HDAC Inhibitors in Triple Negative Breast Cancer

Subjects: Chemistry, Medicinal

Contributor: Cristina Maccallini, Alessandra Ammazzalorso, Barbara De Filippis, Marialuigia Fantacuzzi, Letizia

Giampietro, Rosa Amoroso

Triple negative breast cancer (TNBC) is an urgent as well as huge medical challenge, which is associated with poor prognosis and responsiveness to chemotherapies. Since epigenetic changes are highly implicated in TNBC tumorigenesis and development, inhibitors of histone deacetylases (HDACIs) could represent a promising therapeutic strategy.

anticancer

benzamide

chimeric compounds

hydroxamate

1. Introduction

Breast cancer (BC) is the most prevalent cancer in females worldwide. In 2020, 2.3 million women were diagnosed with BC and 685,000 deaths were recorded [1], which represents a public health issue. Among the BC subtypes, the hormone receptor (HR) positive is the most common, followed by the human epidermal growth factor receptor 2 (HER2) positive and the so-called triple negative (TN) phenotype. Triple negative breast cancers are the most aggressive subtype, since they present high-grade invasiveness, high tendency to give rise to distant metastasis, and are poorly responsive to the neoadjuvant anthracycline- and taxane-based chemotherapies. Dramatically, the overall TNBC patients' survival is about 1 year [2]. Therefore, the identification of new therapeutic options to treat this cancer is an urgent as well as huge medical challenge.

Epigenetic changes are highly implicated in TNBC tumorigenesis and development, and they are affected by the chromatin remodeling [3]. Histone deacetylases (HDACs, **Table 1**) are a family of specialized enzymes responsible for the removal of acetyl groups from the ε-amino group of the proteinogenic histone lysine residues, leading to a compressed chromatin structure and to the consequent gene transcription suppression [4]. Based on the homology with yeast deacetylases, cellular localization, size, and number of active sites, they can be divided into four classes, i.e., Class-I (HDAC isoforms 1, 2, 3, and 8); Class-IIa (HDAC isoforms 4, 5, 7, and 9); Class-IIb (HDAC isoforms 6 and 10); and Class-IV (HDAC isoforms 11). On the other hand, histone acetyltransferases (HATs) are responsible for the inverse modification, i.e., they catalyze the lysine acetylation, leading to chromatin relaxation and active gene transcription [5]. Considering their crucial role in gene transcription and translation, and their overexpression in a variety of human cancers, HDACs have received considerable attention from researchers as therapeutic targets to manage cancer initiation and progression [6]. Different HDAC inhibitors (HDACIs) have been approved by the Food and Drug Administration (FDA), particularly against hematological diseases, such as Vorinostat, Romidepsin, Belinostat, and Panobinostat (**Figure 1**). Moreover, HDACIs have been investigated for

the treatment of solid tumors, including TNBC, although clinical trials involving single agents provided poor results. Nevertheless, recent studies emphasize the high potential impact for HDACIs in controlling TNBC, since these agents can regulate the tumor angiogenesis and metastasis [7][8].

Figure 1. Chemical structure of FDA approved HDAC inhibitors.

2. HDAC Inhibitors and their Mode of Action against TNBC

2.1. Pharmacophoric Model of HDAC Inhibitors

A pharmacophoric model has been established for HDAC inhibitors based on the common structure of the different 11 zinc-dependent isoforms of the HDAC family. The deacetylase domain of these enzymes shows a surface cavity, which is connected to the inner active site by means of an intermediate channel, where the four carbons of the lysine side chain of the natural substrate are accommodated. The active site contains a Zn²⁺ ion, which binds to the histone N-ε acetylated lysine, with the subsequent attack to the N-acetyl group by a water molecule and the production of an N-3 free lysine and acetic acid ^[9]. In some HDAC isoforms, there is an additional hydrophobic cavity referred to as the "foot pocket", where acyl side chains longer than the acetyl are accommodated ^[10]. Based on this enzyme architecture, HDACIs are typically endowed with a zinc binding group (ZBG) and an aromatic tail occupying the entrance area (CAP group), connected by a central lipophilic linker (**Figure 2**). According to the ZBG, HDAC inhibitors are classified in four classes: carboxylic acids, thiols, benzamides, and hydroxamic acids (**Figure 2**).

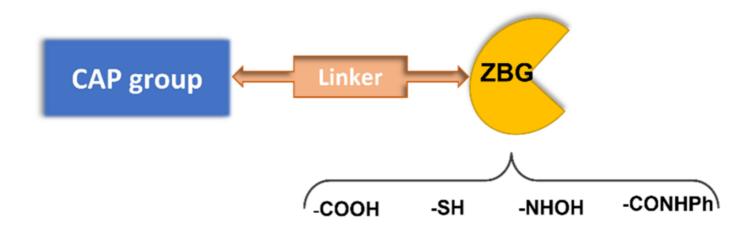


Figure 2. Pharmacophoric model for HDAC inhibitors.

The first generation of compounds obtained based on this pharmacophoric model, not exploiting the differences among the HDAC isoforms, resulted in pan-HDAC inhibitors that are often associated with numerous side effects. Indeed, the biological consequences of targeting multiple HDACs can be unpredictable, given the pleiotropic activities of these enzymes. Thanks to the increasing availability of crystallographic data and comprehensive SAR studies, significant advances in the research of HDACs isoform-selective inhibitors have been carried out [11][12]. The major differences among the HDACs are found at the entry of the active site and in the channel length and size. Moreover, the presence or the absence of the "foot-pocket" is another important difference among these metallo-enzymes, which could be exploited in the design of selective agents. For example, Entinostat and Tucidinostat are benzamide-based Class-I selective HDAC inhibitors (Figure 3) currently under clinical evaluation in both the US and Europe for the treatment of both cancer and nononcological diseases [13][14]. Ricolinostat and Citarinostat (Figure 3) are hydroxamates, which are guite selective toward the Class-II HDAC6 (HDAC1/HDAC6 selectivity index = 13 and 12, respectively) and are under evaluation against different malignancies [15][16]. In general, new and more selective HDAC6 inhibitors have been disclosed as useful tools to counteract tumor progression [17][18], although it was also reported that the selective inhibition of HDAC6 is not necessarily related to an anticancer activity, as observed for Tubathian A (Figure 3) and related compounds [19]. Therefore, increasing attention is required for the evaluation of HDAC inhibitors as anticancer drugs, especially when used as single agents. In recent years, several researches in the field of HDAC inhibitors have pointed to the hydroxamic acid and benzamide type scaffolds that selectively target HDAC. Currently, hydroxamates hold clinical success with respect to benzamides, with three FDA approved molecules (Vorinostat, Belinostat, and Panobinostat), while only Tucidinostat has been approved by the Chinese FDA to treat patients with recurrent or refractory peripheral T-cell lymphoma [20].

Figure 3. Class-selective HDAC inhibitors.

2.2. HDACIs Biological Action against TNBC Progression

Both unselective and selective HDACIs have been investigated against TNBC. Additionally, these studies have disclosed the different pathways influenced by HDACIs that trigger the observed antitumoral effects (**Table 2**).

Table 2. Summary of the main biological effects of HDACIs in TNBC cells.

HDACIS	Action	Ref.
Pan-HDACIs (Vorinostat, Panobinostat) and Class-I Selective (Entinostat)	Antiproliferative, pro-apoptotic, and immunomodulatory by: • ↑p21 and p27 • ↓FOXA1	[7][21][22][23][24] [25][26][27]

HDACIS	Action	Ref.
	• ↓BCL2	
	• ↑PD-L1	
	• ↓DNA repair	
	• ↓MMP9	
Class-I HDAC Selective (Entinostat)	ER- to ER+ conversion; compromise vasculogenic mimicry data	[<u>8</u>][<u>28</u>]
HDAC6 Selective (BAS-6)	Alteration of the glycolytic metabolism	[<u>29</u>]

apoptotic

agents against TNBC cell lines, such as MDA-MB-231, 4T1 and BT-549, by the modulation of multiple pathways and factors. The following factors can be mentioned: the activation of the epithelial-mesenchymal transition (EMT) phenotype and the downregulation of the growth factor FOXA1 [21], the impairment of the homologous recombination repair pathway of DNA [30], the upregulation of tumor suppressor factors p21 and p27, and the downregulation of the survival protein Bcl-2 [22]. Moreover, Vorinostat has the ability to impair BC cell migration and invasion via the inhibition of matrix metallo-proteinase 9 (MMP9) activity [23], both alone and in combination with ionizing radiation.

Interestingly, the Class-I selective HDACI Entinostat, but not pan HDACIs, had the ability to convert ER-negative tumors to ER-positive tumors, sensitizing TNBC to letrozole treatment [28][31]. Moreover, Entinostat demonstrated the ability to upregulate genes involved in the development of the vasculogenic mimicry, i.e., a tumor blood supply system independent from angiogenesis [8]. In addition, a Class-II selective inhibitor directed against the HDAC6, the BAS-6 (**Figure 3**), selectively kills apoptotic-resistant TNBC cells inducing their glycolytic metabolism alteration [29]

To date, the outcomes obtained in the TNBC management by HDACIs point to the significant potential of these compounds in controlling cancer proliferation and chemo-resistance.

References

- 1. World Health Organization. Breast Cancer. Available online: https://www.who.int/news-room/fact-sheets/detail/breast-cancer (accessed on 7 March 2022).
- 2. Duranti, S.; Fabi, A.; Filetti, M.; Falcone, R.; Lombardi, P.; Daniele, G.; Franceschini, G.; Carbognin, L.; Palazzo, A.; Garganese, G.; et al. Breast Cancer Drug Approvals Issued by EMA: A Review of Clinical Trials. Cancers 2021, 13, 5198.

- 3. Zolota, V.; Tzelepi, V.; Piperigkou, Z.; Kourea, H.; Papakonstantinou, E.; Argentou, M.-I.; Karamanos, N.K. Epigenetic Alterations in Triple-Negative Breast Cancer—The Critical Role of Extracellular Matrix. Cancers 2021, 13, 713.
- 4. Yoshida, M.; Kudo, N.; Kosono, S.; Ito, A. Chemical and structural biology of protein lysine deacetylases. Proc. Jpn. Acad. Ser. B 2017, 93, 297–321.
- 5. Grunstein, M. Histone acetylation in chromatin structure and transcription. Nature 1997, 389, 349–352.
- 6. Jenke, R.; Reßing, N.; Hansen, F.; Aigner, A.; Büch, T. Anticancer Therapy with HDAC Inhibitors: Mechanism-Based Combination Strategies and Future Perspectives. Cancers 2021, 13, 634.
- 7. Terranova-Barberio, M.; Thomas, S.; Ali, N.; Pawlowska, N.; Park, J.; Krings, G.; Rosenblum, M.D.; Budillon, A.; Munster, P.N. HDAC inhibition potentiates immunotherapy in triple negative breast cancer. Oncotarget 2017, 8, 114156–114172.
- 8. Maiti, A.; Qi, Q.; Peng, X.; Yan, L.; Takabe, K.; Hait, N.C. Class I histone deacetylase inhibitor suppresses vasculogenic mimicry by enhancing the expression of tumor suppressor and anti-angiogenesis genes in aggressive human TNBC cells. Int. J. Oncol. 2019, 55, 116–130.
- 9. Vannini, A.; Volpari, C.; Filocamo, G.; Casavola, E.C.; Brunetti, M.; Renzoni, D.; Chakravarty, P.; Paolini, C.; De Francesco, R.; Gallinari, P.; et al. Crystal structure of a eukaryotic zinc-dependent histone deacetylase, human HDAC8, complexed with a hydroxamic acid inhibitor. Proc. Natl. Acad. Sci. USA 2004, 101, 15064–15069.
- 10. Bressi, J.C.; Jennings, A.J.; Skene, R.; Wu, Y.; Melkus, R.; De Jong, R.; O'Connell, S.; Grimshaw, C.E.; Navre, M.; Gangloff, A.R. Exploration of the HDAC2 foot pocket: Synthesis and SAR of substituted N-(2-aminophenyl)benzamides. Bioorg. Med. Chem. Lett. 2010, 20, 3142–3145.
- 11. Roche, J.; Bertrand, P. Inside HDACs with more selective HDAC inhibitors. Eur. J. Med. Chem. 2016, 121, 451–483.
- 12. Ho, T.C.S.; Chan, A.H.Y.; Ganesan, A. Thirty Years of HDAC Inhibitors: 2020 Insight and Hindsight. J. Med. Chem. 2020, 63, 12460–12484.
- 13. Connolly, R.M.; Rudek, M.A.; Piekarz, R. Entinostat: A promising treatment option for patients with advanced breast cancer. Future Oncol. 2017, 13, 1137–1148.
- 14. Zhang, M.-C.; Fang, Y.; Wang, L.; Cheng, S.; Fu, D.; He, Y.; Zhao, Y.; Wang, C.-F.; Jiang, X.-F.; Song, Q.; et al. Clinical efficacy and molecular biomarkers in a phase II study of tucidinostat plus R-CHOP in elderly patients with newly diagnosed diffuse large B-cell lymphoma. Clin. Epigenet. 2020, 12, 160.
- 15. Santo, L.; Hideshima, T.; Kung, A.; Tseng, J.-C.; Tamang, D.; Yang, M.; Jarpe, M.; Van Duzer, J.H.; Mazitschek, R.; Ogier, W.C.; et al. Preclinical activity, pharmacodynamic, and

- pharmacokinetic properties of a selective HDAC6 inhibitor, ACY-1215, in combination with bortezomib in multiple myeloma. Blood 2012, 119, 2579–2589.
- 16. Huang, P.; Almeciga-Pinto, I.; Jarpe, M.; Van Duzer, J.H.; Mazitschek, R.; Yang, M.; Jones, S.S.; Quayle, S.N. Selective HDAC inhibition by ACY-241 enhances the activity of paclitaxel in solid tumor models. Oncotarget 2017, 8, 2694–2707.
- 17. Nawar, N.; Bukhari, S.; Adile, A.A.; Suk, Y.; Manaswiyoungkul, P.; Toutah, K.; Olaoye, O.O.; Raouf, Y.S.; Sedighi, A.; Garcha, H.K.; et al. Discovery of HDAC6-Selective Inhibitor NN-390 with In Vitro Efficacy in Group 3 Medulloblastoma. J. Med. Chem. 2022, 65, 3193–3217.
- 18. Li, S.; Zhao, C.; Zhang, G.; Xu, Q.; Liu, Q.; Zhao, W.; Chou, C.J.; Zhang, Y. Development of selective HDAC6 inhibitors with in vitro and in vivo anti-multiple myeloma activity. Bioorg. Chem. 2021, 116, 105278.
- 19. Depetter, Y.; Geurs, S.; De Vreese, R.; Goethals, S.; Vandoorn, E.; Laevens, A.; Steenbrugge, J.; Meyer, E.; De Tullio, P.; Bracke, M.; et al. Selective pharmacological inhibitors of HDAC6 reveal biochemical activity but functional tolerance in cancer models. Int. J. Cancer 2019, 145, 735–747.
- 20. Chan, T.S.; Tse, E.; Kwong, Y.-L. Chidamide in the treatment of peripheral T-cell lymphoma. OncoTargets Ther. 2017, 10, 347–352.
- 21. Wu, S.; Luo, Z.; Yu, P.J.; Xie, H.; He, Y.W. Suberoylanilide hydroxamic acid (SAHA) promotes the epithelial mesenchymal transition of triple negative breast cancer cells via HDAC8/FOXA1 signals. Biol. Chem. 2016, 397, 75–83.
- 22. Carlisi, D.; Lauricella, M.; D'Anneo, A.; Buttitta, G.; Emanuele, S.; di Fiore, R.; Martinez, R.; Rolfo, C.; Vento, R.; Tesoriere, G. The Synergistic Effect of SAHA and Parthenolide in MDA-MB231 Breast Cancer Cells. J. Cell. Physiol. 2015, 230, 1276–1289.
- 23. Chiu, H.-W.; Yeh, Y.-L.; Wang, Y.-C.; Huang, W.-J.; Chen, Y.-A.; Chiou, Y.-S.; Ho, S.-Y.; Lin, P.; Wang, Y.-J. Suberoylanilide Hydroxamic Acid, an Inhibitor of Histone Deacetylase, Enhances Radiosensitivity and Suppresses Lung Metastasis in Breast Cancer In Vitro and In Vivo. PLoS ONE 2013, 8, e76340.
- 24. Min, A.; Im, S.A.; Kim, D.K.; Song, S.H.; Kim, H.J.; Lee, K.H.; Kim, T.Y.; Han, S.W.; Oh, D.Y.; Kim, T.Y.; et al. Histone deacetylase inhibitor, suberoylanilide hydroxamic acid (SAHA), enhances antitumor effects of the poly (ADP-ribose) polymerase (PARP) inhibitor olaparib in triple-negative breast cancer cells. Breast Cancer Res. 2015, 17, 33.
- 25. Bali, P.; Pranpat, M.; Swaby, R.; Fiskus, W.; Yamaguchi, H.; Balasis, M.; Rocha, K.; Wang, H.G.; Richon, V.; Bhalla, K. Activity of suberoylanilide hydroxamic Acid against human breast cancer cells with amplification of her-2. Clin. Cancer Res. 2005, 11, 6382–6389.
- 26. Gammoh, N.; Lam, D.; Puente, C.; Ganley, I.; Marks, P.A.; Jiang, X. Role of autophagy in histone deacetylase inhibitor-induced apoptotic and nonapoptotic cell death. Proc. Natl. Acad. Sci. USA

- 2012, 109, 6561-6565.
- 27. Bellucci, L.; Dalvai, M.; Kocanova, S.; Moutahir, F.; Bystricky, K. Activation of p21 by HDAC Inhibitors Requires Acetylation of H2A.Z. PLoS ONE 2013, 8, e54102.
- 28. Sabnis, G.J.; Goloubeva, O.; Chumsri, S.; Nguyen, N.; Sukumar, S.; Brodie, A.M.H. Functional Activation of the Estrogen Receptor-α and Aromatase by the HDAC Inhibitor Entinostat Sensitizes ER-Negative Tumors to Letrozole. Cancer Res. 2011, 71, 1893–1903.
- 29. Dowling, C.M.; Hollinshead, K.E.R.; Di Grande, A.; Pritchard, J.; Zhang, H.; Dillon, E.T.; Haley, K.; Papadopoulos, E.; Mehta, A.K.; Bleach, R.; et al. Multiple screening approaches reveal HDAC6 as a novel regulator of glycolytic metabolism in triple-negative breast cancer. Sci. Adv. 2021, 7, eabc4897.
- 30. Woo, Y.M. Epigenetic regulation in cystogenesis. Adv. Exp. Med. Biol. 2016, 933, 59-68.
- 31. De Cremoux, P.; Dalvai, M.; N'Doye, O.; Moutahir, F.; Rolland, G.; Chouchane-Mlik, O.; Assayag, F.; Lehmann-Che, J.; Kraus-Berthie, L.; Nicolas, A.; et al. HDAC inhibition does not induce estrogen receptor in human triple-negative breast cancer cell lines and patient-derived xenografts. Breast Cancer Res. Treat. 2015, 149, 81–89.

Retrieved from https://encyclopedia.pub/entry/history/show/59589