

# Bioactive Based Nanocarriers against SARS-CoV-2

Subjects: **Biology**

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Several plants and essential oils, as well as isolated bioactive compounds, such as phenolic acids, flavonoids, terpenes, lignans, coumarins, alkaloids, or proteins, showed a potential role as anti-viral agents. Nanoparticles have also been utilized in noninvasive imaging techniques to diagnose and monitor illnesses, as well as treatment responses. Nanomaterials can prevent viral binding to the host cell-surface receptor, which is crucial in the context of CoVs.

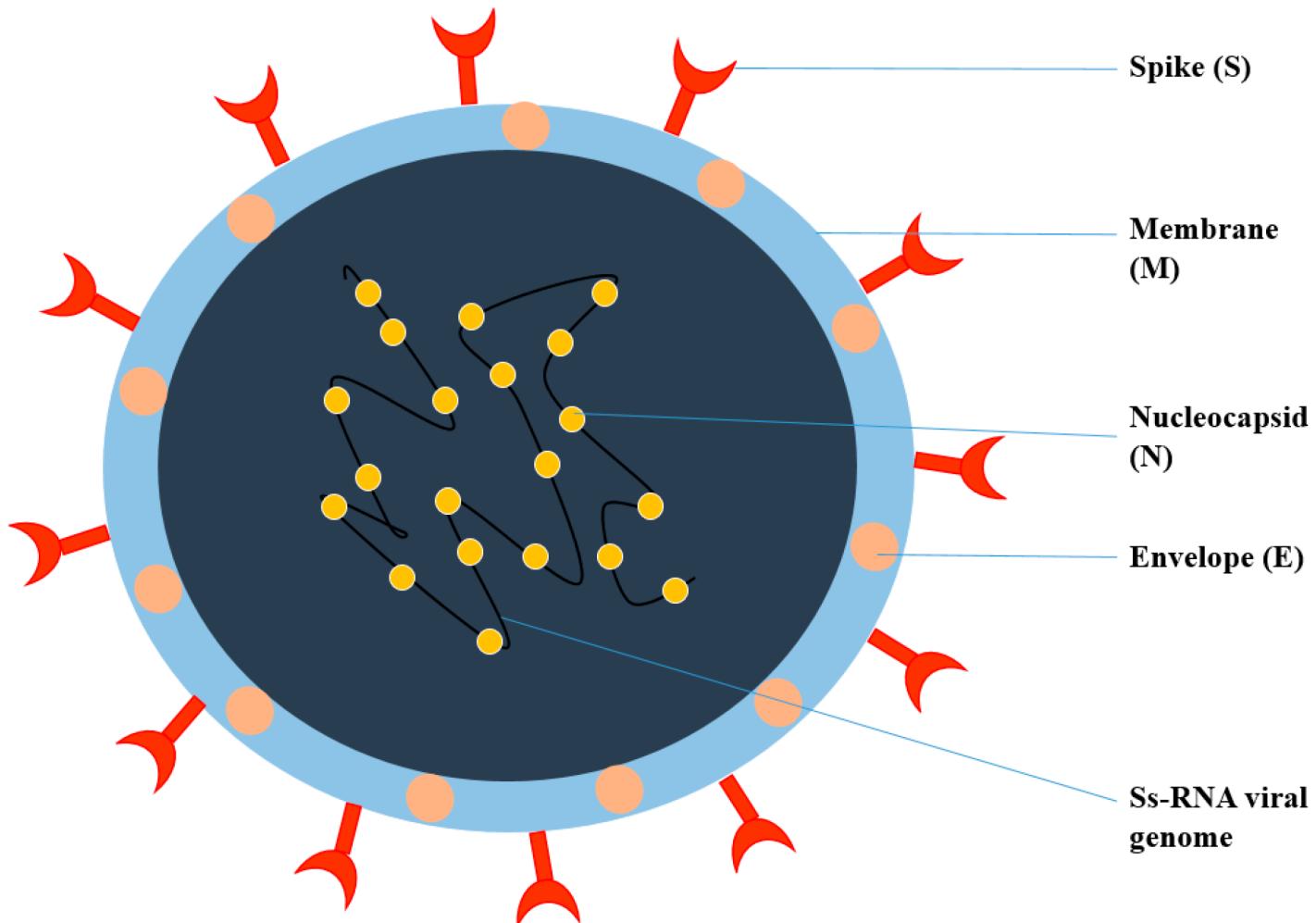
bioavailability

coronavirus

nano-formulations

## 1. General Structure of Viruses

Viruses are intracellular parasites, as depicted in **Figure 1**, composed of DNA or RNA inside a protein coat that lacks cell walls and cell membranes and does not carry out metabolic processes, so they must attach and enter the host's cells to use their energy for protein, DNA and RNA synthesis for their survival. It is difficult to kill and manage viral infections, as they live inside the host's cells and an anti-viral drug that kills viruses may also damage and kill the host's cells; therefore, for pharmaceutical scientists, it is quite challenging to develop an effective anti-viral drug and vaccine. Therapy for viral diseases using anti-viral drugs is further complicated by the fact that clinical symptoms appear late and until then, most viral particles have been replicated [1]. The key points that need to be considered for the successful development of anti-viral therapy include the development of a molecule that can enter the infected cells to interfere with the nucleic acid synthesis, or to prevent the binding and entry of the virus into the host cells, Most herbal bioactive compounds are known to work efficiently using this strategy, in addition to the synthesis of molecules that strengthen the body's immune system to combat infection. [2].



**Figure 1.** Structure of Coronavirus.

Coronavirus is an enveloped, single-stranded RNA virus, known to cause different disorders, such as hepatic, enteric, neurological, and respiratory diseases [3]. Coronaviruses are classified into four genera according to the taxonomic analysis, which includes  $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$  coronavirus. The  $\alpha$  and  $\beta$  coronavirus assemblies trigger respiratory track infection. SARS-CoV-2 falls under  $\beta$  and shares approximately 79.6% of SARS-CoV genome identity [4]. SARS-CoV-2 consists of structural proteins (S, E, M, and N) and non-structural proteins (nsp1–16). The S protein of the virus is a trimeric S glycoprotein that facilitates adhesion to the host cell's receptor. In most coronaviruses, the S protein is fragmented into two polypeptides called S1 and S2 via the host cell's furin-like protease. The S1 is a component of the S protein's large RBD, while the S2 is the stalk of the spike protein structure [4]. The non-structural protein Nsp1 is involved in RNA processing and replication. Nsp2 influences the host cell's survival signaling system. Nsp3 is thought to be involved in the separation of the translated protein. Nsp4 is a transmembrane domain 2 (TM2) protein that alters ER membranes. In replication, Nsp5 participates in the polyprotein process. Nsp6 is a transmembrane domain. The existence of nsp7 and nsp8 boosted the combination of nsp12 with template-primer RNA, considerably. Nsp9 is a protein that binds to ssRNA. Nsp10 is required for viral mRNA cap methylation. The COVID-19 genome codes four-wide structural and five accessory proteins, including ORF3a, ORF6, ORF-7, OR8, and ORF9, with a size of approximately 29 kb [5]. Coronaviruses replicate their genomes and transcribe their genes via an RNA-dependent RNA polymerase (RdRp) complex. The

RdRp complex of SARS-CoV-2 consists of subunits nsp12, nsp7, and nsp8, which enhance their processes and template binding [5].

## 2. Pathophysiology of COVID-19

The three major COVID-19 routes are as follows: (1) transmission by aerosol, (2) transmission by droplets, and (3) transmission by contact [6].

The incidence of COVID-19 infection in individuals is highly impacted by disorders, such as diabetes, hypertension, and lung diseases. This may be attributable to the increased ACE2 receptor expression in several organs, including the kidney, lungs, heart, and host epithelial cells. As COVID-19 enters the human body, it interacts with the ACE2 receptor and releases the RNA inside epithelial cells (EC). It then replicates and is released for further infection and spread via a nasal passage to the alveolar area of the lung [7]. The alveoli are generally the arbitration for gaseous exchange but, because of COVID-19 infection, there is increased permeability, pulmonary edema, disseminated intravascular coagulation (ICD) activation, pulmonary ischemia, hypoxia, respiratory failure, and severe lung damage [8]. It is then transported across the body, including GIT, brain, kidney, liver, and heart, through the blood from the respiratory tract, leading to various neurological disorders, comas, cerebral blood clots, ischemic strokes, and eventually death [9]. COVID-19 infects endothelial cells through ACE-2 binding, causing localized inflammation, endothelial activation, damaging tissue, and impaired release of cytokine. This severe aggregation of cytokine storm through vascular growth factor secretion, monocyte protein-1, interleukin-8, and decreased E-cadherin expression in epithelial cells contribute to vascular permeability and leakages, which are part of the acute respiratory distress syndrome pathophysiology (ARDS). Most Coronavirus-infected patients die from ARDS, in which pulmonary epithelial cells help to begin and transmit ARDS with a shift in the veracity of a vessel's barrier, encourage the condition of pro coagulation, induce vascular inflammation and reconcile inflammatory cell infiltration [10]. The epithelial cells are the key cause of pathogenesis of ARDS and multi-organ failure in COVID-19 patients, according to the hypothesis. Extreme COVID-19 infection stimulates coagulation mechanisms, which can potentially induce angiogenesis and possible epithelial cell hyperplasia, with the formation of DICs and blockages of small capillaries by the inflammatory cell, as well as significant thrombosis in larger vessels [11][12]. According to Teuwen et al., there are multiple mechanisms proposed for increased vascular permeability and vascular leakage in severely infected patients. The virus may have a direct effect on epithelial cells with widespread epithelial dysfunction, lysis, and death. It binds to ACE-2, decreasing ACE2 activity, indirectly turns on a kallikrein-bradykinin route with enhanced vascular permeability to reach the host cells, recruited in pulmonary epithelial cells, stimulated neutrophils to generate cytotoxic mediators, such as reactive oxygen species, immune cells, inflammatory cytokines, and vasoactive molecules contribute to improved contractility and a weakening of inter-endothelial connections in the epithelial cells. The cytokines IL-1 $\beta$  and tumor necrosis factor cause glucuronidases that both degrade glycocalyx and activate hyaluronic acid synthase 2, which results in increased hyaluronic acid deposition within the extracellular matrix and encourages fluid retention [13].

## 3. Natural Anti-Viral Plants and Phytochemicals

Phytoconstituents are naturally isolated bioactive molecules from plant parts, such as vegetables, fruits, flowers, leaves, and roots, that together with nutrients and fibers act as a defense system against disease or, more specifically, help to fight against the disease. Polyphenols, alkaloids, flavonoids, saponins, quinones, terpenes, proanthocyanidins, lignans, tannins, polysaccharides, steroids, organosulfur compounds, and coumarins are protruding bioactive phytochemicals, which have been used in experiments to combat viral infections and have attracted the attention of formulators because of their considerable advantages over synthetic molecules including low toxicity, side effects and low cost and have less potential to develop anti-viral resistance [14]. Phytochemicals are classified into primary and secondary constituents according to their metabolic activities. Primary constituents comprise common sugars, amino acids, proteins, and chlorophyll, while secondary constituents include alkaloids, terpenoids, phenolic compounds, and many more, such as flavonoids, tannins, and so on [15]. Plants have been established naturally over the years in various climatic environments on the planet and are enriched with a wide pharmacokinetic complexity of secondary metabolites. Some of the plants containing natural bioactive constituents are discussed below and play specific roles in combating diverse viral infections and diseases.

### 3.1. *Scutellaria baicalensis*

*Scutellaria baicalensis* has been used for over 200 years in TCM as a remedy for viral infection and inflammation. The roots of this plant have been used for viral infections, anti-inflammation, and anti-cancer activities. This plant also possesses diuretic, chalagogic and cathartic actions. It contains a variety of sterols, flavonoids, phenylethanoids, essential oils, etc. Its dried roots contain over 30 kinds of flavonoids, including baicalein, baicalin, wagonin, oroxylin A, etc. Baicalin is the most important component and has an anti-SARS coronavirus effect, anti-HIV, free radicle scavenging [16][17][18], etc. Another constituent named oroxylin A has anti-respiratory syncytial viral activity [19]. Current research confirmed that baicalin has in-vitro and in-vivo activity against dengue virus, influenza viruses, and enter virus-71 [20]. The compounds particularly hit cell attachment and intracellular replication of H3N2 and H1N1. In the murine RSV infection model, baicalin significantly reduced macrophages and T-lymphocyte infiltration to the lungs.

### 3.2. *Tanacetum vulgare*

*Tanacetum vulgare* L. also acknowledged as tansy is an herbaceous plant found in the temperate region of North Africa and Europe. It has been practiced as a traditional medicine that exhibits various properties, such as carminative, anti-diabetic, diuretic, anti-hypertensive, emmenagogue, and so on [21]. The extracts and active compounds isolated from this plant have shown several therapeutic applications, such as anti-bacterial, anti-fungal, anti-oxidant, immunomodulatory, anti-viral, etc. The chemical constituent of this plant includes flavonoids, vacuolar flavonoids (such as apigenin) glycosides, sterols (such as cholesterol and campesterol), triterpenes (taraxasterol and amyrin), and caffeic acid [22]. In 2009, Álvarez demonstrated the anti-viral activity of tansy against the HSV-1 virus from ethyl acetate extract from its aerial parts corresponding to parthenolide isolation [23]. Furthermore, the study revealed the anti-viral properties of methanol extract of the tansy aerial parts and was active against both HSV-1 and HSV-2 viruses [24]. Methanol extract from the blossom of the tansy plant showed anti-tumor properties against potato virus Y (PVY) and cucumber mosaic virus (CMV) [25].

### 3.3. *Ruta Angustifolia*

*Ruta angustifolia* Pers has been used as a traditional remedy for treating inflammation, curing mal conditions during pregnancy, respiratory problems, musculoskeletal systems, and so on [26]. In Indonesia, it was used prominently for treating liver disease and jaundice. Its chemical constituent consists of angustifolin and aromatic derivatives, including moskachan A, B, C, and D. Other constituents include ostruthin, xanthotoxin, dictamnine, xanthyletin, limonoid, psoralen, etc. [27]. The extract from the leaves of this plant exhibited potent anti-HCV properties. The constituent chalepin and pseudo IX from its leaves inhibited post-entry of HSV, RNA replication and synthesis of viral proteins.

### 3.4. *Liriope platyphylla*

*Liriope platyphylla* is a perennial plant present across the areas of China, Korea, and Japan and was demonstrated to exhibit biological activities against chronic diseases, such as cough, sputum, and neurodegenerative disorders, obesity, and diabetes [28]. Its chemical constituent spicatoside A interferes with lipopolysaccharide-induced (LPS) activation of extracellular signal-regulated kinases, c-Jun N-terminal kinase (JNK), and NF- $\kappa$ B [29]. It was found that *L. platyphylla* roots contain a principal active moiety that inhibits HBV viral promoter activity via interfering with the NF- $\kappa$ B signaling pathway [30].

### 3.5. *Citrus reticulata*

*Citrus reticulata* is an evergreen tree with aromatic flowers and glossy leaves that belongs to the family Rutaceae. It is referred to as tangerine, mandarin, or Kamla lebu in Bengal. The existence of different bioactive compounds confirms the uses of this plant for various ailments by traditional practitioners [31]. In 2009, Kirbaslar et al. demonstrated the various chemical components in this plant, which include limonene, myrcene,  $\gamma$ -terpinene, sabinene, and so on [32]. Guo et al. in 2016 and Choi et al. in 2011 demonstrated that, in China, it has been used for treating gastrointestinal and respiratory disorders [33][34]. Tangerine has also been documented to express an inhibitory effect against the respiratory syncytial virus (RSV) [35] and rotavirus replication [36]. Nobiletin, a key bioactive constituent isolated from its pericarps, affects the intracellular replication of RSV. The efficacy of tangerine was also reported against VHF-causing arenavirus entry.

### 3.6. *Tinospora cardifolia*

*Tinospora cardifolia*, also known as giloya, is an Indian native medicinal plant that belongs to the family Menispermaceae. It is extensively used in Ayurveda because of its therapeutic potential (anti-inflammatory, anti-allergic, anti-diabetic, and immunomodulatory activity). The aqueous extract of *Tinospora cardifolia* boosts immunity in children and is adjuvant to vaccination. In a patient with HIV infection, treatment with *Tinospora cardifolia* extract is found to reduce the TLC, neutrophile, and eosinophil count, suggesting anti- HIV activity [37]. S. Krupanidhi et al. carried out screening of the phytochemicals present in *Tinospora cardifolia* for an inhibitory effect against SARS-CoV-2. Based on the study, the phytochemicals berberine, tinosponone, xanosporic acid, and tembeterine were identified as the possible lead compounds that can be considered to have activity against SARS-

Cov-2. The research proved tinosponone as a potent, selective, and non-toxic inhibitor of 3 CL protease of SARS-CoV-2 [38].

### 3.7. *Andrographis paniculata*

*Andrographis paniculata*, generally known by the name of kalmegh, is one of the most commonly used medications in Ayurveda, belonging to the family Acanthaceae. The principal constituents present in this plant are flavonoids, diterpenoids, and polyphenols. The major bioactive compound responsible for its therapeutic activity is andrographolide [39]. Both 25 $\mu$ g/mL of ethanolic extract of *Andrographis paniculata* and 5  $\mu$ g/mL of Andrographolide demonstrated anti-viral activity against a variety of viruses, including influenza A virus, hepatitis B and C, herpes simplex virus, Epstein–Barr virus, and human immunodeficiency virus. They inhibit viral entry inside the viral cell and prevent the replication of genetic material by DNA and RNA polymerase, protein synthesis, and functional mature proteins [40]. Du et al. used alcohol as a co-surfactant, tween 80 as a surfactant, isopropyl alcohol as an oil phase, and water to prepare an andrographolide-loaded microemulsion [41]. Sermkaew et al. used caproyl, cremphor, and labrasol [42], while Syukri et al. used caproyl, tween 20, and polyethylene glycol 400 for the formulation of microemulsions. It was concluded in all the studies that the solubility and bioavailability of andrographolide were increased significantly [43].

### 3.8. *Saururus chinensis*

*Saururus chinensis*, belonging to the family Saururaceae, is a perennial herbaceous plant found in Korea and China. Traditionally, it has been used as an anti-pyretic, anti-inflammatory, and diuretic agent. A phytochemical investigation led to the discovery of various compounds, including flavonoids, aristolactamus, furanoditerpens, anthraquinones, etc. [44]. Lignans are the main bioactive constituent of plants that exhibit various activities, such as HIV-1 protease, NF- $\kappa$ B, and HIF1 inhibitory effects. Manassantin B, a bioactive compound extracted from the roots of *Saururus*, reported inhibitory activity against Epstein–Barr virus (EBV) lytic replication [45]. Some of the plants with anti-viral activities, along with their phytochemicals, have been summarized in **Table 1**.

**Table 1.** Plants with anti-viral activities, along with their phytochemicals.

Plant	Part	Phytochemical	Class	Active against Virus	Reference
<i>Ziziphus jujuba</i>	Roots	Jubanines	Alkaloids	PEDV	[46]
<i>Rheum palmatum</i>	Roots	Sennoside A	Glycoside	HIV-1	[47]
<i>Swietenia macrophylla</i>	Stem	Limonoids	Lignin	HSV	[48]
<i>Embelia ribes</i>	Seeds	Quercetin	Flavonoid	HSV	[49]
<i>Humulus lupulus</i>	Whole plant	Xanthohumol	Chalcone	BVDV	[50]
<i>Glycyrrhiza inflate</i>	Roots	Chalcones	Ketone	Influenza A	[51]

## 4. Anti-Viral Bioactive-Based Nanocarrier Systems

## References

Recent advances can give beneficial and promising therapeutic results when incorporated into pharmaceutical products.

nanotechnology. Boosted clinical and therapeutic responses are possible with nanoformulations used as carriers for delivering poorly soluble phytoconstituents and plant extracts [52]. Various delivery systems, such as self-nano COVID-19 Outbreak. *Curr. Biol.* 2020, **30**, 1346–1351.

emulsifying drug delivery systems (SNEDDS), hydrogels, phytosomes, microspheres, transferases, etc., have been used for the delivery of phytoconstituents with anti-viral potential. These nanoformulations displayed numerous effects such as improved oral solubility, systemic bioavailability, delayed metabolism, and enhanced therapeutic activity. For example, chitosan nanoparticles containing cathecol and EGCG resulted in an enhanced rate of

3. Sharma, A.; Tiwari, S.; Deb, M.K.; Marty, J.L. Severe acute respiratory syndrome coronavirus-2 particle carrier, which significantly increased its solubility profile [54]. The encapsulation of flavonoids into red blood cells can boost anti-viral activity and bioavailability. The flavonoids showed positive impacts in reducing oxidative 106054.

damage to the erythrocyte membrane [55]. Several researchers reported that RBCs play a vital role in the distribution and bioavailability of circulating quercetin [56]. Surprisingly, flavonoids in chitosan particles retain their

C. Li et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* 2020, 579, 270–273.

release of quercetin [58]. Kim et al. [59] boosted the oral bioavailability of artenitin by incorporating artenitin into the

5. Haraji, M.; Parashar, A.; Ranjbar, R.; Hetzel, M.; Verkord, F.S. Emerging coronaviruses: First into the water in oil emulsion system. Due to the poor solubility and permeability of baicalin, Chang et al. [60] used a SARS, second MERS and third SARS-CoV-2, epidemiological updates of COVID-19. *Le Infez* micellar composition including the carriers Pluronic P123 copolymer and sodium taurocholate, which significantly

increased oral absorption of baicalein. Oleanolic acid was incorporated into SMEEDS, which enhanced its systemic bioavailability.<sup>61</sup> Andrographolide has poor absorption properties and limited oral bioavailability. To circumvent this, IPC Precaution Recommendations: Scientific Brief; World Health Organization: Geneva, Switzerland.

methanolic extracts of strawberries (*Fragaria ananassa* Duch.) and ginger (*Zingiber officinale*) were utilized to [22]

are carried out on the development of nanotechnology for Indonesian Jamun to fight against SARS-CoV-2

[64] Various other formulations containing anti-viral phytoconstituents are summarized in Table 2.  
8. Teuwen, L.A.; Geldhof, V.; Pasut, A.; Carmeliet, P. Author correction: COVID-19, the vasculature

unleashed. *Nat. Rev. Immunol.* 2020, **20**, 389–391. **Table 2.** Various other formulations contain anti-viral phytoconstituents.

## 9. Zaim, S.; Chong, J.H.; Sankaranarayanan, V.; Harky, A. COVID-19 and multiorgan response. **Active Formulation Applications References**

## Phytoconstituent Formulation Applications References

1	Oxymatrine	Phytosome	Enhanced bioavailability	[65]	ocińska, S.
	<i>Artemisia arborescens</i>	Liposomal	Raised anti-viral activity and improved stability	[66]	
1	Hypocrellins	Nanoparticulate	Enhanced hydrophilicity and stability	[67]	el, A.;
	Matrine	Emulsion	Enhanced sustained released activity	[68]	nd
1	Quercetin	Microsphere	Enhanced bioavailability and sustained release formulation	[69]	

coagulation. *Lancet Respir. Med.* 2020, 8, 46–47.

1	Active Phytoconstituent	Formulation	Applications	References	
1	Curcumin	Nanostructured solid lipid carrier systems	Improved mucoadhesion and mucus penetration	[70]	M.T. technol.
	Honokiol	Cyclodextrin inclusion complexes	Enhanced solubility and bioavailability	[71]	

15. Vicas, S.I.; Cavalu, S.; Laslo, V.; Tocai, M.; Costea, T.O.; Moldovan, L. Growth, Photosynthetic Pigments, Phenolic, Glucosinolates Content and Antioxidant Capacity of Broccoli Sprouts in Response to Nanoselenium Particles Supply. *Not. Bot. Horti Agrobot. Cluj-Napoca* 2019, 47, 821–828.

16. Tai, D.Y. Pharmacologic treatment of SARS: Current knowledge and recommendations. *Ann. Acad. Med.* 2007, 36, 438–443.

17. Li, B.Q.; Fu, T.; Dongyan, Y.; Mikovits, J.A.; Ruscetti, F.W.; Wang, J.M. Flavonoid baicalin inhibits HIV-1 infection at the level of viral entry. *Biochem. Biophys. Res. Commun.* 2000, 276, 534–538.

18. Li, H.B.; Jiang, Y.; Chen, F. Separation methods used for *Scutellaria baicalensis* active components. *J. Chromatogr. B* 2004, 812, 277–290.

19. Ma, S.C.; Du, J.; But, P.P.; Deng, X.L.; Zhang, Y.W.; Ooi, V.E.; Xu, H.X.; Lee, S.H.; Lee, S.F. Antiviral Chinese medicinal herbs against respiratory syncytial virus. *J. Ethnopharmacol.* 2002, 79, 205–211.

20. Shieh, D.E.; Liu, L.T.; Lin, C.C. Antioxidant and free radical scavenging effects of baicalein, baicalin and wogonin. *Anticancer Res.* 2000, 20, 2861–2865.

21. Stevovic, S.; Surinski, V.; Calic-Dragosavac, D. Environmental adaptability of tansy (*Tanacetum vulgare* L.). *Afr. J. Biotechnol.* 2009, 8, 6290–6294.

22. Chandler, R.F.; Hooper, S.N.; Hoopper, D.L.; Jamieson, W.D.; Lewis, E. Herbal remedies of the maritime Indians: Sterols and triterpenes of *Tanacetum vulgare* L. (Tansy). *Lipids* 1982, 17, 102–106.

23. Onozato, T.; Nakamura, C.V.; Cortez, D.A.; Dias Filho, B.P.; Ueda-Nakamura, T. *Tanacetum vulgare*: An *in vitro* herpes virus activity of crude extract and the purified compound parthenolide. *Phytother. Res.* 2009, 23, 791–796.

24. Álvarez, Á.L.; Habtemariam, S.; Juan-Badaturuge, M.; Jackson, C.; Parra, F. In vitro anti HSV-1 and HSV-2 activity of *Tanacetum vulgare* extracts and isolated compounds: An approach to their mechanisms of action. *Phytother. Res.* 2011, 25, 296–301.

25. Petrov, N.; Stoyanova, M.; Valkova, M. Antiviral activity of plant extract from *Tanacetum vulgare* against Cucumber Mosaic Virus and Potato Virus Y. *J. Bio. Sci. Biotechnol.* 2016, 5, 189–194.

26. Pollio, A.; De Natale, A.; Appetiti, E.; Aliotta, G.; Touwaide, A. Continuity and change in the Mediterranean medical tradition: *Ruta* spp. (rutaceae) in Hippocratic medicine and present practices. *J. Ethnopharmacol.* 2008, 116, 469–482.

27. Wijeratne, E.M.K.; Bandara, B.M.R.; Gunatilaka, A.A.L.; Tezuka, Y.; Kikuchi, T. Chemical constituents of three Rutaceae species from Sri Lanka. *J. Nat. Prod.* 1992, 55, 1261–1269.

28. Han, Y.; Jung, H.W.; Lee, D.H.; Kwon, S.Y.; Son, K.H.; Park, Y.K. Anti-inflammatory effects of prosapogenin III from the dried roots of *Liriope platyphylla* in LPS-stimulated RAW264.7 cells. *J. Asian Nat. Prod. Res.* 2013, 15, 1038–1049.

29. Sindhu, R.K.; Verma, R.; Salgotra, T.; Rahman, M.H.; Shah, M.; Akter, R.; Murad, W.; Mubin, S.; Bibi, P.; Qusti, S.; et al. Impacting the Remedial Potential of Nano Delivery-Based Flavonoids for Breast Cancer Treatment. *Molecules* 2021, 26, 5163.

30. Huang, T.J.; Tsai, Y.C.; Chiang, S.Y.; Wang, G.J.; Kuo, Y.C.; Chang, Y.C.; Wu, Y.Y.; Wu, Y.C. Anti-viral effect of a compound isolated from *Liriope platyphylla* against hepatitis B virus in vitro. *Virus. Res.* 2014, 192, 16–24.

31. Kumar, P.; Bhaskar, A. Determination of bioactive components from the ethanolic peel extract of *Citrus reticulata* by gas chromatography–mass spectrometry. *Int. J. Drug Dev. Res.* 2012, 4, 166–174.

32. Kirbaslar, F.G.; Tavman, A.; Dülger, B.; Türker, G. Antimicrobial activity of Turkish Citrus peel oils. *Pak. J. Bot.* 2009, 41, 3207–3212.

33. Guo, J.; Tao, H.; Cao, Y.; Ho, C.T.; Jin, S.; Huang, Q. Prevention of Obesity and Type 2 Diabetes with Aged Citrus Peel (Chenpi) Extract. *J. Agric. Food Chem.* 2016, 64, 2053–2061.

34. Choi, M.Y.; Chai, C.; Park, J.H.; Lim, J.; Lee, J.; Kwon, S.W. Effects of storage period and heat treatment on phenolic compound composition in dried Citrus peels (Chenpi) and discrimination of Chenpi with different storage periods through targeted metabolomic study using HPLC-DAD analysis. *J. Pharm. Biomed. Anal.* 2011, 54, 638–645.

35. Xu, J.J.; Wu, X.; Li, M.M.; Li, G.Q.; Yang, Y.T.; Luo, H.J.; Huang, W.H.; Chung, H.Y.; Ye, W.C.; Wang, G.C.; et al. Antiviral activity of polymethoxylated flavones from “Guangchenpi”, the edible and medicinal pericarps of *Citrus reticulata* ‘Chachi’. *J. Agric. Food Chem.* 2014, 62, 2182–2189.

36. Savi, L.A.; Caon, T.; de Oliveira, A.P.; Sobottka, A.M.; Werner, W.; Reginatto, F.H.; Schenkel, E.P.; Barardi, C.R.; Simões, C.M. Evaluation of antirotavirus activity of flavonoids. *Fitoterapia* 2010, 81, 1142–1146.

37. Zhu, Z.; Lian, X.; Su, X.; Wu, W.; Marraro, G.A.; Zeng, Y. From SARS and MERS to COVID-19, a brief summary and comparison of severe acute respiratory infections caused by three highly pathogenic human coronaviruses. *Respir. Res.* 2020, 21, 224.

38. Krupanidhi, S.; Abraham Peele, K.; Venkateswarulu, T.C.; Ayyagari, V.S.; Nazneen Bobby, M.; John Babu, D.; Venkata Narayana, A.; Aishwarya, G. Screening of phytochemical compounds of *Tinospora cordifolia* for their inhibitory activity on SARS-CoV-2, an in silico study. *J. Biomol. Struct. Dyn.* 2021, 39, 5799–5803.

39. Vellingiri, B.; Jayaramayya, K.; Iyer, M.; Narayanasamy, A.; Govindasamy, V.; Giridharan, B.; Ganesan, S.; Venugopal, A.; Venkatesan, D.; Ganesan, H.; et al. COVID-19, a promising cure for the global panic. *Sci. Total Environ.* 2020, 725, 138277.

40. Dudani, T.; Saraogi, A. Use of herbal medicines on coronavirus. *Acta. Sci. Pharm. Sci.* 2020, 11, 416–419.

41. Hwang, B.Y.; Lee, J.H.; Nam, J.B.; Hong, Y.S.; Lee, J.J. Lignans from *Saururus chinensis* inhibiting the transcription factor NF- $\kappa$ B. *Phytochemistry* 2003, 64, 765–771.

42. Du, H.; Yang, X.; Li, H.; Han, L.; Li, X.; Dong, X.; Zhu, Q.; Ye, M.; Feng, Q.; Niu, X. Preparation and evaluation of andrographolide- loaded microemulsion. *J. Microencapsul.* 2012, 29, 657–665.

43. Sermkaew, N.; Ketjinda, W.; Boonme, P.; Phadoongsombut, N.; Wiwattanapatapee, R. Liquid and solid self-microemulsifyin drug delivery systems for improving the oral bioavailability of andrographolide from a crude extract of *Andrographis paniculata*. *Eur. J. Pharm. Sci.* 2013, 50, 459–466.

44. Syukri, Y.; Martien, R.; Lukitaningsih, E.; Nugroho, A.E. Novel Self-Nano Emulsifying Drug Delivery System (SNEDDS) of andrographolide isolated from *Andrographis paniculata* Nees: Characterization, in-vitro and in-vivo assessment. *J. Drug Deliv. Sci. Technol.* 2018, 47, 514–520.

45. Masters, P.S. The molecular biology of coronaviruses. *Adv. Virus Res.* 2006, 66, 193–292.

46. Kang, K.B.; Ming, G.; Kim, G.J.; Ha, T.K.Q.; Cho, H.; Sung, S.H. Jubanines F–J, cyclopeptide alkaloids from the roots of *Ziziphus jujuba*. *Phytochemistry* 2015, 43, 264–267.

47. Esposito, F.; Carli, I.; Vecchio, C.D.; Xu, L.; Corona, A.; Grandi, N.; Piano, D.; Maccioni, E.; Distinto, S.; Parolin, C.; et al. Sennoside A, derived from the traditional Chinese medicine plant *Rheum* L., is a new dual HIV-1 inhibitor effective on HIV-1 replication. *Phytomedicine* 2016, 23, 1383–1391.

48. Wu, S.F.; Lin, C.K.; Chuang, Y.S.; Chang, F.R.; Tseng, C.K.; Wu, Y.C.; Lee, J.C. Anti-hepatitis C virus activity of 3-hydroxy caruilmigan C from *Swietenia macrophylla* stems. *J. Viral Hepat.* 2012, 19, 364–370.

49. Bachmetov, L.; Tanamy, M.G.; Shapira, A.; Vorobeychik, M.; Giterman-Galam, T.; Sathiyamoorthy, P.; Golan-Goldhirsh, A.; Benhar, I.; Tur-Kaspa, R.; Zemel, R. Suppression of hepatitis C virus by the flavonoid quercetin is mediated by inhibition of NS3 protease activity. *J. Viral Hepat.* 2012, 19, e81–e88.

50. Zhang, N.; Liu, Z.; Han, Q.; Chen, J.; Lv, Y. Xanthohumol enhances antiviral effect of interferon  $\alpha$ -2b against bovine viral diarrhea virus, a surrogate of hepatitis C virus. *Phytomedicine* 2010, 17, 310–316.

51. Dao, T.T.; Nguyen, P.H.; Lee, H.S.; Kim, E.; Park, J.; Lim, S.I.; Oh, W.K. Chalcones as novel influenza A (H1N1) neuraminidase inhibitors from *Glycyrrhiza inflata*. *Bioorg. Med. Chem. Lett.* 2011, 21, 294–298.

52. Yin, Y.; Wunderink, R.G. MERS, SARS and other coronaviruses as causes of pneumonia. *Respirology* 2017, 23, 130–137.

53. Dube, A.; Nicolazzo, J.A.; Larson, I. Chitosan nanoparticles enhance the intestinal absorption of the green tea catechins (+)-catechin and (-)-epigallocatechin gallate. *Eur J Pharm. Sci.* 2010, 9, 219–225.

54. Sindhu, R.K.; Gupta, R.; Wadhera, G.; Kumar, P. Modern Herbal Nanogels: Formulation, Delivery Methods, and Applications. *Gels* 2022, 8, 97.

55. Sims, K.R.; He, B.; Koo, H.; Benoit, D.S.W. Electrostatic Interactions Enable Nanoparticle Delivery of the Flavonoid Myricetin. *ACS Omega* 2020, 28, 12649–12659.

56. Kumari, A.; Kumar, V.; Yadav, S.K. Plant extract synthesized PLA nanoparticles for controlled and sustained release of quercetin: A green approach. *PLoS ONE* 2012, 7, e41230.

57. Fiorani, M.; Accorsi, A.; Cantoni, O. Human Red Blood Cell as a Natural Flavonoid Reservoir. *Free Radic. Res.* 2003, 37, 1331–1338.

58. Zhang, H.; Yang, X.; Zhao, L.; Jiao, Y.; Liu, J.; Zhai, G. In vitro and in vivo study of Baicalin-loaded mixed micelles for oral delivery. *Drug Deliv.* 2016, 23, 1933–1939.

59. Dokania, S.; Joshi, A.K. Self-microemulsifying drug delivery system (SMEDDS)--challenges and road ahead. *Drug Deliv.* 2015, 22, 675–690.

60. Feng, R.; Zhang, Z.; Li, Z.; Huang, G. Preparation and in vitro evaluation of etoposide-loaded PLGA microspheres for pulmonary drug delivery. *DrugDeliv.* 2014, 21, 185–192.

61. Yue, P.F.; Yuan, H.L.; Xie, H.; Xiao, X.H.; Yang, M.; Liao, M.X.; Zhu, W.F.; Cai, P.L. Preparation, characterization, and bioavailability of ursodeoxycholic acid-phospholipid complex in vivo. *Drug Dev. Ind. Pharm.* 2008, 34, 708–718.

62. Al-Sanea, M.M.; Abelyan, N.; Abdelgawad, M.A.; Musa, A.; Ghoneim, M.M.; Al-Warhi, T.; Aljaeed, N.; Alotaibi, O.J.; Alnusaire, T.S.; Abdelwahab, S.F.; et al. Strawberry and ginger silver nanoparticles as potential inhibitors for SARS-CoV-2 assisted by in silico modeling and metabolic profiling. *Antibiotics* 2021, 10, 824.

63. Kurniawan, D.W.; Ikhsanudin, A. Potential of Jamun in nanotechnology perspective as an alternative treatment for COVID-19. *J. Pharm. Sci.* 2020, 7, 1.

64. Yang, R.; Huang, X.; Dou, J.; Zhai, G.; Su, L. Self-microemulsifying drug delivery system for improved oral bioavailability of oleanolic acid: Design and evaluation. *Int. J. Nanomed.* 2013, 8, 2917–2926.

65. Jiang, Y.; Wang, F.; Xu, H.; Liu, H.; Meng, Q.; Liu, W. Development of andrographolide loaded PLGA microspheres: Optimization, characterization and in vitro-in vivo correlation. *Int. J. Pharm.* 2014, 475, 475–484.

66. Yue, P.F.; Yuan, H.L.; Li, X.Y.; Yang, M.; Zhu, W.F. Process optimization, characterization and evaluation in vivo of oxymatrine-phospholipid complex. *Int. J. Pharm.* 2010, 387, 139–146.

67. Fadda, A.M.; Sinico, C.; Lai, F.; Logu, A.D. Liposomal incorporation of artimisia arborescenceL. Essential oil and in vitro antiviral activity. *Eur. J. Pharma. Pharm.* 2005, 59, 161–168.

68. Wang, F.; Zhou, L.; Gu, F. Characterization of anticancer hypocrellin A encapsulated with silica nanoparticles. *J. Therm. Anal. Calorim.* 2010, 102, 69–74.

69. Sun, S.W.; Yeh, P.C. Analysis of rhubarb anthraquinones and bianthrone by microemulsion electrokinetic chromatography. *J. Pharm. Biomed. Anal.* 2005, 36, 995–1001.

70. Natrajan, V.; Madhan, B.; Sehgal, P. Formulation and evalution of quercetin polycaprolactone microsphere for the treatment of Rheumatoid arthritis. *J. Pharm. Sophora Alopecuroides Sci.* 2010, 100, 195–205.

71. Casettari, L.; Gennari, L.; Angelino, D.; Ninfali, P.; Castagnino, E. ORAC of chitosan and its derivatives. *Food Hydrocoll.* 2012, 28, 243–247.

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