Lignin Biopolymer in Biomaterials Synthesis

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Contributor: Francisco Vásquez

Lignin is one of the wood and plant cell wall components that is available in large quantities in nature. Its polyphenolic chemical structure has been of interest for valorization and industrial application studies. Lignin can be obtained from wood by various delignification chemical processes, which give it a structure and specific properties that will depend on the plant species. Due to the versatility and chemical diversity of lignin, the chemical industry has focused on its use as a viable alternative of renewable raw material for the synthesis of new and sustainable biomaterials.

 $Keywords: lignin \ ; \ pulping \ processes \ ; \ lignin \ valorization \ ; \ biocomposites \ ; \ sustainable \ materials \ ; \ biomaterials \ ; \ 3D-processes \ ; \ biomaterials \ ;$

printing; biopolymer

1. Introduction

Lignin is an amorphous biopolymer with a phenolic macromolecular structure, that can be extracted from wood, annual plants, and agricultural residues [1][2]. The lignin content varies according to the lignocellulosic source, plant, species, genetics, and environmental conditions, but in general, its amount is in the range of 20%–30% of the biomass dry weight [3]

Industrial processes are designed to extract lignin from lignocellulosic materials to convert the other components, such as cellulose, into accessible and purified fibers for different purposes. The industrial sector that uses these processes with great economic profit is the pulp and paper, which employing chemical cooking of wood, solubilizes lignin and converts cellulose into a commodity of high commercial value [4]. The production of lignin worldwide reaches 70 million tons per year, which would make its use feasible for several industrial applications [5].

Most large-scale industrial processes that use plant polysaccharides have burned lignin to generate the energy needed for the industrial operations $^{[\underline{6}][\underline{7}]}$. Only 2% of residual lignin has been used for other higher value-added applications $^{[\underline{6}]}$. This underutilization motivates the development of engineer lignin-based products of increased commercial value $^{[\underline{8}]}$. However, valorization of lignin has some difficulties that are mainly attributed to the complexity and recalcitrance of this biopolymer $^{[\underline{7}][\underline{9}]}$ and the high heterogeneity, being challenging to predict a stable behavior in advanced material syntheses

Lignin, having a polyphenolic structure, has properties as antioxidant activity, biodegradability, high thermal stability, reactivity, and adhesive properties $^{[10]}$. Therefore, its use as a versatile chemical would give rise to additional economic benefits for the pulp and paper industry as it can be available in large volumes and at a relatively low cost, compared to phenolic compounds derived from oil $^{[11][12]}$. However, the wood type and the pulping processes are responsible for the final structure of lignin. A lignin structure highly homogenous is favorable to its integration in novel materials $^{[5]}$.

Currently, there are several applications aimed to take advantage of the chemical properties of lignin such as bioplastics, carbon fibers, lignin-based composites as high-performance materials, polymers with adhesive properties for wood panels, hydrogels, polymer for synthesis of polyurethane foams, and as an additive in mixtures to confer emulsifying or protective properties [1][10][11][13]. Otherwise, there is also an interest of lignin conversion in oligomeric or monomeric phenolic compounds, using biotechnological or chemical techniques for depolymerization [4][13][14].

2. Lignin Chemistry and Structure

In the plant cell walls, cellulose microfibrils are organized in defined oriented layers embedded in a hemicellulose and lignin matrix [15]. Within this arrangement in the cell wall, the existence of strong covalent bonds between phenolic compounds and carbohydrates has been suggested [16], where lignin acts as a cementing agent, providing mechanical strength, rigidity, and support for the plant growth. Furthermore, it plays a protective role for polysaccharides, hindering their degradation and acting as a barrier to degradation by bacteria and fungi [2][17][18]. Lignin biosynthesis is part of the phenylpropanoid pathway that also produces secondary metabolites such as tannins, flavonoids, and lignans. The lignin

formation process, its biosynthesis and polymerization have been reviewed in detail by Boerjan et al. [19] and Bonawitz and Chapple [20]. The biosynthesis of lignin occurs in vivo via an enzyme-mediated dehydrogenation polymerization resulting in a cross-linked amorphous material with ether and carbon-carbon linkage types as described by Upton and Kasko [8]. Lignin is mainly made up of three phenylpropanoid precursors, known as monolignols: p-coumaryl alcohol, coniferyl alcohol, and sinapyl alcohol [2][4][11][12] as seen in Figure 1. The main difference between these three monomers is in the number of methoxy groups attached to the phenolic ring. The p-hydroxyphenyl unit (H) has no methoxy group, the guaiacyl unit (G) has one methoxy group, while the syringyl unit (S) has two methoxy groups [2][17][21]. These units are connected by carbon-carbon (5-5, β -5, β -1 y β - β), and ether (β -O-4, α -O-4, 4-O-5) linkages [22][23][24]. The amount and composition of intermonomeric linkage units of lignin varies between different plant taxa, cell types, and cell wall layers [25] [26]. Softwoods contain mainly G-type lignin, hardwoods contain SG-type lignin, and grasses have HGS-type lignin [27][28]. The β -O-4 linkage is the most abundant in native lignin, typically accounts 50% of bonds formed during the biosynthesis reactions, and is one of the most labile bonds, and is readily cleaved to by most pretreatment or depolymerization mechanisms [9]]. Depolymerized lignin fractions are highly reactive under depolymerization reactions, which leads to condensation or repolymerization of lignin [2]. A representative structure of lignin is show in Figure 2.

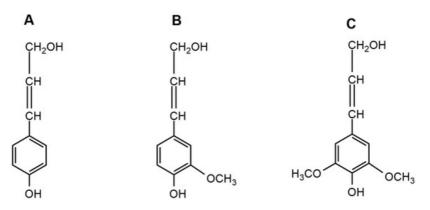


Figure 1. Main monolignoles found in the lignin structure. **(A)** p-coumaryl alcohol; **(B)** coniferyl alcohol; **(C)** sinapyl alcohol.

Figure 2. A representation of lignin structure.

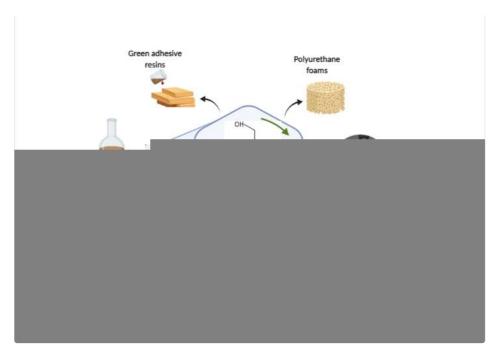
3. Industrial Sources of Lignin

Different methods have been applied to isolate lignin from lignocellulosic material for industrial applications [11]. Due to the different delignification techniques available and the different plant species used, the lignin recovered from these processes presents differences in terms of its physical and chemical properties, since the structure, size, and molecular weight of the resulting lignin fragments are different [3][4][13][17][29]. The most used processes for biomass delignification are kraft, sulfite, and organosolv as seen in Scheme 1. Other less commercial alternatives are pyrolysis, steam explosion, enzymatic hydrolysis, acidolysis, and alkaline hydrolysis [3][12][13][17][29][30][31]. These methods are different in several parameters that influence the extraction yield, lignin functionality, molar mass, and solubility, among other properties according to the temperature, pH, chemical load, and pressure of the system [8][30].

Scheme 1. Main lignin extraction processes.

4. Lignin Applications

The conversion of lignin to high value-added products plays a key role in the economic viability of a biorefinery. The study of lignin conversion mechanisms has established the catalytic routes for the generation of products such as aromatic chemical reagents and biobased materials. Some of these applications are detailed below, emphasizing phenolic adhesive resins, epoxy resins, polyurethane foams, carbon fibers, hydrogels, and 3D printed biocomposites as shown in <u>Scheme 2</u>.



Scheme 2. Summary scheme of the main biomaterials synthesis using lignin.

4.1. Lignin-Based Phenolic Resins

Phenol-formaldehyde (PF) resins are the most widely used adhesive in the wood panel industry. These resins gave wood products excellent adhesive strength, thermal stability, moisture resistance, preventing from delamination or degradation [32][33][34][35]. PF resins can be synthesized under acidic or basic conditions, each being named as novolac or resol resins,

respectively [36]. As phenol is a petrochemical compound, PF resins costs are highly dependent on fluctuations in oil prices. Under this scenario, the search for the partial or total substitution of phenol by a cheaper compound from renewable origin has been the interest of several investigations [32][33][34][35][36][37][38].

The use of lignin for the synthesis of adhesives has been one of the most studied applications due to its polyphenolic structure and large number of available hydroxyl groups that provided reactivity and adhesive properties [34][35][32][38][39][40]. Lignin-based phenolic resins have been studied for application in plywood, medium density fiberboard (MDF), particle board (PB), and oriented strand board (OSB) [32][33]. However, lignin has been shown to have low reactivity towards formaldehyde compared to phenol and only can be used to partially replace phenol in adhesive formulations. Çetin and Özmen [32] were able to synthesize lignin-phenol-formaldehyde (LPF) resin with 20–30% of organosolv lignin replacing phenol, obtaining viscosities of 200–250 cP and curing temperature of 150 °C, the desired range for a resol-type resin. Similar results were obtained by Wang et al. [37] that replaced between 25–75% of phenol by softwood organosolv lignin in a resol type resin at 150–175 °C curing temperature and thermal stability below 60%. Zhang et al. [33] also prepared LPF resins substituting from 10% to 70% of phenol for lignin from biorefinery residues and optimal results were achieved when 50% substitution was used.

On the other hand, some investigations indicate the higher percentages of substitution could be reached only after a chemical modification of lignin, as is hydroxymethylation with glyoxal in an alkaline medium [41]. In addition, demethylation and phenolation were identified as strategies to increase lignin reactivity towards formaldehyde [1][21]. Kalami et al. [35] showed that a total replacement of phenol by lignin in PF resol type resins is possible using lignin from corn stover pretreated with dilute acid and extracted with alkali. The dry mechanical resistance and curing temperature of the resins with total replacement were 3.4 MPa and 203 °C, comparable to 3.6 MPa and 193 °C of the commercial PF resin, respectively.

Formaldehyde-based wood adhesives such as urea-formaldehyde and phenol-formaldehyde are the resins mostly used in the production of wood boards, such as plywood and particle boards $\frac{[42][43]}{}$. However, there are concerns regarding formaldehyde use since it causes skin irritation and respiratory discomfort. Prolonged exposure may result in spontaneous abortion, memory injury, nasopharyngeal cancer, and leukemia. High formaldehyde prices also have a negative impact on the wood board manufacturing costs [42][43][44]. The development of formaldehyde-free adhesives using lignin would be also of great benefit to the wood-board industry and human health. Liu and Li [45] developed a formaldehyde-free adhesive using demethylated Kraft lignin (DKL) and polyethyleneimine (PEI), where it was proposed that the hydroxyl groups of DKL can be oxidized to guinones during the curing process generating covalent links with PEI. The synthesized resins were optimally cured at 120 °C within 5 min and 1:1 ratio of DKL/PEI, resulting in a shear strength value on dry measurement of 7 MPa versus 6 MPa when compared with a commercial phenol-formaldehyde resin. Furthermore, a molecular weight between 70,000 to 750,000 g/mol PEIs could have a small effect on the strength and water resistance in the lap-shear specimens resulting in values within a range of 4.5 and 7 MPa. Recently, Gong et al. [44] reported a highperformance lignin-based adhesive without formaldehyde using acetonitrile extracting lignin, modified by phenol (PAEL) and polyvinylpyrrolidone (K30). The optimal conditions for this preparation were using a mole ratio of lignin to phenol 1:2, mole ratio of PAEL to K30 1:0.4, at 80 °C for 3 h. Mechanical tests of shear and water resistance in plywood (after soaking for 24 h), where 1.70 and 1.02 MPa, respectively, complying standards for plywood properties. Lately, Younesi-Kordkheili and Pizzi [46] have described a method to synthesis a resin using urea, lignin, and glyoxal for plywood panels. The resin of urea-lignin-glyoxal (ULG), a phenolic resin, was improved by adding small epoxy resin quantities (1, 3, and 5% w/w). The results described in this work show that the plywood bonded with this resin presented a better tensile shear strength than the control resin without epoxy content. The resin that contained 5% of epoxy resin exhibited the highest panel dry strength with 1.7 vs. 0.43 MPa observed in the plywood bonded with the control resin. The resin with 5% of epoxy content presented the highest panel wet strength with 1.29 vs. 0.2 MPa in the plywood bonded with the control resin. The addition of epoxy in the ULG resin decreased water adsorption from 15% to 6% [46].

4.2. Lignin-Based Epoxy Resins

Epoxy resins are used in different applications as they have excellent properties of hardness and dimensional stability; therefore, they are used in fiber-reinforced composite materials, adhesives, additive in lubricating oils, polyurethane foams, surface coatings, among others $\frac{[36][47][48][49]}{[48][49]}$. Epoxy resins are thermoset materials, where epoxy structures are reactive towards the other chemical forms, mainly hydroxyl groups $\frac{[36]}{[48]}$. Epoxy resins are synthesized using bisphenol A (BPA), a compound obtained from the petrochemical industry and also used to produce polycarbonates and plastics BPA with a worldwide production of approximately 6.8 million tons $\frac{[50][51][52][53]}{[53]}$. The synthesis of this resin is based on the reaction between bisphenol A (BPA) and epichlorohydrin, giving rise to bisphenol A diglycidyl ether (DGEBA), the main precursor of epoxy resins $\frac{[36][48][54]}{[50]}$. Unfortunately, according to authors such as Vandenberg et al. $\frac{[50]}{[50]}$, Aouf et al. $\frac{[53]}{[53]}$, and

Huang et al. [51], BPA can be a highly harmful compound for the environment and human health, which has been proven as carcinogen. When absorbed by the organism, BPA could increase the risk of obesity, diabetes, and cardiac disorders [53]. Therefore, replacing BPA with compounds of natural origin could become a desirable and viable alternative.

El Mansouri et al. [41] carried out chemical epoxidation with epichlorohydrin and modified Kraft lignin by methylolation and glyoxalation in replacement of BPA. It was shown that using methylolated lignin, higher epoxidation indexes (3.85%) were achieved than with glyoxylated (3.1%) or unmodified Kraft lignin (2.45%) with 82% epoxidation yield using lignin/NaOH ratio 1/3, at 70 °C for 3 h of reaction. Jablonskis et al. [55] used softwood Kraft lignin with epichlorohydrin and KOH as alkaline catalyst in dimethyl sulfoxide solution. Up to 10% of a BPA was replaced by lignin, resulting in a resin with high thermal stability, with 50% weight degradation at 390 °C, similar value to that obtained with a reference BPA-based epoxy resin. In addition, an increase in the Young's modulus was observed in epoxidized resins cured with lignin, resulting in a value of 3.36 GPa using 10% of lignin versus 2.91 GPa with BPA-based epoxy resin.

Aouf et al. [48] proposed a novel chemo-enzymatic mechanism using immobilized enzyme lipase B (Novozyme 435), hydrogen peroxide, and caprylic acid to carry out the epoxidation of vanillic acid hydroxyl groups in replacement of BPA, achieving conversion efficiency of 87% after 48 h at 40 °C. This catalytic method demonstrated a good performance in the epoxidation of the allylic double bonds compared to with the chemical mechanism using the hazardous peracid mCPBA, where the highest epoxidation value reached was 75% using vanillic acid as raw material. Kraft and organosolv lignins were also used for chemo-enzymatic epoxidation, where the first step was the allylation of the available hydroxyl groups of lignin, followed by the enzymatic reaction with immobilized lipase B (Novozyme 435) and a fatty acid to produce oxiranes rings (epoxy groups) [49]. This study reports conversion yields to epoxy groups of 90% and 55% for organosolv and Kraft lignin, respectively. Regarding the residual enzymatic activity, it was determined that it is mostly reversible when Kraft lignin is used as starting material, however, the exposure of the enzyme to concentrations of 6 M of hydrogen peroxide during 12 h reaction would cause the loss of enzymatic activity through denaturation. These studies reinforce the potential use of low molar mass lignin to produce epoxy resins, where it could even play an important role in the curing process as hardening agent [12][49].

4.3. Lignin-Based Polyurethane Foams

Polyurethane or polyurethane foams are the most versatile type of materials used, with applications as an interior material for transport media, roof and floor insulation, medical use material, as co-reactive adhesive, use in thermoplastics, as sealants of flats, among others. These foams are synthesized through the reaction between isocyanate and hydroxyl groups. It has been proposed that due to its phenolic nature, lignin could provide the necessary hydroxyl groups and replace polyols conventionally used in the industry of these foams [12][56][36][57]. Pan and Saddler [58] replaced the commercial polyol (Voranol 270) with organosolv lignin from hardwood and Kraft lignin from softwood. Authors carried out preparations to replace the use of Voranol 270 from 19% to 23% (*w/w*) with Kraft lignin. It was shown that lignin effectively generated crosslinking to obtain rigid polyurethane foams, however, the densities obtained were higher than those obtained with commercial foams. The use of 19% (*w/w*) of Kraft lignin in the formulations resulted in the desired decrease in the density of the product (100 kg/m³). The density of the foam increased with the higher proportion of Kraft lignin, generating the formation of unwanted granules in the foaming process [58].

Various authors have also suggested that lignin should be modified for use in foams, with the aim of improving its reactivity and solubility in the synthesis media. Li and Ragauskas [59] described a method for chemical modification of pine Kraft lignin to obtain a liquid polyol by means of oxypropylation with propylene oxide (PO) for synthesis of rigid polyurethane foam, where a total OH concentration of 6.9 mmol/g lignin was obtained compared to a concentration of 5.62 mmol/g lignin without oxypropylation. The oxypropylation of lignin allowed incorporating a higher lignin content in the formulation of the foams, even only using the polyols from lignin in reaction with the MDI di-isocyanate polymer. The mechanical properties of the rigid PU foams obtained using lignin improved considerably in strength index (0.14 MPa) and compression modulus (3.41 MPa). Ligno-polyurethane materials have been developed by de Oliveira et al. [60], using mixtures of oxypropylated lignosulfonates and castor oil rich in hydroxyl groups and where it was hypothesized that this modification would increase its reactivity towards methylene diphenyl isocyanate (MDI). The synthesized lignopolyurethane product presented high impact resistance and modulus of elasticity compared to composites prepared with lignosulfonates without oxypropyl [60]. In the same way, authors have prepared foams using different molar proportions of NCO:OH, and different fractions of lignins for the production of polyols from lignin modified by esterification with chlorinated fatty acid in a solvent-free system [1][10]. Increasing the use of lignin in foam preparations has been shown to increase the tensile strength and deformability of synthesized polyurethanes. This confirms the importance of the chemical and structural properties of lignin, such as available hydroxyl groups, molar mass, reactivity for the optimization of the stoichiometry in polyurethane foam preparations [10][60][61].

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4.4. Lignin-Based Carbon Fibers

Carbon fibers (CFs) are one of the most advanced engineering products currently produced. CFs are composite products that are used in the transportation industry, manufacture of sports equipment, and construction $\frac{[1][62]}{}$. The carbon fibers exhibit a unique combination of properties, such as lightness, high strength, flexibility, and resistance to fatigue, which makes them suitable for a wide range of applications.

Polyacrylonitrile (PAN) and regenerated cellulose are the primary precursors of current CFs production. Its manufacture involves melting or wet spinning, oxidative stabilization, and carbonization under an inert atmosphere, followed by surface and dimension treatment. Due to its low cost, lignin has a significant interest as a raw material for CFs production [1][21][62]. Different authors have been able to synthesize products based mainly on mixtures of Kraft lignin and polyethylene oxide (PO) through thermal spinning and subsequent carbonization [12][62][63]. Kadla et al. [62] manufactured carbon fibers with PO and Kraft lignin where the tensile strength obtained was 300-450 MPa and the modulus of elasticity 30-60 GPa, properties that where similar to the conventional carbon fibers. Liu et al. [64] propose the synthesis of the carbon fiber precursor prepared from lignosulfonate, acrylonitrile, and itaconic acid, called LS-AN-IA terpolymer. The thermal stability of the precursor fibers could lead to a decrease in the temperature and reaction times of the CFs synthesis process. The fibers obtained from the precursor LS-AN-IA resulted in good tensile strength (1.74 GPa) and tensile modulus (210 GPa). Moreover, different proportions of organosolv lignin were tested by Hosseinaei et al. [63] to improve the spinning process, stabilization, and the properties of the CFs, obtaining thermal stabilization even with the highest proportions of lignin (50%). However, the glass transition temperatures were decreased using higher proportions than 50% of lignin, starting at 127 °C (50%) until reaching 122 °C (85%). Liu et al. [65] prepared mixtures of carbon fibers containing alkaline lignin and PAN (1:2 ratio), and carbon nanotubes through electric spinning. The carbonization process was carried out at 1100 °C. The fibers obtained had a resistance of 1720 MPa and a modulus of elasticity of 230 GPa, characteristics comparable to those that use PAN (1600 MPa and 223 GPa, respectively). In general, the use of lignin for CFs preparations would have the advantages of reducing production costs, synthesis times, and negative impacts on the environment. However, there is a need to obtain high-purity and high-quality lignin to reach high-performance CF results to compete with products synthesized that using PAN [12].

4.5. Lignin-Based Hydrogels

Polymeric materials as hydrogels have potential application in environmental, biological, and pharmaceutical fields [29][61]. Hydrogels are polymeric structures that have the capacity to swell up to a several times their weight without dissolving. These structural networks have hydrophilic groups to absorb water giving the form of a hydrogel. The most widely methods used for the synthesis of hydrogels are free radical polymerization, crosslinking covalent, and physical crosslinking. The polymers mainly used in hydrogels include poly (2-polyhydroxyethyl methacrylate) (PHEMA), polyethyl methacrylate (PEMA), polyacrylamide (PAAm), polymethacrylic acid (PMA), polyacrylic acid (PAA), polyglucoesiletil methacrylate (PGEMA), and polyhydroxypropyl methacrylamide (PHPMA) [21][29].

The interest in using lignin in hydrogels is mainly based on the biopolymer advantages, as biodegradability, low toxicity, environmentally friendly, and suitable for enzymatic degradation [29]. Some authors have carried out the application of organosolv lignin in the synthesis of hydrogels, based on the polymerization with N-isopropylacrylamide in the presence of N,N'-methylenebisacrylamide, as well as the use of Kraft lignin for the incorporation of diazobenzene groups [12].

Lignin-based hydrogels have also been studied for the supply of polyphenolic molecules. Ciolacu et al. [66] proposed superabsorbent hydrogels based on cellulose/lignin mixtures. The synthesis process consisted of dissolving cellulose in an alkaline medium and then incorporating lignin, according to the crosslinking process using epichlorohydrin. Through DSC analysis, the crosslinking was determined between lignin and cellulose using a formulation of 50% of each component at 80 °C. These synthesized hydrogels would have potential applications in the biomedical area for the supply of polyphenolic structures [60][66]. Another application of lignin-based hydrogels was developed for agricultural use where lignin-based alginate hydrogels were studied for the controlled release of the insecticide azadirachtin [67]. The modified lignin hydrogels had a trapping efficiency of the insecticide of 99.5%. Adding lignin to the hydrogel protects the insecticide from photodegradation, helping to maintain it active for a higher time [67]. In the field of water resources, the work developed by Mazloom et al. [68] describe a solution method to reduce the impact of water scarcity on agricultural soils, using a superabsorbent and biodegradable hydrogel based on alkali lignin to retain moisture from irrigation or rain, releasing water depending on the water demand of the soil and crops. In the results reported by Mazloom et al. [68], it was shown that maize crop plants were taller, had a higher amount of phosphorus, and a higher biomass when they were grown in soils with lignin hydrogels, compared to synthetic hydrogels and no hydrogels. Additionally, it was suggested that the addition of alkaline lignin to the soil of some crops could promote the activity of the roots due to the stimulation of specific phytohormones in plants, improving growth and elongation of plant stems [68][69].

4.6. Lignin-Based Composites and 3D Printing

There has been a growing interest in the incorporation of lignin in thermoplastic materials, that can be used for the formulation of composites or biocomposites by 3D-printing in combination with other biopolymers such as polylactic acid (PLA), polyhydroxybutyrate (PHB), acrylonitrile butadiene styrene (ABS), and polyethylene (PE), with the aim to provide biodegradable properties, antimicrobial capacity, UV radiation stabilizer, and antioxidant activity properties [5][6][70][71][72][73]. Antioxidant materials can contribute to reduce the specific concentration of free radicals and active oxygen species, and lignin can provide some of these antioxidant activity properties. Domínguez-Robles et al. [74] developed composites made from PLA and lignin and demonstrated that the manufactured materials presented antioxidant activity achieving a decrease in the DPPH (2,2-diphenyl-1-pycrylhydrazyl) compound of up to 80% after 5 h of application. Recent research reported the use of modified or unmodified lignin as reinforcing material in thermoplastic composites. Gordobil et al. [70] used acetylated and non-acetylated Kraft lignin for the elaboration of composites with PLA through extrusion of the components at 200 °C. The presence of lignin in the composites increased the thermal stability of PLA and its hydrophobicity measured by contact angle. It was determined that the presence of low lignin contents (<10%) did not affect the mechanical properties of the composites, rather there was an increase in the elongation index at break using the two types of lignin [70].

The FDM method (fused deposition modeling) has been widely used for the development of additive manufacturing for 3D printing $^{[74]}$. Polylactic acid (PLA) is one of the biopolymers used for 3D printing, which is obtained from natural sources, being biodegradable and biocompatible $^{[5][6][73]}$. However, the use of PLA has some disadvantages such as sensitivity to humidity, low glass transition temperature, low mechanical performance at elongation. Incorporate other biopolymers, like lignin, into the formulation of filaments for 3D printing emerges as an excellent alternative. Results reported by Tanase-Opedal et al. $^{[6]}$ described a method for obtaining filaments based on PLA and softwood lignin from soda pulping process in proportions between 20–40%. Authors described a nucleating phenomenon of lignin in the mixture with PLA, which facilitated the crystallization of PLA in the elaboration of the filaments. In mechanical tests, PLA/lignin biocomposites were shown to have a similar tensile elastic modulus index (Et = 2695 MPa) vs. biocomposites that only had PLA (Et = 2890 MPa) using a printing at 215 °C. Although the integration of lignin into the biocomposites could reduce the tensile strength

modulus, this mechanical property can be improved optimizing the 3D printing temperatures. In the biocomposites based on PLA/lignin, an important antioxidant activity was observed, increased by 50% in the composite made with 40% of lignin. Furthermore, the developed PLA/lignin biocomposites exhibited good extrudability and flowability without lignin agglomerations ^[G].

Nguyen et al. ^[72] developed an additive manufacturing system using organosolv and Kraft lignin in different mixtures of acrylonitrile-butadiene-styrene (ABS) polymer and nylon. These mixtures were used to elaborate materials by 3D printing using the FDM method, where a proportion of lignin between 40–60% in weight was employed. The resulting composite with the best performance and with promising properties was obtained in a mixture of nylon 12 and hardwood organosolv lignin. This composite had high stiffness and tensile strength at room temperature. Vaidya et al. ^[71] prepared composites based on polyhydroxybuirate (PHB) softwood lignin as a non-reactive filler without any derivatization or chemical modification. The biocomposite was developed by extrusion to form filaments for 3D-printing by FDM. Biocomposites with PHB and 50% lignin presented an increased contact angle to 72.8° vs. 46.4° in the pure PHB biocomposite, improving the hidrofobicity of the biocomsites. Besides, the incorporation from 20% lignin as filler in the filament could benefit the printability and reduction of the shrinkage during the filament impression.

Mimini et al. ^[5] studied the compatibility of three types of lignins (organosolv, Kraft, and lignosulfonate) in mixtures with PLA to produce biocomposites by 3D-printing. The results showed that the mixtures between PLA and organosolv lignin exhibited better flexural resistance than those made with Kraft lignin or lignosulfonate with a maximum of 100 MPa using 5% organosolv lignin, a value almost identical to that obtained only with PLA. In addition, the mixture between PLA/organosolv lignin had better thermal resistance compared with the other lignins determined by thermogravimetric analysis (TGA). The high compatibility between organosolv lignin and PLA could be attributed to its low molecular weight (Mw 1200 g/mol vs. Mw 4000 g/mol in the other lignins) being a suitable compound to be incorporated in extrusion and 3D printing systems ^[5].

References

- 1. Laurichesse, S.; Avérous, L. Chemical Modification of Lignins: Towards Biobased Polymers. Prog. Polym. Sci. 2014, 3 9, 1266–1290.
- 2. Abdelaziz, O.Y.; Brink, D.P.; Prothmann, J.; Ravi, K.; Sun, M.; García-Hidalgo, J.; Sandahl, M.; Hulteberg, C.P.; Turner, C.; Lidén, G.; et al. Biological Valorization of Low Molecular Weight Lignin. Biotechnol. Adv. 2016, 34, 1318–1346.
- 3. Lange, H.; Decina, S.; Crestini, C. Oxidative Upgrade of Lignin—Recent Routes Reviewed. Eur. Polym. J. 2013, 49, 11 51–1173.
- 4. Gall, D.L.; Ralph, J.; Donohue, T.J.; Noguera, D.R. Biochemical Transformation of Lignin for Deriving Valued Commodit ies from Lignocellulose. Curr. Opin. Biotechnol. 2017, 45, 120–126.
- 5. Mimini, V.; Sykacek, E.; Hashim, S.N.A.; Holzweber, J.; Hettegger, H.; Fackler, K.; Potthast, A.; Mundigler, N.; Rosena u, T. Compatibility of Kraft Lignin, Organosolv Lignin and Lignosulfonate with PLA in 3D Printing. J. Wood Chem. Techn ol. 2019, 39, 14–30.
- Tanase-Opedal, M.; Espinosa, E.; Rodríguez, A.; Chinga-Carrasco, G. Lignin: A Biopolymer from Forestry Biomass for Biocomposites and 3D Printing. Materials 2019, 12, 3006.
- 7. Wang, H.; Pu, Y.; Ragauskas, A.; Yang, B. From Lignin to Valuable Products—Strategies, Challenges, and Prospects. Bioresour. Technol. 2019, 271, 449–461.
- 8. Upton, B.M.; Kasko, A.M. Strategies for the Conversion of Lignin to High-Value Polymeric Materials: Review and Persp ective. Chem. Rev. 2016, 116, 2275–2306.
- 9. Constant, S.; Wienk, H.L.J.; Frissen, A.E.; de Peinder, P.; Boelens, R.; van Es, D.S.; Grisel, R.J.H.; Weckhuysen, B.M.; Huijgen, W.J.J.; Gosselink, R.J.A.; et al. New Insights into the Structure and Composition of Technical Lignins: A Comparative Characterization Study. Green Chem. 2016, 18, 2651–2665.
- 10. Naseem, A.; Tabasum, S.; Zia, K.M.; Zuber, M.; Ali, M.; Noreen, A. Lignin-Derivatives Based Polymers, Blends and Composites: A Review. Int. J. Biol. Macromol. 2016, 93, 296–313.
- 11. Ragauskas, A.J.; Beckham, G.T.; Biddy, M.J.; Chandra, R.; Chen, F.; Davis, M.F.; Davison, B.H.; Dixon, R.A.; Gilna, P.; Keller, M.; et al. Lignin Valorization: Improving Lignin Processing in the Biorefinery. Science 2014, 344.
- 12. Kun, D.; Pukánszky, B. Polymer/Lignin Blends: Interactions, Properties, Applications. Eur. Polym. J. 2017, 93, 618-641.
- 13. Zinovyev, G.; Sumerskii, I.; Korntner, P.; Sulaeva, I.; Rosenau, T.; Potthast, A. Molar Mass-Dependent Profiles of Functi onal Groups and Carbohydrates in Kraft Lignin. J. Wood Chem. Technol. 2017, 37, 171–183.

- 14. Hodásová, L.; Jablonský, M.; Škulcová, A.; Ház, A. Lignin, Potential Products and Their Market Value. Wood Res. 201 5, 60, 973–986.
- 15. Gauss, C.; Pickering, K.L.; Muthe, L.P. The Use of Cellulose in Bio-derived Formulations for 3D/4D printing: A review. C ompos. Part C Open Access 2021, 4, 100113.
- 16. Limayem, A.; Ricke, S.C. Lignocellulosic Biomass for Bioethanol Production: Current Perspectives, Potential Issues and Future Prospects. Prog. Energy Combust. Sci. 2012, 38, 449–467.
- 17. Doherty, W.O.S.; Mousavioun, P.; Fellows, C.M. Value-Adding to Cellulosic Ethanol: Lignin Polymers. Ind. Crop. Prod. 2 011, 33, 259–276.
- 18. Xu, C.; Arancon, R.A.D.; Labidi, J.; Luque, R. Lignin Depolymerisation Strategies: Towards Valuable Chemicals and Fu els. Chem. Soc. Rev. 2014, 43, 7485–7500.
- 19. Boerjan, W.; Ralph, J.; Baucher, M. Lignin Biosynthesis. Annu. Rev. Plant Biol. 2003, 54, 519-546.
- 20. Bonawitz, N.D.; Chapple, C. The Genetics of Lignin Biosynthesis: Connecting Genotype to Phenotype. Annu. Rev. Gen et. 2010, 44, 337–363.
- 21. Peretti, S.; Barton, R.; Mendonça, R. Lignin as Feedstock for Fibers and Chemicals. In Commercializing Biobased Products: Opportunities, Challenges, Benefits, and Risks; Royal Society of Chemistry: Cambridge, UK, 2016; pp. 132–165.
- 22. Ralph, J.; Lundquist, K.; Brunow, G.; Lu, F.; Kim, H.; Schatz, P.F.; Marita, J.M.; Hatfield, R.D.; Ralph, S.A.; Christensen, J.H.; et al. Lignins: Natural Polymers from Oxidative Coupling of 4-Hydroxyphenyl- Propanoids. Phytochem. Rev. 2004, 3, 29–60.
- 23. Ralph, J. Hydroxycinnamates in Lignification. Phytochem. Rev. 2010, 9, 65–83.
- 24. Abdel-Hamid, A.M.; Solbiati, J.O.; Cann, I.K.O. Insights into Lignin Degradation and Its Potential Industrial Applications. In Advances in Applied Microbiology; Academic Press: Cambridge, MA, USA, 2013; Volume 82, pp. 1–28.
- 25. Rencoret, J.; Gutiérrez, A.; Nieto, L.; Jiménez-Barbero, J.; Faulds, C.B.; Kim, H.; Ralph, J.; Martínez, Á.T.; del Río, J.C. Lignin Composition and Structure in Young versus Adult Eucalyptus globulus Plants on JSTOR. Plant. Physiol. 2011, 1 55, 667–682.
- 26. Saito, K.; Horikawa, Y.; Sugiyama, J.; Watanabe, T.; Kobayashi, Y.; Takabe, K. Effect of Thermochemical Pretreatment on Lignin Alteration and Cell Wall Microstructural Degradation in Eucalyptus globulus: Comparison of Acid, Alkali, and Water Pretreatments. J. Wood Sci. 2016, 62, 276–284.
- 27. Calvo-Flores, F.G.; Dobado, J.A. Lignin as Renewable Raw Material. ChemSusChem 2010, 3, 1227-1235.
- 28. Li, C.; Zhao, X.; Wang, A.; Huber, G.W.; Zhang, T. Catalytic Transformation of Lignin for the Production of Chemicals an d Fuels. Chem. Rev. 2015, 115, 11559–11624.
- 29. Thakur, V.K.; Thakur, M.K. Recent Advances in Green Hydrogels from Lignin: A Review. Int. J. Biol. Macromol. 2015, 7 2, 834–847.
- 30. Zakzeski, J.; Bruijnincx, P.C.A.; Jongerius, A.L.; Weckhuysen, B.M. The Catalytic Valorization of Lignin for the Productio n of Renewable Chemicals. Chem. Rev. 2010, 110, 3552–3599.
- 31. Galkin, M.V.; Samec, J.S.M. Lignin Valorization through Catalytic Lignocellulose Fractionation: A Fundamental Platform for the Future Biorefinery. ChemSusChem 2016, 9, 1544–1558.
- 32. Çetin, N.S.; Özmen, N. Use of Organosolv Lignin in Phenol-Formaldehyde Resins for Particleboard Production: II. Parti cleboard Production and Properties. Int. J. Adhes. Adhes. 2002, 22, 481–486.
- 33. Zhang, W.; Ma, Y.; Wang, C.; Li, S.; Zhang, M.; Chu, F. Preparation and Properties of Lignin-Phenol-Formaldehyde Res ins Based on Different Biorefinery Residues of Agricultural Biomass. Ind. Crop. Prod. 2013, 43, 326–333.
- 34. Qiao, W.; Li, S.; Guo, G.; Han, S.; Ren, S.; Ma, Y. Synthesis and Characterization of Phenol-Formaldehyde Resin Usin g Enzymatic Hydrolysis Lignin. J. Ind. Eng. Chem. 2015, 21, 1417–1422.
- 35. Kalami, S.; Arefmanesh, M.; Master, E.; Nejad, M. Replacing 100% of Phenol in Phenolic Adhesive Formulations with Lignin. J. Appl. Polym. Sci. 2017, 134, 45124.
- 36. Raquez, J.M.; Deléglise, M.; Lacrampe, M.F.; Krawczak, P. Thermosetting (Bio)Materials Derived from Renewable Resources: A Critical Review. Prog. Polym. Sci. 2010, 35, 487–509.
- 37. Wang, M.; Leitch, M.; Xu, C. Synthesis of Phenol-Formaldehyde Resol Resins Using Organosolv Pine Lignins. Eur. Pol ym. J. 2009, 45, 3380–3388.
- 38. Jing, Z.; Lihong, H.; Bingchuan, L.; Caiying, B.; Puyou, J.; Yonghong, Z. Preparation and Characterization of Novolac P henol-Formaldehyde Resins with Enzymatic Hydrolysis Lignin. J. Taiwan Inst. Chem. Eng. 2015, 54, 178–182.

- 39. El Mansouri, N.E.E.; Salvadó, J. Structural Characterization of Technical Lignins for the Production of Adhesives: Applic ation to Lignosulfonate, Kraft, Soda-Anthraquinone, Organosolv and Ethanol Process Lignins. Ind. Crop. Prod. 2006, 2 4, 8–16.
- 40. Pang, B.; Yang, S.; Fang, W.; Yuan, T.Q.; Argyropoulos, D.S.; Sun, R.C. Structure-Property Relationships for Technical Lignins for the Production of Lignin-Phenol-Formaldehyde Resins. Ind. Crop. Prod. 2017, 108, 316–326.
- 41. El Mansouri, N.E.; Yuan, Q.; Huang, F. Synthesis and Characterization of Kraft Lignin-Based Epoxy Resins. BioResour ces 2011, 6, 2492–2503.
- 42. Li, K.; Geng, X. Formaldehyde-Free Wood Adhesives from Decayed Wood. Macromol. Rapid Commun. 2005, 26, 529–532.
- 43. Jang, Y.; Huang, J.; Li, K. A New Formaldehyde-Free Wood Adhesive from Renewable Materials. Int. J. Adhes. Adhes. 2011, 31, 754–759.
- 44. Gong, X.; Liu, T.; Yu, S.; Meng, Y.; Lu, J.; Cheng, Y.; Wang, H. The Preparation and Performance of a Novel Lignin-Bas ed Adhesive without Formaldehyde. Ind. Crop. Prod. 2020, 153, 112593.
- 45. Liu, Y.; Li, K. Preparation and Characterization of Demethylated Lignin-Polyethylenimine Adhesives. J. Adhes. 2006, 8 2, 593–605.
- 46. Younesi-Kordkheili, H.; Pizzi, A. Improving the Properties of Urea-Lignin-Glyoxal Resin as a Wood Adhesive by Small A ddition of Epoxy. Int. J. Adhes. Adhes. 2020, 102, 102681.
- 47. Törnvall, U.; Orellana-Coca, C.; Hatti-Kaul, R.; Adlercreutz, D. Stability of Immobilized Candida antarctica Lipase B duri ng Chemo-Enzymatic Epoxidation of Fatty Acids. Enzym. Microb. Technol. 2007, 40, 447–451.
- 48. Aouf, C.; Lecomte, J.; Villeneuve, P.; Dubreucq, E.; Fulcrand, H. Chemo-Enzymatic Functionalization of Gallic and Vanil lic Acids: Synthesis of Bio-Based Epoxy Resins Prepolymers. Green Chem. 2012, 14, 2328–2336.
- 49. Vásquez-Garay, F.; Mendonça, R.T.; Peretti, S.W. Chemoenzymatic Lignin Valorization: Production of Epoxidized Pre-P olymers Using Candida Antarctica Lipase B. Enzym. Microb. Technol. 2018, 112, 6–13.
- 50. Vandenberg, L.N.; Hauser, R.; Marcus, M.; Olea, N.; Welshons, W.V. Human Exposure to Bisphenol A (BPA). Reprod. T oxicol. 2007, 24, 139–177.
- 51. Huang, Y.Q.; Wong, C.K.C.; Zheng, J.S.; Bouwman, H.; Barra, R.; Wahlström, B.; Neretin, L.; Wong, M.H. Bisphenol A (BPA) in China: A Review of Sources, Environmental Levels, and Potential Human Health Impacts. Environ. Int. 2012, 4 2, 91–99.
- 52. Aouf, C.; Nouailhas, H.; Fache, M.; Caillol, S.; Boutevin, B.; Fulcrand, H. Multi-Functionalization of Gallic Acid. Synthesis of a Novel Bio-Based Epoxy Resin. Eur. Polym. J. 2013, 49, 1185–1195.
- 53. Maćczak, A.; Bukowska, B.; Michalowicz, J. Comparative Study of the Effect of BPA and Its Selected Analogues on He moglobin Oxidation, Morphological Alterations and Hemolytic Changes in Human Erythrocytes. Comp. Biochem. Physi ol. Part C Toxicol. Pharmacol. 2015, 176, 62–70.
- 54. Chua, S.C.; Xu, X.; Guo, Z. Emerging Sustainable Technology for Epoxidation Directed toward Plant Oil-Based Plastici zers. Process. Biochem. 2012, 47, 1439–1451.
- 55. Jablonskis, A.; Arshanitsa, A.; Arnautov, A.; Telysheva, G.; Evtuguin, D. Evaluation of Ligno BoostTM Softwood Kraft Li gnin Epoxidation as an Approach for Its Application in Cured Epoxy Resins. Ind. Crop. Prod. 2018, 112, 225–235.
- 56. Mahmood, N.; Yuan, Z.; Schmidt, J.; Xu, C. Depolymerization of Lignins and Their Applications for the Preparation of P olyols and Rigid Polyurethane Foams: A Review. Renew. Sustain. Energy Rev. 2016, 60, 317–329.
- 57. Gómez-Fernández, S.; Ugarte, L.; Calvo-Correas, T.; Peña-Rodríguez, C.; Corcuera, M.A.; Eceiza, A. Properties of Fle xible Polyurethane Foams Containing Isocyanate Functionalized Kraft Lignin. Ind. Crop. Prod. 2017, 100, 51–64.
- 58. Pan, X.; Saddler, J.N. Effect of Replacing Polyol by Organosolv and Kraft Lignin on the Property and Structure of Rigid Polyurethane Foam. Biotechnol. Biofuels 2013, 6, 12.
- 59. Li, Y.; Ragauskas, A.J. Kraft Lignin-Based Rigid Polyurethane Foam. J. Wood Chem. Technol. 2012, 32, 210-224.
- 60. de Oliveira, F.; Ramires, E.C.; Frollini, E.; Belgacem, M.N. Lignopolyurethanic Materials Based on Oxypropylated Sodiu m Lignosulfonate and Castor Oil Blends. Ind. Crop. Prod. 2015, 72, 77–86.
- 61. Figueiredo, P.; Lintinen, K.; Hirvonen, J.T.; Kostiainen, M.A.; Santos, H.A. Properties and Chemical Modifications of Lignin: Towards Lignin-Based Nanomaterials for Biomedical Applications. Prog. Mater. Sci. 2018, 93, 233–269.
- 62. Kadla, J.F.; Kubo, S.; Venditti, R.A.; Gilbert, R.D.; Compere, A.L.; Griffith, W. Lignin-Based Carbon Fibers for Composit e Fiber Applications. Carbon 2002, 40, 2913–2920.

- 63. Hosseinaei, O.; Harper, D.P.; Bozell, J.J.; Rials, T.G. Improving Processing and Performance of Pure Lignin Carbon Fibers through Hardwood and Herbaceous Lignin Blends. Int. J. Mol. Sci. 2017, 18, 1410.
- 64. Liu, D.; Ouyang, Q.; Jiang, X.; Ma, H.; Chen, Y.; He, L. Thermal Properties and Thermal Stabilization of Lignosulfonate-Acrylonitrile-Itaconic Acid Terpolymer for Preparation of Carbon Fiber. Polym. Degrad. Stab. 2018, 150, 57–66.
- 65. Liu, H.C.; Chien, A.T.; Newcomb, B.A.; Liu, Y.; Kumar, S. Processing, Structure, and Properties of Lignin- and CNT-Inco rporated Polyacrylonitrile-Based Carbon Fibers. ACS Sustain. Chem. Eng. 2015, 3, 1943–1954.
- 66. Ciolacu, D.; Oprea, A.M.; Anghel, N.; Cazacu, G.; Cazacu, M. New Cellulose–Lignin Hydrogels and Their Application in Controlled Release of Polyphenols. Mater. Sci. Eng. C 2012, 32, 452–463.
- 67. Flores-Céspedes, F.; Martínez-Domínguez, G.P.; Villafranca-Sánchez, M.; Fernández-Pérez, M. Preparation and Chara cterization of Azadirachtin Alginate-Biosorbent Based Formulations: Water Release Kinetics and Photodegradation Study. J. Agric. Food Chem. 2015, 63, 8391–8398.
- 68. Mazloom, N.; Khorassani, R.; Zohury, G.H.; Emami, H.; Whalen, J. Lignin-Based Hydrogel Alleviates Drought Stress in Maize. Environ. Exp. Bot. 2020, 175, 104055.
- 69. Lv, T.; Yang, H.; Zhang, R.; Fan, W.; Xu, Y.; Cao, H.; Ning, L.; Zhou, C.; Wang, L. Effects of Lignin on Root Activity and Soil Nutrients of Malus hupehensis. Var. Pingyiensis under the Use of Organic Fertilizer. Agric. Sci. 2017, 8, 341–347.
- 70. Gordobil, O.; Delucis, R.; Egüés, I.; Labidi, J. Kraft Lignin as Filler in PLA to Improve Ductility and Thermal Properties. I nd. Crop. Prod. 2015, 72, 46–53.
- 71. Vaidya, A.A.; Collet, C.; Gaugler, M.; Lloyd-Jones, G. Integrating Softwood Biorefinery Lignin into Polyhydroxybutyrate Composites and Application in 3D Printing. Mater. Today Commun. 2019, 19, 286–296.
- 72. Nguyen, N.A.; Barnes, S.H.; Bowland, C.C.; Meek, K.M.; Littrell, K.C.; Keum, J.K.; Naskar, A.K. A Path for Lignin Valori zation via Additive Manufacturing of High-Performance Sustainable Composites with Enhanced 3D Printability. Sci. Ad v. 2018, 4, eaat4967.
- 73. Cataldi, A.; Rigotti, D.; Nguyen, V.D.H.; Pegoretti, A. Polyvinyl Alcohol Reinforced with Crystalline Nanocellulose for 3D Printing Application. Mater. Today Commun. 2018, 15, 236–244.
- 74. Domínguez-Robles, J.; Martin, N.; Fong, M.; Stewart, S.; Irwin, N.; Rial-Hermida, M.; Donnelly, R.; Larrañeta, E. Antioxi dant PLA Composites Containing Lignin for 3D Printing Applications: A Potential Material for Healthcare Applications. P harmaceutics 2019, 11, 165.

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