Cistus ladanifer as a Potential Feedstock for Biorefineries

Subjects: Energy & Fuels

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Cistus ladanifer (rockrose) is a widespread shrub species in the Mediterranean region well known due to its production of labdanum gum, especially in the hot season. Its leaves and branches can be subjected to different extraction and distillation processes to produce various types of extracts. The natural extracts of *C. ladanifer* have several applications, especially in the perfumery and cosmetics sector. *C. ladanifer* extracts, in addition to presenting interesting odoriferous properties, are also known for their bioactive properties, such as antioxidant and antimicrobial. The use of this species in animal feed or phytostabilisation of mining areas has also been successfully applied. Furthermore, the lignin and polysaccharides that are the major fractions from *Cistus* residues can be relevant sources of high-value products in a biorefinery framework.

added-value products bioeconomy biofuels essential oils integrated upgrade

rockrose

1. Introduction

Cistus ladanifer L. (crimson-spot rockrose) is a wild perennial shrub species of the *Cistaceae* family and the *Cistus* genus that is mainly distributed in Mediterranean countries, such as France, Greece, Spain, Portugal, Morocco, Algeria, and Cyprus ^{[1][2][3][4][5][6]}.

The *C. ladanifer* species includes three subspecies: *ladanifer* (...), *africanus* (Dans), and *sulcatus* (Demoly). The subsp. *ladanifer* is mainly distributed in the Iberian Peninsula, France, and northern Africa; the subsp. *sulcatus* is endemic to southwestern Portugal along the coastal cliff tops of Costa Vicentina; and the subsp. *africanus* is commonly found in northern Africa and also spread in southern Spain ^{[7][8]}. Subsp. *ladanifer* has an erect habit, generally with linear–lanceolate and lanceolate leaves ^{[5][9]}; subspecies *sulcatus* is a prostrated habit shrub (50 cm) when it grows near the sea or up to 200 cm and erect when protected from the wind, with white flowers and sessile leaves, generally elliptical or oblanceolate, with accentuated nervures in the upper surface ^[9]; and subsp. *africanus* has leaves with an apparent petiole, lanceolate–elliptic, or oblong to linear–generally lanceolate–with little apparent nerves on the upper side, and other leaves oval or obovate, with well visible nerves on the upper side.

C. ladanifer is a shrub, generally erect, with a reddish-brown stem, hardwood, and sticky bark with striking vivid flowers (**Figure 1**). The 40–80 mm long leaves are impregnated by the labdanum, which makes them sticky and with a strong and specific smell ^[10]. It is considered a species of rapid growth and development that reproduces

easily ^[11] by natural seed propagation during winter and autumn ^{[5][12]}. In gardening, *Cistus* spp. can be multiplied by stem cuttings and layering ^[13]. Vegetative propagation of *C. ladanifer* is carried out preferably in September and October using lateral cuttings of 1 to 15 cm in length (without flowering) with five or six pairs of leaves or also 8 to 12 cm long mature wooden stem stakes with a heel or knot ^[14]. Micropropagation by germinating seeds of *C. ladanifer* and other species of Cistus has also been successfully studied ^[15]. The exploitation of wild populations to produce essential oil and labdanum has been the main practice.

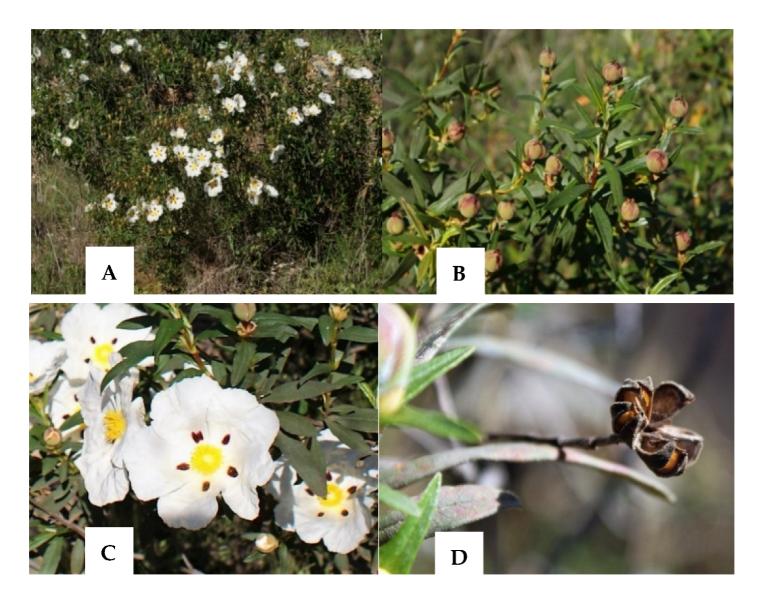


Figure 1. Cistus ladanifer L.: (A) whole plant; (B) flower buds; (C) flowers; (D) open cysts.

C. ladanifer occurs in a wide range of altitudes, latitudes, climates, and soil types, but it prefers acidic and siliceous soils ^{[16][17]} and dry areas with high insolation where it can give rise to large and dense populations. It has high-stress tolerance and, consequently, is competitive under various environmental conditions, including poor soils with low organic matter content, low pH, and high concentrations of trace elements, as well as hydric stress and high temperature and solar radiation ^[18]. This tolerance may be associated with the activity of different isoenzymes, namely superoxide dismutase (SOD), peroxidase (POD), and catalase (CAT) ^[19], which are the major enzymes involved in the detoxification of active oxygen species (AOS) ^[20]. An evaluation was made of As, Cu, Pb, and Zn

concentrations and activity of the soluble and cell wall ionically bound forms of enzymes SOD, POD, and CAT in leaves of *C. ladanifer*. The enzymatic activity of these enzymes varied with populations of *C. ladanifer*, as well as with the seasons of the year, with specificity for each trace element and the location of the enzyme in the cells ^[19].

C. ladanifer shows an important adaptative mechanism in post-fire plant dynamics, and its recovery after burning is faster when compared with other shrub species (e.g., *Erica australis* and *Calluna vulgaris*) ^[21]. *Cistus* species tend to recover by an autosuccession process in areas affected by cutting and burning, being dominant after the first or second year ^[22]. Its strong heat resistance suggests that there is a break in seed dormancy by the high temperatures during a fire; e.g., *Quercus* woodlands are occupied by *C. ladanifer* after fire ^[5]. A study under controlled conditions demonstrated an increase in the germination of seeds preheated at 100 °C when compared to seeds stored at room temperature ^[23]. *C. ladanifer* seeds and plantlets generally show rapid germination and growth ^[24].

The fruits are globular lignified capsules (cysts) that can produce 500 to 1000 seeds, and a single plant may produce 250 thousand seeds annually ^{[25][26]}. Mature fruits remain attached to the plant and release the seeds gradually when they open, allowing short-distance dissemination for a long time ^[5]. The seeds are small, allowing easy penetration and accumulation in the soil, and have a stiff and impermeable cover important for their longevity; however, they contain few nutrient reserves, which requires a quick start for the photosynthesis process ^[25]. The number of valves per fruit (five to twelve) can vary between populations, between plants, and within the same plant. Rockrose is a highly polymorphic plant and the only species of the *Cistaceae* family with a variable number of valves per fruit. This variation may be a result of natural selection, phenotypic plasticity, and developmental instability of the plant ^[27].

Rockrose flowers are white with a crimson spot on the base of the petals. There are two color varieties: *C. ladanifer* var. *albiflorus* with white petals and *C. ladanifer* var. *maculatus* with red stains at the petal base ^[28]. The flowers are large (ca. 64 mm in diameter), appear during spring (March–May), and produce abundant pollen and nectar ^{[5][29]}. The size and longevity of the flowers positively influence the incidence of folivores, mainly ants and beetles ^[30].

The ecosystem of *C. ladanifer* also provides high proliferation of edible mushroom species, some of which are in high demand due to their gastronomic interest ^[31]. Rockrose is also considered promising for phytoremediation and revegetation of contaminated soil in semi-arid climates as it is capable of colonizing these soils with a great capacity for tolerance and adaptability to adverse climatic conditions, e.g., droughts and high temperatures ^{[17][32]}.

Cistus ladanifer (rock rose) is widespread throughout the Mediterranean basin, where it is a naturally occurring shrub. It is estimated that it extends to circa 2 million hectares in the south and southwest of the Iberian Peninsula ^[33], where it is the main shrub species with an important ecological role but also an actor in some challenging situations, such as regarding rural fires that take place in summer in these regions. Rockrose is an example of an underused species without any regular and complete exploitation, and it is currently mainly exploited by the perfumery industry or as an ornamental plant ^[14], generally associated with small family businesses or linked to rural organizations. There is still a lack of robust economic exploitation as well as developed value chains.

2. Traditional Products from Rockrose

Rockrose plants have been traditionally used as an important resource for primary health care given their low cost, accessibility, and accumulated ancestral experience. The aerial parts of rockrose are used to produce extracts, obtaining exudates and essential oils with complex composition and pharmacological properties that make interesting further application targeted investigation ^[34]. Traditional use of rockrose has contributed to socio-economic development of rural communities ^[35]. Both labdanum and essential oil from *C. ladanifer* are much appreciated in perfumery, cosmetics, aromatherapy, and food flavors (restricted use), being used as ingredients in about 30% of modern perfumes due to their excellent fixative properties ^{[36][37]}. The *C. ladanifer* products are described in detail below.

2.1. Aromatic Extracts

Figure 2 summarizes the major conventional products obtained from rockrose for the perfumery/cosmetic industry and the main extraction methods used. Three main products can be obtained directly from the *C. ladanifer* plant: essential oil, using steam distillation or hydrodistillation, labdanum gum, employing hot alkaline extraction followed by acidulation, and cistus concrete, by extraction with apolar solvents, such as hexane and isopropanol. From these extracts, other products with interesting properties odoriferous and/or commercial can be further obtained, e.g., labdanum resinoid, labdanum oil, labdanum absolute from labdanum gum, and absolute from concrete. However, the portfolio of natural extracts obtained exclusively from *C. ladanifer* can still be more diversified for use in many applications, such as fragrance and flavoring ingredients, among which are hydrolate, labdanum resinoid 50%/DPG (dipropylene glycol synthetic), hydrocarboresine, dynamone, cistus absolute SIS, cistus by-absolute, cistus by-colorless, cistus organic oil, cistus water concentrate, and others ^[37].

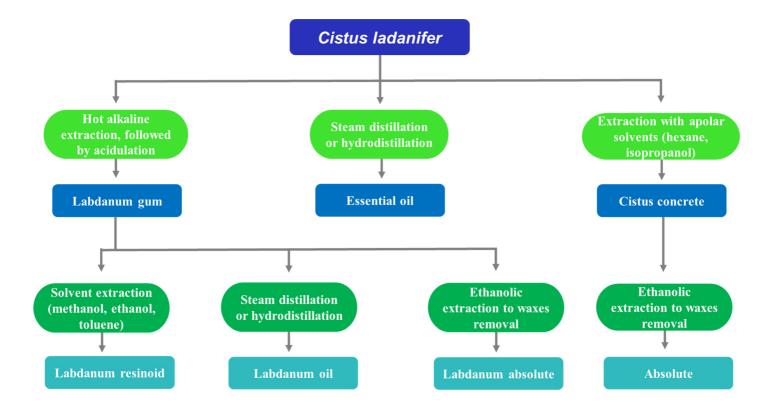


Figure 2. Main conventional aromatic extracts obtained from *C. ladanifer* and respective extraction methods.

Table 1 shows the major constituents detected in *C. ladanifer* essential oil from plants grown in different regions. The diversity observed can be due to several factors, such as climatic and soil variations, the stage of the vegetative cycle, seasonal factors, part of the plant analyzed, and the method used to obtain the essential oil, among others ^[38].

Table 1. Chemical composition (relative % of the peak area) of *C. ladanifer* essential oils from plants grown in different countries: center-interior of Portugal ^[29], central Spain ^[39], Corsica with plants of Spanish origin ^[40], and eastern Morocco ^[41].

Component	Centre Interior	Central Spain a	Corsica ^b	Eastern
	of Portugal ^a			Morocco ^a
Monoterpene hydrocarbons				
Tricyclene	-	-	-	2.7
α-Pinene	2.1	4.70	39	4.2
Camphene	0.3	0.64	2.1	15.5
Pinocarvone	1.1	-	0.9	-
Limonene	-	0.37	1.7	-
y-Terpinene	-	0.10	0.4	3.8
α-Terpinene	-	-	0.1	1.8
<i>p</i> -cymenene	-	1.17	1.7	2.3

Component	Centre Interior of Portugal ^a	Central Spain a	Corsica ^b	Eastern Morocco ^a
Oxygenated monoterpenes				
Bornyl acetate	1.6	7.03	3.1	
Terpinen-4-ol	1.0	6.37	1.1	6.3
α-Terpineol	-	2.20	-	1.2
trans-pinocarveol	2.1	20.00	1.9	
Borneol	0.7	-	0.8	11.1
Myrtenal	0.7	2.26	0.5	-
<i>cis-</i> Pinocamphone	-	3.84	-	-
2 (10)-Pinen-3-one	-	5.05	-	-
Verbonene	-	0.85	0.3	0.8
Camphor	-	0.86	-	1.5
p-Mentha-1,5-dien-8-ol	-	4.78	-	-
Sesquiterpene hydrocarbons				
Viridiflorene	1.3	0.41	-	-

Component	Centre Interior	Central Spain	Corsica ^b	Eastern
Component	of Portugal ^a	a	COISICa	Morocco ^a
C ₁₅ H ₂₆ O sesquiterpene alcohol	6.0	-	-	-
Cyclosativene	-	0.70	0.7	0.6
Aromadendrene	-	1.77	-	-
Allo-aromadendrene	0.8		1.9	-
α-Copaene	-	0.62	0.8	-
α-Cubebene	-	-	-	2.2
δ-cadinene	1.0	-	0.8	6.4
Oxygenated sesquiterpenes				
Viridiflorol	17.4	13.59	11.8	2.8
Spathulenol	0.8	0.53	0.5	-
Globulol	5.0	-	0.3	-
Ledol	-	4.36	3.3	-
Caryophyllene oxide	1.8	-	-	-

Component	Centre Interior of Portugal ^a	Central Spain ^a	Corsica ^b	Eastern Morocco ^a	
Palustrol	-	0.50	-	-	
Others					mercia
2,2,6-trimethylcyclohexanone	2.8	-	0.9	7.3	atic
Phthalates					retero,
Diethyl phthalate	-	-	-	2.9)
Bis (2-ethylhexyl) phthalate	-	-	-	0.2	Plant
Material used for	dry leaves and small branches	fresh leaves	leaves and stems	dry leaves	án, C.; 3 for
hydrodistillation					US

ladanifer (Cistaceae) despite the Absence of Special Dispersal Mechanisms. J. Biogeogr. 2009, 36, 954-968.

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PLoS ONE 2021, 16, e0258976. **2.2. Extractives**

9. Carlier, J.; LeitÃo, J.; Fonseca, F. Population Genetic Structure of Cistus ladanifer L. (Cistaceae)

Botande Canter a Diffesentiation from a concerning indicates selectives a primation and the concerning of the concerning in the concerning of the concerning

highlighted in this section correspond to a wider set of non-structural compounds that can be removed from the

plant. Extractives comprise a wide variety of chemical compounds, generally of low molecular weight, that are not 10. Demoly, J.-P.; Montserrat, P. Cistus; Castroviejo, S., Aedo, C., Laínz, M., Muñoz Garmendia, F., structural compounds of the plant cell walls ⁴². They can come from two general sources: (1) primary metabolites Nieto Feliner, G., Paiva, J., Benedí, C., Eds.; Flora iber; CSIC: Madrid, Spain, 1993. that include sugars, amino acids, simple fats, and various carboxylic acids, among others; and secondary

12 he Bouges, Aveidh Genteibuigão Plana co Estudo Dan Anatosnia Da Eollisa e Castoste Cistus la daniferdida,

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or Exernicial estel Madsidur Sesainuch 25 1 Actabyli 1288 (r pporoblean 1378s or lichens [43].

12. Alías, J.C.; Sosa, T.; Escudero, J.C.; Chaves, N. Autotoxicity Against Germination and Seedling The usual procedures for isolating extractives include traditional techniques (maceration, decoction, digestion, Emergence in Cistus ladanifer L. Plant Soil 2006, 282, 327–332. infusion, boiling under reflux, Soxhlet) and a wide range of modern techniques that have been introduced in recent

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14. Raimundo, J.R.; Frazão, D.F.; Domingues, J.L.: Quintela-Sabarís, C.; Dentinho, T.P.; Anjos, O.; Table 2 shows the major phytochemical categories of volatile and non-volatile compounds isolated from extracts Alves, M.; Delgado, F. Neglected Mediterranean Plant Species Are Valuable Resources: The solubilized from different parts of *C. ladanifer* that were identified using various chromatographic and spectroscopic Example of Cistus ladanifer. Planta 2018, 248, 1351–1364.

15. Iriondo, J.M.; Moreno, C.; Pérez, C. Micropropagation of Six Rockrose (Cistus) Species.

HTALLE Contracts obtained from various parts of the plant.

1	Compound	Chemical Group	Part of the Plant	References	tals in
1	Volatile compounds				Native
1	α-Pinene		Aerial part; shoots	[<u>45][46]</u>	ntion
-	Camphene		Aerial part; shoots	[<u>45][46]</u>	וונסוו ופ 578–
1	Pinocarvone		Aerial part; shoots	[<u>45][46]</u>	lanifer
2	Limonene	Monoterpene	Aerial part	[45]	Grass
2	α-Phellandrene	hydrocarbons	Leaves	[<u>47</u>]	–1570. ting and
	y-Terpinene		Aerial part	[45]	L80,
2	α -Thujene		Aerial part	[<u>45]</u>	-283.
2	<i>p</i> -cymene		Aerial part	[45]	. 1997,
2	Bornyl acetate	Oxygenated	Aerial part; shoots	[<u>45][46]</u>	<u>z,</u> Spain,
2	Terpinen-4-ol	monoterpenes	Aerial part; shoots	[<u>45][46]</u>	uestos ain,
	2006.				

26. Delgado, J.A.; Serrano, J.M.; López, F.; Acosta, F.J. Seed Size and Seed Germination in the Mediterranean Fire-Prone Shrub Cistus Ladanifer. Plant Ecol. 2007, 197, 269–276.

2Compound	Chemical Group	Part of the Plant	References	erspect
α-Campholenal		Aerial part	[45]	
2 trans-pinocarveol		Aerial part	[<u>45]</u>	f the nd Their
2 Borneol		Leaves; aerial part	[<u>45]</u>	IS
3 Myrtenal		Aerial part	[<u>45</u>]	the
3 (cis)-Verbenol		Leaves; shoots	[<u>46][47]</u>	⊃. Is. Agric
Verbonene 3		Leaves; aerial part; shoots	[<u>45][46][47]</u>	r L. 2016,
Camphor		Leaves; shoots	[<u>46][47]</u>	
3 Viridiflorol	al 2011, 331, 66–7	Shoots	[<u>46]</u>	ación de
3 Globulol		Shoots	[<u>46]</u>	adanife
3 Ledol	Oxygenated sesquiterpenes	Leaves	[<u>47</u>]	⁻ L.) En pañol,
Caryophyllene oxide		Shoots	[<u>46]</u>	ra-
Eugenol	Phenylpropene	Leaves	[<u>47</u>]	Flavor
Benzenepropanoic acid 3	Phenylpropanoid	Shoots	[<u>46]</u>	alogue-
2-Phenylethanol 3	Alcohol	Leaves; aerial part	[<u>45][47]</u>	J.

Identification of Flavonoid Content and Chemical Composition of the Essential Oils of Moroccan Herbs: Myrtle (Myrtus communis L.), Rockrose (Cistus ladanifer L.) and Montpellier Cistus (Cistus monspeliensis L.). J. Essent. Oil Res. 2011, 23, 1–9.

			.602–
Aromatic ketone	Leaves	[47]	
	Aerial part	[<u>45]</u>	ial Oil
Others	Shoots	[<u>46]</u>	cal Idanife
	Leaves; aerial part; shoots	[<u>45][46][47]</u>	ıality UK,
			of
Flavonoids	Leaves; aerial part; whole plant	[<u>48][49][50]</u>	he
	Leaves	[<u>51</u>]	án, C.
	Whole plant	[48]	s for
	Aerial part; leaves; whole plant	[<u>48][50][51]</u>	r L. 2016,
	Whole plant	[48]	s Of
	Leaves	[49]	retero
	Leaves	[49]	/onoid
	Leaves	[49]	
	Others	Aerial part Others Aerial part Shoots Leaves; aerial part; whole plant Flavonoids Leaves; aerial part; whole plant Leaves Aerial part; leaves; whole plant Aerial part; leaves; whole plant Leaves Leaves Leaves Leaves	Arrial part Arrial part Image: Shoots Image: Shoots

Compounds from a Cistus ladanifer Aqueous Extract. Phytochem. Anal. 2010, 21, 307–313.

5 C	ompound	Chemical Group	Part of the Plant	References	I.C. tracts.
	4´-dimethyl-kaempferol		Leaves	[<u>49]</u>	
5	3,7-dimethyl-kaempferol		Leaves	[49]	Genus ce of em.
5	3,7,4´-trimethyl-kaempferol		Leaves	[<u>49]</u>	เทt
5	Kaempferol methylether		Aerial part; leaves	[<u>50][51]</u>	enolic
	Quercetin-O-hexoside-Ohexoside		leaves	[<u>51]</u>	mpd.
5	Epigallocatechin	: Liquid Chromatog	Aerial part; leaves	[<u>50][51]</u>	ed to
	Gallic acid		Aerial part	[<u>50]</u>	3.
5	Glucogallin (isomer)		Aerial part	[<u>50]</u>	id
5	Gentisoyl glucoside	Phenolic acids and	Aerial part; whole plant	[<u>48][50]</u>	/onoid
0	Digaloil-β-D-glucopiranose	derivatives	Aerial part	[<u>50]</u>	
	Galloyl glucose		Leaves	[<u>51]</u>	it Ecol
5	Mirciaphenone B		Aerial part	[<u>50]</u>	۹.; N S. Foo
5	Punicalagin isomer 1	Ellagic acid and derivatives	Aerial part, leaves	[<u>50][51]</u>	5.100
	Punicalagin isomer 2		Aerial part, leaves	[<u>50][51]</u>	onal
	Punicalagin gallate 1		Leaves	[<u>51</u>]	onal 603.

62. Guerreiro, O.; Dentinho, M.T.P.; Moreira, O.C.; Guerra, A.R.; Ramos, P.A.B.; Bessa, R.J.B.; Duarte, M.F.; Jerónimo, E. Potential of Cistus ladanifer L. (Rockrose) in Small Ruminant Diets-

Compound	Chemical Group	Part of the Plant	References	dant
9 Punicalagin gallate 2		Leaves	[<u>51]</u>	on of c. 2007,
Punicalin		Aerial part; whole plant	[<u>48][50]</u>	tic
Cornusiin		Aerial part	[<u>50]</u>	us 427– terpenes,
Ellagic acid-7-xyloside		Aerial part	[<u>50]</u> [<u>52</u>]	rub Is Je to their
Ellagic acid		Aerial part [<mark>54</mark>]	[<u>50]</u>	ie to their ^[53] . They ∰e⊛fsuch
Ducheside A		Aerial part	[<u>50]</u>	<u>}</u> .
5 Shikimic acid		Aerial part	[<u>48][50]</u>	variety of bberellins tracts
	[<u>52</u>]	Whole plant		ince their
Quinic acid	[<u>56</u>]	Aerial part [<u>57]</u>	[<u>48][50]</u>	i 3,7,49- acid, <i>p</i> -
3	Others	Whole plant	[<u>58</u>]	ave been Effects
Hexahydroxydiphenoyl-D-glucose 7 (isomer)		Aerial part	[50]	15–149. part of C.
	[59]]		sugsnificant
Phenethyl-β-primeveroside		Aerial part	[<u>50]</u>	IS es occurs

In winter and the minimum in spring-summer, but the maximum production of flavonoids is detected in summer and 712e Tainaneen 16. Sintellarowse Aperatories invite as et house house house for the asteries. Sind Gamericember at Reserce as ethe confeigureined of AaConBiop 1991 urtes from Stores te Bionatasety Essential Dills grown Hydrolatees afreen Waaste shose grownpressensalvesisamicese Millingrod rolisture text atalifes from the Orange IP estel c 20270. VI. 44 diation 34 diation is

the main inductor in the production of flavonoids. This induction may be synergistically increased by drought, where 73. Hooda, V. Phytoremediation of Toxic Metals from Soil and Waste Water. J. Environ. Biol. 2007, there is higher production of methylated flavonoids (kaempferols and 7-methylated apigenins), suggesting that the 28, 367–376. methylated form is part of the defense mechanism of the plant against hydric stress [61]. Seasonal variations in the 712valvareneano Re NonAraulias Mara; callvansea tan Filemental disatake raneb Breat-lea a kesu Tranaster in Gisturic ladanifer L. Growing in a Contaminated Pyrite Mining Area (Aljustrel-Portugal). Water Air Soil

sub**βtalhutes2004**triðu52tjis£Le96and their deposition in cortical cells defend the plants from predators and protect the _ipternal tissues against UV-B radiation, respectively [63] 75. ADreu, M.M., Santos, E., Fernandes, E., Joao, M.; Ferreira, M. Acumulação e Translocação de Elementos Vestigiais Em Cistus ladanifer L. de Áreas Mineiras Da FPI Portuguesa. Rev. Ciênc. Vitamins, reducing sugars, and polyunsaturated fatty acids (PUFA) were also detected in rockrose. *C. ladanifer* Agrár. 2011, 34, 44–56. leaves presented a very high level of ascorbic acid (647.6 μg/g dry weight of the plant) and of sugars such as 76uc5æetogluEose; Albrese, MuM.rattalbais, f0.ctd/agaelingetse Mc&tFatTradoanE(estaentgD);strilbutiony investgitisof the platDeveltopedidensGosaareil/disecil/Adactescian.dr@istdis.laddarifer IlnoTexterabicevand iBioaticeul/inudiationextracts [64].GeedthemstEcoploent220162, df23JF45e6cturs in winter and spring and that of branched-chain fatty acids (BCFA) _in summer [65]. 77. Abreu, M.M.; Magalhães, M.C. Phytostabilization of Soils in Mining Areas. Case Studies from Portugal. In Soil Remediation; Aachen, L., Eichmann, P., Eds.; Nova Science Publishers Inc.: New **3**or**Bioactivity**9; pp. 297–344.

78 aribers normalized the second strength of the second strength of

799. Jerionainand, Eatuanated Companioes, S.P.; Dentinho, M.T.; Prates, J.A.; Vasta, V.; Santos-Silva, J.;

Bessa, R.J. Effect of Dietary Grape Seed Extract and Cistus ladanifer L. in Combination with **Table 3.** Biological activities of *C. ladanifer* extracts obtained with different solvents from different plant parts. Vegetable Oil Supplementation on Lamb Meat Quality. Meat Sci. 2012, 92, 841–847.

Plant Part	Type of Extract	Biological Activities	References
Fresh leaves from flowering stems	Methanol/water	Antifungal	<u>(67</u>)
Leaves	Aqueous	Autotoxicity	[<u>12</u>]
Wood/stalks, bark, and leaves	Ethanol and Acetone	Antioxidant	n: <u>(68)</u> - S- fr
Leaves	Aqueous	Allelopathic	[<u>58]</u> رازن 9
Aerial parts	Aqueous	Antihypertensive	(<u>69</u>) ta

of Meat through Dietary Cistus iadanifer L. In Lamp Fed increasing Levels of Polyunsaturated Fatty Acid Rich Vegetable Oils. Meat Sci. 2020, 164, 108092.

Plant Part	Type of Extract	Biological Activities	References	S
Whole plant	Hydroalcoholic and spray- dried/spray-dried aqueous extract	Antibacterial	[<u>4]</u>	Wor
		Antioxidant)	omic
Leaves	Aqueous	Antimicrobial,	[<u>59]</u>	d.
		Cytotoxic activity against human cancer cells		al Was
Leaves	Flavonoids	Allelopathic	[<u>70]</u>	
Shoots	Water-soluble and volatile compounds	Phytotoxic	[71]	oene
Leaves and small branches	Essential oil	Antitungical,	[28]	so, S 37.
Whole plant	Labdanum	Antibacterial		vier,
Aerial parts	Essential oil	Herbicidal activity	[<u>39]</u>	ellulos
	Essential oil			
Fruits, stems, flowers, and leaves	water, ethanol, ethanol: water (50:50), methanol, methanol: water (50:50), acetonitrile	Antioxidant		.; Alkal
	Hydrolates volatiles	Antioxidant		D- y
Aerial parts	Essential oil	Anti-inflammatory	[<u>72</u>]	ISS
		Antimicrobial		ion o

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4. Novel and Potential Applications

Other potential applications have been suggested for *C. ladanifer* besides the traditional use of extracts in their already well established application in the perfumery industry as a fixative of perfumes. Some of the new directions regarding the valorization of rockrose include phytoremediation of soils contaminated by heavy metals, use of the plant for feed with nutritional benefits for animal health, and use as a lignocellulosic raw material for added value products, namely the production of bioethanol.

4.1. Phytoremediation

Phytoremediation is an emerging environmentally friendly and low-cost technology that uses plants and their associated microorganisms to remove pollutants from contaminated sites, especially heavy metals ^[73]. *C. ladanifer* was reported to survive and grow in soils with high concentrations of toxic elements, such as Mn, Cu, Zn, Pb, and As ^{[74][75][76]}. Observations in mining areas suggest selectivity in the absorption and translocation of metals. Therefore, the tolerant behavior of this species in soils with toxic elements can expand the possibilities of phytoremediation in environments with significant rates of contamination ^[74].

One study on rockrose resistance to heavy metals was conducted in hydroponic experiments ^[16]. Higher tolerance to metals, such as Cd, Co, Cr, Mn, and Ni, was observed in plant populations originating from ultramafic soils or soils developed on basic rock, while populations originating from acid rock soils exhibited higher tolerance to Cu and Zn. Different patterns were observed in the accumulation of metals in the plant: Cd, Co, and Mn accumulated in the aerial part, while Cu and Pb were not transported efficiently through the roots up to the shoots. In general, *C. ladanifer* can accumulate heavy metals in the aerial parts without inhibiting plant growth ^[16].

Thus, its potential for phytostabilization in mine soil can be considered suitable considering trace element concentrations in leaves and seeds and seed germination rates ^[76]. Beyond immobilization of chemical elements, phytostabilization with autochthonous species also increases organic matter and water retention capacity, improving soil structure and reducing erosion ^[77].

It is worth mentioning that hexane extracts from *C. ladanifer* plants growing in mining areas did not exhibit potentially hazardous heavy metals, suggesting no human health risks ^[32]. Therefore, the high resistance of *C. ladanifer* to nutrient unbalanced soils, with potentially toxic elements and adverse climatic conditions, makes this species appropriate for phytoremediation and revegetation of contaminated soils ^[17].

4.2. Animal Feed

The use of *C. ladanifer* (soft stems and leaves or extracts) as a food supplement for animal nutrition and productivity increase has shown interesting results. The inclusion of leaves and soft stems of *C. ladanifer* in diets of lambs supplemented with an oil blend (sunflower and linseed oils) increased intramuscular fatty acids content ^[78]. The fatty acids provided by the diet with *C. ladanifer* benefited the health and did not jeopardize animal performance.

Another approach using a *C. ladanifer* diet, with or without oil supplementation, reduced lipid oxidation of lamb meat in pro-oxidant conditions and did not affect the meat's sensory properties [79].

Incorporation of ethanolic *C. ladanifer* extracts in rabbit feed showed also to be possible since it did not affect productivity, although the consumption rate was higher due to excess fiber and low protein ^[80]. Phenolic crude extracts from *C. ladanifer* were also employed in the treatment of soybean meal to reduce rumen degradation, which may be advantageous to increase the flux of potential feed protein into the post-ruminal compartments ^[81].

Several studies evaluated the effect of adding rockrose-condensed tannins in diets for lambs, showing that they can improve the digestive efficiency of soybean meal protein without compromising growth performance, blood metabolites, carcass characteristics, and meat quality, thus being able to reduce feed costs by reducing the content of protein used in diets ^[82]. Inclusion of low levels of *C. ladanifer* extracts of condensed tannins (1.25%) in the diet of lambs also showed favorable results in the pattern of ruminal biohydrogenation, while addition of 2.5% of condensed tannins negatively affected lamb growth, with no beneficial effect on the fatty acids composition of intramuscular and subcutaneous fat ^[83]. An increase in the α -tocopherol content in the muscle, with a reduction in the lipid oxidation of the meat, has also been reported in studies with the incorporation of leaves and soft stems of *C. ladanifer* in lamb diets ^[84].

4.3. Added-Value Products from Lignocellulosic Material

Development of industrial activity, fluctuations in the fossil fuel markets, and the need to minimize global climate change impacts have led to increasing interest in the use of natural resources, in particular lignocellulosic residues and byproducts in the context of a transition to a circular bio-based economy. In this sense, the valorization of biomass within the biorefinery concept has been gaining increasing relevance since a biorefinery integrates biomass conversion processes to obtain energy, materials, and chemical products, namely of added value ^[85]. These include biofuels, biochemical, and biobased compounds. Among these, organic acids, as well as bioactive compounds, are quite relevant. The phenolic compounds, both derived from extractives and from lignin, have strong potential for application in various industries, such as cosmetics, pharmaceuticals, and food, due to their potential functional activities.

Oligosaccharides, in particular xylooligosaccharides (XOS), are potential functional products that can be obtained from hydrolysis of hemicelluloses. These compounds have already been used as food ingredients (sweetener, weight control agents, humectants, etc.) and pharmacological supplements (prebiotic, anti-cariogenic, immunostimulant, antioxidant, antibiotic alternative, glycemic regulators, etc.), as well as in the cosmetic industry, animal and fish food, and agriculture ^[86]. In a study involving the techno-economic and environmental assessment of lignocellulosic-based small-scale biorefineries, it was demonstrated that the market price for products, such as XOS (USD 4.05/kg), for example, can give rise to significant economic profits, taking into account that the associated production costs are rather competitive (USD 1.18/kg) ^[87].

Rockrose lignocellulosic residues, from the production of essential oils and labdanum gum, are produced in significant amounts as these traditional products represent a small percentage of the total raw material. Thus, the products to be derived from the lignocellulosic fraction, even if they have a cheaper commercial cost, can be obtained in larger quantities. These residues are rich in polysaccharides and lignin, and, when they result from distilleries as extracted solid residues, they still contain an important fraction of extractives ^{[88][89][90]}. According to the literature, the chemical composition of *C. ladanifer* presents some variations depending on the plant parts and plant age. In ten years old whole plants, contents of polysaccharides, Klason lignin, and extractives of 41.5%, 15.6%, and 6.2%, respectively, were reported ^[91]. For distillery residues, composed mainly of the aerial part of the plant, the polysaccharide values are around 29.2–33.3%, Klason lignin 17–19.2%, and extractives 39.2–43.7% ^[88]. ^[90]. For extractive-free biomass, polysaccharide and lignin values increase to about 50% and 30%, respectively. Some abundant agro-industrial residues, such as olive pomace, sugar cane bagasse, or rice straw, have lower lignin contents than those found in extractive-free cistus biomass (16–26% vs. 29%) but higher polysaccharide contents (55–67% vs. 50%) ^[92].

Recently, the researchers presented a set of works that suggest an integrated upgrading strategy for *C. ladanifer* distillery biomass residues obtained after essential oil steam distillation. The strategy started with the removal of extractives (40% of the dry biomass), producing an extract rich in phenolic compounds (mainly gallic acid and flavonoids) with antioxidant properties ^{[89][90]}. The remaining lignocellulosic material, containing mainly polysaccharides (51%) and lignin (33%), was subjected to selective fractionation processes for sequential recovery of hemicelluloses by an autohydrolysis process ^[89] and lignin by organosolv and alkaline processes ^[93], producing a solid enriched in cellulose that had increased enzymatic digestibility (approximately four times higher than the initial feedstock) ^[94].

Optimization of the autohydrolysis process enabled to obtain XOS with potential prebiotic activity, in a maximum concentration of 16 g/L, corresponding to a yield of 10.2 g/100 g extracted feedstock ^[89]. The alkaline and organosolv delignification processes affected the monomeric composition of the residual lignin, with a decrease in the S/G ratio (quantified by analytic pyrolysis, Py-GC/MS) and solubilization and recovery of several phenolic compounds with high added value, namely vanillic acid, *p*-coumaric acid, and epicatechin ^[93]. The alkaline treatments lead to higher delignification (87%) and subsequent higher cellulose enzymatic saccharification yields (79%) ^[93]. Glucan-rich solids and pentoses-rich hemicellulosic hydrolysates were used, separately or together, for the selective production of the D-lactic acid isomer (D-LA) by the recombinant strain *E. coli* JU15 in different fermentation modes: simultaneous saccharification and co-fermentation (SSCF), SHF, and SSF. In this study, high lactic acid yields, in the range of 92–99 g D-LA/100 g sugars were achieved ^[95].

5. Future Perspectives

The use of *C. ladanifer*, not only as a source of essential oils or labdanum gum but also as a feedstock to obtain other products, namely from its industrial residues, can be strategic in the expansion of essential oil distilleries and consequently in the development of rural areas where endogenous biomass potential is still poorly explored, promoting the bioeconomy and circular economy models.

The possible integration of processes and products for the valorization of *C. ladanifer* in a biorefinery framework is shown in Figure 3 with full resource use and exploitation to obtain added-value products or novel applications, conveying the biorefinery goals, i.e., integral and sustainable use of biomass for concomitant production of biofuels, energy, materials, and chemicals, preferably with added value ^[85].

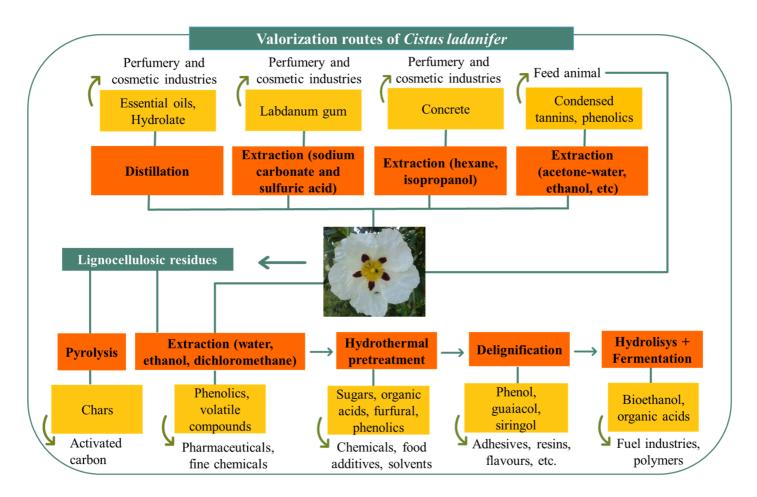


Figure 3. Potential path for use of *Cistus ladanifer* with an integrated valorization in a biorefinery framework for obtaining different added-value products.

Valorization of *C. ladanifer* may contribute to better use of an underexploited endogenous resource while promoting residue management, reducing pressure on the environment, and promoting sustainable development through the creation of new products and the generation of new jobs. Nevertheless, to shift biorefineries into industrial reality, the development of a distribution map highlighting local potential availability and aspects related to logistics of transport must be taken into consideration to build a complete sustainability analysis. Additionally, a comparative techno-economic analysis of the different processes, namely with the help of modeling tools, as well as a life cycle assessment in terms of environmental, social, and economic sustainability should also be carried out before selecting biomass commercial valorization pathways.