

# Lycorine Ameliorates Thioacetamide-Induced Hepatic Fibrosis in Rats

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Liver fibrosis is a foremost medical concern worldwide. Lycorine—a natural alkaloid—has antioxidant, anti-inflammatory, and antitumor activities. Lycorine hinders TAA-induced liver fibrosis in rats, due to—at least partly—its antioxidative and anti-inflammatory properties, along with its ability to inhibit Signal Transducer and Activator of Transcription factor (STAT3) signaling.

Keywords: lycorine ; thioacetamide ; liver fibrosis ; STAT3

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## 1. Introduction

Liver damage could lead to deleterious consequences to the human body, including death <sup>[1]</sup>. Liver disease is responsible for two million deaths every year globally, with cirrhosis representing the 11th most common cause of mortality worldwide <sup>[2]</sup>. Metabolic disorders, including insulin resistance, diabetes mellitus, obesity, and dyslipidemia, contribute to liver damage <sup>[3]</sup>. Liver fibrosis is a healing process where extracellular matrix (ECM) proteins accumulate as a substituent of hepatocytes after persistent liver injury <sup>[4][5]</sup>. Both oxidative stress and inflammation were conveyed in all fibrotic disorders with chronic damage and remodeling <sup>[6]</sup>. Interestingly, Signal Transducer and Activator of Transcription factor (STAT3) is physiologically stimulated during tissue repair as a result of inflammation, participating in the initial steps of the healing process <sup>[7]</sup>. However, if it is insistently activated, it could contribute to damaging effects and various disorders, including organ fibrosis <sup>[8][9]</sup>. STAT3 has been well-known to confer potent proliferative actions and apoptotic resistance, which could contribute to the replication of myofibroblasts, with a subsequent buildup of connective tissue in fibrotic states <sup>[10]</sup>. Therefore, blocking STAT3 could be a favorable aim for antifibrotic action.

Inopportune, there is no reasonable therapy for hepatic fibrosis, which requires chronic use of safe and effective drugs <sup>[3]</sup>. On the other hand, respectable recognition of natural products has been increasing among the scientific and public communities <sup>[11]</sup>. In this regard, lycorine—a natural alkaloid extracted from the genus Amaryllidaceae—was formerly reported to inhibit the growth and cell division in yeasts, algae, and higher plants <sup>[12]</sup>. Then, studies were continued to detect other activities of lycorine, including antitumor, anti-inflammatory, and antioxidant effects <sup>[13][14][15]</sup>. Another study <sup>[16]</sup> indicated the hepatoprotective effects of lycorine in an animal model of CCl<sub>4</sub>-induced acute hepatotoxicity. More recently, lycorine has been reported to activate the mitochondrial apoptosis pathway via targeting STAT3, which was effective at inhibiting the malignant growth of colorectal cancer both in-vivo and in-vitro <sup>[17]</sup>. Although the antifibrotic activity of lycorine has been investigated in experimental models like bleomycin-induced pulmonary fibrosis <sup>[18]</sup>, as well as several experimentally-induced cardiac dysfunctions <sup>[19][20][21]</sup>, there is a paucity of information regarding the antifibrotic activity of lycorine against experimentally-induced hepatic fibrosis with respect to STAT3 activity.

## 2. The Potential Antifibrotic Effect of Lycorine against Thioacetamide-Induced Liver Fibrosis in Rats

Presently, there is no reasonable remedy for liver fibrosis <sup>[22]</sup>. Lycorine is a natural alkaloid that possesses antitumor <sup>[13]</sup> <sup>[14]</sup>, antioxidant <sup>[23]</sup>, and anti-inflammatory activities <sup>[24]</sup>, in addition to being hepatoprotective in CCl<sub>4</sub>-induced acute hepatotoxicity <sup>[16]</sup>. Moreover, it has been reported to activate the mitochondrial apoptosis pathway via targeting STAT3 <sup>[17]</sup>.

For decades, TAA has been known for its ability to induce liver fibrosis <sup>[25]</sup>. Yet, its molecular mechanism is still not fully unstated. Indeed, TAA undergoes bioactivation in the liver via oxidation processes via hepatic CYP2E1 <sup>[26][27]</sup>. This leads to the formation of reactive metabolites, namely, S-oxide and SS-dioxide, which are apparently responsible for TAA-induced hepatic injury <sup>[28]</sup>. Basically, liver injury activates hepatic stellate cells (HSCs) to multiply and secrete extracellular matrix (ECM) components like interstitial collagens <sup>[29][30]</sup>. Moreover, HSCs are converted into myofibroblasts expressing  $\alpha$ -SMA, which induces fibrogenesis and tissue stiffness <sup>[31]</sup>. By mechanisms including autocrine and paracrine pathways,

ECM components promote growth factor signaling, principally by TGF- $\beta$ , which in-turn add to the activation of HSCs, generating a positive feedback circle [32]. It is worth mentioning that the TGF- $\beta$  family comprises three isoforms TGF- $\beta$  1, 2, and 3 with different biological activities [33]. The isoform TGF- $\beta$ 1 gained the most importance due to its pleiotropic nature, extending from tissue fibrosis to preparing the microenvironment for carcinogenesis [34][35]. With regard to hepatic pathogenesis, the assessed TGF- $\beta$ 1 has been reported to have a critical role in hepatic fibrosis through direct activation of HSCs and accumulation of ECM [36]. Lycorine treatment significantly diminished TAA-induced hepatic injury by significantly ameliorating the rise of hepatic transaminases “ALT and AST” and the fibrotic marker “hydroxyproline”. This was confirmed by its ability to improve histopathological-fibrotic changes, as evidenced by H&E, MT, and SR staining, and to guard against the excessive immunohistochemical expression of  $\alpha$ SMA and TGF $\beta$ 1. These findings are in line with recent studies investigating the antifibrotic activity of lycorine against bleomycin-induced pulmonary fibrosis [18], as well as experimentally-induced cardiac dysfunction [19][20][21].

Oxidative stress and inflammation have crucial roles in HSCs activation, which ultimately ends in fibrogenesis [31]. Lycorine treatment significantly improved TAA-induced oxidative stress in hepatic tissues, as manifested by the enhanced GSH content, SOD activity, and reduced lipid peroxidation. This is in agreement with previous reports, confirming the antioxidant activity of lycorine as a DPPH scavenger [15] and its capability of protecting human erythrocytes against 2-amidinopropane-induced oxidative hemolysis [23]. In addition, lycorine exhibited hepatoprotective effects against CCl<sub>4</sub>-induced oxidative stress in mice [37]. Concerning inflammatory cascade, it is notorious that activated HSCs express cytokines that are key mediators of fibrogenesis. Specifically, the proinflammatory cytokine TNF- $\alpha$  is released from HSCs secondary to TGF- $\beta$ 1, which in turn inaugurates inflammatory responses via stimulating the secretion of other proinflammatory cytokines, such as IL-1 $\beta$  [38]. In this, lycorine treatment significantly ameliorated TAA-induced hepatic inflammation, as manifested by diminishing the rise of TNF- $\alpha$ , IL-1 $\beta$ , and IL-6. Notably, lycorine was found to possess an efficient anti-inflammatory activity in numerous experimental models such as rat adjuvant arthritis [39] carrageen-induced rat paw edema [16]; and lipopolysaccharide (LPS) challenge in RAW264.7 cells via inhibiting iNOS, PGE<sub>2</sub>, TNF- $\alpha$ , IL-6, and JAK-STAT3 signaling pathways [24].

### 3. The Possible Causal Mechanisms with Respect to the STAT3 Pathway

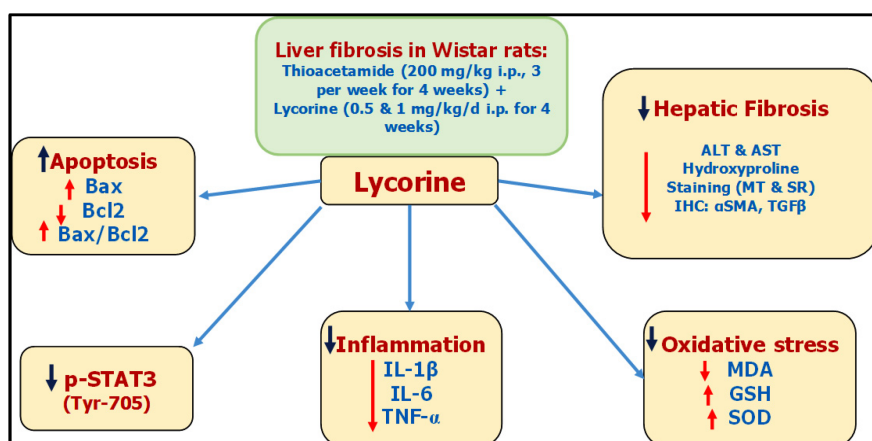
Recently, STAT3 has been documented to be closely linked to the existence and progress of liver fibrosis, triggered by numerous factors [40]. It is noted that STAT3 activation could cross-talk with TGF- $\beta$ 1 signaling in HSCs, exacerbating liver injury. In addition, the knockdown of STAT3 mRNA by siRNA could suppress the expression of TGF- $\beta$ 1 [36]. Many synthetic STAT3 inhibitors have been designed as chemical probes to endorse the impact of STAT3 in chemical-induced liver injury. For instance, STAT3 dimerization inhibitor “STX-0119” is suggested to play a role in controlling the development of CCl<sub>4</sub> and TAA-induced liver fibrosis through lessening the activated HSCs [41]. Another synthetic inhibitor, “HJC0123”, has been shown to render the liver more resistant to fibrosis by inhibiting the phosphorylation, nuclear translocation, and transcriptional activity of STAT3 [42]. Moreover, some herbal medicines have shown an inhibitory effect on STAT3 and offered protection against liver fibrosis induced by CCl<sub>4</sub>, for instance; cucurbitacin-B from many plants of the family Cucurbitaceae [43]; asiatic acid extracted from *Centella asiatica* [44]; S-allyl-cysteine (SAC) from aged garlic extract [45]; and “CCM111” extracted from *Antrodia cinnamomea* [46].

Here, lycorine was able to diminish STAT3 activation, as evidenced by decreasing the phospho-STAT3 (Tyr-705) expression induced by TAA in hepatic tissue, which in turn protected against liver fibrosis. This was corroborated by the lowered antiapoptotic Bcl-2 in conjunction with amplified apoptotic Bax mRNA expressions, leading to the rise of the calculated Bax/Bcl-2 ratio. The observed findings are in accordance with the study of Kang et al. [24], in which lycorine inhibited STAT3 activation in LPS-challenged RAW264.7 cells in-vitro. Furthermore, Wu et al. [17] indicated—by molecular docking—that lycorine inactivates phospho-STAT3 (Tyr-705) by directly binding to its SH2 domain. Accordingly, lycorine stimulated apoptosis in human colorectal cancer cells in-vitro, as evidenced by the activation of caspase and the increase in the ratio of Bax/Bcl-2. In this regard, lycorine is considered to be an apoptosis inducer of both mitochondrial and death receptor-mediated pathways in cancer cells, like breast and bladder cancer, in addition to hematological malignancies, including leukemia and myeloma. It was found that lycorine downregulates the expression of antiapoptotic Bcl-2 family proteins and increases the expression of proapoptotic BAX [47][48]. Recent studies have indicated that the induction of apoptosis in HSCs could ameliorate the progression of liver fibrosis [49][50].

### 4. Conclusions

It could be concluded that lycorine hinders TAA-induced liver fibrosis in rats, due to—at least partly—its antioxidative and anti-inflammatory properties along with its ability to inhibit STAT3 signaling. These proposed mechanisms have been summarized as a collective diagram, as shown in **Figure 1**. However, more studies are needed in order to establish the

clinical applicability of lycorine treatment in patients with active liver fibrogenesis. The experimental beneficial effects of lycorine in ameliorating liver fibrosis require additional confirmatory studies in the clinical setting to assess its safety and efficacy.



**Figure 1.** A collective diagram of lycorine antifibrotic effects against TAA-induced hepatic fibrosis in rats.

## References

1. Rouiller, C. The Liver: Morphology, Biochemistry, Physiology; Academic Press: Cambridge, MA, USA, 2013.
2. Asrani, S.K.; Devarbhavi, H.; Eaton, J.; Kamath, P.S. Burden of liver diseases in the world. *J. Hepatol.* 2019, 70, 151–171.
3. Bataller, R.; Brenner, D.A. Liver fibrosis. *J. Clin. Investig.* 2005, 115, 209–218.
4. Pinzani, M. Pathophysiology of Liver Fibrosis. *Dig. Dis.* 2015, 33, 492–497.
5. Seki, E.; Brenner, D.A. Recent advancement of molecular mechanisms of liver fibrosis. *J. Hepato-Biliary-Pancreat. Sci.* 2015, 22, 512–518.
6. Novo, E.; Parola, M. Redox mechanisms in hepatic chronic wound healing and fibrogenesis. *Fibrogenes. Tissue Repair* 2008, 1, 5.
7. Dauer, D.J.; Ferraro, B.; Song, L.; Yu, B.; Mora, L.; Buettner, R.; Enkemann, S.; Jove, R.; Haura, E.B. Stat3 regulates genes common to both wound healing and cancer. *Oncogene* 2005, 24, 3397–3408.
8. Levy, D.E.; Darnell, J.E. STATs: Transcriptional control and biological impact. *Nat. Rev. Mol. Cell Biol.* 2002, 3, 651–662.
9. Ogata, H.; Chinen, T.; Yoshida, T.; Kinjyo, I.; Takaesu, G.; Shiraishi, H.; Iida, M.; Kobayashi, T.; Yoshimura, A. Loss of SOCS3 in the liver promotes fibrosis by enhancing STAT3-mediated TGF-β1 production. *Oncogene* 2006, 25, 2520–2530.
10. Hirano, T.; Ishihara, K.; Hibi, M. Roles of STAT3 in mediating the cell growth, differentiation and survival signals relayed through the IL-6 family of cytokine receptors. *Oncogene* 2000, 19, 2548–2556.
11. Hiraganahalli, B.D.; Chinampudur, V.C.; Dethe, S.; Mundkinajeddu, D.; Pandre, M.K.; Balachandran, J.; Agarwal, A. Hepatoprotective and antioxidant activity of standardized herbal extracts. *Pharmacogn. Mag.* 2012, 8, 116–123.
12. De Leo, P.; Dalessandro, G.; De Santis, A.; Arrigoni, O. Inhibitory effect of lycorine on cell division and cell elongation. *Plant Cell Physiol.* 1973, 14, 481–486.
13. Liu, X.S.; Jiang, J.; Jiao, X.Y.; Wu, Y.E.; Lin, J.H.; Cai, Y.M. Lycorine induces apoptosis and down-regulation of Mcl-1 in human leukemia cells. *Cancer Lett.* 2009, 274, 16–24.
14. McNulty, J.; Nair, J.J.; Bastida, J.; Pandey, S.; Griffin, C. Structure-activity studies on the lycorine pharmacophore: A potent inducer of apoptosis in human leukemia cells. *Phytochemistry* 2009, 70, 913–919.
15. Oleyede, K.G.; Oke, M.J.; Raji, Y.; Olugbade, T. Antioxidant and Anticonvulsant Alkaloids in *Crinum ornatum* Bulb Extract. *World J. Chem.* 2010, 5, 26–31.
16. Çitoğlu, G.S.; Acikara, O.B.; Yilmaz, B.S.; Özbek, H. Evaluation of analgesic, anti-inflammatory and hepatoprotective effects of lycorine from *Sternbergia fisheriana* (Herbert) Rupr. *Fitoterapia* 2012, 83, 81–87.

17. Wu, S.; Qiu, Y.; Shao, Y.; Yin, S.; Wang, R.; Pang, X.; Ma, J.; Zhang, C.; Wu, B.; Koo, S.; et al. Lycorine Displays Potent Antitumor Efficacy in Colon Carcinoma by Targeting STAT3. *Front. Pharmacol.* 2018, 9, 881.
18. Liang, Q.; Cai, W.; Zhao, Y.; Xu, H.; Tang, H.; Chen, D.; Qian, F.; Sun, L. Lycorine ameliorates bleomycin-induced pulmonary fibrosis via inhibiting NLRP3 inflammasome activation and pyroptosis. *Pharmacol. Res.* 2020, 158, 104884.
19. Schimmel, K.; Jung, M.; Foinquinos, A.; José, G.S.; Beaumont, J.; Bock, K.; Grote-Levi, L.; Xiao, K.; Bär, C.; Pfanne, A.; et al. Natural compound library screening identifies new molecules for the treatment of cardiac fibrosis and diastolic dysfunction. *Circulation* 2020, 141, 751–767.
20. Ni, T.; Huang, X.; Pan, S.; Lu, Z. Dihydrolycorine Attenuates Cardiac Fibrosis and Dysfunction by Downregulating Runx1 following Myocardial Infarction. *Oxid. Med. Cell. Longev.* 2021, 2021, 8528239.
21. Wu, J.; Fu, Y.; Wu, Y.X.; Wu, Z.X.; Wang, Z.H.; Li, P. Lycorine ameliorates isoproterenol-induced cardiac dysfunction mainly via inhibiting inflammation, fibrosis, oxidative stress and apoptosis. *Bioengineered* 2021, 12, 5583–5594.
22. Lam, P.; Cheung, F.; Tan, H.Y.; Wang, N.; Yuen, M.F.; Feng, Y. Hepatoprotective effects of chinese medicinal herbs: A focus on anti-inflammatory and anti-oxidative activities. *Int. J. Mol. Sci.* 2016, 17, 465.
23. Ilavenil, S.; Kaleeswaran, B.; Sumitha, P.; Tamilvendan, D.; Ravikumar, S. Protection of human erythrocyte using *Crinum asiaticum* extract and lycorine from oxidative damage induced by 2-amidinopropane. *Saudi J. Biol. Sci.* 2011, 18, 181–187.
24. Kang, J.; Zhang, Y.; Cao, X.; Fan, J.; Li, G.; Wang, Q.; Diao, Y.; Zhao, Z.; Luo, L.; Yin, Z. Lycorine inhibits lipopolysaccharide-induced iNOS and COX-2 up-regulation in RAW264.7 cells through suppressing P38 and STATs activation and increases the survival rate of mice after LPS challenge. *Int. Immunopharmacol.* 2012, 12, 249–256.
25. Crespo Yanguas, S.; Cogliati, B.; Willebrords, J.; Maes, M.; Colle, I.; van den Bossche, B.; de Oliveira, C.P.M.S.; Andraus, W.; Alves, V.A.; Leclercq, I.; et al. Experimental models of liver fibrosis. *Arch. Toxicol.* 2016, 90, 1025–1048.
26. Chilakapati, J.; Korrapati, M.C.; Shankar, K.; Hill, R.A.; Warbritton, A.; Latendresse, J.R.; Mehendale, H.M. Role of CYP2E1 and saturation kinetics in the bioactivation of thioacetamide: Effects of diet restriction and phenobarbital. *Toxicol. Appl. Pharmacol.* 2007, 219, 72–84.
27. Kang, J.S.; Wanibuchi, H.; Morimura, K.; Wongpoomchai, R.; Chusiri, Y.; Gonzalez, F.J.; Fukushima, S. Role of CYP2E1 in thioacetamide-induced mouse hepatotoxicity. *Toxicol. Appl. Pharmacol.* 2008, 228, 295–300.
28. Hajovsky, H.; Hu, G.; Koen, Y.; Sarma, D.; Cui, W.; Moore, D.S.; Staudinger, J.L.; Hanzlik, R.P. Metabolism and toxicity of thioacetamide and thioacetamide S-Oxide in rat hepatocytes. *Chem. Res. Toxicol.* 2012, 25, 1955–1963.
29. Mederacke, I.; Hsu, C.C.; Troeger, J.S.; Huebener, P.; Mu, X.; Dapito, D.H.; Pradere, J.P.; Schwabe, R.F. Fate tracing reveals hepatic stellate cells as dominant contributors to liver fibrosis independent of its aetiology. *Nat. Commun.* 2013, 4, 2823.
30. Iwaisako, K.; Jiang, C.; Zhang, M.; Cong, M.; Moore-Morris, T.J.; Park, T.J.; Liu, X.; Xu, J.; Wang, P.; Paik, Y.H.; et al. Origin of myofibroblasts in the fibrotic liver in mice. *Proc. Natl. Acad. Sci. USA* 2014, 111, E3297–E3305.
31. Friedman, S.L. Mechanisms of Hepatic Fibrogenesis. *Gastroenterology* 2008, 134, 1655–1669.
32. Ricard-Blum, S.; Baffet, G.; Théret, N. Molecular and tissue alterations of collagens in fibrosis. *Matrix Biol.* 2018, 68–69, 122–149.
33. Voisin, A.; Damon-Soubeyrand, C.; Bravard, S.; Saez, F.; Drevet, J.R.; Guiton, R. Differential expression and localisation of TGF- $\beta$  isoforms and receptors in the murine epididymis. *Sci. Rep.* 2020, 10, 995.
34. Prud'homme, G.J. Pathobiology of transforming growth factor  $\beta$  in cancer, fibrosis and immunologic disease, and therapeutic considerations. *Lab. Investig.* 2007, 87, 1077–1091.
35. Chung, J.Y.F.; Chan, M.K.K.; Li, J.S.F.; Chan, A.S.W.; Tang, P.C.T.; Leung, K.T.; To, K.F.; Lan, H.Y.; Tang, P.M.K. TGF- $\beta$  Signaling: From Tissue Fibrosis to Tumor Microenvironment. *Int. J. Mol. Sci.* 2021, 22, 7575.
36. Xu, M.Y.; Hu, J.J.; Shen, J.; Wang, M.L.; Zhang, Q.Q.; Qu, Y.; Lu, L.G. Stat3 signaling activation crosslinking of TGF- $\beta$ 1 in hepatic stellate cell exacerbates liver injury and fibrosis. *Biochim. Biophys. Acta-Mol. Basis Dis.* 2014, 1842, 2237–2245.
37. Ilavenil, S.; Karthik, D.; Arasu, M.V.; Vijayakumar, M.; Srigopalram, S.; Arokiyaraj, S.; Ravikumar, S.; Choi, K.C.; Ravikumar, S. Hepatoprotective mechanism of lycorine against carbon tetrachloride induced toxicity in swiss albino mice—A proteomic approach Keywords: Lycorine CCI 4 Oxidative stress 2D gel MALDI-TOF-MS ATP synthase Regucalcin HSP 60. *Asian Pac. J. Reprod.* 2015, 4, 123–128.
38. Robert, S.; Gicquel, T.; Bodin, A.; Lagente, V.; Boichot, E. Characterization of the MMP/TIMP imbalance and collagen production induced by IL-1  $\beta$  or TNF- $\alpha$  release from human hepatic stellate cells. *PLoS ONE* 2016, 11, e0153118.

39. Mikami, M.; Kitahara, M.; Kitano, M.; Ariki, Y.; Mimaki, Y.; Sashida, Y.; Yamazaki, M.; Yui, S. Suppressive activity of lycoricidinol (narciclasine) against cytotoxicity of neutrophil-derived calprotectin, and its suppressive effect on rat adjuvant arthritis model. *Biol. Pharm. Bull.* 1999, 22, 674–678.
40. Zhao, J.; Qi, Y.F.; Yu, Y.R. STAT3: A key regulator in liver fibrosis. *Ann. Hepatol.* 2021, 21, 100224.
41. Choi, S.; Jung, H.J.; Kim, M.W.; Kang, J.H.; Shin, D.; Jang, Y.S.; Yoon, Y.S.; Oh, S.H. A novel STAT3 inhibitor, STX-0119, attenuates liver fibrosis by inactivating hepatic stellate cells in mice. *Biochem. Biophys. Res. Commun.* 2019, 513, 49–55.
42. Lopez, O.N.; Bohanon, F.J.; Wang, X.; Ye, N.; Corsello, T.; Rojas-Khalil, Y.; Chen, H.; Chen, H.; Zhou, J.; Radhakrishnan, R.S. STAT3 Inhibition Suppresses Hepatic Stellate Cell Fibrogenesis: HJC0123, a Potential Therapeutic Agent for Liver Fibrosis. *RSC Adv.* 2016, 6, 100652.
43. Sallam, A.M.; Esmat, A.; Abdel-Naim, A.B. Cucurbitacin-B attenuates CCL4-induced hepatic fibrosis in mice through inhibition of STAT-3. *Chem. Biol. Drug Des.* 2018, 91, 933–941.
44. Fan, J.; Chen, Q.; Wei, L.; Zhou, X.; Wang, R.; Zhang, H. Asiatic acid ameliorates CCL4-induced liver fibrosis in rats: Involvement of Nrf2/ARE, NF- $\kappa$ B/ $\text{I}\kappa$ B $\alpha$ , and JAK1/STAT3 signaling pathways. *Drug Des. Dev. Ther.* 2018, 12, 3595.
45. Gong, Z.; Ye, H.; Huo, Y.; Wang, L.; Huang, Y.; Huang, M.; Yuan, X. S-allyl-cysteine attenuates carbon tetrachloride-induced liver fibrosis in rats by targeting STAT3/SMAD3 pathway. *Am. J. Transl. Res.* 2018, 10, 1337–1346.
46. Lin, I.Y.; Chiou, Y.S.; Wu, L.C.; Tsai, C.Y.; Chen, C.T.; Chuang, W.C.; Lee, M.C.; Lin, C.C.; Lin, T.T.; Chen, S.C.; et al. CCM111 prevents hepatic fibrosis via cooperative inhibition of TGF- $\beta$ , Wnt and STAT3 signaling pathways. *J. Food Drug Anal.* 2019, 27, 184–194.
47. Liu, J.; Hu, W.X.; He, L.F.; Ye, M.; Li, Y. Effects of lycorine on HL-60 cells via arresting cell cycle and inducing apoptosis. *FEBS Lett.* 2004, 578, 245–250.
48. Li, L.; Dai, H.-J.; Ye, M.; Wang, S.-L.; Xiao, X.-J.; Zheng, J.; Chen, H.-Y.; Luo, Y.-H.; Liu, J. Lycorine induces cell-cycle arrest in the G0/G1 phase in K562 cells via HDAC inhibition. *Cancer Cell Int.* 2012, 12, 49.
49. He, J.; Hong, B.; Bian, M.; Jin, H.; Chen, J.; Shao, J.; Zhang, F.; Zheng, S. Docosahexaenoic acid inhibits hepatic stellate cell activation to attenuate liver fibrosis in a PPAR $\gamma$ -dependent manner. *Int. Immunopharmacol.* 2019, 75, 105816.
50. Koda, Y.; Teratani, T.; Chu, P.-S.; Hagihara, Y.; Mikami, Y.; Harada, Y.; Tsujikawa, H.; Miyamoto, K.; Suzuki, T.; Taniki, N.; et al. CD8 $^{+}$  tissue-resident memory T cells promote liver fibrosis resolution by inducing apoptosis of hepatic stellate cells. *Nat. Commun.* 2021, 12, 4474.