

Dermo-Cosmetic Benefits of Marine Macroalgae-Derived Phenolic Compounds

Subjects: **Biochemistry & Molecular Biology**

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Marine macroalgae have an interesting profile of bioactive compounds and have gained tremendous attention in cosmeceuticals with negligible toxicity effects (cytotoxicity, reproductive toxicity, genotoxicity, mutagenicity, carcinogenicity, etc.) on humans and exhibit strong benefits for the skin. Among the diversified compounds, phenolic compounds are the group of phytochemicals found in high amounts with great structural diversity. Phlorotannin is the most studied polyphenol compound in brown algae, but besides there are some other phenolic compounds observed and studied in macroalgae such as terpenoids, bromophenols, mycosporine amino acids (MAAs), and flavonoids. These compounds are already characterized and studied for their full range of cosmeceutical benefits such as skin whitening, moisturizing, photoprotection, antiaging, antiwrinkle, anti-melanogenic, and antioxidant activities as well as in the treatment of pruritus (caused by acne, eczema, dermatitis, hives, psoriasis), photoaging, and skin pigmentation disorders (hypopigmentation due to the absence of melanocytes and hyperpigmentation caused by skin irritation or metabolic disorders).

cosmetics

marine algae

polyphenols

1. Introduction

Cosmeceutical ingredients are active compounds that are used to improve the appearance of the human body and represent a new category of preparations placed between cosmetics and pharmaceuticals. Cosmeceutical formulations intend the improvements of skin health and beauty ^{[1][2][3]}. Globally, the cosmeceutical sector is growing each year due to increasing modern beauty trends. To meet consumer demand, industries are moving towards the excessive use of synthetic cosmetic ingredients in formulations listed as Hydroquinone (HQ), Phthalates, Para-aminobenzoic acid (PABA), Benzophenones, Butylated Hydroxyanisole (BHA), Butylated Hydroxytoluene (BHT), and Dibenzoylmethane (DBM). According to SCCS (Scientific Committee on Consumer Safety) opinion (SCCS/1564/15), the excessive use of synthetic ingredients in cosmeceutical formulations may lead to different types of toxicities such as acute toxicity, corrosion and irritation, skin sensation, dermal/percutaneous absorption, repeated dose toxicity, reproductive toxicity, mutagenicity/genotoxicity, carcinogenicity, and photoinduced toxicity on the skin as well as human health. Hydroxyanisole, widely used in skin-whitening creams, has reported many harmful effects such as ochronosis and potential mutagenicity ^{[4][5][6]}. Benzophenones, DBM, and PABA have shown allergic phototoxicities, dermatitis, and skin irritations ^{[7][8]}. Besides, BHA and BHT are applied in moisturization and lipstick preparations that cause allergic reactions, irritation, and corrosivity in the skin. Another ingredient, parabens, are highly carcinogenic and neurotoxic among other harmful

health effects. Around 75 to 90 percent of commercially available products contain parabens, which are mostly used as a mixture in cosmetic formulations. Parabens have been reported to have a high risk of breast cancer and the development of malignant melanoma in women [9]. However, in the ACDS Contact Allergen Management Program (CAMP) report, about 19% of products contained different types of parabens, mainly methylparaben, ethylparaben, propylparaben, and butylparaben. According to them, these components have little allergenicity compared to other preservatives, with no adverse reactions, and low toxicity, safety, and cost [10].

Various natural resources can be used in skin cosmetic products such as terrestrial plants, fungi, marine algae, bacteria, animals, etc. [11][12][13][14][15]. Among them, marine macroalgae are widely utilized for their skin benefits nowadays. Marine macroalgae are also known as seaweed: eukaryotic, aquatic photosynthetic macroscopic, multicellular organisms that are ubiquitously found along the seacoast and in seawater. They belong to the Eukaryota domain and are classified into three major taxonomic groups, red algae, brown algae, and green algae, belonging to the Rhodophyta phylum, Ochrophyta phylum, and Chlorophyta phylum, respectively [16][17][18][19]. These different types of marine macroalgae are illustrated in **Figure 1**. There is an increasing demand for bioactive constituents in cosmetic and cosmeceutical applications from macroalgae. The applications of macroalgae-derived compounds to the cosmetic industry are based on their potential biological activities [20][21][22]. These are lipids, fatty acids, polysaccharides, vitamins, minerals, amino acids, phenolic compounds, proteins, pigments, etc., which have attracted attention for their skin cosmeceutical benefits [23][24][25].



(a)



(b)



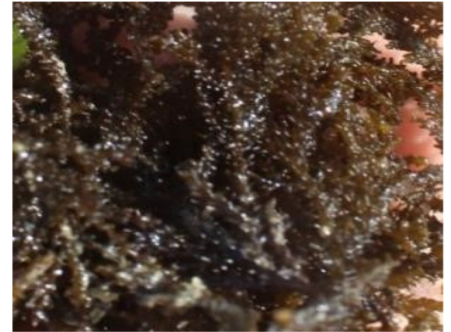
(c)



(d)



(e)



(f)



(g)



(h)



(i)



(j)



(k)



(l)



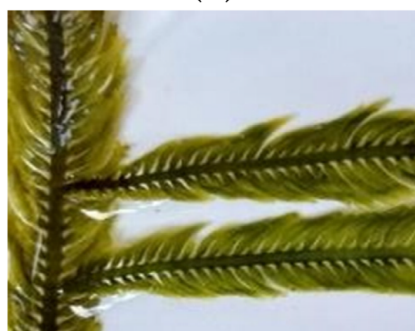
(m)



(n)



(o)



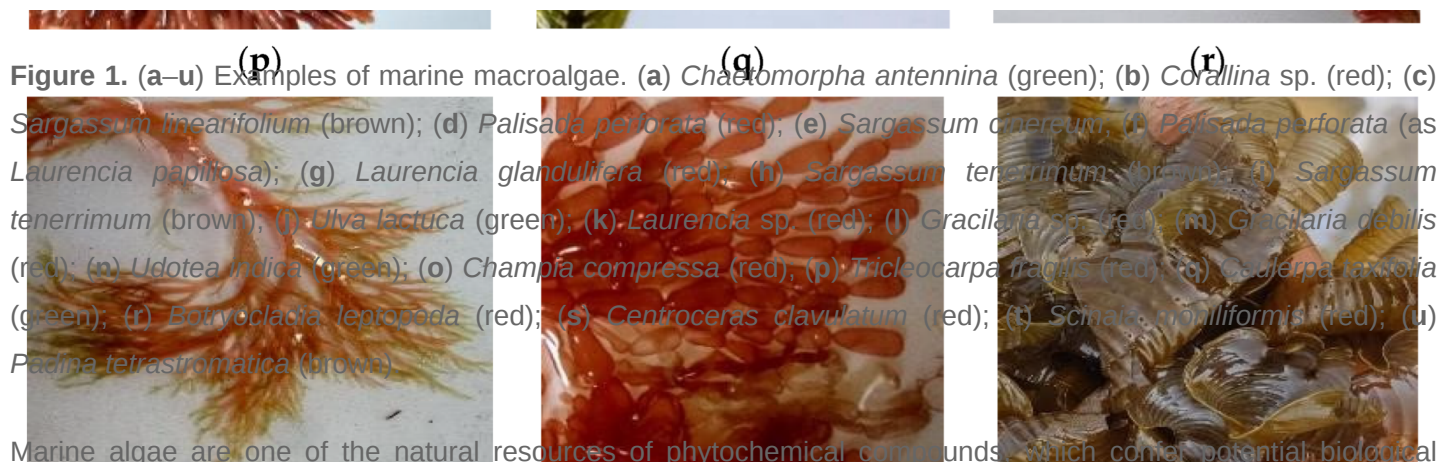


Figure 1. (a–u) Examples of marine macroalgae. (a) *Chaetomorpha antennina* (green); (b) *Corallina* sp. (red); (c) *Sargassum linearifolium* (brown); (d) *Palisada perforata* (red); (e) *Sargassum cinereum*; (f) *Palisada perforata* (as *Laurencia papillosa*); (g) *Laurencia glandulifera* (red); (h) *Sargassum tenerrimum* (brown); (i) *Sargassum tenerrimum* (brown); (j) *Ulva lactuca* (green); (k) *Laurencia* sp. (red); (l) *Gracilaria* sp. (red); (m) *Gracilaria debilis* (red); (n) *Udotea indica* (green); (o) *Champia compressa* (red); (p) *Tricleocarpa fragilis* (red); (q) *Codium taxifolia* (green); (r) *Botryocladia leptopoda* (red); (s) *Centroceras clavulatum* (red); (t) *Schmida moniliformis* (red); (u) *Padina tetrastrum* (brown).

Marine algae are one of the natural resources of phytochemical compounds, which confer potential biological activities [26][27]. Phenolic compounds are one of the bioactive compounds produced in seaweeds, are made of an

aromatic ring with one or more hydroxyl groups, and their structures diversify from simple to complex, higher molecular weight compounds [28][29]. Many previous studies have been carried out in which phenolic compounds were isolated from marine algae and they include simple phenolic compounds or polyphenols such as flavonoids, phlorotannins, mycosporine-like amino acids (MAAs), bromophenols, and terpenoids [30]. The biological action of phenolic compounds is determined by the position of the hydroxyl groups, and the number of phenyl rings in the structure [31]. Brown algal species contain a high amount of phlorotannins whereas green and red algae mainly produce flavonoids, bromophenols, terpenoids, and mycosporine amino acids in response to environmental conditions [30][31][32][33][34]. Marine algae-derived phenolic compounds have a wide variety of applications such as enzyme inhibitory effects (for example, tyrosinase inhibition, elastase inhibition, collagenase inhibition, matrix metalloproteinase inhibition in photoprotection, inhibition of angiotensin-converting enzyme-1 (ACE-1), pro-inflammatory cyclooxygenase and lipoxygenase (COX-1, 2 and 5-LOX) as well as dipeptidyl peptidase-4 (DPP-4) inhibition, and hydroxymethyl glutaryl coenzyme A reductase (hMGCR) inhibition) antibacterial, antifungal, antioxidant, and anti-inflammatory properties, which can be very attractive when utilized in cosmetics and cosmeceutical product preparations [35][36][37][38][39][40][41]. In cosmetics, phlorotannin provides hyaluronidase activation, antiallergic, anti-wrinkle, anti-aging, skin whitening, photoprotection, and skin health improvement benefits [42][43].

2. Characterization and Types of Phenolic Compounds

2.1. Polyphenolic Compounds

Polyphenol is mainly of two types, phlorotannin, and phloroglucinol. The former is a polymer of phloroglucinol with an additional halogen or hydroxyl group whereas the latter contains an aromatic ring structure with three hydroxyl groups [44][45][46]. These can be subclassified into six different groups: (i) Eckols; (ii) Fucophlorethols; (iii) Fucols; (iv) Phlorethols; (v) Carmalols; and (vi) Fuhalols.

2.2. Lignans

Lignans are a type of phenolic compound, a dimer or oligomer, formed due to the union of monolignols, coniferyl alcohol, and sinapyl alcohol. Freile-Pelegrín and Robledo [47] reported the presence of lignans in calcified red marine algae *Calliarthron cheilosporioides* (Rhodophyta). Another polymeric phenol, lignin, is the most abundant organic polymer found in nature but not extensively studied in marine algae, which are structurally composed of monolignols (coniferyl alcohol, sinapyl alcohol), and lignan units randomly linked forming a polymeric network. Tannins are usually divided into three different chemical structures: hydrolyzable tannins, flavonoid-based tannins, and phlorotannins. The first one is derived from simple phenolic acids and their carbohydrate hydroxyl groups that are partially or completely esterified with phenolic groups. The second, flavonoid-based tannins, synthesize through flavins and catechins whereas the last, phlorotannins, are oligomers of phloroglucinol that are exclusively found in brown algae [48].

2.3. Phlorotannins

Phlorotannins, a group of compounds that majorly include dioxinodehydroeckol (eckstolonol), dieckol, eckol, phlorofucofuroeckol A, 7-phloroeckol, and fucofuroeckol A and 8,8'-bieckol, exhibit antioxidant-inhibitory effects against melanin synthesis, skin whitening (tyrosinase inhibition), and UV protection [49][50][51][52][53][54]. Phlorotannins are the most deeply studied phenolic compounds from algae [55]. Their antioxidant power is 2 to 10 times higher when compared to ascorbic acid or tocopherol [56][57], which demonstrates their role as an anti-inflammatory agent [58]. They can act as an anti-UCB protector; Ryu et al. [59] suggested UVB protection by dioxinodehydroeckol from *E. cava* on the HaCat cells that reduce the provoked apoptosis. Moreover, phlorotannins such as dieckol, dioxinodehydroeckol, eckol, eckstolonol, phlorofucofuroeckol A, and 7-phloroeckol isolated from different marine algae are being researched in cosmetics as whiteners and antiwrinkle agents. They have been shown as promising tyrosinase inhibitors and hyaluronidase inhibitors [60][61][62][63][64][65][66].

2.4. Bromophenols

Phenolic compounds such as bromophenol and benzoic acids have been fully isolated and characterized from red seaweeds [67]. Pérez et al. [68], Duan et al. [69], and Choi et al. [70] studied the antioxidant activity of *Vertebrata constricta* (formerly *Polysiphonia stricta* or *P. urceolata*) (Rhodophyta)-derived phenolic compounds, but that depends on the brominated units and degree of bromination. In the same study, *Symphycladia latiuscula*-derived bromophenols reported antioxidant activity that was studied by DPPH assay.

2.5. Flavonoids

Other classes of phenolic compounds have been investigated for varieties of applications in cosmetics. Tanna et al. [71] found the antioxidant activity of various flavonoids such as kaempferol and quercetin from *Caleurpa* spp. (Chlorophyta). *Acanthophora spicifera* (Rhodophyta)-derived flavonoid demonstrates a mixture of chlorogenic acid (69.64%), caffeic acid (12.86%), vitexin-rhamnose (12.35%), quercetin (1.41%), and catechol (0.59%), and this flavonoid-enriched extract has revealed antioxidant activity [72][73]. These molecules are multi-active components that play a role in UV radiation absorption, the neutralization of ROSs, and the inhibition of radical reactions, etc., which makes them important contributors to cosmeceuticals [74].

2.6. Phenolic Terpenoids

Makkar and Chakraborty [75] studied a chromene-based phenolic compound from *Gracilaria opuntia* (Rhodophyta) that has been reported to have antioxidant activity in in vitro assays. Pillai et al. [76] reported the role of antioxidants in the prevention of extracellular matrix damage, the activation of MMPs, and inhibition of their expression. These molecules scavenge and quench radical oxygen species (ROS).

2.7. Mycosporine-like Amino Acid

Various marine algal species such as *Asparagopsis armata*, *Chondrus crispus*, *Mastocarpus stellatus*, *Palmaria palmata*, *Gelidium* sp., *Pyropia* sp. (formerly known as *Porphyra* sp.), *Gracilaria cornea*, *Solieria chordalis*, *Grateloupia lanceola*, and *Curdiea racovitzae* (Rhodophyta) have been investigated for this exclusive class of phenolic compounds. This class of compounds is more commonly found free in the intracellular space and around cell organelles sensitive to ultraviolet rays. Mycosporine-like amino acids (MAAs) are formed by cyclohexenone or cycloheximide chromophore conjugated to imino alcohol or an amino acid residue [77][78]. Various MAAs (palythine, shinorine, asterina-330, Porphyra-334, palythanol, and usujirene) have already been studied that have high antioxidant, photoprotection, and anti-proliferative (HeLa cancer cell line, human cervical adenocarcinoma cell line) and HaCat (human immortalized keratinocyte) activity [79][80].

3. Extraction of Phenolic Compounds

There are several extraction techniques available for obtaining phenolic compounds; two general techniques are found: conventional and nonconventional extraction techniques. The conventional techniques include simple solid solvent extraction, whereas nontraditional techniques include microwave-assisted extraction, subcritical CO₂ extraction, ultrasound-assisted extraction, and pressurized liquid extraction, among others.

The most important step is to select an appropriate extraction method, since many procedures of extraction are available nowadays. Traditional methods include heat-assisted extraction or maceration, percolation, and Soxhlet extraction as reported by Aires [81]. One of the classical methods is maceration, in which the components are extracted by submerging marine algae in an appropriate solvent or solvent combinations [82]. On a large scale, at the industrial level, ethanol is preferred as a solvent for extraction because of its economic benefit [83]. This procedure is widely applicable in current practice. In this method, methanol, ethanol, acetone, water, and ethyl ethanoate in different proportions are commonly utilized for extraction. The selection can be done based on polarity. Due to the hydrophilic nature of these compounds, hydroalcoholic solvent is the most effective for this process. Some previous studies have mentioned the combination of solvents, with acids such as citric acid, tartaric acid, or HCl potentially improving the extraction of phenolic compounds [84][85]. In traditional procedures, Soxhlet extraction provides better results of extraction in terms of yield, although this technique also presents some demerits such as the degradation of temperature-sensitive compounds as some phenolic acids, tannins, and anthocyanins require a large number of solvents and are time-consuming. Besides, this classical Soxhlet extraction

method is a continuous process; the solvent can easily be recycled, and less time and less solvent are used than in maceration and percolation [86].

A non-conventional technique, Pressurized Liquid Extraction (PLE), also known as extraction with pressurized solvent, includes high pressure (1 to 15 MPa), short processing time, and temperature ranges of about 50 to 200 °C using a low volume of nontoxic solvent and thus being considered a green technology. Otero et al. [87] observed a highest extraction yield of 37% for 80 °C and 52% for 160 °C using diluted ethanol from the brown alga *Laminaria ochroleuca* (Phaeophyceae) at 100 bars. Microwave-assisted extraction is mainly used for the extraction of polyphenols and polysaccharides. This method can be performed in open (at atmospheric pressure) or closed (higher than atmospheric pressure) vessels. In this method, electromagnetic waves cause changes in cell structures. Two mechanisms, ionic conduction, and dipole rotation, transform electromagnetic energy into calorific energy [88][89].

References

1. Martin, K.I.; Glaser, D.A. Cosmeceuticals: The new medicine of beauty. *Mo. Med.* 2011, 108, 60.
2. Dureja, H.; Kaushik, D.; Gupta, M.; Kumar, V.; Lather, V. Cosmeceuticals: An emerging concept. *Indian J. Pharmacol.* 2005, 37, 155.
3. Draelos, Z.D. The cosmeceutical realm. *Clin. Dermatol.* 2008, 26, 627–632.
4. Yin, S.N.; Hayes, R.B.; Linet, M.S.; Li, G.L.; Dosemeci, M.; Travis, L.B.; Zhang, Z.N.; Li, D.G.; Chow, W.H.; Wacholder, S.; et al. An expanded cohort study of cancer among benzene-exposed workers in China. Benzene Study Group. *Environ. Health Perspect.* 1996, 104 (Suppl. S6), 1339–1341.
5. Briganti, S.; Camera, E.; Picardo, M. Chemical and instrumental approaches to treat hyperpigmentation. *Pigment. Cell Res.* 2003, 16, 101–110.
6. Zhang, L.; Robertson, M.L.; Kolachana, P.; Davison, A.J.; Smith, M.T. Benzene metabolite, 1,2,4-benzenetriol, induces micronuclei and oxidative DNA damage in human lymphocytes and HL60 cells. *Environ. Mol. Mutagen.* 1993, 21, 339–348.
7. Fernández-Álvarez, M.; Llompart, M.; Sánchez-Prado, L.; García-Jares, C.; Lores, M. Photochemical behavior of UV filter combinations. In *Cosmetics: Types, Allergies and Applications*; Nova Science Publishers, Inc.: Hauppauge, NY, USA, 2010; p. 1.
8. Knowland, J.; McKenzie, E.A.; McHugh, P.J.; Cridland, N.A. Sunlight-induced mutagenicity of a common sunscreen ingredient. *FEBS Lett.* 1993, 324, 309–313.
9. Kerdudo, A.; Burger, P.; Merck, F.; Dingas, A.; Rolland, Y.; Michel, T.; Fernandez, X. Development of a natural ingredient–Natural preservative: A case study. *Comptes Rendus. Chim.* 2016, 19,

1077–1089.

10. Mowad, C.M. Allergic contact dermatitis caused by parabens: 2 case reports and a review. *Am. J. Contact Dermat.* 2000, 11, 53–56.
11. Kim, A.R.; Shin, T.S.; Lee, M.S.; Park, J.Y.; Park, K.E.; Yoon, N.Y.; Kim, J.S.; Choi, J.S.; Jang, B.C.; Byun, D.S.; et al. Isolation and identification of phlorotannins from *Ecklonia stolonifera* with antioxidant and anti-inflammatory properties. *J. Agric. Food Chem.* 2009, 57, 3483–3489.
12. Pereira, L. *Therapeutic and Nutritional Uses of Algae*; CRC Press: Boca Raton, FL, USA, 2018; ISBN 9781498755382.
13. Maqsood, S.; Benjakul, S.; Shahidi, F. Emerging role of phenolic compounds as natural food additives in fish and fish products. *Crit. Rev. Food Sci. Nutr.* 2013, 53, 162–179.
14. Panzella, L.; Napolitano, A. Natural phenol polymers: Recent advances in food and health applications. *Antioxidants* 2017, 6, 30.
15. Leandro, A.; Pereira, L.; Gonçalves, A.M. Diverse applications of marine macroalgae. *Mar. Drugs* 2019, 18, 17.
16. Pereira, L. *Algae. Litoral of Viana do Castelo*; Câmara Municipal de Viana do Castelo: Viana do Castelo, Portugal, 2010; pp. 7–8. ISBN 978-972-588-217-7.
17. Pereira, L. *Guia Ilustrado das Macroalgas—Conhecer e Reconhecer Algumas Espécies da Flora Portuguesa*; Universityde Coimbra Press: Coimbra, Portugal, 2009; p. 91. ISBN 978-989-26-0002-4.
18. Pereira, L. Chapter 4—Cytological and cytochemical aspects in selected carrageenophytes (Gigartinales, Rhodophyta). In *Advances in Algal Cell Biology*; Heimann, K., Katsaros, C., Eds.; De Gruyter: Berlin, Germany, 2012; pp. 81–104. ISBN 978-3-11-022960-8.
19. González-Minero, F.J.; Bravo-Díaz, L. The use of plants in skin-care products, cosmetics and fragrances: Past and present. *Cosmetics* 2018, 5, 50.
20. Ibañez, E.; Herrero, M.; Mendiola, J.A.; Castro-Puyana, M. Extraction and characterization of bioactive compounds with health benefits from marine resources: Macro and micro algae, cyanobacteria, and invertebrates. In *Marine Bioactive Compounds*; Springer: Boston, MA, USA, 2012; pp. 55–98.
21. Vo, T.S.; Kim, S.K. Fucoidans as a natural bioactive ingredient for functional foods. *J. Funct. Foods* 2013, 5, 16–27.
22. Venkatesan, J.; Kim, S.K. Osteoporosis treatment: Marine algal compounds. *Adv. Food Nutr. Res.* 2011, 64, 417–427.

23. Berthon, J.Y.; Nachat-Kappes, R.; Bey, M.; Cadoret, J.P.; Renimel, I.; Filaire, E. Marine algae as attractive source to skin care. *Free. Radic. Res.* 2017, 51, 555–567.
24. Gam, D.H.; Park, J.H.; Hong, J.W.; Jeon, S.J.; Kim, J.H.; Kim, J.W. Effects of *Sargassum thunbergii* extract on skin whitening and anti-wrinkling through inhibition of TRP-1 and MMPs. *Molecules* 2021, 26, 7381.
25. Querellou, J.; Børresen, T.; Boyen, C.; Dobson, A.; Höfle, M.; Ianora, A.; Jaspars, M.; Kijjoo, A.; Olafsen, J.; Rigos, G. Marine biotechnology: Realising the full potential of Europe. *VLIZ Spec. Publ.* 2010, 47, 21.
26. Acosta-Estrada, B.A.; Gutiérrez-Urbe, J.A.; Serna-Saldívar, S.O. Bound phenolics in foods, a review. *Food Chem.* 2014, 152, 46–55.
27. Komes, D.; Belščak-Cvitanović, A.; Horžić, D.; Rusak, G.; Likić, S.; Berendika, M. Phenolic composition and antioxidant properties of some traditionally used medicinal plants affected by the extraction time and hydrolysis. *Phytochem. Anal.* 2011, 22, 172–180.
28. Jesumani, V.; Du, H.; Aslam, M.; Pei, P.; Huang, N. Potential use of seaweed bioactive compounds in skincare—A review. *Mar. Drugs* 2019, 17, 688.
29. Chisté, R.C.; Godoy, H.T.; Prado, M.A. The phenolic compounds and the antioxidant potential of infusion of herbs from the Brazilian Amazonian region. *Food Res. Int.* 2013, 53, 875–881.
30. Nurilmala, M.; Hidayat, T.; Sudirdjo, F. Characteristics of seaweed as raw materials for cosmetics. *Aquat. Procedia* 2016, 7, 177–180.
31. Pereira, L. Seaweeds as source of bioactive substances and skin care therapy—Cosmeceuticals, algothoraphy, and thalassotherapy. *Cosmetics* 2018, 5, 68.
32. Gómez-Guzmán, M.; Rodríguez-Nogales, A.; Algieri, F.; Gálvez, J. Potential role of seaweed polyphenols in cardiovascular-associated disorders. *Mar. Drugs* 2018, 16, 250.
33. Moraes, T.; Inácio, A.; Coutinho, T.; Ministro, M.; Cotas, J.; Pereira, L.; Bahcevandziev, K. Seaweed potential in the animal feed: A review. *J. Mar. Sci. Eng.* 2020, 8, 559.
34. Costa, P.; Gonçalves, S.; Valentão, P.; Andrade, P.B.; Almeida, C.; Nogueira, J.M.; Romano, A. Metabolic profile and biological activities of *Lavandula pedunculata* subsp. *lusitanica* (Chaytor) Franco: Studies on the Essential Oil and Polar Extracts. *Food Chem.* 2013, 141, 2501–2506.
35. Wijesinghe, W.A.J.P.; Jeon, Y.J. Biological activities and potential cosmeceutical applications of bioactive components from brown seaweeds: A review. *Phytochem. Rev.* 2011, 10, 431–443.
36. Kizhakkekalam, V.K.; Chakraborty, K. Pharmacological properties of marine macroalgae-associated heterotrophic bacteria. *Arch. Microbiol.* 2019, 201, 505–518.

37. Holmquist, B.; Bunning, P.; Riordan, J.F. A continuous spectrophotometric assay for angiotensin converting enzyme. *Anal. Biochem.* 1979, 95, 540–548.
38. Brand-Williams, W.; Cuvelier, M.; Berset, C. Use of free radical method to evaluate antioxidant activity. *LWT-Food Sci. Technol.* 1995, 28, 25–30.
39. Re, R.; Pellegrini, N.; Proteggente, A.; Pannala, A.; Yang, M.; Rice Evans, C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radic. Biol. Med.* 1999, 26, 1231–1237.
40. Ademiluyi, A.O.; Oboh, G. Soybean phenolic-rich extracts inhibit key-enzymes linked to type-2 diabetes (α -amylase and α -glucosidase) and hypertension (angiotensin-I converting enzyme) in-vitro. *Exp. Toxicol. Pathol.* 2013, 65, 305–309.
41. Charlier, C.; Michaux, C. Dual inhibition of cyclooxygenase-2 (COX-2) and 5-lipoxygenase (5-LOX) as a new strategy to provide safer non-steroidal anti-inflammatory drugs. *Eur. J. Med. Chem.* 2003, 38, 645–659.
42. Kalasariya, H.S.; Yadav, V.K.; Yadav, K.K.; Tirth, V.; Algahtani, A.; Islam, S.; Gupta, N.; Jeon, B.-H. Seaweed-Based Molecules and Their Potential Biological Activities: An Eco-Sustainable Cosmetics. *Molecules* 2021, 26, 5313.
43. Nasab, S.B.; Homaei, A.; Pletschke, B.I.; Salinas-Salazar, C.; Castillo-Zacarias, C.; Parra-Saldívar, R. Marine resources effective in controlling and treating diabetes and its associated complications. *Process Biochem.* 2020, 92, 313–342.
44. Gupta, S.; Abu-Ghannam, N. Recent developments in the application of seaweeds or seaweed extracts as a means for enhancing the safety and quality attributes of foods. *Innov. Food Sci. Emerg. Technol.* 2011, 12, 600–609.
45. Handique, J.G.; Baruah, J.B. Polyphenolic compounds: An overview. *React. Funct. Polym.* 2002, 52, 163–188.
46. Mouritsen, O.G. The science of seaweeds: Marine macroalgae benefit people culturally, industrially, nutritionally, and ecologically. *Am. Sci.* 2013, 101, 458–466.
47. Freile-Pelegrín, Y.; Robledo, D. Bioactive phenolic compounds from algae. In *Bioactive Compounds from Marine Foods: Plant and Animal Sources*; John Wiley & Sons Ltd.: Chichester, UK, 2013; pp. 113–129.
48. Kim, M.M.; Kim, S.K. Effect of phloroglucinol on oxidative stress and inflammation. *Food Chem. Toxicol.* 2010, 48, 2925–2933.
49. Charoensiddhi, S.; Franco, C.; Su, P.; Zhang, W. Improved antioxidant activities of brown seaweed *Ecklonia radiata* extracts prepared by microwave-assisted enzymatic extraction. *J. Appl. Phycol.* 2015, 27, 2049–2058.

50. Chang, M.Y.; Byon, S.H.; Shin, H.C.; Han, S.E.; Kim, J.Y.; Byun, J.Y.; Lee, J.D.; Park, M.K. Protective effects of the seaweed phlorotannin polyphenolic compound dieckol on gentamicin-induced damage in auditory hair cells. *Int. J. Pediatr. Otorhinolaryngol.* 2016, 83, 31–36.
51. Piao, M.J.; Hewage, S.R.; Han, X.; Kang, K.A.; Kang, H.K.; Lee, N.H.; Hyun, J.W. Protective Effect of Diphlorethohydroxycarmalol against Ultraviolet B Radiation-Induced DNA Damage by Inducing the Nucleotide Excision Repair System in HaCaT Human Keratinocytes. *Mar. Drugs* 2015, 13, 5629–5641.
52. Kang, S.M.; Heo, S.J.; Kim, K.N.; Lee, S.H.; Yang, H.M.; Kim, A.D.; Jeon, Y.J. Molecular docking studies of a phlorotannin, dieckol isolated from *Ecklonia cava* with tyrosinase inhibitory activity. *Bioorg. Med. Chem.* 2012, 20, 311–316.
53. Kirke, D.A.; Smyth, T.J.; Rai, D.K.; Kenny, O.; Stengel, D.B. The chemical and antioxidant stability of isolated low molecular weight phlorotannins. *Food Chem.* 2017, 221, 1104–1112.
54. de Lima Cherubim, D.J.; Buzanello Martins, C.V.; Oliveira Fariña, L.; da Silva de Lucca, R.A. Polyphenols as natural antioxidants in cosmetics applications. *J. Cosmet. Dermatol.* 2020, 19, 33–37.
55. Gager, L.; Lalegerie, F.; Connan, S.; Stiger-Pouvreau, V. Marine Algal Derived Phenolic Compounds and their Biological Activities for Medicinal and Cosmetic Applications. In *Recent Advances in Micro and Macroalgal Processing: Food and Health Perspectives*; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2021; pp. 278–334.
56. Shibata, T.; Ishimaru, K.; Kawaguchi, S.; Yoshikawa, H.; Hama, Y. Antioxidant activities of phlorotannins isolated from Japanese Laminariaceae. In *Nineteenth International Seaweed Symposium*; Springer: Dordrecht, The Netherlands, 2007; pp. 255–261.
57. Besednova, N.N.; Zvyagintseva, T.N.; Kuznetsova, T.A.; Makarenkova, I.D.; Smolina, T.P.; Fedyanina, L.N.; Kryzhanovsky, S.P.; Zaporozhets, T.S. Marine algae metabolites as promising therapeutics for the prevention and treatment of HIV/AIDS. *Metabolites* 2019, 9, 87.
58. Kim, A.R.; Lee, M.S.; Choi, J.W.; Utsuki, T.; Kim, J.I.; Jang, B.C.; Kim, H.R. Phlorofuocuroeckol A suppresses expression of inducible nitric oxide synthase, cyclooxygenase-2, and pro-inflammatory cytokines via inhibition of nuclear factor- κ B, c-Jun NH₂-terminal kinases, and Akt in microglial cells. *Inflammation* 2013, 36, 259–271.
59. Ryu, B.; Ahn, B.N.; Kang, K.H.; Kim, Y.S.; Li, Y.X.; Kong, C.S.; Kim, S.K.; Kim, D.G. Dioxinodehydroeckol protects human keratinocyte cells from UVB-induced apoptosis modulated by related genes Bax/Bcl-2 and caspase pathway. *J. Photochem. Photobiol. B Biol.* 2015, 153, 352–357.
60. Kumar, L.R.; Paul, P.T.; Anas, K.K.; Tejpal, C.S.; Chatterjee, N.S.; Anupama, T.K.; Mathew, S.; Ravishankar, C.N. Phlorotannins—bioactivity and extraction perspectives. *J. Appl. Phycol.* 2022,

- 34, 2173–2185.
61. Shibata, T.; Fujimoto, K.; Nagayama, K.; Yamaguchi, K.; Nakamura, T. Inhibitory activity of brown algal phlorotannins against hyaluronidase. *Int. J. Food Sci. Technol.* 2002, 37, 703–709.
 62. Kang, H.S.; Kim, H.R.; Byun, D.S.; Son, B.W.; Nam, T.J.; Choi, J.S. Tyrosinase inhibitors isolated from the edible brown alga *Ecklonia stolonifera*. *Arch. Pharmacol Res.* 2004, 27, 1226–1232.
 63. Yoon, N.Y.; Eom, T.K.; Kim, M.M.; Kim, S.K. Inhibitory effect of phlorotannins isolated from *Ecklonia cava* on mushroom tyrosinase activity and melanin formation in mouse B16F10 melanoma cells. *J. Agric. Food Chem.* 2009, 57, 4124–4129.
 64. Lee, S.H.; Kang, S.M.; Sok, C.H.; Hong, J.T.; Oh, J.Y.; Jeon, Y.J. Cellular activities and docking studies of eckol isolated from *Ecklonia cava* (Laminariales, Phaeophyceae) as potential tyrosinase inhibitor. *Algae* 2015, 30, 163–170.
 65. Heo, S.J.; Ko, S.C.; Cha, S.H.; Kang, D.H.; Park, H.S.; Choi, Y.U.; Kim, D.; Jung, W.K.; Jeon, Y.J. Effect of phlorotannins isolated from *Ecklonia cava* on melanogenesis and their protective effect against photo-oxidative stress induced by UV-B radiation. *Toxicol. In Vitro* 2009, 23, 1123–1130.
 66. Nurrochmad, A.; Wirasti, W.; Dirman, A.; Lukitaningsih, E.; Rahmawati, A.; Fakhrudin, N. Effects of Antioxidant, Anti-Collagenase, Anti-Elastase, Anti-Tyrosinase of The Extract and Fraction From *Turbinaria decurrens* Bory. *Indones. J. Pharm.* 2018, 29, 188.
 67. De Almeida, C.L.F.; Falcão, H.D.S.; Lima, G.R.D.M.; Montenegro, C.D.A.; Lira, N.S.; de Athayde-Filho, P.F.; Rodrigues, L.C.; De Souza, M.D.F.V.; Barbosa-Filho, J.M.; Batista, L.M. Bioactivities from marine algae of the genus *Gracilaria*. *Int. J. Mol. Sci.* 2011, 12, 4550–4573.
 68. Abu-Ghannam, N.; Rajauria, G. Antimicrobial activity of compounds isolated from algae. In *Functional Ingredients from Algae for Foods and Nutraceuticals*; Woodhead Publishing: Sawston, UK, 2013; pp. 287–306.
 69. Duan, X.J.; Li, X.M.; Wang, B.G. Highly brominated mono-and bis-phenols from the marine red alga *Symphyocladia latiuscula* with radical-scavenging activity. *J. Nat. Prod.* 2007, 70, 1210–1213.
 70. Choi, J.S.; Park, H.J.; Jung, H.A.; Chung, H.Y.; Jung, J.H.; Choi, W.C. A cyclohexanonyl bromophenol from the red alga *Symphyocladia latiuscula*. *J. Nat. Prod.* 2000, 63, 1705–1706.
 71. Tanna, B.; Choudhary, B.; Mishra, A. Metabolite profiling, antioxidant, scavenging and anti-proliferative activities of selected tropical green seaweeds reveal the nutraceutical potential of *Caulerpa* spp. *Algal Res.* 2018, 36, 96–105.
 72. Jeyaprakash, R.R.K. HPLC Analysis of flavonoids in *Acanthophora specifera* (red seaweed) collected from Gulf of Mannar, Tamilnadu, India. *Int. J. Sci. Res.* 2017, 6, 69–72.

73. Cotas, J.; Leandro, A.; Monteiro, P.; Pacheco, D.; Figueirinha, A.; Gonçalves, A.M.; da Silva, G.J.; Pereira, L. Seaweed phenolics: From extraction to applications. *Mar. Drugs* 2020, 18, 384.
74. Arct, J.; Pytkowska, K. Flavonoids as components of biologically active cosmeceuticals. *Clin. Dermatol.* 2008, 26, 347–357.
75. Makkar, F.; Chakraborty, K. Highly oxygenated antioxidative 2 H-chromen derivative from the red seaweed *Gracilaria opuntia* with pro-inflammatory cyclooxygenase and lipoxygenase inhibitory properties. *Nat. Prod. Res.* 2018, 32, 2756–2765.
76. Pillai, S.; Oresajo, C.; Hayward, J. Ultraviolet radiation and skin aging: Roles of reactive oxygen species, inflammation and protease activation, and strategies for prevention of inflammation-induced matrix degradation—A review. *Int. J. Cosmet. Sci.* 2005, 27, 17–34.
77. Carreto, J.I.; Carignan, M.O. Mycosporine-like amino acids: Relevant secondary metabolites. Chemical and ecological aspects. *Mar. Drugs* 2011, 9, 387–446.
78. Rosic, N.N.; Braun, C.; Kvaskoff, D. Extraction and Analysis of Mycosporine-Like Amino Acids in Marine Algae. In *Natural Products from Marine Algae: Methods and Protocols*; Stengel, D.B., Connan, S., Eds.; Springer: New York, NY, USA, 2015.
79. Guihéneuf, F.; Gietl, A.; Stengel, D.B. Temporal and spatial variability of mycosporine-like amino acids and pigments in three edible red seaweeds from western Ireland. *J. Appl. Phycol.* 2018, 30, 2573–2586.
80. Suh, S.S.; Oh, S.K.; Lee, S.G.; Kim, I.C.; Kim, S. Porphyra-334, a mycosporine-like amino acid, attenuates UV-induced apoptosis in HaCaT cells. *Acta Pharm.* 2017, 67, 257–264.
81. Padilla, M.; Palma, M.; Barroso, C.G. Determination of phenolics in cosmetic creams and similar emulsions. *J. Chromatogr. A* 2005, 1091, 83–88.
82. Kim, S.M.; Kang, S.W.; Jeon, J.S.; Jung, Y.J.; Kim, W.R.; Kim, C.Y.; Um, B.H. Determination of major phlorotannins in *Eisenia bicyclis* using hydrophilic interaction chromatography: Seasonal variation and extraction characteristics. *Food Chem.* 2013, 138, 2399–2406.
83. Stengel, D.B.; Connan, S. Natural products from marine algae: Methods and protocols. *Nat. Prod. Mar. Algae Methods Protoc.* 2015, 1308, 1–439.
84. Santos-Buelga, C.; Gonzalez-Manzano, S.; Dueñas, M.; Gonzalez-Paramas, A.M. Extraction and isolation of phenolic compounds. In *Natural Products Isolation*; Springer: Berlin/Heidelberg, Germany, 2012; pp. 427–464.
85. Vieira, V.; Prieto, M.A.; Barros, L.; Coutinho, J.A.; Ferreira, I.C.; Ferreira, O. Enhanced extraction of phenolic compounds using choline chloride based deep eutectic solvents from *Juglans regia* L. *Ind. Crops Prod.* 2018, 115, 261–271.

86. Ospina, M.; Castro-Vargas, H.I.; Parada-Alfonso, F. Antioxidant capacity of Colombian seaweeds: 1. extracts obtained from *Gracilaria mammillaris* by means of supercritical fluid extraction. *J. Supercrit. Fluids* 2017, 128, 314–322.
87. Otero, P.; López-Martínez, M.I.; García-Risco, M.R. Application of pressurized liquid extraction (PLE) to obtain bioactive fatty acids and phenols from *Laminaria ochroleuca* collected in Galicia (NW Spain). *J. Pharm. Biomed. Anal.* 2019, 164, 86–92.
88. Kalil, S.J.; Moraes, C.C.; Sala, L.; Burkert, C.A. Bioproduct extraction from microbial cells by conventional and nonconventional techniques. In *Food Bioconversion*; Academic Press: Cambridge, MA, USA, 2017; pp. 179–206.
89. Cikoš, A.M.; Jokić, S.; Šubarić, D.; Jerković, I. Overview on the application of modern methods for the extraction of bioactive compounds from marine macroalgae. *Mar. Drugs* 2018, 16, 348.

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