Carcinoma. Lancet 2018, 391, 1301–1314.
- 5. Younossi, Z.; Stepanova, M.; Ong, J.P.; Jacobson, I.M.; Bugianesi, E.; Duseja, A.; Eguchi, Y.; Wong, V.W.; Negro, F.; Yilmaz, Y.; et al. Nonalcoholic Steatohepatitis Is the Fastest Growing Cause of Hepatocellular Carcinoma in Liver Transplant Candidates. Clin. Gastroenterol. Hepatol. 2019, 17, 748–755.e3.
- 6. Lekakis, V.; Papatheodoridis, G.V. Natural History of Metabolic Dysfunction-Associated Steatotic Liver Disease. Eur. J. Intern. Med. 2023. ahead of print.
- 7. Younossi, Z.M.; Koenig, A.B.; Abdelatif, D.; Fazel, Y.; Henry, L.; Wymer, M. Global Epidemiology of Nonalcoholic Fatty Liver Disease—Meta-analytic Assessment of Prevalence, Incidence, and Outcomes. Hepatology 2016, 64, 73–84.
- Liu, Y.; Zhang, X.; Chen, S.; Wang, J.; Yu, S.; Li, Y.; Xu, M.; Aboubacar, H.; Li, J.; Shan, T.; et al. Gut-Derived Lipopolysaccharide Promotes Alcoholic Hepatosteatosis and Subsequent Hepatocellular Carcinoma by Stimulating Neutrophil Extracellular Traps through Toll-like Receptor 4. Clin. Mol. Hepatol. 2022, 28, 522–539.
- 9. Raghav, P.K.; Mann, Z. Cancer Stem Cells Targets and Combined Therapies to Prevent Cancer Recurrence. Life Sci. 2021, 277, 119465.
- Raghav, P.K.; Kumar, R.; Kumar, V.; Raghava, G.P.S. Docking-based Approach for Identification of Mutations That Disrupt Binding between Bcl-2 and Bax Proteins: Inducing Apoptosis in Cancer Cells. Mol. Genet. Genom. Med. 2019, 7, e910.
- Vantourout, P.; Hayday, A. Six-of-the-Best: Unique Contributions of Γδ T Cells to Immunology. Nat. Rev. Immunol. 2013, 13, 88–100.
- Di Marco Barros, R.; Roberts, N.A.; Dart, R.J.; Vantourout, P.; Jandke, A.; Nussbaumer, O.; Deban, L.; Cipolat, S.; Hart, R.; Iannitto, M.L.; et al. Epithelia Use Butyrophilin-like Molecules to Shape Organ-Specific Γδ T Cell Compartments. Cell 2016, 167, 203–218.e17.
- Davey, M.S.; Willcox, C.R.; Hunter, S.; Kasatskaya, S.A.; Remmerswaal, E.B.M.; Salim, M.; Mohammed, F.; Bemelman, F.J.; Chudakov, D.M.; Oo, Y.H.; et al. The Human Vδ2+ T-Cell Compartment Comprises Distinct Innate-like Vy9+ and Adaptive Vy9-Subsets. Nat. Commun. 2018, 9, 1760.
- Morita, C.T.; Jin, C.; Sarikonda, G.; Wang, H. Nonpeptide Antigens, Presentation Mechanisms, and Immunological Memory of Human Vγ2Vδ2 T Cells: Discriminating Friend from Foe through the Recognition of Prenyl Pyrophosphate Antigens. Immunol. Rev. 2007, 215, 59–76.
- Pitard, V.; Roumanes, D.; Lafarge, X.; Couzi, L.; Garrigue, I.; Lafon, M.-E.; Merville, P.; Moreau, J.-F.; Déchanet-Merville, J. Long-Term Expansion of Effector/Memory Vδ2– Γδ T Cells Is a Specific Blood Signature of CMV Infection. Blood 2008, 112, 1317–1324.
- 16. Davey, M.S.; Willcox, C.R.; Hunter, S.; Oo, Y.H.; Willcox, B.E. Vδ2+ T Cells-Two Subsets for the Price of One. Front. Immunol. 2018, 9, 2106.
- 17. Makkouk, A.; Yang, X.C.; Barca, T.; Lucas, A.; Turkoz, M.; Wong, J.T.S.; Nishimoto, K.P.; Brodey, M.M.; Tabrizizad, M.; Gundurao, S.R.Y.; et al. Off-the-Shelf Vδ1 Gamma Delta T Cells Engineered with Glypican-3 (GPC-3)-Specific Chimeric Antigen Receptor (CAR) and Soluble IL-15 Display Robust Antitumor Efficacy against Hepatocellular Carcinoma. J. Immunother. Cancer 2021, 9, e003441.
- Jia, Z.-H.; Li, Y.-Y.; Wang, J.-Y.; Zhang, J.-Y.; Huang, A.; Guo, X.-D.; Zhu, Z.-Y.; Wang, F.-S.; Wu, X.-L. Activated Γδ T Cells Exhibit Cytotoxicity and the Capacity for Viral Clearance in Patients with Acute Hepatitis B. Clin. Immunol. 2019, 202, 40–48.
- Zakeri, N.; Hall, A.; Swadling, L.; Pallett, L.J.; Schmidt, N.M.; Diniz, M.O.; Kucykowicz, S.; Amin, O.E.; Gander, A.; Pinzani, M.; et al. Characterisation and Induction of Tissue-Resident Gamma Delta T-Cells to Target Hepatocellular Carcinoma. Nat. Commun. 2022, 13, 1372.

- 20. Agrati, C.; D'Offizi, G.; Narciso, P.; Abrignani, S.; Ippolito, G.; Colizzi, V.; Poccia, F. Vδ1T Lymphocytes Expressing a Th1 Phenotype Are the Major Γδ T Cell Subset Infiltrating the Liver of HCV-Infected Persons. Mol. Med. 2001, 7, 11–19.
- 21. Seo, W.; Eun, H.S.; Kim, S.Y.; Yi, H.; Lee, Y.; Park, S.; Jang, M.; Jo, E.; Kim, S.C.; Han, Y.; et al. Exosome-mediated Activation of Toll-like Receptor 3 in Stellate Cells Stimulates Interleukin-17 Production by Γδ T Cells in Liver Fibrosis. Hepatology 2016, 64, 616–631.
- Ribot, J.C.; Ribeiro, S.T.; Correia, D.V.; Sousa, A.E.; Silva-Santos, B. Human Γδ Thymocytes Are Functionally Immature and Differentiate into Cytotoxic Type 1 Effector T Cells upon IL-2/IL-15 Signaling. J. Immunol. 2014, 192, 2237–2243.
- 23. Hammerich, L.; Bangen, J.M.; Govaere, O.; Zimmermann, H.W.; Gassler, N.; Huss, S.; Liedtke, C.; Prinz, I.; Lira, S.A.; Luedde, T.; et al. Chemokine Receptor CCR6-Dependent Accumulation of Γδ T Cells in Injured Liver Restricts Hepatic Inflammation and Fibrosis. Hepatology 2014, 59, 630–642.
- Ibidapo-obe, O.; Stengel, S.; Köse-Vogel, N.; Quickert, S.; Reuken, P.A.; Busch, M.; Bauer, M.; Stallmach, A.; Bruns, T. Mucosal-Associated Invariant T Cells Redistribute to the Peritoneal Cavity During Spontaneous Bacterial Peritonitis and Contribute to Peritoneal Inflammation. Cell Mol. Gastroenterol. Hepatol. 2020, 9, 661–677.
- 25. Ibidapo-Obe, O.; Bruns, T. Tissue-Resident and Innate-like T Cells in Patients with Advanced Chronic Liver Disease. JHEP Rep. 2023, 5, 100812.
- Queck, A.; Rueschenbaum, S.; Kubesch, A.; Cai, C.; Zeuzem, S.; Weigert, A.; Brüne, B.; Nour-Eldin, N.-E.A.; Gruber-Rouh, T.; Vogl, T.; et al. The Portal Vein as a Distinct Immunological Compartment—A Comprehensive Immune Phenotyping Study. Hum. Immunol. 2018, 79, 716–723.
- 27. Manou, M.; Kanakoglou, D.S.; Loupis, T.; Vrachnos, D.M.; Theocharis, S.; Papavassiliou, A.G.; Piperi, C. Role of Histone Deacetylases in the Pathogenesis of Salivary Gland Tumors and Therapeutic Targeting Options. Int. J. Mol. Sci. 2023, 24, 10038.
- Palamaris, K.; Tzimou, L.-M.; Levidou, G.; Masaoutis, C.; Theochari, I.; Rontogianni, D.; Theocharis, S. Histone Deacetylases (HDACs): Promising Biomarkers and Potential Therapeutic Targets in Thymic Epithelial Tumors. Int. J. Mol. Sci. 2023, 24, 4263.
- 29. Psilopatis, I.; Pergaris, A.; Vrettou, K.; Theocharis, S.; Troungos, C. Thymic Epithelial Neoplasms: Focusing on the Epigenetic Alterations. Int. J. Mol. Sci. 2022, 23, 4045.
- 30. Goutas, D.; Theocharis, S.; Tsourouflis, G. Unraveling the Epigenetic Role and Clinical Impact of Histone Deacetylases in Neoplasia. Diagnostics 2021, 11, 1346.
- Xiong, D.; Zhang, L.; Sun, Z.-J. Targeting the Epigenome to Reinvigorate T Cells for Cancer Immunotherapy. Mil. Med. Res. 2023, 10, 59.
- 32. Arts, R.J.W.; Carvalho, A.; La Rocca, C.; Palma, C.; Rodrigues, F.; Silvestre, R.; Kleinnijenhuis, J.; Lachmandas, E.; Gonçalves, L.G.; Belinha, A.; et al. Immunometabolic Pathways in BCG-Induced Trained Immunity. Cell Rep. 2016, 17, 2562–2571.
- Giamarellos-Bourboulis, E.J.; Tsilika, M.; Moorlag, S.; Antonakos, N.; Kotsaki, A.; Domínguez-Andrés, J.; Kyriazopoulou, E.; Gkavogianni, T.; Adami, M.-E.; Damoraki, G.; et al. Activate: Randomized Clinical Trial of BCG Vaccination against Infection in the Elderly. Cell 2020, 183, 315–323.e9.
- 34. Suen, T.K.; Moorlag, S.J.C.F.M.; Li, W.; de Bree, L.C.J.; Koeken, V.A.C.M.; Mourits, V.P.; Dijkstra, H.; Lemmers, H.; Bhat, J.; Xu, C.-J.; et al. BCG Vaccination Induces Innate Immune Memory in Γδ T Cells in Humans. J. Leukoc. Biol. 2023, 115, 149–163.
- 35. Roels, J.; Kuchmiy, A.; De Decker, M.; Strubbe, S.; Lavaert, M.; Liang, K.L.; Leclercq, G.; Vandekerckhove, B.; Van Nieuwerburgh, F.; Van Vlierberghe, P.; et al. Distinct and Temporary-Restricted Epigenetic Mechanisms Regulate Human Aβ and Γδ T Cell Development. Nat. Immunol. 2020, 21, 1280–1292.
- 36. Sagar; Pokrovskii, M.; Herman, J.S.; Naik, S.; Sock, E.; Zeis, P.; Lausch, U.; Wegner, M.; Tanriver, Y.; Littman, D.R.; et al. Deciphering the Regulatory Landscape of Fetal and Adult Γδ T-cell Development at Single-cell Resolution. EMBO J. 2020, 39, e104159.
- 37. Schmolka, N.; Serre, K.; Grosso, A.R.; Rei, M.; Pennington, D.J.; Gomes, A.Q.; Silva-Santos, B. Epigenetic and Transcriptional Signatures of Stable versus Plastic Differentiation of Proinflammatory Γδ T Cell Subsets. Nat. Immunol. 2013, 14, 1093–1100.
- Fernández-Sánchez, A.; Baragaño Raneros, A.; Carvajal Palao, R.; Sanz, A.B.; Ortiz, A.; Ortega, F.; Suárez-Álvarez, B.; López-Larrea, C. DNA Demethylation and Histone H3K9 Acetylation Determine the Active Transcription of the NKG2D Gene in Human CD8 + T and NK Cells. Epigenetics 2013, 8, 66–78.

- Hu, Y.; Chen, D.; Hong, M.; Liu, J.; Li, Y.; Hao, J.; Lu, L.; Yin, Z.; Wu, Y. Apoptosis, Pyroptosis, and Ferroptosis Conspiringly Induce Immunosuppressive Hepatocellular Carcinoma Microenvironment and Γδ T-Cell Imbalance. Front. Immunol. 2022, 13, 1–14.
- 40. Arvanitakis, K.; Mitroulis, I.; Germanidis, G. Tumor-Associated Neutrophils in Hepatocellular Carcinoma Pathogenesis, Prognosis, and Therapy. Cancers 2021, 13, 2899.
- 41. Wang, J.; Ling, S.; Ni, J.; Wan, Y. Novel Γδ T Cell-Based Prognostic Signature to Estimate Risk and Aid Therapy in Hepatocellular Carcinoma. BMC Cancer 2022, 22, 638.
- 42. Zhao, N.; Dang, H.; Ma, L.; Martin, S.P.; Forgues, M.; Ylaya, K.; Hewitt, S.M.; Wang, X.W. Intratumoral Γδ T-Cell Infiltrates, Chemokine (C-C Motif) Ligand 4/Chemokine (C-C Motif) Ligand 5 Protein Expression and Survival in Patients With Hepatocellular Carcinoma. Hepatology 2021, 73, 1045–1060.
- 43. Cai, X.Y.; Wang, J.X.; Yi, Y.; He, H.W.; Ni, X.C.; Zhou, J.; Cheng, Y.F.; Jin, J.J.; Fan, J.; Qiu, S.J. Low Counts of Γδ T Cells in Peritumoral Liver Tissue Are Related to More Frequent Recurrence in Patients with Hepatocellular Carcinoma after Curative Resection. Asian Pac. J. Cancer Prev. 2014, 15, 775–780.
- Minaei, N.; Ramezankhani, R.; Tamimi, A.; Piryaei, A.; Zarrabi, A.; Aref, A.R.; Mostafavi, E.; Vosough, M. Immunotherapeutic Approaches in Hepatocellular Carcinoma: Building Blocks of Hope in near Future. Eur. J. Cell Biol. 2023, 102, 151284.
- 45. Baglieri, J.; Brenner, D.A.; Kisseleva, T. The Role of Fibrosis and Liver-Associated Fibroblasts in the Pathogenesis of Hepatocellular Carcinoma. Int. J. Mol. Sci. 2019, 20, 1723.
- 46. Papadakos, S.P.; Arvanitakis, K.; Stergiou, I.E.; Lekakis, V.; Davakis, S.; Christodoulou, M.-I.; Germanidis, G.; Theocharis, S. The Role of TLR4 in the Immunotherapy of Hepatocellular Carcinoma: Can We Teach an Old Dog New Tricks? Cancers 2023, 15, 2795.
- Di Blasi, D.; Boldanova, T.; Mori, L.; Terracciano, L.; Heim, M.H.; De Libero, G. Unique T-Cell Populations Define Immune-Inflamed Hepatocellular CarcinomaUnique T-Cell Populations Define Immune-Inflamed Hepatocellular Carcinoma. Cell Mol. Gastroenterol. Hepatol. 2020, 9, 195–218.
- 48. Papadakos, S.P.; Dedes, N.; Kouroumalis, E.; Theocharis, S. The Role of the NLRP3 Inflammasome in HCC Carcinogenesis and Treatment: Harnessing Innate Immunity. Cancers 2022, 14, 3150.
- Papadakos, S.P.; Arvanitakis, K.; Stergiou, I.E.; Vallilas, C.; Sougioultzis, S.; Germanidis, G.; Theocharis, S. Interplay of Extracellular Vesicles and TLR4 Signaling in Hepatocellular Carcinoma Pathophysiology and Therapeutics. Pharmaceutics 2023, 15, 2460.
- 50. Kang, Y.; Han, M.; Kim, M.; Hwang, H.J.; Ahn, B.C.; Tak, E.; Song, G.-W.; Hwang, S.; Koh, K.-N.; Jung, D.-H.; et al. Cytotoxicity of Human Hepatic Intrasinusoidal Gamma/Delta T Cells Depends on Phospho-Antigen and NK Receptor Signaling. Anticancer Res. 2023, 43, 63–73.
- 51. Sun, R.; Li, J.; Lin, X.; Yang, Y.; Liu, B.; Lan, T.; Xiao, S.; Deng, A.; Yin, Z.; Xu, Y.; et al. Peripheral Immune Characteristics of Hepatitis B Virus-Related Hepatocellular Carcinoma. Front. Immunol. 2023, 14, 1–16.
- 52. Xi, X.; Guo, Y.; Zhu, M.; Qiu, F.; Lei, F.; Li, G.; Du, B. Identification of New Potential Antigen Recognized by ΓδT Cells in Hepatocellular Carcinoma. Cancer Immunol. Immunother. 2021, 70, 1917–1927.
- 53. Yi, Y.; He, H.W.; Wang, J.X.; Cai, X.Y.; Li, Y.W.; Zhou, J.; Cheng, Y.F.; Jin, J.J.; Fan, J.; Qiu, S.J. The Functional Impairment of HCC-Infiltrating Γδ T Cells, Partially Mediated by Regulatory T Cells in a TGFβ- and IL-10-Dependent Manner. J. Hepatol. 2013, 58, 977–983.
- Santana Carrero, R.M.; Beceren-Braun, F.; Rivas, S.C.; Hegde, S.M.; Gangadharan, A.; Plote, D.; Pham, G.; Anthony, S.M.; Schluns, K.S. IL-15 Is a Component of the Inflammatory Milieu in the Tumor Microenvironment Promoting Antitumor Responses. Proc. Natl. Acad. Sci. USA 2019, 116, 599–608.
- 55. Guo, M.; Zhang, H.; Zheng, J.; Liu, Y. Glypican-3: A New Target for Diagnosis and Treatment of Hepatocellular Carcinoma. J. Cancer 2020, 11, 2008–2021.
- 56. Zhang, T.; Chen, J.; Niu, L.; Liu, Y.; Ye, G.; Jiang, M.; Qi, Z. Clinical Safety and Efficacy of Locoregional Therapy Combined with Adoptive Transfer of Allogeneic Γδ T Cells for Advanced Hepatocellular Carcinoma and Intrahepatic Cholangiocarcinoma. J. Vasc. Interv. Radiol. 2022, 33, 19–27.e3.
- 57. Townsend, M.H.; Bennion, K.; Robison, R.A.; O'Neill, K.L. Paving the Way towards Universal Treatment with Allogenic T Cells. Immunol. Res. 2020, 68, 63–70.
- 58. Sugai, S.; Yoshikawa, T.; Iwama, T.; Tsuchiya, N.; Ueda, N.; Fujinami, N.; Shimomura, M.; Zhang, R.; Kaneko, S.; Uemura, Y.; et al. Hepatocellular Carcinoma Cell Sensitivity to Vy9Vδ2 T Lymphocyte-Mediated Killing Is Increased by Zoledronate. Int. J. Oncol. 2016, 48, 1794–1804.

- 59. Hoh, A.; Dewerth, A.; Vogt, F.; Wenz, J.; Baeuerle, P.A.; Warmann, S.W. The Activity of Cd T Cells against Paediatric Liver Tumour Cells and Spheroids in Cell Culture. Liver Int. 2012, 33, 127–136.
- 60. Tian, W.E.I.; Ma, J.U.N.; Shi, R.; Ren, C.; He, J.; Zhao, H. Γδ T Cell-Mediated Individualized Immunotherapy for Hepatocellular Carcinoma Considering Clinicopathological Characteristics and Immunosuppressive Factors. Oncol. Lett. 2018, 15, 5433–5442.
- Qian, P.; Zhang, Y.; Zhou, Z.; Liu, J.; Yue, S.; Guo, X.; Sun, L.; Lv, X.; Chen, J. Artesunate Enhances Γδ T-Cell-Mediated Antitumor Activity through Augmenting Γδ T-Cell Function and Reversing Immune Escape of HepG2 Cells. Immunopharmacol. Immunotoxicol. 2018, 40, 107–116.
- 62. Labanieh, L.; Majzner, R.G.; Mackall, C.L. Programming CAR-T Cells to Kill Cancer. Nat. Biomed. Eng. 2018, 2, 377–391.
- 63. Liaskou, E.; Hirschfield, G.M.; Gershwin, M.E. Mechanisms of Tissue Injury in Autoimmune Liver Diseases. Semin. Immunopathol. 2014, 36, 553–568.
- 64. Huang, H.; Tsui, Y.; Ng, I.O. Fueling HCC Dynamics: Interplay Between Tumor. Cell Mol. Gastroenterol. Hepatol. 2023, 15, 1105–1116.
- 65. D'Alessio, A.; Pinato, D.J. Dissecting the Tumor Microenvironment to Predict Immunotherapy Response in Hepatocellular Cancer. Gastroenterology 2022, 163, 1712–1713.
- 66. Nguyen, P.H.D.; Ma, S.; Phua, C.Z.J.; Kaya, N.A.; Lai, H.L.H.; Lim, C.J.; Lim, J.Q.; Wasser, M.; Lai, L.; Tam, W.L.; et al. Intratumoural Immune Heterogeneity as a Hallmark of Tumour Evolution and Progression in Hepatocellular Carcinoma. Nat. Commun. 2021, 12, 227.
- 67. Jiang, H.; Yang, Z.; Song, Z.; Green, M.; Song, H.; Shao, Q. Γδ T Cells in Hepatocellular Carcinoma Patients Present Cytotoxic Activity but Are Reduced in Potency Due to IL-2 and IL-21 Pathways. Int. Immunopharmacol. 2019, 70, 167– 173.
- 68. Jin, X.; Zhang, S.; Wang, N.; Guan, L.; Shao, C.; Lin, Y.; Liu, J.; Li, Y. High Expression of TGF-B1 Contributes to Hepatocellular Carcinoma Prognosis via Regulating Tumor Immunity. Front. Oncol. 2022, 12, 1–11.
- 69. Xu, P.; Sun, Z.; Wang, Y.; Miao, C. Long-Term Use of Indomethacin Leads to Poor Prognoses through Promoting the Expression of PD-1 and PD-L2 via TRIF/NF-KB Pathway and JAK/STAT3 Pathway to Inhibit TNF-α and IFN-γ in Hepatocellular Carcinoma. Exp. Cell Res. 2015, 337, 53–60.
- 70. Ren, H.; Liu, X.; Xu, Q.; Li, W.; Zhao, N. Interleukin-35 Expression Promotes Hepatocellular Carcinogenesis by Inducing Γδ T-Cell Exhaustion. Genomics 2023, 115, 110639.
- 71. He, W.; Hu, Y.; Chen, D.; Li, Y.; Ye, D.; Zhao, Q.; Lin, L.; Shi, X.; Lu, L.; Yin, Z.; et al. Hepatocellular Carcinomainfiltrating Γδ T Cells Are Functionally Defected and Allogenic Vδ2 + Γδ T Cell Can Be a Promising Complement. Clin. Transl. Med. 2022, 12, e800.
- 72. Fan, J.; Li, J.; Han, J.; Zhang, Y.; Gu, A.; Song, F.; Duan, J.; Yin, D.; Wang, L.; Yi, Y. Expression of Leukocyte Immunoglobulin-like Receptor Subfamily B Expression on Immune Cells in Hepatocellular Carcinoma. Mol. Immunol. 2021, 136, 82–97.

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