Microfluidic High-Throughput Platforms

Subjects: Nanoscience & Nanotechnology

Contributor: Jianhua Zhou

High-throughput screening is a potent technique to accelerate the discovery and development of new materials. By performing massive synthesis and characterization processes in parallel, it can rapidly discover materials with desired components, structures and functions. Among the various approaches for high-throughput screening, microfluidic platforms have attracted increasing attention. Compared with many current strategies that are generally based on robotic dispensers and automatic microplates, microfluidic platforms can significantly increase the throughput and reduce the consumption of reagents by several orders of magnitude.

Keywords: Microfluidic High-Throughput Platforms, High-throughput screening, nanomaterials

1. Introduction

Compared with traditional microplate-based HTPs that require samples of at least several microliters in each well, microfluidic platforms consume much less reagents with the scale of nanoliters to picoliters, which significantly reduces the cost and is beneficial to save rare samples. Microarray is one of the major microfluidic platforms, which integrates a large quantity of isolated reactors on one substrate. Additionally, each reactor is microscaled with volumes ranging from nanoliters to picoliters. It allows multiple parameters to be tested in parallel by simultaneously performing tens to thousands of experiments per batch. For example, Zhang and his coworkers developed a hydrogel microarray (Figure 1a), in which 2000 individual microgels with varying bioactivities were regularly patterned on a standard microscope slide, providing a high-throughput platform to rapidly screen desired polymers with thermal-responsive properties [1]. Perera et al. developed an automatic synthetic platform for drug discovery, which integrated commercially available components into a highly integrated module unit to perform both nanomole-scale reactions and micromole-scale syntheses [2]. This setup allows screenings of more than 1500 homogeneous reactions within 24 h under different temperature, pressure, and solvent, which has the advantages of real-time analysis, sufficient mixing, and avoidance of solvent evaporation. Due to the application of microarray-based HTPs, reactions were performed in parallel under a broad range of experimental parameters so that appropriate conditions for generating nanostructures with specific morphologies can be rapidly identified. Moreover, Duffy et al. described a hydrogel microarray that integrated 80 unique holes on a single microscope slide using three-dimensional (3D) printing [3]. By filling the holes with double network hydrogels, the novel platform offered a powerful tool to screen hydrogels with desired compressive and tensile properties, which could be further optimized for drug delivery, cell encapsulation, and tissue engineering. Microarrays have also been widely applied in a wide range of biomedical applications, such as pharmaceutical discovery, small molecule and protein screening, toxicity tests, etc. [4][5][6][7][8][9]. For example, Hay et al. used the polymer microarray with high content screening system and Pathfinder software to screen and discover new extracellular substrates, which can promote hepatic endoderm, druginducible metabolism and toxicology [5]. Additionally, Khan et al. proposed a microarray platform combined with a highthroughput screening approach to screen and analyze the biological functionality of 135 polymer blends, leading to the identification of cell-compatible biopolymers permissive for human skeletal stem cell growth in both in vitro and in vivo applications [7].

Figure 1. (a) A representative high-throughput platform (HTP) based on microarrays. A polymer hydrogel microarray with 2436 polymers (28×87 hydrogel spots) were prepared by inkjet printing. Subsequently, the microarray was incubated with HeLa cells for 48 h. Mosaic (the middle column in Figure 1a), and enlarged fluorescent images of cells were obtained. Reproduced with permission from [$^{[1]}$. Copyright Elsevier, 2009. (b) A representative HTP based on microdroplets. A microfluidic device was used to prepare colloid-filled hydrogel granules with different sizes and shapes by changing the flow rate of reagent 1 (Q1), reagent 2 (Q2) and oil (Q3). Reproduced with permission from $^{[10]}$. Copyright American Chemical Society, 2006. (c) A representative continuous-flow microfluidic system was used to produce Au-Pd dumbbell nanoparticles. Reproduced with permission from $^{[11]}$. Copyright American Chemical Society, 2017. (d) A digital microfluidic circuit and the four fundamental droplet operations: creating, cutting, joining and transporting. Reproduced with permission from $^{[12]}$. Copyright IEEE Xplore, 2002.

Despite the improvement in throughput, microarray-based HTPs are still limited in many cases that required higher screening efficiency. To address the issue, microdroplet technology has drawn increasing attention and been developed for high-throughput screenings [13]. Microfluidic droplet chips can be divided into continuous microfluidic chips (Figure 1b,c) [14][15][16][17][18] and digital microfluidic chips (Figure 1d) [19][20][21]. Shepherd's group provided a continuous microfluidic device (Figure 1b) to generate monodisperse colloid-filled hydrogel particles with different shapes and compositions [10]. Additionally, Jensen et al. described a new device for the production of Au-Pd dumbbell-like nanostructure with high electrocatalytic activity [11]. This device was integrated with a sequential-addition microfluidic reactor and an ultrasonic to control the growth of Au onto the both sides of Pd nanorods (Figure 1c). As the key platform of microdroplet technology, continuous microfluidic chip can generate monodisperse droplets (usually at nano- or picoliters) at very high frequencies (from tens to thousands of droplets per second) [22][23]. Additionally, each microdroplet serves as an independent microreactor, in which synthesis of materials can be carried out without interference under certain conditions. Digital microfluidics employed electrowetting to control and discretize the continuous flow into individual droplets. Sung et al. fully reported the functional digital microfluidic circuits and the four fundamental droplet operations mechanisms [12]. It provides a promising experimental platform with advantages of a fast response, high precision, and digital readouts. Microdroplet-based HTPs has many advantages [24][25][26]. Firstly, it consumes much less reactants since the working volume of a plate well (e.g., 10 µL for each well of a 384-well plate) is ten million times that of a single droplet (1.0 pL) [27]. Secondly, the high surface-to-volume ratio of microdroplets and short diffusion distance in microdroplets result in pronounced acceleration of reactions and thus can significantly shorten the screening time. Thirdly, it provides chemical and physical confinement to avoid cross-contamination. Using this technique, a large quantity of independent experiments can be easily performed within a very short period and only a small amount of reagents are consumed.

2. Basic Principles

Both microarrays- and microdroplets-based HTPs can significantly increase the screening throughput and accelerate development of material science. For microarray-based HTPs, the synthetic parameters of each reaction can be precisely encoded by spatial coordinates, but the throughput is limited by device area and density of reaction sites. For microdroplet-based HTPs, the throughput greatly increases owing to the continuous and rapid generation of microdroplets. However, it remains challenging to accurately encode the synthetic parameters of each microdroplet, thus limiting the further increasement of its throughput. Recently, a novel high-throughput method called "droplet library", which combines a microfluidic droplet generator with microarrays, are proposed [28][29]. The basic principles are shown in Figure 2. Firstly, droplets containing small compounds are prepared by parallel microfluidic devices and subsequently transported to microarray plates. Then the following droplets with different compounds could be gathered in one tube as a droplet library. The droplet library was then reinjected into another device to mix with a target for screening the compounds with

optimal antimicrobial activities. This integrated platform takes significantly less time than conventional microdroplet-based HTPs. Although mainly applied for biological experiments, such as investigations of antimicrobial activities, pharmacological screening, drug-resistance analysis, etc., the novel integrated approach shows great potentials in screening materials with ultra-high throughput, providing a promising approach towards the development of next-generation HTPs.

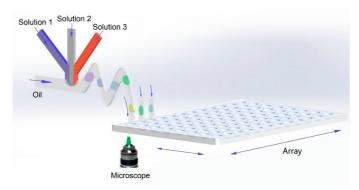


Figure 2. A demonstration of high-throughput-screening platform that combine droplet generator, microarray and high-throughput characterization for screening material structures.

Moreover, to achieve truly high-throughput screening, it is necessary to establish highly integrated HTPs with multiple functions of material synthesis, characterization and data analysis. Zhou et al. [30] have proposed a high-throughput screening system. It combined a microfluidic reactor to generate hydrogel droplets with different crystals of drugs, a camera to capture the optical images of the droplets, and deep learning to analyze and classify the obtained images. Additionally, the microfluidic chip was fabricated with a flow-focusing geometry to produce droplets. Their system offered a new high-throughput platform and could be applied to quickly synthesize the massive materials and accurately analyze the data. With massive materials informatics and databases, it offers a potent platform to accelerate the development of the new materials. Despite the great advancements in material synthesis, the performance of current HTPs in high-throughput characterization is still far from satisfactory. Therefore, developing compatible high-throughput characterization techniques to combine with synthetic modules is one of the important trends of future HTPs. Additionally, as HTPs usually produce massive data, approaches for high-throughput data processing are also in great demand. Machine learning is a powerful tool to process and analyze massive information, which shows promising applications in future HTPs. Since the application of HTPs has gradually played a critical role in new material preparation, it will show significant impact on the development of material science, biological science, biomedical engineering and military science in the future.

References

- 1. Zhang, R.; Liberski, A.; Sanchez-Martin, R.M.; Bradley, M. Microarrays of over 2000 hydrogels–Identification of substrat es for cellular trapping and thermally triggered release. Biomaterials 2009, 30, 6193–6201.
- 2. Perera, D.; Tucker, J.W.; Brahmbhatt, S.; Helal, C.J.; Chong, A.; Farrell, W.; Richardson, P.; Sach, N.W. A platform for a utomated nanomole-scale reaction screening and micromole-scale synthesis in flow. Science 2018, 359, 429–434.
- 3. Duffy, C.; Venturato, A.; Callanan, A.; Lilienkampf, A.; Bradley, M. Arrays of 3D double-network hydrogels for the high-th roughput discovery of materials with enhanced physical and biological properties. Acta Biomater. 2016, 34, 104–112.
- 4. Hansen, A.; McMillan, L.; Morrison, A.; Petrik, J.; Bradley, M. Polymers for the rapid and effective activation and aggreg ation of platelets. Biomaterials 2011, 32, 7034–7041.
- 5. Hay, D.C.; Pernagallo, S.; Díaz-Mochón, J.J.; Medine, C.N.; Greenhough, S.; Hannoun, Z.; Schrader, J.; Black, J.R.; Fl etcher, J.; Dalgetty, D.; et al. Unbiased screening of polymer libraries to define novel substrates for functional hepatocyt es with inducible drug metabolism. Stem Cell Res. 2011, 6, 92–102.
- 6. Hook, A.L.; Scurr, D.J.; Anderson, D.G.; Langer, R.; Williams, P.; Davies, M.C.; Alexander, M.R. High throughput discov ery of thermo-responsive materials using water contact angle measurements and time-of-flight secondary ion mass spe ctrometry. Surf. Interface Anal. 2012, 45, 181–184.
- 7. Khan, F.; Tare, R.S.; Kanczler, J.M.; Oreffo, R.; Bradley, M. Strategies for cell manipulation and skeletal tissue engineering using high-throughput polymer blend formulation and microarray techniques. Biomater. 2010, 31, 2216–2228.
- 8. Moraes, C.; Wang, G.; Sun, Y.; Simmons, C.A. A microfabricated platform for high-throughput unconfined compression of micropatterned biomaterial arrays. Biomaterials 2010, 31, 577–584.
- 9. Du, W.; Li, L.; Nichols, K.P.; Ismagilov, R.F. SlipChip. Lab. Chip 2009, 9, 2286-2292.

- 10. Shepherd, R.F.; Conrad, J.C.; Rhodes, S.K.; Link, D.R.; Marquez, M.; Weitz, D.A.; Lewis, J.A. Microfluidic Assembly of Homogeneous and Janus Colloid-Filled Hydrogel Granules. Langmuir 2006, 22, 8618–8622.
- 11. Sebastian, V.; Zaborenko, N.; Gu, L.; Jensen, K.F. Microfluidic Assisted Synthesis of Hybrid Au–Pd Dumbbell-like Nano structures: Sequential Addition of Reagents and Ultrasonic Radiation. Cryst. Growth Des. 2017, 17, 2700–2710.
- 12. Cho, S.K.; Fan, S.K.; Moon, H.; Kim, C.J. Towards Tigital Microfluidic Circuits: Creating, Transporting, Cutting and Merg ing Liquid Droplets by Electrowetting-based Actuation. In Proceedings of the Fifteenth IEEE International Conference on Micro Electro Mechanical Systems, Las Vegas, NV, USA, 24 January 2002.
- 13. Marre, S.; Jensen, K.F. Synthesis of micro and nanostructures in microfluidic systems. Chem. Soc. Rev. 2010, 39, 1183 –1202.
- 14. Hakala, T.A.; Bialas, F.; Toprakcioglu, Z.; Bräuer, B.; Bohndiek, S.E.; Levin, A.; Bernardes, G.J.L.; Becker, C.F.W.; Know les, T.P. Continuous Flow Reactors from Microfluidic Compartmentalization of Enzymes within Inorganic Microparticles. ACS Appl. Mater. Interfaces 2020, 12, 32951–32960.
- 15. Carneiro, J.; Campos, J.B.L.M.; Miranda, J.M. High viscosity polymeric fluid droplet formation in a flow focusing microfl uidic device Experimental and numerical study. Chem. Eng. Sci. 2019, 195, 442–454.
- 16. Juthani, N.; Doyle, P.S. A platform for multiplexed colorimetric microRNA detection using shape-encoded hydrogel particles. Analyst 2020, 145, 5134–5140.
- 17. Parthiban, P.; Doyle, P.S.; Hashimoto, M. Self-assembly of droplets in three-dimensional microchannels. Soft Matter 20 19, 15, 4244–4254.
- 18. Hao, N.; Nie, Y.; Xu, Z.; Jin, C.; Fyda, T.J.; Zhang, J.X.J. Microfluidics-enabled acceleration of Fenton oxidation for degradation of organic dyes with rod-like zero-valent iron nanoassemblies. J. Colloid Interface Sci. 2020, 559, 254–262.
- 19. Cooney, C.G.; Chen, C.-Y.; Emerling, M.R.; Nadim, A.; Sterling, J.D. Electrowetting droplet microfluidics on a single pla nar surface. Microfluid. Nanofluidics 2006, 2, 435–446.
- 20. Prakash, S.; Ashley, B.K.; Doyle, P.S.; Hassan, U. Design of a Multiplexed Analyte Biosensor using Digital Barcoded Particles and Impedance Spectroscopy. Sci. Rep. 2020, 10, 1–10.
- 21. Fang, Z.; Ding, Y.; Zhang, Z.; Wang, F.; Wang, Z.; Wang, H.; Pan, T. Digital microfluidic meter-on-chip. Lab. Chip 2020, 20, 722–733.
- 22. Yobas, L.; Martens, S.; Ong, W.-L.; Ranganathan, N. High-performance flow-focusing geometry for spontaneous gener ation of monodispersed droplets. Lab. Chip 2006, 6, 1073–1079.
- 23. Zhou, C.; Zhu, P.; Tian, Y.; Xu, M.; Wang, L. Engineering Micromotors with Droplet Microfluidics. ACS Nano 2019, 13, 6 319–6329.
- 24. Millington, D.; Norton, S.; Singh, R.; Sista, R.; Srinivasan, V.; Pamula, V. Digital microfluidics comes of age: High-throug hput screening to bedside diagnostic testing for genetic disorders in newborns. Expert Rev. Mol. Diagn. 2018, 18, 701–712.
- 25. Scheler, O.; Postek, W.; Garstecki, P. Recent developments of microfluidics as a tool for biotechnology and microbiolog y. Curr. Opin. Biotechnol. 2019, 55, 60–67.
- 26. Yang, D.; Priest, C. Microfluidic Platform for High-Throughput Screening of Leach Chemistry. Anal. Chem. 2018, 90, 85 17–8522.
- 27. Brouzes, E.; Medkova, M.; Savenelli, N.; Marran, D.; Twardowski, M.; Hutchison, J.B.; Rothberg, J.M.; Link, D.R.; Perri mon, N.; Samuels, M.L. Droplet microfluidic technology for single-cell high-throughput screening. Proc. Natl. Acad. Sci. USA 2009, 106, 14195–14200.
- 28. Liu, Y.; Du, J.; Choi, J.-S.; Chen, K.-J.; Hou, S.; Yan, M.; Lin, W.-Y.; Chen, K.S.; Ro, T.; Lipshutz, G.S.; et al. A High-Thr oughput Platform for Formulating and Screening Multifunctional Nanoparticles Capable of Simultaneous Delivery of Genes and Transcription Factors. Angew. Chem. Int. Ed. 2015, 55, 169–173.
- 29. Guo, M.T.; Rotem, A.; Heyman, J.A.; Weitz, D.A. Droplet microfluidics for high-throughput biological assays. Lab. Chip 2012, 12, 2146–2155.
- 30. Su, Z.; He, J.; Zhou, P.; Huang, L.; Zhou, J. A high-throughput system combining microfluidic hydrogel droplets with dee p learning for screening the antisolvent-crystallization conditions of active pharmaceutical ingredients. Lab. Chip 2020, 20, 1907–1916.