Marine Endoperoxide Norterpenes

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Organic extracts of marine invertebrates, mainly sponges, from seas all over the world are well known for their high in vitro anticancer and antibiotic activities which make them promising sources of compounds with potential use as pharmaceutical leads. Most of the structures discovered so far have a peculiar structural feature in common: a 1,2-dioxane ring. This is a highly reactive heterocycle that can be considered as an endoperoxide function. Together with other structural features, this group could be responsible for the strong biological activities of the substances present in the extracts. Numerous research programs have focused on their structural elucidation and total synthesis since the seventies. As a consequence, the number of established chiral centres and the similarity between different naturally occurring substances is increasingly higher. Most of these compounds have a terpenoid nature, mainly diterpene and sesterterpene, with several peculiar structural features, such as the loss of one carbon atom. Although there are many reviews dealing with the occurrence of marine peroxides, their activities, or potential pharmaceutical uses, no one has focused on those having a terpene origin and the endoperoxide function. We present here a comprehensive review of these compounds paying special attention to their structural features and their biological activity.

marine terpenoid endoperoxide norseterterpene

1. Introduction

Marine water covers a great proportion of the earth's surface and is proposed to contain a high percentage of the world's plant and animal species; although, ninety one percent of marine species remain undiscovered ^[1]. Organic extracts of these marine species often exhibit promising specialized biological activities, prompting the search for new marine metabolites ^[2]. Between these compounds, a huge number of peroxides have been isolated, mainly from marine invertebrates, particularly in sponges and soft corals.

Numerous peroxides have shown confirmed activity in vitro against tumour cells and they are considered an important source of leads for drug discovery ^[3]. In fact, there are actually three approved marine drugs derived from sponges and several more have entered clinical trial ^[4]. On the other hand, terpenoids have a great chemical structural diversity and are also important secondary metabolites of marine species presenting diverse bioactivities ^[5].

This entry summarises marine endoperoxides of terpene origin and it is focused on their structure and the structural revision of newly-established stereochemistry, as well as in their biological activity. This entry is structured into two main sections: the first one summarises endoperoxides with a norterpene skeleton (norsesteterpenes and norditerpenes, principally) and the second one summarises endoperoxides with a terpene

skeleton (sesquiterpenes and diterpenes). The first section is also subdivided for a better understanding, based on the type of cyclic skeleton. A final overview of all reported biological activities for these systems isalso included.

In all structural formulae, the stereocentres are depicted using the following convention: solid wedged bonds and hashed wedged bonds represent the known absolute configuration. In addition, *R* or *S* letters are written near the centre. When only the relative stereochemistry of the compound is known, it is represented with bold bonds and hashed bonds.

2. Marine Endoperoxide Norterpenes

2.1. Endoperoxide Norterpenes with an Acyclic Carbon Skeleton





2.2. Endoperoxide Norterpenes with a Monocyclic Carbon Skeleton







(-)-13,14-epoxymuqubilin A (27)

(-)-9,10-epoxymuqubilin A (28)





2.3. Endoperoxide Norterpenes with a Bicyclic Carbon Skeleton





deoxydiacarnoate B benzyl ester (55) R= CH_2Ph , X= H diacarnoate B methyl ester (56) R= Me, X,X= O

COOCH₃

COOH

R) соон



R = H trunculin A (57) R = CH_3 trunculin A methyl ester (57a)



R =H trunculin B (58)

0. 0

R = CH₃ trunculin B methyl ester (58a)

Ο

trunculin D (60)

0

trunculin F (62)

trunculin H (64)

н

Ĥ

н

н



trunculin C (59)















contrunculin B (66)









diacaperoxide D (75)

diacaperoxide E (76)

diacaperoxide F (77)



diacaperoxide G (78)



diacaperoxide S or megaspinoxide A (79)

3. Marine Endoperoxide Terpenes

3.1. Endoperoxide Sesquiterpenes





3.2. Endoperoxide Diterpenes



4. Biological Activity Overview

So far, more than 90 marine terpenic endoperoxides have been described, the vast majority being norsesterterpenes (59 compounds) and norditerpenes (17 compounds). Other terpenoids with an endoperoxidic

moiety have also been isolated, although these are less frequent, like those having a sesqui- or diterpene structure (14 compounds).

There are many reports on the biological activity, possibly stimulated by the attractive endoperoxide moiety present in these substances. The studies have mainly focused on their activity against microorganisms, such as bacteria, yeasts, or viruses, as well as against protozoa that cause diseases, especially *Plasmodium*, which causes malaria, or *Trypanosoma* (**Table 1**). In addition, measurements of in vitro activity against different tumour cell lines are well documented (see table 1 for references). To a lesser extent, the anti-inflammatory activity has also been measured.

Both norsterterpenes and norditerpenes exhibit a remarkable range of bioactivities, although the number of studies with the first type is superior, possibly due to the fact that norsesterterpenes are more abundant. In all cases, the bioactive compounds are the free carboxylic acids, although sometimes natural methyl esters have been isolated. For example, although the acids **57** and **58** show activity against the bacteria and yeasts tested, their corresponding methyl esters, **57a** and **58a**, are inactive ^[6].

It is interesting to notice that the presence of the free acid form of these metabolites and the configuration of the methyl group in C6, may be critical for their growth of inhibitory activity in various cancer cell lines in vitro ^{[Z][8]}. In the case of the NO inhibition related to inflammatory processes, both the presence of a free carboxylic acid and an equatorial orientation of the methyl group attached to C6 are critical for increased activity ^[9].

Of all the isolated compounds, the most tested and the one with the greatest range of activities is (+)-muqubilin (**12**) and its enantiomer **21**. This compound is active against *Plasmodium*, and this is the reason why it has been used as a prototype for the development of new antimalarial agents ^{[10][11]}. In addition, it is effective against *Troxoplasma gondii*. Moderate activity against the *Nicotiana tabacum* plant has also been described, as a result of a program devoted to the search of natural marine products as prototypes for agrochemical agents ^[12].

Finally, it is also worth mentioning that diterpene **89** and the acid fraction of *Sigmosceptrella laevis*, from which **50**, **51**, and **52** were isolated, also have ichthyotoxic properties as a defence method against predators ^[13].

Target		Norsesterterpe		Securi and	
	Acyclics a	Monocyclics ^b	Bicyclics ^c	Norditerpenes ^d	Diterpenes ^e
Bacteria	1 ^[14] , 3a ^[15] , 4 ^[15]	12 ^{[<u>16]</u>}	57 ^[6] , 58 ^[6] , 61 ^[17] , 67 ^[18] , 68 ^[18] , 79 ^[19]		80 ^[20] , 81 ^[20] , 84 ^[21] , 86 ^[22]
Yeasts	1 ^[14]	12 ^{[<u>16]</u>}	57 ^[6] , 58 ^[6] , 61 [<u>17]</u> , 79 ^[19]		

Table 1. Biological activities of marine terpene endoperoxides.

Target		Norsesterterpe		Securi- and	
	Acyclics a	Monocyclics ^b	Bicyclics ^c	Norditerpenes ^d	Diterpenes ^e
Viruses	5 ^[23]	12 ^[23]	67 ^{[<u>18]</u>, 68 ^{[<u>18]</u>}}		
Plasmodia		12 ^[11] , 15 ^{[11][24]} , 20 ^{[11][24]} , 21 ^[25] , 45 ^[25] , 45a ^[25] , 46 ^[25] , 47 ^[25]	50 ^[10] , 55 ^[24]	$8 \frac{[11][24]}{[25]}, 9 \frac{[26]}{[11][24]}, 30$ $\frac{[25]}{[25]}, 31 \frac{[26]}{[26]}, 32 \frac{[26]}{[26]}$	
Trypanosoma	10 ^[27] , 11 ^[27]	22 ^[27]	50 ^[27] , 50a ^[27] , 51 ^[27]	26 ^[27]	
Toxoplasma		12 ^[23]	51 ^[23]		
Inflammation		22 ^{[9][28]}			86 ^[22] , 89 ^[29] [<u>30][31][32</u>]
Citotoxicity	5 ^{[33][7]}	15 ^[34] , 16 ^[34] , 17 ^[34] , 21 ^[8] , 38 ^[35]	50 ^[9] , 51 ^[7] , 51a ^[7] , 67 ^[18] , 68 ^[18] , 74 ^[36] , 75 ^[37] , 76 ^[37] , 77 ^[37] , 78 ^[37] , 79 ^[19]	6 ^[33] , 7 ^[33] , 13 ^[38] , 18 ^[7] , 25 ^[39]	80 ^[40] , 83 ^[21] , 84 ^{[41][21]} , 86 ^{[22][42]} , 89 ^{[13][43][29]} [30][31]

^a 6 described structures, 6 bioactive. ^b 22 described structures, 11 bioactive. ^c 31 described structures, 14 bioactive. ^d 17 described structures, 12 bioactive. ^e 14 described structures, 6 bioactive.

a) 6 described structures, 6 bioactive. b) 22 described structures, 11 bioactive. c) 31 described structures, 14 bioactive. d) 17 described structures, 12 bioactive. e) 14 described structures, 6 bioactive.

5. Conclusions

Marine macro- and microorganisms, especially algae, soft corals, sponges and fungi, have a metabolism that produces a great diversity of bioactive compounds, many of them unparalleled in the terrestrial environment. The composition of many species of sponges from tropical and temperate waters has been investigated, with a wide diversity of compounds found in their extracts. Among them, a group of substances stands out: those with a norterpene skeleton, that is, terpenoids that have one carbon atom less than what would be produced through an ordinary biosynthesis, and that additionally incorporate an endoperoxide function. These substances not only have structures of moderate complexity, but they also present numerous asymmetric centres. The structural identification of these compounds is very complex, especially from the point of view of the stereochemical assignment of the chiral centres, that has required the development of remarkable special techniques. The group of substances presented in this review, originating from marine invertebrates, such as sponges, is exceptional due to the range of potential pharmacological applications. The antitumor activity of some of these substances stands out, some of which are capable of inhibiting the tumour cell growth in in vitro cultures at concentrations between 10 and 100

 μ g/mL. It is certainly a very promising field of research that will continue to provide exceptional developments in the near future.

References

- 1. Sweetlove, L. Number of Species on Earth Tagged at 8.7 Million. Nature 2011. Available online: https://doi.org/10.1038/news.2011.498 (accessed on 19 November 2021).
- 2. Sato, A. The Search for New Drugs from Marine Organisms. J. Toxicol. Toxin Rev. 1996, 15, 171– 198.
- 3. Dembitsky, V.M.; Gloriozova, T.A.; Poroikov, V.V. Natural peroxy anticancer agents. Mini-Rev. Med. Chem. 2007, 7, 571–589.
- 4. He, Q.; Miao, S.; Ni, N.; Man, Y.; Gong, K. A Review of the Secondary Metabolites From the Marine Sponges of the Genus Aaptos. Nat. Prod. Commun. 2020, 15, 1934578X20951439.
- 5. Jiang, M.; Wu, Z.; Guo, H.; Liu, L.; Chen, S. A Review of Terpenes from Marine-Derived Fungi: 2015–2019. Mar. Drugs 2020, 18, 321.
- 6. Capon, R.J.; MacLeod, J.K.; Willis, A.C. Trunculins A and B, norsesterterpene cyclic peroxides from a marine sponge, Latrunculia brevis. J. Org. Chem. 1987, 52, 339–342.
- Al-Tarabeen, M.; El-Neketi, M.; Albohy, A.; Mueller, W.E.G.; Rasheed, M.; Ebrahim, W.; Proksch, P.; Ebada, S.S. Isolation and Molecular Docking of Cytotoxic Secondary Metabolites from Two Red Sea Sponges of the Genus Diacarnus. ChemistrySelect 2021, 6, 217–220.
- Lefranc, F.; Nuzzo, G.; Hamdy, N.A.; Fakhr, I.; Moreno, Y.; Banuls, L.; Van Goietsenoven, G.; Villani, G.; Mathieu, V.; van Soest, R.; et al. In Vitro Pharmacological and Toxicological Effects of Norterpene Peroxides Isolated from the Red Sea Sponge Diacarnus erythraeanus on Normal and Cancer Cells. J. Nat. Prod. 2013, 76, 1541–1547.
- Cheenpracha, S.; Park, E.-J.; Rostama, B.; Pezzuto, J.M.; Chang, L.C. Inhibition of nitric oxide (NO) production in lipopolysaccharide (LPS)-activated murine macrophage RAW 264.7 cells by the norsesterterpene peroxide, epimuqubilin A. Mar. Drugs 2010, 8, 429–437.
- El Sayed, K.A.; Dunbar, D.C.; Goins, D.K.; Cordova, C.R.; Perry, T.L.; Wesson, K.J.; Sanders, S.C.; Janus, S.A.; Hamann, M.T. The marine environment: A resource for prototype antimalarial agents. J. Nat. Toxins 1996, 5, 261–285.
- 11. Laurent, D.; Pietra, F. Antiplasmodial marine natural products in the perspective of current chemotherapy and prevention of malaria. A Review. Mar. Biotechnol. 2006, 8, 433–447.
- 12. Peng, J.; Shen, X.; El Sayed, K.A.; Dunbar, D.C.; Perry, T.L.; Wilkins, S.P.; Hamann, M.T.; Bobzin, S.; Huesing, J.; Camp, R.; et al. Marine natural products as prototype agrochemical agents. J.

Agric. Food Chem. 2003, 51, 2246–2252.

- Uchio, Y.; Eguchi, S.; Kuramoto, J.; Nakayama, M.; Hase, T. Denticulatolide, an ichthyotoxic peroxide-containing cembranolide from the soft coral Lobophytum denticulatum. Tetrahedron Lett. 1985, 26, 4487–4490.
- 14. Capon, R.J.; Macleod, J.K. Structural and stereochemical studies on marine norterpene cyclic peroxides. Tetrahedron 1985, 41, 3391–3404.
- Ovenden, S.P.B.; Capon, R.J. Nuapapuin A and Sigmosceptrellins D and E: New Norterpene Cyclic Peroxides from a Southern Australian Marine Sponge, Sigmosceptrella sp. J. Nat. Prod. 1999, 62, 214–218.
- 16. Sokoloff, S.; Halevy, S.; Usieli, V.; Colorni, A.; Sarel, S. Prianicin A and B, nor-sesterterpenoid peroxide antibiotics from Red Sea sponges. Experientia 1982, 38, 337–338.
- 17. He, H.Y.; Faulkner, D.J.; Lu, H.S.M.; Clardy, J. Norsesterterpene peroxides from the sponge Latrunculia sp. J. Org. Chem. 1991, 56, 2112–2115.
- Tanaka, J.-I.; Higa, T.; Suwanborirux, K.; Kokpol, U.; Bernardinelli, G.; Jefford, C.W. Bioactive norsesterterpene 1,2-dioxanes from a Thai sponge, Mycale sp. J. Org. Chem. 1993, 58, 2999– 3002.
- 19. Ibrahim, S.R.M. Diacarperoxide S, new norterpene cyclic peroxide from the sponge Diacarnus megaspinorhabdosa. Nat. Prod. Commun. 2012, 7, 9–12.
- Vairappan, C.S.; Suzuki, M.; Ishii, T.; Okino, T.; Abe, T.; Masuda, M. Antibacterial activity of halogenated sesquiterpenes from Malaysian Laurencia spp. Phytochemistry 2008, 69, 2490– 2494.
- 21. Ishii, T.; Ueoka, R.; Matsunaga, S.; Vairappan, C.S. Bioactive secondary metabolites from the Borneon soft corals of the genus Nephthea. Malays. J. Sci. 2010, 29, 262–268.
- 22. Roy, P.K.; Ashimine, R.; Miyazato, H.; Taira, J.; Ueda, K. Endoperoxy and hydroperoxy cadinanetype sesquiterpenoids from an Okinawan soft coral, Sinularia sp. Arch. Pharm. Res. 2016, 39, 778–784.
- 23. El Sayed, K.A.; Hamann, M.T.; Hashish, N.E.; Shier, W.T.; Kelly, M.; Khan, A.A. Antimalarial, antiviral, and antitoxoplasmosis norsesterterpene peroxide acids from the Red Sea sponge Diacarnus erythraeanus. J. Nat. Prod. 2001, 64, 522–524.
- D'Ambrosio, M.; Guerriero, A.; Deharo, E.; Debitus, C.; Munoz, V.; Pietra, F. New types of potentially antimalarial agents. Epidioxy-substituted norditerpene and norsesterterpenes from the marine sponge Diacarnus levii. Helv. Chim. Acta 1998, 81, 1285–1292.
- 25. Yang, F.; Wang, R.-P.; Xu, B.; Yu, H.-B.; Ma, G.-Y.; Wang, G.-F.; Dai, S.-W.; Zhang, W.; Jiao, W.-H.; Song, S.-J.; et al. New antimalarial norterpene cyclic peroxides from Xisha Islands sponge

Diacarnus megaspinorhabdosa. Bioorg. Med. Chem. Lett. 2016, 26, 2084–2087.

- 26. Yang, F.; Zou, Y.; Wang, R.-P.; Hamann, M.T.; Zhang, H.-J.; Jiao, W.-H.; Han, B.-N.; Song, S.-J.; Lin, H.-W. Relative and absolute stereochemistry of diacarperoxides: Antimalarial norditerpene endoperoxides from marine sponge Diacarnus megaspinorhabdosa. Mar. Drugs 2014, 12, 4399– 4416.
- 27. Rubio, B.K.; Tenney, K.; Ang, K.-H.; Abdulla, M.; Arkin, M.; McKerrow, J.H.; Crews, P. The Marine Sponge Diacarnus bismarckensis as a Source of Peroxiterpene Inhibitors of Trypanosoma brucei, the Causative Agent of Sleeping Sickness. J. Nat. Prod. 2009, 72, 218–222.
- 28. Park, E.-J.; Cheenpracha, S.; Chang, L.C.; Pezzuto, J.M. Suppression of cyclooxygenase-2 and inducible nitric oxide synthase expression by epimuqubilin A via IKK/IκB/NF-κB pathways in lipopolysaccharide-stimulated RAW 264.7 cells. Phytochem. Lett. 2011, 4, 426–431.
- 29. Kobayashi, M.; Ishizaka, T.; Miura, N.; Mitsuhashi, H. Marine terpenes and terpenoids. III. Isolation and structures of two cembrane diols from the soft coral Sinularia mayi. Chem. Pharm. Bull. 1987, 35, 2314.
- 30. Kamada, T.; Kang, M.-C.; Phan, C.-S.; Zanil, I.I.; Jeon, Y.-J.; Vairappan, C.S. Bioactive Cembranoids from the Soft Coral Genus Sinularia sp. in Borneo. Mar. Drugs 2018, 16, 99.
- 31. Chao, C.-H.; Wen, Z.-H.; Wu, Y.-C.; Yeh, H.-C.; Sheu, J.-H. Cytotoxic and Anti-inflammatory Cembranoids from the Soft Coral Lobophytum crassum. J. Nat. Prod. 2008, 71, 1819–1824.
- 32. Kapojos, M.M.; Lee, J.-S.; Oda, T.; Nakazawa, T.; Takahashi, O.; Ukai, K.; Mangindaan, R.E.P.; Rotinsulu, H.; Wewengkang, D.S.; Tsukamoto, S.; et al. Two unprecedented cembrene-type terpenes from an indonesian soft coral Sarcophyton sp. Tetrahedron 2010, 66, 641–645.
- 33. Youssef, D.T.A.; Yoshida, W.Y.; Kelly, M.; Scheuer, P.J. Cytotoxic cyclic norterpene peroxides from a Red Sea sponge Diacarnus erythraenus. J. Nat. Prod. 2001, 64, 1332–1335.
- D'Ambrosio, M.; Guerriero, A.; Debitus, C.; Waikedre, J.; Pietra, F. Relative contributions to antitumoral activity of lipophilic vs. polar reactive moieties in marine terpenoids. Tetrahedron Lett. 1997, 38, 6285–6288.
- 35. Dai, J.; Liu, Y.; Zhou, Y.-D.; Nagle, D.G. Hypoxia-Selective Antitumor Agents: Norsesterterpene Peroxides from the Marine Sponge Diacarnus levii Preferentially Suppress the Growth of Tumor Cells under Hypoxic Conditions. J. Nat. Prod. 2007, 70, 130–133.
- Phuwapraisirisan, P.; Matsunaga, S.; Fusetani, N.; Chaitanawisuti, N.; Kritsanapuntu, S.; Menasveta, P. Mycaperoxide H, a new cytotoxic norsesterterpene peroxide from a Thai marine sponge Mycale sp. J. Nat. Prod. 2003, 66, 289–291.
- 37. Ibrahim, S.R.M.; Ebel, R.; Wray, V.; Muller, W.E.G.; Edrada-Ebel, R.; Proksch, P. Diacarperoxides, Norterpene Cyclic Peroxides from the Sponge Diacarnus megaspinorhabdosa. J. Nat. Prod.

2008, 71, 1358-1364.

- 38. Miao, X.-X.; Chen, M.-X.; Zhu, H.-R.; Sun, F.; Hong, L.-L.; Wu, W.-H.; Lin, H.-W.; Yang, F. New polyketides and norterpenoids from the marine sponge Diacarnus megaspinorhabdosa. Tetrahedron 2020, 76, 131062.
- Chao, C.-H.; Chou, K.-J.; Wang, G.-H.; Wu, Y.-C.; Wang, L.-H.; Chen, J.-P.; Sheu, J.-H.; Sung, P.-J. Norterpenoids and Related Peroxides from the Formosan Marine Sponge Negombata corticata. J. Nat. Prod. 2010, 73, 1538–1543.
- 40. Erickson, K.L.; Beutler, J.A.; Gray, G.N.; Cardellina, J.H.; Boyd, M.R. Majapolene A, a Cytotoxic Peroxide, and Related Sesquiterpenes from the Red Alga Laurencia majuscula. J. Nat. Prod. 1995, 58, 1848–1860.
- 41. Cheng, S.-Y.; Dai, C.-F.; Duh, C.-Y. Sesquiterpenoids and Artificial 19-Oxygenated Steroids from the Formosan Soft Coral Nephthea erecta. J. Nat. Prod. 2007, 70, 1449–1453.
- 42. Miyazato, H.; Taira, J.; Ueda, K. Hydrogen peroxide derived from marine peroxy sesquiterpenoids induces apoptosis in HCT116 human colon cancer cells. Bioorg. Med. Chem. Lett. 2016, 26, 4641–4644.
- 43. Kusumi, T.; Ohtani, I.; Inouye, Y.; Kakisawa, H. Absolute configurations of cytotoxic marine cembranolides; Consideration of mosher's method. Tetrahedron Lett. 1988, 29, 4731–4734.

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