

# Immune Checkpoint Gene Regulation by microRNA in Cancer

Subjects: Oncology

Contributor: Fatimat Kipkeeva, Tatyana Muzaffarova, Alexandra Korotaeva, Danzan Mansorunov, Pavel Apanovich, Maxim Nikulin, Olga Malikhova, Ivan Stilidi, Alexander Karpukhin

Currently, the search for new promising tools of immunotherapy continues. In this regard, microRNAs that influence immune checkpoint gene expression in tumor and T-cells. An important feature of miRNA is its ability to affect the expression of several genes simultaneously, which corresponds to the trend toward the use of combination therapy. MiRNAs regulate gene expression by blocking mRNA translation. An important feature of miRNA is its ability to affect the expression of several genes simultaneously, which corresponds to the trend toward the use of combination therapy.

Keywords: microRNA ; immune checkpoint ; immunotherapy

## 1. Introduction

Immunotherapy is an innovative method of cancer treatment. As a result of experiments and clinical trials, it has been found that immunotherapy can increase progressionfree survival and overall survival. However, this method of treatment is effective in a limited number of patients, and in addition, it can cause severe adverse reactions due to hyperreactivity of the immune system [1]. In this regard, research is underway to develop new therapeutic approaches based on targeting immune checkpoints (ICs). Tumor cells have the ability to generate ligands that can bind to co-inhibitory receptor molecules. This interaction suppresses the antitumor immune response, allowing the tumor to "escape" from the immune system. In order to increase the effectiveness of immunotherapy, the FDA approved a number of regimens, including a combination of two IC inhibitors, a combination of IC inhibitors and targeted therapy drugs, as well as antitumor bispecific antibodies [2][3]. It has been shown that in combination therapy regimens, patients experienced a higher response rate compared to monotherapy [4]. In addition, the search for a more promising immunotherapy approach is currently ongoing. In this regard, microRNAs (miRNAs) are considered. According to recent studies, miRNAs influence IC gene expression and are important regulators in both T-cells and tumor cells [5]. MiRNAs regulate gene expression by binding to the 3'-UTR of their mRNA [6][7][8]. MiRNAs can also affect IC expression indirectly, through molecules of different signaling pathways, such as PTEN, IFR-1, and others [9]. It is also important that one miRNA can affect several genes [5][10]. Here presents a entry of miRNAs that interact with IC genes, analyzes their regulating IC expression in tumors of various types of cancer, and identifies miRNAs that act on several IC genes simultaneously. Due to these properties, miRNA-based therapy may become an alternative to the combination of targeted drugs in the future. In addition, miRNAs are considered that are capable of simultaneously regulating the expression of targeted therapy genes along with IC genes. These issues have not been previously analyzed in existing reviews of miRNAs as IC regulators [11][12][13][14][15]. The researchers have reviewed more than 200 miRNAs that regulate ICs in tumors of various types.

**Table 1.** The miRNAs interacting with IC genes in different types of cancer.

Immune Checkpoint	microRNA	Cancer	Reference
PD-1	miR-374b, miR-4717 miR-183	Liver cancer	[16][17]
PD-1/PD-L1	miR-138-5p, miR-200b, miR-429, miR-508	RCC	[18]
		Lung cancer	[19][20]

Immune Checkpoint	microRNA	Cancer	Reference
	miR-142-5p	PC, OC	[21][22]
	miR-497-5p	ccRCC	[23]
	miR-20-b, miR-21, miR-130b, miR-138-5p, miR-148a-3p, miR-191-5p	CRC	[24][25][26] [27]
	miR-195, miR-424-5p, miR-497, miR-873, miR-3609	BC	[28][29][30] [31]
	miR-17-5p, miR-146a	Melanoma	[32][33]
	miR-15a, miR-15b, miR-16, miR-193a-3p, miR-320a	Pleural Mesothelioma	[34][35]
	miR-155, miR-195, miR-214	B-cell lymphoma	[36][37][38]
	miR-16, miR-195	Prostate cancer	[39]
	miR-34a, miR-34b, miR-34c, miR-140, miR-200, miR-200a-3p, miR-3127-5p	Lung cancer	[40][41][42] [43][44]
PD-L1	miR-34a	AML	[45]
	miR-23a-3p, miR-570	Liver cancer	[46][47]
	miR-375	HNSCC	[48]
	miR-145	OC, bladder cancer	[49][50]
	miR-513a-5p	Retinoblastoma	[51]
	miR-105-5p, miR-152, miR-200b, miR-200c, miR-570	GC	[52][53][54] [55][56]
	miR-18a, miR-140, miR-142, miR-340, miR-383	Cervical cancer	[57]
	miR-217	Laryngeal cancer	[58]
	miR-20b-5p	Models of lung and BC	[59]
	miR-194-5p	PC	[60]
PD-L1+B7-H3	miR-326	Lung cancer	[61]
PD-1, CTLA-4	miR-424	OC	[62]
	miR-138-5p	Glioma	[63]
CD80/CTLA-4	miR-424	CRC	[64]
PD-1, PD-L1, CTLA-4	miR-33a	Lung cancer	[65]
PD-1, BTLA, Tim-3	miR-28	Melanoma mouse model	[66]
BTLA	miR-32	OC	[67]
Tim-3	miR-498	AML	[68]
IDO1	miR-153, miR-448	CRC	[69][70]
Gal-3	miR-424-3p	OC	[71]
	miR-128	CRC	[72]
	miR-22	Liver cancer	[73]
Gal-9	miR-15b-5p, miR-455-5p, miR-1237, miR-1246	CRC	[74][75]
ICOS (B7-H2)/ICOSL	miR-24	GC	[76]

Immune Checkpoint	microRNA	Cancer	Reference
	miR-29 (a, b and c)	Neuroblastoma, sarcoma, brain tumors	[77]
	miR-145	Lung cancer	[78]
	miR-28-5p, miR-29a, miR-128, miR-145, miR-155/miR-143, miR-187, miR-192, miR-335-5p, miR-378, miR-1301-3p	CRC	[79][80][81] [82][83]
	miR-187	ccRCC	[84]
	miR-29c	Melanoma, CRC	[85][86]
B7-H3	miR-29c, miR-34b, miR-124a, miR-125b-2, miR-214, miR-297, miR-326, miR-363, miR-380-5p, miR-506, miR-555, miR-567, miR-593, miR-601, miR-665, miR-708, miR-885-3p, miR-940	BC	[87]
	miR-539	Glioma	[88]
	miR-124	Osteosarcoma	[89]
	miR-506	Mantle cell lymphoma	[90]
	miR-214	Multiple myeloma	[91]
	miR-29, miR-1253	Medulloblastoma	[92][93]
	miR-199a	Cervical cancer	[94]
B7-H5 (VISTA, BTNL2)	miR-125a-5p	GC	[95]
	miR-155/miR-143, miR-1207	CRC	[80][96]
B7-H4 (VTCN1)	miR-7-5p, hsa-let-7c, hsa-let-7f-5p, miR-17-3p, miR-21-3p, miR-21-5p, miR-24-1-5p, miR-27b-3p, miR-31-3p, miR-31-5p, miR-33a-5p, miR-33b-5p, miR-122-3p, miR-130b-3p, miR-138-1-3p, miR-148a-3p, miR-149-3p, miR-183-3p, miR-186-5p, miR-196a-5p, hsa-miR-204-3p, miR-299-5p, miR-302a-3p, miR-302e, miR-335-3p, miR-335-5p, miR-361-5p, miR-374c-5p, miR-483-3p, miR-513a-5p, miR-519e-3p, miR-520d-5p, miR-525-5p, miR-615-3p, miR-642a-5p, miR-744-5p, miR-937, miR-1246, miRPlus-G1246-3p, miR-1260a, miR-1265, miR-1284, miR-1290, miR-1973, miR-2115-3p, miR-2116-5p, miR-3178, miR-3202, miR-3646, miR-3651, miR-3676-3p, miR-3685, miR-3686, miR-4258, miR-4279, miR-4284, miR-4288, miR-4290, miR-4306, miR-4324	PC	[97]
B7-H6 (NCR3LG1)	miR-93, miR-195, miR-340	BC	[29]
B7-H7 (HHLA2)	miR-3116, miR-6870-5p	ccRCC	[98]

Footnotes: RCC—renal cell cancer; PC—pancreatic cancer; OC—ovarian cancer; CRC—colorectal cancer; BC—breast cancer; AML—acute myeloid leukemia; HNSCC—head and neck squamous cell cancer; GC—gastric cancer.

## 2. The Features of Immune Checkpoint Gene Regulation by microRNA in Cancer

The results of accumulated data analysis demonstrate a significant relationship between the action of miRNAs on ICs genes and the type of tumor—only about 14% (95% CI: 9.8–20.1%) of the studied miRNAs regulate the expression of specific IC in more than one type of cancer.

Currently, there are numerous studies underway to identify miRNAs that are the most promising as immunotherapy agents. In vivo experiments have repeatedly shown that miRNA-based therapy leads to significant tumor regression. Although miRNA has not yet entered the arsenal of antitumor agents used in practice, some results are encouraging. Thus, the miR-155 inhibitor has performed well in clinical trials. The study of miR-138 is promising. Ongoing research on miR-34a may also lead to a positive result. Thus, there is the prospect of using miRNA as a therapeutic agent in cancer immunotherapy regimens. At the same time, the ability of miRNAs to inhibit several genes can lead to adverse events. To

overcome this, it is important to expand data of the spectrum of miRNA targets in a particular type of cancer. Additional studies of the miRNA–genes interaction features and the search for an optimal miRNA mimic structure are necessary, thus allowing an increase in the efficiency and selectivity of interaction with the mRNA of target genes. It can increase the effectiveness of therapy, as well as reduce the dose of the drug, thereby reducing its side effects.

## References

1. Tan, S.; Li, D.; Zhu, X. Cancer immunotherapy: Pros, cons and beyond. *Biomed. Pharmacother.* 2020, **124**, 109821.
2. Vafaei, S.; Zekiy, A.O.; Khanamir, R.A.; Zaman, B.A.; Ghayourvahdat, A.; Azimizonuzi, H.; Zamani, M. Combination therapy with immune checkpoint inhibitors (ICIs); a new frontier. *Cancer Cell Int.* 2022, **22**, 2.
3. Esfandiari, A.; Cassidy, S.; Webster, R.M. Bispecific antibodies in oncology. *Nat. Rev. Drug Discov.* 2022, **21**, 411–412.
4. Feng, Y.; Jin, H.; Guo, K.; Xiang, Y.; Zhang, Y.; Du, W.; Shen, M.; Ruan, S. Results from a Meta-analysis of Combination of PD-1/PD-L1 and CTLA-4 Inhibitors in Malignant Cancer Patients: Does PD-L1 Matter? *Front. Pharmacol.* 2021, **12**, 572845.
5. Zhang, Y.; Tanno, T.; Kanelloupolou, C. Cancer therapeutic implications of microRNAs in the regulation of immune checkpoint blockade. *ExRNA* 2019, **1**, 19.
6. Shao, L.; He, Q.; Wang, J.; He, F.; Lin, S.; Wu, L.; Gao, Y.; Ma, W.; Dong, J.; Yang, X.; et al. MicroRNA-326 attenuates immune escape and prevents metastasis in lung adenocarcinoma by targeting PD-L1 and B7-H3. *Cell Death Discov.* 2021, **7**, 145.
7. Xu, W.; Atkins, M.B.; McDermott, D.F. Checkpoint inhibitor immunotherapy in kidney cancer. *Nat. Rev. Urol.* 2020, **17**, 137–150.
8. Soleimani, M.; Thi, M.; Saxena, N.; Khalaf, D.J.; Eigl, B.J.; Chi, K.N.; Kollmannsberger, C.K.; Nappi, L. 693P Plasma exosome microRNA-155-3p expression in patients with metastatic renal cell carcinoma treated with immune checkpoint inhibitors: Potential biomarker of response to systemic therapy. *Ann. Oncol.* 2021, **32**, S708.
9. Zhu, J.; Chen, L.; Zou, L.; Yang, P.; Wu, R.; Mao, Y.; Zhou, H.; Li, R.; Wang, K.; Wang, W.; et al. MiR-20b, -21, and -130b inhibit PTEN expression resulting in B7-H1 over-expression in advanced colorectal cancer. *Hum. Immunol.* 2014, **75**, 348–353.
10. Cortez, M.A.; Anfossi, S.; Ramapriyan, R.; Menon, H.; Atalar, S.C.; Aliru, M.; Welsh, J.; Calin, G.A. Role of miRNAs in immune responses and immunotherapy in cancer. *Genes Chromosomes Cancer* 2019, **58**, 244–253.
11. Nguyen, M.-H.T.; Luo, Y.-H.; Li, A.-L.; Tsai, J.-C.; Wu, K.-L.; Chung, P.-J.; Ma, N. miRNA as a Modulator of Immunotherapy and Immune Response in Melanoma. *Biomolecules* 2021, **11**, 1648.
12. Yi, M.; Xu, L.; Jiao, Y.; Luo, S.; Li, A.; Wu, K. The role of cancer-derived microRNAs in cancer immune escape. *J. Hematol. Oncol.* 2020, **13**, 25.
13. Omar, H.A.; El-Serafi, A.T.; Hersi, F.; Arafa, E.A.; Zaher, D.M.; Madkour, M.; Arab, H.H.; Tolba, M.F. Immunomodulatory MicroRNAs in cancer: Targeting immune checkpoints and the tumor microenvironment. *FEBS J.* 2019, **286**, 3540–3557.
14. Skafi, N.; Fayyad-Kazan, M.; Badran, B. Immunomodulatory role for MicroRNAs: Regulation of PD-1/PD-L1 and CTLA-4 immune checkpoints expression. *Gene* 2020, **754**, 144888.
15. Eichmüller, S.B.; Osen, W.; Mandelboim, O.; Seliger, B. Immune Modulatory microRNAs Involved in Tumor Attack and Tumor Immune Escape. *JNCI J. Natl. Cancer Inst.* 2017, **109**, djx034.
16. Huang, F.; Wang, B.; Zeng, J.; Sang, S.; Lei, J.; Lu, Y. MicroRNA-374b inhibits liver cancer progression via down regulating programmed cell death-1 expression on cytokine-induced killer cells. *Oncol. Lett.* 2018, **15**, 4797–4804.
17. Zhang, G.; Li, N.; Li, Z.; Zhu, Q.; Li, F.; Yang, C.; Han, Q.; Lv, Y.; Zhou, Z.; Liu, Z. microRNA-4717 differentially interacts with its polymorphic target in the PD1 3' untranslated region: A mechanism for regulating PD-1 expression and function in HBV-associated liver diseases. *Oncotarget* 2015, **6**, 18933–18944.
18. Zhang, Q.; Di, W.; Dong, Y.; Lu, G.; Yu, J.; Li, J.; Li, P. High serum miR-183 level is associated with poor responsiveness of renal cancer to natural killer cells. *Tumor Biol.* 2015, **36**, 9245–9249.
19. Grenda, A.; Krawczyk, P.; Błach, J.; Chmielewska, I.; Kubiatowski, T.; Kieszko, S.; Wojas-Krawczyk, K.; Kucharczyk, T.; Jarosz, B.; Paśnik, I.; et al. Tissue MicroRNA Expression as a Predictor of Response to Immunotherapy in NSCLC Patients. *Front. Oncol.* 2021, **10**, 563613.

20. Song, N.; Li, P.; Song, P.; Li, Y.; Zhou, S.; Su, Q.; Li, X.; Yu, Y.; Li, P.; Feng, M.; et al. MicroRNA-138-5p Suppresses Non-small Cell Lung Cancer Cells by Targeting PD-L1/PD-1 to Regulate Tumor Microenvironment. *Front. Cell Dev. Biol.* 2020, 8, 540.
21. Jia, L.; Xi, Q.; Wang, H.; Zhang, Z.; Liu, H.; Cheng, Y.; Guo, X.; Zhang, J.; Zhang, Q.; Zhang, L.; et al. miR-142-5p regulates tumor cell PD-L1 expression and enhances anti-tumor immunity. *Biochem. Biophys. Res. Commun.* 2017, 488, 425–431.
22. Aichen, Z.; Kun, W.; Xiaochun, S.; Lingling, T. LncRNA FGD5-AS1 promotes the malignant phenotypes of ovarian cancer cells via targeting miR-142-5p. *Apoptosis* 2021, 26, 348–360.
23. Qu, F.; Ye, J.; Pan, X.; Wang, J.; Gan, S.; Chu, C.; Chu, J.; Zhang, X.; Liu, M.; He, H.; et al. MicroRNA-497-5p down-regulation increases PD-L1 expression in clear cell renal cell carcinoma. *J. Drug Target.* 2019, 27, 67–74.
24. Zhu, J.; Chen, L.; Zou, L.; Yang, P.; Wu, R.; Mao, Y.; Zhou, H.; Li, R.; Wang, K.; Wang, W.; et al. MiR-20b, -21, and -130b inhibit PTEN expression resulting in B7-H1 over-expression in advanced colorectal cancer. *Hum. Immunol.* 2014, 75, 348–353.
25. Zhao, L.; Yu, H.; Yi, S.; Peng, X.; Su, P.; Xiao, Z.; Liu, R.; Tang, A.; Li, X.; Liu, F.; et al. The tumor suppressor miR-138-5p targets PD-L1 in colorectal cancer. *Oncotarget* 2016, 7, 45370–45384.
26. Ashizawa, M.; Okayama, H.; Ishigame, T.; Thar Min, A.K.; Saito, K.; Ujiie, D.; Murakami, Y.; Kikuchi, T.; Nakayama, Y.; Noda, M.; et al. miRNA-148a-3p Regulates Immunosuppression in DNA Mismatch Repair–Deficient Colorectal Cancer by Targeting PD-L1. *Mol. Cancer Res.* 2019, 17, 1403–1413.
27. Chen, X.-Y.; Zhang, J.; Hou, L.-D.; Zhang, R.; Chen, W.; Fan, H.-N.; Huang, Y.-X.; Liu, H.; Zhu, J.-S. Upregulation of PD-L1 predicts poor prognosis and is associated with miR-191-5p dysregulation in colon adenocarcinoma. *Int. J. Immunopathol. Pharmacol.* 2018, 32, 205873841879031.
28. Gao, L.; Guo, Q.; Li, X.; Yang, X.; Ni, H.; Wang, T.; Zhao, Q.; Liu, H.; Xing, Y.; Xi, T.; et al. MiR-873/PD-L1 axis regulates the stemness of breast cancer cells. *EBioMedicine* 2019, 41, 395–407.
29. Yang, L.; Cai, Y.; Zhang, D.; Sun, J.; Xu, C.; Zhao, W.; Jiang, W.; Pan, C. miR-195/miR-497 Regulate CD274 Expression of Immune Regulatory Ligands in Triple-Negative Breast Cancer. *J. Breast Cancer* 2018, 21, 371.
30. Li, D.; Wang, X.; Yang, M.; Kan, Q.; Duan, Z. miR3609 sensitizes breast cancer cells to adriamycin by blocking the programmed death-ligand 1 immune checkpoint. *Exp. Cell Res.* 2019, 380, 20–28.
31. Dastmalchi, N.; Hosseinpourfeizi, M.A.; Khojasteh, S.M.B.; Baradaran, B.; Safaralizadeh, R. Tumor suppressive activity of miR-424-5p in breast cancer cells through targeting PD-L1 and modulating PTEN/PI3K/AKT/mTOR signaling pathway. *Life Sci.* 2020, 259, 118239.
32. Audrito, V.; Serra, S.; Stingi, A.; Orso, F.; Gaudino, F.; Bologna, C.; Neri, F.; Garaffo, G.; Nassini, R.; Baroni, G.; et al. PD-L1 up-regulation in melanoma increases disease aggressiveness and is mediated through miR-17-5p. *Oncotarget* 2017, 8, 15894–15911.
33. Mastroianni, J.; Stickel, N.; Andrllova, H.; Hanke, K.; Melchinger, W.; Duquesne, S.; Schmidt, D.; Falk, M.; Andrieux, G.; Pfeifer, D.; et al. miR-146a Controls Immune Response in the Melanoma Microenvironment. *Cancer Res.* 2019, 79, 183–195.
34. Kao, S.C.; Cheng, Y.Y.; Williams, M.; Kirschner, M.B.; Madore, J.; Lum, T.; Sarun, K.H.; Linton, A.; McCaughan, B.; Klebe, S.; et al. Tumor Suppressor microRNAs Contribute to the Regulation of PD-L1 Expression in Malignant Pleural Mesothelioma. *J. Thorac. Oncol.* 2017, 12, 1421–1433.
35. Costa, C.; Indovina, P.; Mattioli, E.; Forte, I.M.; Iannuzzi, C.A.; Luzzi, L.; Bellan, C.; De Summa, S.; Bucci, E.; Di Marzo, D.; et al. P53-regulated miR-320a targets PDL1 and is downregulated in malignant mesothelioma. *Cell Death Dis.* 2020, 11, 748.
36. He, B.; Yan, F.; Wu, C. Overexpressed miR-195 attenuated immune escape of diffuse large B-cell lymphoma by targeting PD-L1. *Biomed. Pharmacother.* 2018, 98, 95–101.
37. Zheng, Z.; Sun, R.; Zhao, H.-J.; Fu, D.; Zhong, H.-J.; Weng, X.-Q.; Qu, B.; Zhao, Y.; Wang, L.; Zhao, W.-L. MiR155 sensitized B-lymphoma cells to anti-PD-L1 antibody via PD-1/PD-L1-mediated lymphoma cell interaction with CD8+T cells. *Mol. Cancer* 2019, 18, 54.
38. Sun, J.-R.; Zhang, X.; Zhang, Y. MiR-214 prevents the progression of diffuse large B-cell lymphoma by targeting PD-L1. *Cell. Mol. Biol. Lett.* 2019, 24, 68.
39. Tao, Z.; Xu, S.; Ruan, H.; Wang, T.; Song, W.; Qian, L.; Chen, K. MiR-195/-16 Family Enhances Radiotherapy via T Cell Activation in the Tumor Microenvironment by Blocking the PD-L1 Immune Checkpoint. *Cell. Physiol. Biochem.* 2018, 48, 801–814.

40. Cortez, M.A.; Ivan, C.; Valdecanas, D.; Wang, X.; Peltier, H.J.; Ye, Y.; Araujo, L.; Carbone, D.P.; Shilo, K.; Giri, D.K.; et al. PDL1 Regulation by p53 via miR-34. *JNCI J. Natl. Cancer Inst.* 2016, 108, djv303.
41. Xie, W.-B.; Liang, L.-H.; Wu, K.-G.; Wang, L.-X.; He, X.; Song, C.; Wang, Y.-Q.; Li, Y.-H. MiR-140 Expression Regulates Cell Proliferation and Targets PD-L1 in NSCLC. *Cell. Physiol. Biochem.* 2018, 46, 654–663.
42. Tang, D.; Zhao, D.; Wu, Y.; Yao, R.; Zhou, L.; Lu, L.; Gao, W.; Sun, Y. The miR-3127-5p/p-STAT3 axis up-regulates PD-L1 inducing chemoresistance in non-small-cell lung cancer. *J. Cell. Mol. Med.* 2018, 22, 3847–3856.
43. Chen, L.; Gibbons, D.L.; Goswami, S.; Cortez, M.A.; Ahn, Y.-H.; Byers, L.A.; Zhang, X.; Yi, X.; Dwyer, D.; Lin, W.; et al. Metastasis is regulated via microRNA-200/ZEB1 axis control of tumour cell PD-L1 expression and intratumoral immunosuppression. *Nat. Commun.* 2014, 5, 5241.
44. Wei, S.; Wang, K.; Huang, X.; Zhao, Z.; Zhao, Z. LncRNA MALAT1 contributes to non-small cell lung cancer progression via modulating miR-200a-3p/programmed death-ligand 1 axis. *Int. J. Immunopathol. Pharmacol.* 2019, 33, 205873841985969.
45. Wang, X.; Li, J.; Dong, K.; Lin, F.; Long, M.; Ouyang, Y.; Wei, J.; Chen, X.; Weng, Y.; He, T.; et al. Tumor suppressor miR-34a targets PD-L1 and functions as a potential immunotherapeutic target in acute myeloid leukemia. *Cell. Signal.* 2015, 27, 443–452.
46. Liu, J.; Fan, L.; Yu, H.; Zhang, J.; He, Y.; Feng, D.; Wang, F.; Li, X.; Liu, Q.; Li, Y.; et al. Endoplasmic Reticulum Stress Causes Liver Cancer Cells to Release Exosomal miR-23a-3p and Up-regulate Programmed Death Ligand 1 Expression in Macrophages. *Hepatology* 2019, 70, hep.30607.
47. Guo, W.; Tan, W.; Liu, S.; Huang, X.; Lin, J.; Liang, R.; Su, L.; Su, Q.; Wang, C. MiR-570 inhibited the cell proliferation and invasion through directly targeting B7-H1 in hepatocellular carcinoma. *Tumor Biol.* 2015, 36, 9049–9057.
48. Wu, Q.; Zhao, Y.; Sun, Y.; Yan, X.; Wang, P. miR-375 inhibits IFN- $\gamma$ -induced programmed death 1 ligand 1 surface expression in head and neck squamous cell carcinoma cells by blocking JAK2/STAT1 signaling. *Oncol. Rep.* 2018, 39, 1461–1468.
49. Sheng, Q.; Zhang, Y.; Wang, Z.; Ding, J.; Song, Y.; Zhao, W. Cisplatin-mediated down-regulation of miR-145 contributes to up-regulation of PD-L1 via the c-Myc transcription factor in cisplatin-resistant ovarian carcinoma cells. *Clin. Exp. Immunol.* 2020, 200, 45–52.
50. Zhu, J.; Li, Y.; Luo, Y.; Xu, J.; Liufu, H.; Tian, Z.; Huang, C.; Li, J.; Huang, C. A Feedback Loop Formed by ATG7/Autophagy, FOXO3a/miR-145 and PD-L1 Regulates Stem-Like Properties and Invasion in Human Bladder Cancer. *Cancers* 2019, 11, 349.
51. Wu, L.; Chen, Z.; Zhang, J.; Xing, Y. Effect of miR-513a-5p on etoposide-stimulating B7-H1 expression in retinoblastoma cells. *J. Huazhong Univ. Sci. Technol. Med. Sci.* 2012, 32, 601–606.
52. Miliotis, C.; Slack, F.J. miR-105-5p regulates PD-L1 expression and tumor immunogenicity in gastric cancer. *Cancer Lett.* 2021, 518, 115–126.
53. Wang, W.; Li, F.; Mao, Y.; Zhou, H.; Sun, J.; Li, R.; Liu, C.; Chen, W.; Hua, D.; Zhang, X. A miR-570 binding site polymorphism in the B7-H1 gene is associated with the risk of gastric adenocarcinoma. *Hum. Genet.* 2013, 132, 641–648.
54. Wang, Y.; Wang, D.; Xie, G.; Yin, Y.; Zhao, E.; Tao, K.; Li, R. MicroRNA-152 regulates immune response via targeting B7-H1 in gastric carcinoma. *Oncotarget* 2017, 8, 28125–28134.
55. Xie, G.; Li, W.; Li, R.; Wu, K.; Zhao, E.; Zhang, Y.; Zhang, P.; Shi, L.; Wang, D.; Yin, Y.; et al. Helicobacter Pylori Promote B7-H1 Expression by Suppressing miR-152 and miR-200b in Gastric Cancer Cells. *PLoS ONE* 2017, 12, e0168822.
56. Qian, L.; Liu, F.; Chu, Y.; Zhai, Q.; Wei, X.; Shao, J.; Li, R.; Xu, Q.; Yu, L.; Liu, B.; et al. MicroRNA-200c Nanoparticles Sensitized Gastric Cancer Cells to Radiotherapy by Regulating PD-L1 Expression and EMT. *Cancer Manag. Res.* 2020, 12, 12215–12223.
57. Dong, P.; Xiong, Y.; Yu, J.; Chen, L.; Tao, T.; Yi, S.; Hanley, S.J.B.; Yue, J.; Watari, H.; Sakuragi, N. Control of PD-L1 expression by miR-140/142/340/383 and oncogenic activation of the OCT4–miR-18a pathway in cervical cancer. *Oncogene* 2018, 37, 5257–5268.
58. Miao, S.; Mao, X.; Zhao, S.; Song, K.; Xiang, C.; Lv, Y.; Jiang, H.; Wang, L.; Li, B.; Yang, X.; et al. miR-217 inhibits laryngeal cancer metastasis by repressing AEG-1 and PD-L1 expression. *Oncotarget* 2017, 8, 62143–62153.
59. Jiang, K.; Zou, H. microRNA-20b-5p overexpression combing Pembrolizumab potentiates cancer cells to radiation therapy via repressing programmed death-ligand 1. *Bioengineered* 2022, 13, 917–929.

60. Wang, C.; Li, X.; Zhang, L.; Chen, Y.; Dong, R.; Zhang, J.; Zhao, J.; Guo, X.; Yang, G.; Li, Y.; et al. miR-194-5p downregulates tumor cell PD-L1 expression and promotes anti-tumor immunity in pancreatic cancer. *Int. Immunopharmacol.* 2021, **97**, 107822.
61. Shao, L.; He, Q.; Wang, J.; He, F.; Lin, S.; Wu, L.; Gao, Y.; Ma, W.; Dong, J.; Yang, X.; et al. MicroRNA-326 attenuates immune escape and prevents metastasis in lung adenocarcinoma by targeting PD-L1 and B7-H3. *Cell Death Discov.* 2021, **7**, 145.
62. Xu, S.; Tao, Z.; Hai, B.; Liang, H.; Shi, Y.; Wang, T.; Song, W.; Chen, Y.; OuYang, J.; Chen, J.; et al. miR-424(322) reverses chemoresistance via T-cell immune response activation by blocking the PD-L1 immune checkpoint. *Nat. Commun.* 2016, **7**, 11406.
63. Wei, J.; Nduom, E.K.; Kong, L.-Y.; Hashimoto, Y.; Xu, S.; Gabrusiewicz, K.; Ling, X.; Huang, N.; Qiao, W.; Zhou, S.; et al. MiR-138 exerts anti-glioma efficacy by targeting immune checkpoints. *Neuro. Oncol.* 2016, **18**, 639–648.
64. Zhao, X.; Yuan, C.; Wangmo, D.; Subramanian, S. Tumor-Secreted Extracellular Vesicles Regulate T-Cell Costimulation and Can Be Manipulated To Induce Tumor-Specific T-Cell Responses. *Gastroenterology* 2021, **161**, 560–574.e11.
65. Boldrini, L.; Giordano, M.; Niccoli, C.; Melfi, F.; Lucchi, M.; Mussi, A.; Fontanini, G. Role of microRNA-33a in regulating the expression of PD-1 in lung adenocarcinoma. *Cancer Cell Int.* 2017, **17**, 105.
66. Li, Q.; Johnston, N.; Zheng, X.; Wang, H.; Zhang, X.; Gao, D.; Min, W. miR-28 modulates exhaustive differentiation of T cells through silencing programmed cell death-1 and regulating cytokine secretion. *Oncotarget* 2016, **7**, 53735–53750.
67. Zhang, R.-R.; Wang, L.-M.; Shen, J.-J. Overexpression of miR-32 inhibits the proliferation and metastasis of ovarian cancer cells by targeting BTLA. *Eur. Rev. Med. Pharmacol. Sci.* 2020, **24**, 4671–4678.
68. Moghaddam, Y.; Andalib, A.; Mohammad-Ganji, M.; Homayouni, V.; Sharifi, M.; Ganjalikhani-Hakemi, M. Evaluation of the effect of TIM-3 suppression by miR-498 and its effect on apoptosis and proliferation rate of HL-60 cell line. *Pathol. Res. Pract.* 2018, **214**, 1482–1488.
69. Lou, Q.; Liu, R.; Yang, X.; Li, W.; Huang, L.; Wei, L.; Tan, H.; Xiang, N.; Chan, K.; Chen, J.; et al. miR-448 targets IDO1 and regulates CD8+ T cell response in human colon cancer. *J. Immunother. Cancer* 2019, **7**, 210.
70. Huang, Q.; Xia, J.; Wang, L.; Wang, X.; Ma, X.; Deng, Q.; Lu, Y.; Kumar, M.; Zhou, Z.; Li, L.; et al. miR-153 suppresses IDO1 expression and enhances CAR T cell immunotherapy. *J. Hematol. Oncol.* 2018, **11**, 58.
71. Bieg, D.; Sypniewski, D.; Nowak, E.; Bednarek, I. MiR-424-3p suppresses galectin-3 expression and sensitizes ovarian cancer cells to cisplatin. *Arch. Gynecol. Obstet.* 2019, **299**, 1077–1087.
72. Lu, W.; Wang, J.; Yang, G.; Yu, N.; Huang, Z.; Xu, H.; Li, J.; Qiu, J.; Zeng, X.; Chen, S.; et al. Posttranscriptional regulation of Galectin-3 by miR-128 contributes to colorectal cancer progression. *Oncotarget* 2017, **8**, 15242–15251.
73. Yang, Q.; Jiang, W.; Zhuang, C.; Geng, Z.; Hou, C.; Huang, D.; Hu, L.; Wang, X. microRNA-22 downregulation of galectin-9 influences lymphocyte apoptosis and tumor cell proliferation in liver cancer. *Oncol. Rep.* 2015, **34**, 1771–1778.
74. Yang, Q.; Hou, C.; Huang, D.; Zhuang, C.; Jiang, W.; Geng, Z.; Wang, X.; Hu, L. miR-455-5p functions as a potential oncogene by targeting galectin-9 in colon cancer. *Oncol. Lett.* 2017, **13**, 1958–1964.
75. Morishita, A.; Nomura, K.; Tani, J.; Fujita, K.; Iwama, H.; Takuma, K.; Nakahara, M.; Tadokoro, T.; Oura, K.; Chiyo, T.; et al. Galectin-9 suppresses the tumor growth of colon cancer in vitro and in vivo. *Oncol. Rep.* 2021, **45**, 105.
76. Yang, P.; Tang, R.; Zhu, J.; Zou, L.; Wu, R.; Zhou, H.; Mao, Y.; Li, R.; Hua, D.; Wang, W.; et al. A functional variant at miR-24 binding site in B7-H2 alters susceptibility to gastric cancer in a Chinese Han population. *Mol. Immunol.* 2013, **56**, 98–103.
77. Xu, H.; Cheung, I.Y.; Guo, H.-F.; Cheung, N.-K.V. MicroRNA miR-29 Modulates Expression of Immunoinhibitory Molecule B7-H3: Potential Implications for Immune Based Therapy of Human Solid Tumors. *Cancer Res.* 2009, **69**, 6275–6281.
78. Huang, L. The expression and clinical significance of B7-H3 and miR-145 in lung cancer patients with malignant pleural effusion. *Eur. Rev. Med. Pharmacol. Sci.* 2020, **24**, 6759–6766.
79. Wang, L.; Zhao, Y.; Xu, M.; Zhou, F.; Yan, J. Serum miR-1301-3p, miR-335-5p, miR-28-5p, and their target B7-H3 may serve as novel biomarkers for colorectal cancer. *J. BUON* 2019, **24**, 1120–1127.
80. Zhou, X.; Mao, Y.; Zhu, J.; Meng, F.; Chen, Q.; Tao, L.; Li, R.; Fu, F.; Liu, C.; Hu, Y.; et al. TGF- $\beta$ 1 promotes colorectal cancer immune escape by elevating B7-H3 and B7-H4 via the miR-155/miR-143 axis. *Oncotarget* 2016, **7**, 67196–67211.

81. Hu, X.; Xu, M.; Hu, Y.; Li, N.; Zhou, L. B7-H3, Negatively Regulated by miR-128, Promotes Colorectal Cancer Cell Proliferation and Migration. *Cell Biochem. Biophys.* 2021, 79, 397–405.
82. Wang, J.; Chen, X.; Xie, C.; Sun, M.; Hu, C.; Zhang, Z.; Luan, L.; Zhou, J.; Zhou, J.; Zhu, X.; et al. MicroRNA miR-29a Inhibits Colon Cancer Progression by Downregulating B7-H3 Expression: Potential Molecular Targets for Colon Cancer Therapy. *Mol. Biotechnol.* 2021, 63, 849–861.
83. Wang, Z.-S.; Zhong, M.; Bian, Y.-H.; Mu, Y.-F.; Qin, S.-L.; Yu, M.-H.; Qin, J. MicroRNA-187 inhibits tumor growth and invasion by directly targeting CD276 in colorectal cancer. *Oncotarget* 2016, 7, 44266–44276.
84. Zhao, J.; Lei, T.; Xu, C.; Li, H.; Ma, W.; Yang, Y.; Fan, S.; Liu, Y. MicroRNA-187, down-regulated in clear cell renal cell carcinoma and associated with lower survival, inhibits cell growth and migration though targeting B7-H3. *Biochem. Biophys. Res. Commun.* 2013, 438, 439–444.
85. Wang, J.; Chong, K.K.; Nakamura, Y.; Nguyen, L.; Huang, S.K.; Kuo, C.; Zhang, W.; Yu, H.; Morton, D.L.; Hoon, D.S.B. B7-H3 Associated with Tumor Progression and Epigenetic Regulatory Activity in Cutaneous Melanoma. *J. Investig. Dermatol.* 2013, 133, 2050–2058.
86. Wang, S.; Mou, J.; Cui, L.; Wang, X.; Zhang, Z. Astragaloside IV inhibits cell proliferation of colorectal cancer cell lines through down-regulation of B7-H3. *Biomed. Pharmacother.* 2018, 102, 1037–1044.
87. Nygren, M.K.; Tekle, C.; Ingebrigtsen, V.A.; Mäkelä, R.; Krohn, M.; Aure, M.R.; Nunes-Xavier, C.E.; Perälä, M.; Tramm, T.; Alsner, J.; et al. Identifying microRNAs regulating B7-H3 in breast cancer: The clinical impact of microRNA-29c. *Br. J. Cancer* 2014, 110, 2072–2080.
88. Li, R.G.; Gao, Z.; Jiang, Y. B7-H3 repression by miR-539 suppresses cell proliferation in human gliomas. *Int. J. Clin. Exp. Pathol.* 2017, 10, 4363–4369.
89. Wang, L.; Kang, F.; Sun, N.; Wang, J.; Chen, W.; Li, D.; Shan, B. The tumor suppressor miR-124 inhibits cell proliferation and invasion by targeting B7-H3 in osteosarcoma. *Tumor Biol.* 2016, 37, 14939–14947.
90. Zhu, X.; Wang, J.; Zhu, M.; Wang, Y.; Yang, S.; Ke, X. MicroRNA-506 inhibits the proliferation and invasion of mantle cell lymphoma cells by targeting B7 H3. *Biochem. Biophys. Res. Commun.* 2019, 508, 1067–1073.
91. Gao, Y.; Fang, P.; Li, W.-J.; Zhang, J.; Wang, G.-P.; Jiang, D.-F.; Chen, F.-P. LncRNA NEAT1 sponges miR-214 to regulate M2 macrophage polarization by regulation of B7-H3 in multiple myeloma. *Mol. Immunol.* 2020, 117, 20–28.
92. Purvis, I.J.; Avilala, J.; Guda, M.R.; Venkataraman, S.; Vibhakar, R.; Tsung, A.J.; Velpula, K.K.; Asuthkar, S. Role of MYC-miR-29-B7-H3 in Medulloblastoma Growth and Angiogenesis. *J. Clin. Med.* 2019, 8, 1158.
93. Kanchan, R.K.; Perumal, N.; Atri, P.; Chiravuri Venkata, R.; Thapa, I.; Klinkebiel, D.L.; Donson, A.M.; Perry, D.; Punsoni, M.; Talmon, G.A.; et al. MiR-1253 exerts tumor-suppressive effects in medulloblastoma via inhibition of CDK6 and CD276 (B7-H3). *Brain Pathol.* 2020, 30, 732–745.
94. Yang, X.; Feng, K.-X.; Li, H.; Wang, L.; Xia, H. MicroRNA-199a Inhibits Cell Proliferation, Migration, and Invasion and Activates AKT/mTOR Signaling Pathway by Targeting B7-H3 in Cervical Cancer. *Technol. Cancer Res. Treat.* 2020, 19, 153303382094224.
95. Oliveira, P.; Carvalho, J.; Rocha, S.; Azevedo, M.; Reis, I.; Camilo, V.; Sousa, B.; Valente, S.; Paredes, J.; Almeida, R.; et al. Dies1/VISTA expression loss is a recurrent event in gastric cancer due to epigenetic regulation. *Sci. Rep.* 2016, 6, 34860.
96. Wu, D.; Tang, R.; Qi, Q.; Zhou, X.; Zhou, H.; Mao, Y.; Li, R.; Liu, C.; Wang, W.; Hua, D.; et al. Five functional polymorphisms of B7/CD28 co-signaling molecules alter susceptibility to colorectal cancer. *Cell. Immunol.* 2015, 293, 41–48.
97. Qian, Y.; Feng, L.; Wu, W.; Weng, T.; Hu, C.; Hong, B.; Wang, F.X.C.; Shen, L.; Wang, Q.; Jin, X.; et al. MicroRNA Expression Profiling of Pancreatic Cancer Cell Line L3.6p1 Following B7-H4 Knockdown. *Cell. Physiol. Biochem.* 2017, 44, 494–504.
98. Wang, B.; Ran, Z.; Liu, M.; Ou, Y. Prognostic Significance of Potential Immune Checkpoint Member HHLA2 in Human Tumors: A Comprehensive Analysis. *Front. Immunol.* 2019, 10, 1573.