Algae Metabolites in Cosmeceutical

Subjects: Dermatology Contributor: Yoon-Yen Yow, Krishnapriya Thiyagarasaiyar

Cosmeceuticals are topical cosmetic-pharmaceutical hybrids which refer to a cosmetic product with active ingredients claiming to have medicinal or drug-like benefits to skin health. Marine algae are rich in bioactive substances that have shown to exhibit strong benefits to the skin, particularly in overcoming rashes, pigmentation, aging, and cancer.

Keywords: marine algae ; cosmeceuticals ; UV-radiation ; anti-aging ; anticancer ; skin whitening

1. Introduction

1.1. Synthetic Versus Natural Ingredients in Cosmetic Industry

Cosmeceuticals are topical cosmetic-pharmaceutical hybrids which refer to a cosmetic product with active ingredients claiming to have medicinal or drug-like benefits to skin health $[\underline{1}][\underline{2}]$. Globally, the cosmeceutical industry is growing each year due to the trend of modern lifestyle. More recently, the cosmeceutical industry is progressively shifting to natural bioactive ingredients because of the ineffectiveness of synthetic cosmetics $[\underline{3}]$.

Ineffectiveness of synthetic cosmetics includes their side effects and low absorption rate. The low absorption rate of cosmetics could be due to the big size of the molecular compounds. A study by Bos and Marcus ^[4] asserted that only compounds with the molecular weight lesser than 500 Dalton (Da) could penetrate through the skin. Cyclosporin (MW 1202 Da), a topical immunosuppressant, was not effective against psoriasis and allergic contact dermatitis as a higher molecular weight of the compounds inhibits skin penetration. Still, it was effective in psoriasis treatment when directly injected into the skin. Some of the side effects include irritation and allergic reaction to the users. According to a case study, hydroxybenzoic acid esters (parabens), which are widely used in cosmetic products, has been reported to mimic oestrogen; hence, increasing the incidence of breast cancer and causing the development of malignant melanoma ^[5].

In addition, a study on a population conducted by the Centers for Disease Control and Prevention reported that 97% of 2540 individuals were exposed to phthalates (a component of plastic that appears in cosmetic products; for instance, dibutyl phthalate in nail polish), which could result in DNA damage in human sperm ^[6]. In 2004, the Environment California, Environmental Working Group, and Friends of the Earth issued reports on cosmetic products containing chemical ingredients that lacked safety data. Some of these chemicals caused adverse effects in animal studies such as male genitalia congenital disabilities, altered pregnancy outcomes, and decreased in sperm counts ^[6]. As a result, consumers have changed their preference and opted for natural cosmetic products. The global market value for natural cosmetics was about \$34.5 billion in 2018, and it is estimated to reach approximately \$54.5 billion in 2027 ^[7]. The everexpanding market for skincare products based on natural bioactive ingredients, which include plants, herbs, and even marine algae ^[8].

Macroalgae are classified into three major classes, namely Phaeophyceae (brown algae), Rhodophyceae (red algae), and Chlorophyceae (green algae). Based on the total culture production, it is estimated that about 59% of brown algae, 40% of red algae, and less than 1% of green algae are produced worldwide ^[9]. Marine algae are rich sources of structurally diverse bioactive compounds, which are absent in other taxonomic groups. Algae contain 10 times greater diversity of compounds than terrestrial plants ^[10] and they have a totally different flavonoid composition from vegetables and fruits. Macroalgae are a rich source of catechins and flavonols ^[11]. Furthermore, algae-derived phlorotannin possesses a unique structure, which is not found in terrestrial plants and this compound may constitute up to 25% of the dry weight of brown algae ^[11]. Algae produce a wide array of primary metabolites, such as unsaturated fatty acids, polysaccharides, vitamins, and essential amino acids ^{[12][13]}. Additionally, many research findings reported that secondary metabolites derived from algae such as fucoidan, fucoxanthin, sulphated polysaccharide, polyphenol and fucosterol were shown to possess anti-inflammation, antioxidant, anticancer, antibacterial and anti-aging effects ^{[14][15][16][17][18]}. The demand for algae bioactive compounds in cosmeceuticals is rapidly increasing as they contain natural extracts which are considered safe; thus,

rendering fewer side effects on humans. In ancient times, marine algae were used as medicine to treat skin-related diseases, such as atopic dermatitis and matrix metalloproteinase (MMP) related disease ^[12]. In a nutshell, marine algae are a promising resource for the development of cosmeceuticals.

Marine algae can survive in harsh conditions (i.e., withstand heat, cold, ultra-violet radiation, salinity, and desiccation) [8](9] ^[19] due to their ability to adapt to physiological changes by producing stress tolerant substances. For example, algae produce organic osmolytes during stress conditions, which also act as antioxidants and heat protectants. Algae grow under desiccation by producing specialized spores which remain dormant during stress conditions and revive once the conditions return to normal. The presence of thick cell walls with protective layers of chemical substances and mucilage sheath helps to delay the process of desiccation. Algae that grow in cold desserts can endure the subzero temperature and protect the cells from UV irradiation by producing spores that have thick cell walls and reserve food as lipid and sugars ^[20]. In addition, marine algae uptake inorganic ions to balance extracellular ion concentration and produce organic osmolytes which protects them from desiccation and UV lights. A study reported that Dunaliella salina has 55 novel membrane-associated proteins that showed changes in the composition and structure of the membranes associated with algae adaptation to salinity [21]. Algae are rich in a wide variety of secondary metabolites to help them adapt and survive in harsh conditions. Algae could also adapt to desiccation stress by producing specialized spores such as aplanspores, which are rich in astaxanthin. Astaxanthin is a carotenoid that protects the cells from photo-oxidation. Algae exposed to UV radiations will produce UV screening compounds such as mycosporine-like amino acids (MAA), which acted as antioxidants and involved in osmotic regulations. Furthermore, algae exposed to high solar radiation and low nitrogen concentration produce more β -carotene, such as Dunaliella ^[20]. Thus, algae that are naturally exposed to oxidative stress develop defense systems that protect them against reactive oxygen species (ROS) and free radicals. These compounds could be used in cosmetics to protect the cells against the adverse effects of UV radiation. Some of the environmental benefits of algae include fixation of carbon dioxide. Studies have reported that large cultivation of microalgae capable of uptaking carbon from the atmosphere; for instance, Spirulina platensis with carbon fixation rate of 318 mg/L⁻¹d⁻¹ and Chlorella vulgaris with carbon fixation rate of 251 mg/L⁻¹d⁻¹ (22)[23].

1.2. Current Applications of Algae-Derived Metabolites in Cosmeceutical Industrial

The transition from synthetic compounds to natural products such as marine algae have been attracting the attention of many researchers since algae possess a wide range of pharmacological activities with negligible cytotoxicity effects in human cells ^[24]. Marine algae are used for different purposes in food, pharmaceutical, biofuel, agriculture, and cosmetic industries. Industries, such as Cyanotech, Fuji Chemical Seambiotic, and Mera Pharmaceuticals are producers of microalgae biomass contributing to products in pharmaceuticals, cosmetics, and nutritious feed ^[25]. Interestingly, phycocyanin (usually found in red algae and cyanobacteria) is accepted as a natural color additive in food and cosmetics by the Food and Drug Administration (FDA) due to its non-toxic, natural, and biodegradable properties. Accordingly, it becomes the major target of the market in the United States ^[26].

Meanwhile, carotenoid such as astaxanthin plays a crucial role in scavenging free radicals in the human body and it is considered a strong antioxidant; hence, its popularity as a human dietary supplement. Leading cosmeceutical industries, such as Unilever, L'Oreal, Henkel, and Beiersdorf are expected to improve the growth of carotenoid market value in the European market ^[27]. The market value for carotenoids is expected to reach about \$1.53 billion by 2021 ^[27]

Furthermore, red algae *Gracilaria* account for most of the raw material for the agar extraction. It is reported by the Food and Agriculture Organization (FAO) of the United Nations that more than 80% of the agar were produced from *Gracilaria* species, which are mainly produced by China and Indonesia ^[29]. *Gracilaria* species have been widely used in cosmetics due to their stabilizing, thickening, and gelling characters. Commercially available products from *Gracilaria* species include hydrogel soap by Sea Laria[®], facial mask by Balinique[®], and hydrating cream by Thalasso[®] ^[29].

A number of algae-based skin products have been marketed, such as Algenist (an anti-aging moisturizer containing microalgae oil and alguronic acid from algae) ^[30], Helionori[®] by Gelyma and Helioguard365[®] (a sunscreen product containing MAAs from red seaweed *Porphyra umbilicalis*) ^[31], Protulines[®] by Exsymol S.A.M., Monaco (an anti-aging agent from protein-rich extract of *Arthrospira*), and Dermochlorella by Codif, St. Malo, France (an anti-wrinkling agent from *Chlorella vulgaris* extract) ^[32]. Therefore, bioactive compounds derived from algae could be considered a potential cosmeceutical agent for skincare.

1.3. UV Radiation and Skin-Related Diseases

Skin is one of the most complex and largest organs that serves as a protective barrier against water losses and environmental stresses, such as ultraviolet radiation (UVR), pathogens, physical agents, and chemicals ^[33]. The skin

comprises three layers—epidermis, dermis, and hypodermis. The presence of keratinocyte cells and melanocyte cells in the epidermis layer plays a vital role in repairing damaged skin and protecting the skin from UV light. The dermis consists of elastin, hyaluronic acid, and collagen which involves tissue repair and stability, whereas hypodermis consists of fats, which involved in body insulation ^[2]. Several skin-related diseases that have been reported include acne, eczema, dermatitis, hives, psoriasis, and pityriasis rosea which cause rashes ^[34]. Other skin diseases include pigmentation disorders, such as hypopigmentation due to the absence of melanocytes and hyperpigmentation caused by a metabolic disorder or skin irritation. In addition, one of the biggest concerns is skin cancers (e.g., squamous, basal, and melanoma) with melanoma being the deadliest form in America because of overexposure to UV radiation ^[35].

In most cases, humans are exposed to UV radiation due to overexposure to sunlight. UV radiation can produce many adverse effects within the cells, including DNA damage, skin pathologies, such as erythema and inflammation, skin aging, and cancer ^[36]. There are three main components of UV radiation, namely UVA (315–400 nm), UVB (280–315 nm), and UVC (100–280 nm) ^[37]. UVA can reach the dermis layer of skin, increasing the level of ROS that indirectly induce DNA mutagenesis, which results in skin aging and wrinkling. UVA can act as a carcinogen by shortening telomere in the DNA strand and it has less ability to stimulate melanin production resulting in redness, sun tanning, and freckles. UVB can penetrate the epidermis layer and damage the DNA in skin cells directly and induce skin cancers. UVC is highly bioactive but humans are not exposed to UVC because it is mostly absorbed by the ozone layer. In addition, UV-induced oxidative stress plays a crucial role in causing aging, inflammation, melanogenesis and even cancer which are shown in **Figure 1**^[9] ^{[12][38][39][40][41]}.

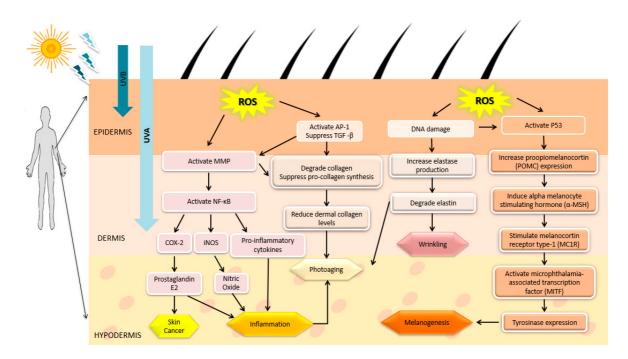
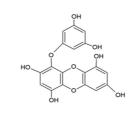
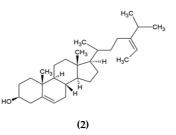


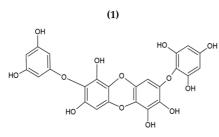
Figure 1. Effect of UV radiation-induced reactive oxygen species (ROS). Accumulation of ROS leads to skin cancer, inflammation, photoaging, wrinkling, and melanogenic through activation of respective signaling pathways.

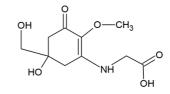
2. Marine Algae-Derived Compounds in Cosmeceutical Application

Based on the evidence from previous studies, brown algae contribute the most in cosmeceuticals. Some bioactive compounds from brown algae exhibit multiple cosmeceutical activities, including phlorotannin, which possesses several activities, such as anti-melanogenic, antioxidant, anti-inflammation, and anti-aging ^{[12][42][43][44]}. Likewise, fucoidan, a sulphated polysaccharide isolated from brown algae, contributes to anti-inflammation, anti-melanogenic and anticancer ^{[45][46][47]}. Fucoxanthin, a carotenoid isolated from brown, red, green and microalgae exhibit anti-melanogenic, anti-aging and antioxidant activities ^{[48][49][50]}. Mycosporine-like amino acids (MAAs), which are commonly found in red and green seaweeds, and microalgae also contribute to antioxidant, anti-inflammation, and anti-aging ^{[51][52][53]}. Other examples of bioactive compounds derived from algae, their applications and mode of actions in cosmeceuticals are presented in **Table 1**. The chemical structures of some prominent bioactive compounds are shown in **Figure 2**.

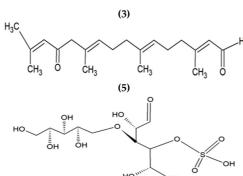


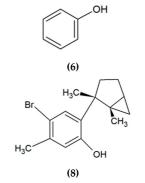


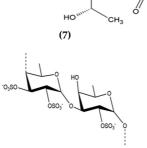




(4)

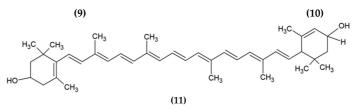


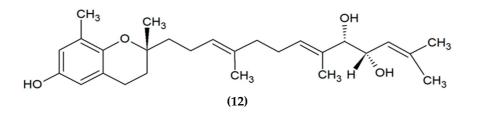


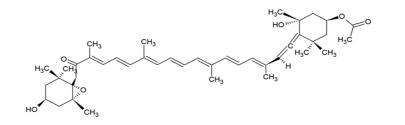


`СН₃

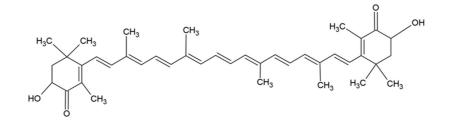




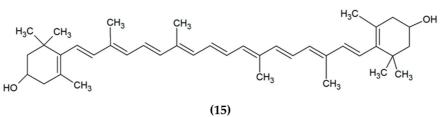


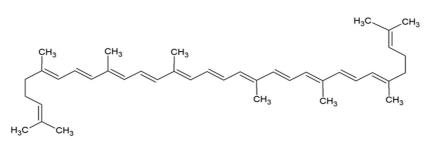


(13)



(14)





(16)

Figure 2. Chemical structures of bioactive compounds derived from algae. (1) Eckol, (2) Fucosterol, (3) Diphlorethohydroxycarmalol, (4) Mycosporine-glycine, (5) Eleganonal, (6) Phenol, (7) Ascophyllan, (8) Laurinterol, (9) Fucoidan, (10) Eicosapentaenoic acid, (11) Lutein, (12) Sargachromanol E, (13) Fucoxanthin, (14) Astaxanthin, (15) Zeaxanthin, and (16) Lycopene.

Table 1	Pionetivo com	nounde dorivod fr	am algae and t	heir applications ir	cosmocouticals
TANIC T.	Dioactive com	ipounus uenveu no	Jili alyae aliu i	inen applications il	i cosmeceuticais.

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
		Brown algae			
Ascophyllum nodosum	Ascophyllan	Anticancer	Inhibit MMP expression	B16 melanoma cells	[54]

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
Bifurcaria bifurcata	Eleganonal	Antioxidant	DPPH inhibition	In vitro	[<u>55]</u>
Chnoospora implexa	Ethanol extract	Antimicrobial	Bacterial growth inhibition	Staphylococcus aureus, Staphylococcus pyogenes	[56]
Chnoospora minima	Fucoidan	Anti- inflammation	Inhibition of LPS-induced NO production, iNOS, COX-2, and PGE2 levels	RAW macrophages	[47]
Cladosiphon okamuranus	Fucoxanthin	Antioxidant	DPPH inhibition	In vitro	[49]
Colpomenia sinuosa	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>
Cystoseira barbata	Fat-soluble vitamin and carotenoids	Antioxidant	High fat-soluble vitamin and carotenoid content	In vitro	[<u>57]</u>
Cystoseira foeniculacea	Polyphenol	Antioxidant	DPPH inhibition (EC ₅₀ = 5.27 mg/mL)	In vitro	[58]
Cystoseira hakodatensis	Phenol and fucoxanthin	Antioxidant	High total phenolic and fucoxanthin content	In vitro	[59]
Cystoseira osmundacea	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. pyogenes	[<u>56]</u>
Dictyopteris delicatula	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>
Dictyota dichotoma	Algae extract	Antimicrobial	Inhibit the synthesis of the peptidoglycan layer of bacterial cell walls	Penicillium purpurescens, Candida albicans, Aspergillus flavus	[60]

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
Ecklonia cava	Dieckol	Anti- inflammation	Suppression of iNOS and COX-2	Murine BV2 microglia	[<u>61]</u>
	Phlorotannin	Anti- melanogenic	Inhibit melanin production	B16F10 melanoma cells	[44]
	Phlorotannin	Antioxidant	ROS scavenging potential	Chinese hamster lung fibroblast (V79- 4)	[62]
Ecklonia kurome	Phlorotannin	Anti- inflammation	Inhibit hyaluronidase	Assay of HAase (In vitro)	[<u>42]</u>
	Phlorotannin	Anti-aging	Inhibit MMP expression	Human dermal fibroblast cell	[43]
Ecklonia Stolonifera	Phlorofucofuroeckol A and B	Anti- inflammation	Inhibition of NO production by downregulating iNOS and prostaglandin E2	LPS stimulated RAW 264.7 cells	[63]
Eisenia arborea	Phlorotannin	Anti- inflammation	Inhibit release of histamine	Rat basophile leukemia cells (RBL-2HE)	[64]
Eisenia bicyclis	Phlorotannin	Anti- inflammation	Inhibit hyaluronidase	Assay of HAase (In vitro)	[42]
Fucus evanescens	Fucoidan	Anticancer	Inhibit cell proliferation	Human malignant melanoma cells	[<u>45]</u>

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
	Extract	Anti-aging	Stimulate collagen production	N/A	8]
Fucus vesiculosus	Fucoidan	Anti- melanogenic	Inhibit tyrosinase and melanin	B16 murine melanoma cells	[46]
	Fucoidan	Anticancer	Decrease melanoma growth	Mice	[<u>65]</u>
	Fucoxanthin	Antioxidant	Prevent oxidation formation	<i>In vitro</i> , RAW 264.7 macrophage, Mouse (ex vivo)	[<u>66]</u>
Halopteris scoparia	Ethanol extract	Anti- inflammation	COX-2 inhibition	COX inhibitory screening assay kit	[<u>67]</u>
Himanthalia elongota	Fatty acid and Phenol	Antimicrobial	Bacterial growth inhibition	Escherichia coli, Staphylococcus aureus	[68]
	Fucosterol	Anti-aging	Inhibit MMP expression	Human dermal fibroblast	[<u>18]</u>
Hizikia fusiformis	Ethyl acetate extract	Anti- melanogenic	Inhibit tyrosinase and melanin	B16F10 mouse melanoma cells	[69]
	Fucoxanthin	Antioxidant	DPPH inhibition	In vitro	[70]
Hydroclathrus clathratus	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>
Ishige foliacea	Phlorotannin	Anti- melanogenic	Downregulation of tyrosinase and melanin synthesis	B16F10 cells Zebrafish embryo	[71][72]

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
lshige okamurae	Diphlorethohydroxycarmalol	Anti- inflammation	Down-regulation of iNOS and COX-2 expression and NF-κβ activation	Human umbilical vein endothelial cells	[73]
Laminaria japonica	Fucoxanthin	Anti- melanogenic	Suppress tyrosinase activity	UVB- irradiated guinea pig	[48]
Laminaria ochroleuca	Polyphenol	Antioxidant	High total phenolic content and antioxidant capacity	In vitro	[74]
Macrocystis	Phlorotannin	Antioxidant	ROS scavenging potential	In vitro	8
pyrifera	Hyaluronic acid	Anti-aging	Enhance the production of syndecan-4	N/A	[<u>75]</u>
Padina concrescens	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>
Padina pavonica	Polyphenol	Antimicrobial	Bacterial growth inhibition	Candida albicans and Mucor ramaniannus	[17]
	Acetone extract	Antioxidant	Free radical scavenging activity (IC ₅₀ = 691.56 μg L ⁻¹)	In vitro	[60]
Padina tetrastromatic	Diterpenes	Antioxidant	DPPH ($IC_{50} =$ 1.73) & ABTS ($IC_{50} = 2.01$) inhibitions	In vitro	[76]
	Sulfated polysaccharide	Anti- inflammation	COX-2 and iNOS inhibitions	Paw edema in rats	[77]

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
Petalonia	Ethanol extract	Anti- melanogenic	Inhibit tyrosinase and melanin	B16F10 murine melanoma cells	[<u>78]</u>
binghamiae	Aqueous extract	Antioxidant Anti- inflammation	DPPH inhibition COX-2 inhibition	In vitro In vitro	[<u>67]</u>
Rosenvingea intrincata	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>
Saccharina latissima	Phenol	Antioxidant	High total phenolic content, DPPH scavenging activity and FRAP	In vitro	[79]
Sargassum fulvellum	Fucoxanthin	Antioxidant	DPPH inhibition	In vitro	[<u>70]</u>
Sargassum furcatum	Methanol extract	Antioxidant	DPPH ($EC_{50} =$ 0.461) & ABTS ($EC_{50} = 0.266$) inhibitions	In vitro	[80]
Sargassum hemiphyllum	Sulfated polysaccharide	Anti- inflammation	Inhibit LPS- induced inflammatory response	RAW 264.7 macrophage cells	[<u>81]</u>
Sargassum henslowianum	Sulfated polysaccharide	Anticancer	Activation of caspase-3	B16 melanoma cells	[82]
Sargassum horridum	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[56]
Sargassum	Sargachromanol.E	Anti-aging	Inhibit MMP expression	UVA irradiated dermal fibroblast	[83]
horneri	Alginic acid	Anti- inflammation	Inhibit inflammatory response	HaCaT cells	[<u>84]</u>

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
Sargassum muticum	Tetraprenyltoluquinol chromane meroterpenoid	Anti-aging	ROS scavenging potential	Human dermal fibroblast	[<u>85]</u>
Sargassum polycystum	Ethanol extract	Anti- melanogenic	Inhibit tyrosinase and melanin production	B16F10 melanoma cells	[39]
Sargassum serratifolium	Sargachromenol	Anti- melanogenic	Downregulation of microphthalmia- associated transcription factor	B16F10 melanoma cells	[39]
Sargassum siliquastrum	Fucoxanthin	Antioxidant	Reduced UVB- induced ROS production	Human fibroblast	[<u>86]</u>
Sargassum thunbergi	Thunbergols	Antioxidant	DPPH inhibition	In vitro	[<u>87]</u>
Sargassum vulgare	Methanol extract	Antioxidant	β-carotene bleaching activity	In vitro	[88]
Stoechospermum marginatum	Spatane diterpenoids	Anticancer	Cell growth inhibition	Murine B16F10 melanoma cells	[<u>89]</u>
Turbinaria conoides	Laminarin, alginate, fucoidan	Antioxidant	ROS scavenging potential	N/A	[33]
Turbinaria ornata	Fucoxanthin	Antioxidant	High FRAP value (>10 μΜ/ μg of extract)	In vitro	[90]

Fucoxanthin Anti-aging MMP expression reduction, VEGF Mouse IED Undaria pinnatifida Ethyl acetate extract Anti- melanogenic Down regulate melanin and inhibit tyrosinase Mouse B16 melanoma cells Bull Polyunsaturated fatty acid Anti- inflammation N/A Mouse ear edema and erythema Bull Fucoxanthin Antioxidant DPPH inhibition In vitro IZO Alsidium corallinum Methanol extract Antimicrobial Bacterial growth inhibition Escherichia coli, Klebsiella pneumoniae, Staphylococcus aureus Bull Bangia Algae extract Antioxidant Induce peroxidase and superoxidase and superoxidase to reduce oxidative stress In vitro Bull Bryothammion triquetrum Methanol extract Antioxidant DPPH (ECso = 0.357) & ABTS (ECso = 0.370) inhibitions In vitro Bull	Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
Undaria pinnatifidaEthyl acetate extractAnti- melanogenicmelanin and mihibit tyrosinaseMouse B16 melanoma cellsB21Polyunsaturated fatty acidAnti- inflammationN/AMouse ear edema and erythemaB21FucoxanthinAntioxidantDPPH inhibitionIn vitroIZ0Alsidium corallinumMethanol extractAntimicrobialBacterial growth inhibitionEscherichia coli, Klebsiella neumoniae, staphylococcusB23BangiaAlgae extractAntioxidantInduce superoxide dismutase to reduce oxidative stressIn vitroI20Bryothamnion triguetrumMethanol extractAntioxidantDPPH (ECs0 = 0.357) & ABTS (CS5 = 0.370) inhibitionsIn vitroB21Burgothamnion triguetrumMethanol extractAntioxidantDPPH (ECs0 = 0.357) & ABTS (CS5 = 0.370) inhibitionsIn vitroB21Burgothamnion triguetrumMethanol extractAntioxidantDPPH (ECs0 = 0.357) & ABTS (CS5 = 0.370) inhibitionsIn vitroB21		Fucoxanthin	Anti-aging	expression reduction,	Mouse	[50]
Polyunsaturated fatty acidAnti- inflammationN/Aedema and erythema1921FucoxanthinAntioxidantDPPH inhibitionIn vitroIZ0Red algaeRed algaeRed algaeIzonIzonAlsidium corallinumMethanol extractAntimicrobialBacterial growth 		Ethyl acetate extract		melanin and inhibit		[91]
Red algae Escherichia coli, Klebsiella pneumoniae, staphylococcus aureus IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII		Polyunsaturated fatty acid		N/A	edema and	[<u>92]</u>
Alsidium corallinumMethanol extractAntimicrobialBacterial growth inhibitionEscherichia coli, Klebsiella 		Fucoxanthin	Antioxidant	DPPH inhibition	In vitro	[70]
Alsidium corallinumMethanol extractAntimicrobialBacterial growth inhibitionColi, Klebsiella pneumoniae, staphylococcus aureus133BangiaAlgae extractAntioxidantInduce peroxidae and superoxide dismutase to reduce oxidative stressIn vitro133Bryothamnion triquetrumMethanol extractAntioxidantDPPH (EC_50 = 0.357) & ABTS (EC_50 = 0.370) inhibitionsDn vitro133			Red algae			
BangiaAlgae extractAntioxidantperoxidase and superoxide dismutase to reduce oxidative stressIn vitro[94]Bryothamnion triquetrumMethanol extractAntioxidantDPPH (EC_{50} = 0.357) & ABTS (EC_{50} = 0.370) inhibitionsIn vitro[94]		Methanol extract	Antimicrobial		coli, Klebsiella pneumoniae, Staphylococcus	[93]
Bryothamnion triquetrum Methanol extract Antioxidant 0.357) & ABTS (EC50 = 0.370) In vitro [80] inhibitions In vitro [80]	Bangia	Algae extract	Antioxidant	peroxidase and superoxide dismutase to reduce oxidative	In vitro	[94]
Escherichia						
Ceramium Methanol extract Antimicrobial Bacterial growth Enterococcus [93] rubrum Staphylococcus aureus		Methanol extract	Antioxidant	0.357) & ABTS (EC ₅₀ = 0.370)	In vitro	[<u>80]</u>
Chondrocanthus acicularis Methanol extract Antimicrobial Bacterial growth pneumoniae, E. [93] [93] inhibition faecalis, S. aureus	triquetrum Ceramium			0.357) & ABTS (EC ₅₀ = 0.370) inhibitions Bacterial growth	Escherichia coli, Enterococcus faecalis, Staphylococcus	

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
Chondrus canaliculatus	Polysaccharide	Antioxidant	DPPH inhibition	In vitro	[<u>95]</u>
Chondrus crispus	Aqueous extract	Antimicrobial	Bacterial growth inhibition	Salmonella Enteritidis	[<u>96]</u>
Corallina pilulifera	Methanol extract	Anti-aging Antioxidant	Reduce the expression of gelatinase Inhibit free radical oxidation	Human dermal fibroblast Human fibrosarcoma (HT-1080)	<u>[97]</u>
Corallina vancouverensis	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>
Ganonema farinosum	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>
Gelidium crinaale	Fat-soluble vitamin and carotenoids	Antioxidant	High fat-soluble vitamin and carotenoid content	In vitro	[57]
Gelidium robustum	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>
Gracilaria gracilis	Phenol	Antioxidant	ROS scavenging potential	In vitro	[<u>98]</u>
Gracilariopsis Iemaneiformis	Sulfated polysaccharide	Antioxidant	DPPH, Superoxide radical assay, hydroxyl radical assay (EC ₅₀ = 2.45 mg/mL)	In vitro	[99]
Gracilaria salicornia	2H- chromenyl	Antioxidant Anti- inflammation	DPPH and ABTS inhibitions COX-1 inhibition	In vitro	[100]
Jania rubens	Glycosaminoglycan	Anti-aging	Collagen synthesis	Unknown	[75]

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
Laurencia caspica	Phenol Ethanol extract	Antioxidant Antimicrobial	DPPH inhibition Bacterial growth inhibition	In vitro Klebsiella pneumonia, Pseudomonas aeroginosa	[101]
Laurencia Iuzonensis	Sesquiterpenes	Antimicrobial	Bacterial growth inhibition	Bacillus megaterium	[12]
Laurenicia obtusa	Polysaccharide	Antioxidant	DPPH (IC_{50} = 24 µg/mL), FRAP (IC_{50} = 92 µg/mL), Hydroxyl radical scavenging activity (IC_{50} = 113 µg/mL)	In vitro	[102]
Laurenicia pacifica	Laurinterol	Antimicrobial	Bacterial growth inhibition	Staphylococcus aureus	[9]
Laurencia rigida	Sesquiterpenes	Antimicrobial	Bacterial growth inhibition	Bacillus megaterium	[<u>12]</u>
Meristotheca dakarensis	Glycosaminoglycan	Anti-aging	Collagen synthesis	Unknown	[<u>75</u>]
Osmundaria obtusilo	Methanol extract	Antioxidant	DPPH ($EC_{50} =$ 0.041 mg/mL), ABTS ($EC_{50} =$ 0.031 mg/mL), Metal chelating ($EC_{50} = 0.1$ mg/mL), folin ciocalteu ($EC_{50} =$ = 0.128 mg/mL)	In vitro	[80]
Palisada flagellifera	Methanol extract	Antioxidant	β-carotene bleaching activity	In vitro	[88]
Palmaria palmata	МАА	Anti-aging	Collagenase inhibition	Clostridium histolyticum	[53]

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
Polysiphonia howei	Fucoxanthin	Antioxidant	High FRAP value (>5 μΜ/μg of extract)	In vitro	[90]
Porphyra haitanensis	Sulfated Polysaccharide	Antioxidant	ROS scavenging potential	Mice	[103]
Porphyra umbilicalis	МАА	Anti-aging	Control expression of MMP	Human dermal fibroblast	[<u>16]</u>
Porphyra sp.	MAA	Anti-aging	Collagenases inhibition	Clostridium histolyticum	[<u>53]</u>
Porphyra yezoensis	MAA Polyphenol Phycoerythrin	Antioxidant Anticancer Anti- inflammation	ROS scavenging potential and MMP expression Induce apoptosis Suppression of mast cells	Human skin fibroblast HaCaT cells Rat	[51]
Pterocladia capillacea	Sulfated polysaccharide	Antimicrobial	N/A	Staphylococcus aureus Escherichia coli	[104]
Pyropia columbia	Phenol	Antioxidant	DPPH, β- carotene bleaching and ABTS inhibitions	Piaractus mesopotamicus	(<u>105)</u>
Pyropia yezoensis	Polysaccharide	Anti-aging	Promote collagen synthesis	Human dermal fibroblast	[106]

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference						
Rhodomela confervoides	Polyphenol	Antimicrobial	Bacterial growth inhibition	Candida albicans Mucor ramaniannus	[17]						
	Bromophenol	Antioxidant	DPPH inhibition	In vitro	[107]						
Schizymenia dubyi	Phenol	Anti- melanogenic	Inhibit tyrosinase activity	In vitro	[39]						
Green algae											
Bryopsis plumose	Polysaccharide	Antioxidant	ROS scavenging potential	In vitro	[<u>108]</u>						
Chaetomorpha antennia	Fucoxanthin	Antioxidant	DPPH inhibition (63.77%)	In vitro	[109]						
Chlamydomonas hedleyi	MAA	Antioxidant Anti-aging Anti- inflammation	ROS scavenging potential Increase UV- suppressed genes (procollagen C proteinase enhancer and elastin) expression Reduce COX-2 and involucrin expression	<i>In vitro</i> HaCaT cells HaCaT cells	[52]						
Cladophora sp.	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>						
Codium amplivesiculatum	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>						
Codium cuneatum	Ethanol extract	Antimicrobial	Bacterial growth inhibition	S. aureus, S. pyogenes	[<u>56]</u>						

Algae Species	Bioactive Compound/Extract	Beneficial Activity	Mechanism of Action	Experimental Model	Reference
		Anti-	Reduces the expression of		
Codium fragile References	Sterol	inflammation	COX-2, iNOS, and TNF-α	Mice	[110]
Kligman, D. Cos	meceuticals. Dermatol. Clin. 2	2000, 18, 609-	-615.		
cอิสแก่จรไก่แล้วร 37, 155–159.	shik _{ethan} GuntacM.; Kumar, V.;	Latheicrybap	Bacterial growth smeceuticals: An inhibition	emerging concep pyogenes	ot. 🌆 dian J. Pharm. 20
. Smit, N.; Vicanc Entromorpha i nRestinalis ; Meina Dermatol. 2000,	va, J.; Pavel, S. The Hunt for Chloroform and methanol ardie Matt. The 500 Dalton rule 9, 165–169.	Natural Skin V Antioxidant for the Skin p	Vhitening Agents. SOD activity is enetration of chen	Int. J. Mol. Sci. 2 Labidochromis nicalecompounds	009, 10, 5325–5349. ^[111] and drugs. Exp.
, Kerdudo, A.; Bu Enteromorpha ingredient–Natu Iinza	rger, P.; Merck, F.; Dingas, A.; ral Pobserchaivid eA case study	Rolland, Y.; N Contripxider Ren	ndasa@migning 2016	ez, X. Developm , 19 ,// 10 77–1089.	ent of a natural
	nical Exposures: The Ugly Sid		potential		
. Global Market V	alue for Natural Cosmetics in	2018–2027 St	atista. Available o	nline:	
https://www.stat	ista.com/statistics/673641/glo	bal-market-val	ue-for-natural-cos	metics/ (accesse	ed on 18 November
Caxtalla 2019). oxysperma	Fucoxanthin	Antioxidant		In vitro	[<u>90]</u>
. Ariede, M.B.; Ca	andido, T.M.; Jacome, A.L.M.; /. Algal Res. 2017, 25, 483–48	Velasco, M.V. 7.	(>6 µM/µg of R.; de Carvalho, J extract)	.C.M.; Baby, A.R	. Cosmetic attributes
. Wang, H.M.D.; (Chen, C.C.; Huynh, P.; Chang	, J.S. Explorin	g the potential of ι	using algae in cos	smetics. Bioresour.
Technol. 2015, 1	184, 355–362.	Antimicrohial	Bacterial growth	Strentococcus	[56]
Eu. W.: Nelson.	D.R.; Yi, Z.; Xu, M.; Khraiwes	h. B.: Jijakli. K	inhibition Chaiboonchoe	A pvAdromenai. A.: A	J-Khairy, D.:
	; et al. Bioactive compounds fi				
Chem. 2017, 54	, 199–225.		DPPH inhibition		
Ulva fasciata Freile-Pelegrín	Fucoxanthin Y.; Robledo, D. Bioactive phe	Antioxidant	1d\$ ⁸⁷⁰⁰⁷ *)gae. In	In vitro Bioactive Compo	[109] Junds from Marine
•	d Animal Sources, 1st ed.; He	•	•	•	
Chichester, UK,	2014; pp. 113–129.	Anti-	N1/A	N1/A	[75]
<i>Ulva lactuca</i> . Thomas. N.V.: K	Phycocolloids Kim, S.K. Beneficial effects of I	inflammation	N/A ompounds in cosn	N/A neceuticals. Mar.	
<u>164.</u>		name algare.			
, Fernando, I.S.; Ulva pertusa Pharm. 2016, 44	Nah, J.W.; Jeon, Y.J. Potentia Polysaccharide 3, 22–30.	anti-inflamma Antioxidant	ROS atory natural produ scavenging potential	icts from marine In vitro	alg <u>ae</u> . Environ. Toxico
. Peng, J.; Yuan,	JP.; Wu, CF.; Wang, JH. F	ucoxanthin, a	marine carotenoi	d present in brow	n seaweeds and
diatoms: Metabo	olism and bioactivities relevan	t to human he	alth. Mar. Drugs 2 DPPH inhibition,	011, 9, 1806–182	28.
i. Talero, E.; Garc Uselatelifatem mi	ía-Mauriño, S.; Ávila-Román, croସାନ୍ତିଶ୍ରଥା ନାର୍ଫାମିଚ୍ୟମିତ୍ୟମିଶ୍ରଣା କାର୍ଯ୍ୟ	J.; Rodríguez- ioA ⁿ tindidantce	Luna, A.; Alcaide,	A.; Motilva, V. Bi 5, <i>1</i> 13; ^{it} 6152–6209	oactive compounds).
	i, X.C.; Lee, D.J.; Chang, J.S				
0	ljoup <mark>Fen</mark> glenabdesselam, F.; To Afr. J. Biotechnol. 2012, 11, 94		Inggalaatikikkabme	ethannlic extracts	of ife ur Algerian mari
Udansp ge in UVB	c, S.Y.; Sun, Z.W.; Shin, H.S.; -irr soliated þolysar od harnde l fibro	b lansit-s .g Mg .r. B	ioteyahnoha2014, 1	Human dermal 6, 361–370.	[114]
. Martins, A.; Viei	ra, H.; Gaspar, H.; Santos, S.	Marketed mar	production ine natural produc	ts in the pharma	ceutical and
	ndustries: Tips for success. Mi				
	<i>Microa</i> po, K.; Verma, S. Adaptation i				Conditions. In Plant
	t egies in Changing Environme				

halotolerant alga Dunaliella salina as revealed by blue native gel electrophoresis and nano-LC-MS/MS analysis. Mol. Cell. Proteom. 2007, 6, 1459–1472.

- 22. Sydney, E.B.; Sydney, A.C.N.; de Carvalho, J.C.: Soccol, C.R. Potential carbon fixation of industrially important Algner Sangaes. In Biof Biosofiver Cangae, Elsevirer Ansteridam, The Netherlands, 2019, pp. 67–88. Reference
- 23. Usher, P.K.; Ross, A.B.; Camargo-Valero, M.A.; Tomlin, A.S.; Gale, W.F. An overview of the potential environmental impacts of large-scale microalgae cultivation. Biofuels 2014, 5, 331–349.
- 24**vájúnicera**Gómez, F.; Korbee, N.; Casas-Arrojo, V.: Abdala-Díaz, R.T.; Figueroa, F.L. UV photoprotection, cytotoxicity and immunology capacity of red algae extracts. Molecules 2019, 24, 341.
- 25. Khanra, S.; Mondal, M.; Halder, G.; Tiwari, O.N.; Gayen, K.; Bhowmick, T.K. Downstream processing of microalgae for Arthrospira pigments, protein and carbohydrate in industria appridation: ANeview. Food Bio Bio Bio 2018, 110, 60–84. exopolysacchandes
- 26. Pagels, F.; Guedes, A.C.; Amaro, H.M.; Kijjoa, A.; Vasconcelos, V. Phycobiliproteins from cyanobacteria: Chemistry and biotechnological applications. Biotechnol. Adv. 2019, 37, 422–463ect cells
- 27^CAlonedat, fr.S.^a, Gogi Setty, POlleAis wathanarayana, A.C., PRIX, P.N.; Bo, C.; Yuepeng, S. Industrial potential of carotenoid pigments from microalgae: Current trends and future prospects. Crit. Rev. Food Sci. Nutr. 2019, 59, 1880–1902.
- 28**C6/akesta**o, C.; Corinaldesi, C.; Sansone, C. Carotenoids from marine organisms: Biological functions and industrial MAA Anti-aging from UV N/A N/A radiation
- 29. Torres, P.; Santos, J.P.; Chow, F.; dos Santos, D.Y. A comprehensive review of traditional uses, bioactivity potential, and chemical diversity of the genus Gracilaria (Gracilariales, Rhodophyta), Algal Res. 2019, 37, 288–306.
- 30. Jahan, A.; Ahmad, M各, Fatima, N.; Ansari, V.A.; Ankina的如此 Alg 如何改进 tive compotition compoting a 2017, 56, 410–422. radiation *Chlorella*
- 31s Siekimana J. Microbial sunscreens. Microb. Biotechnol. 2011, 4, 1–7.

Reduce UV

- 32. Muñoz, R.; Gonzalez, Fernandez, C. (Eds.) Microalgae Based Biofugils and Bioproducts: From Feedstock Cultivation to End-Products; Woodhead Publishing: Duxford, UK, 2017.
- 33. Berthon, J.Y.; Nachat-Kappes, R.; Bey, M.; Cadoret, J.P.; Renimel, I.; Filaire, E. Marine algae as attractive source to skin care. Free Radic. Res. 2017, 51, 555–567. Anti-aging Reduced Human diploid [117]
- 34. Tabassum, N.; Hamdani, M. Plants used to treat skin diseases. Pharm. Rev. 2014, 8, 52–60.
- 35. Skin Cancer |Skin Cancer Facts|Common Skin Cancer Types. Available online: https://www.cancer.org/cancer/skin-Downregulated mRNA
- 36. Tan, L.T.; Mahendra, C.K.; Yow, Y.Y.; Chan, K.G.; Khan, T.M.; Equession Goh, BNtC/Streptionaryces High MUM273b: A inflammation mangrove-derived potential source for antioxidant and UVB radiatison for execution. Microbiol. Open 2019, 8, e859.
- and IFN-y 37. D'Orazio, J.; Jarrett, S.; Amaro-Ortiz, A.; Scott, T. UV radiation and the skin. Int. J. Mol. Sci. 2013, 14, 12222–12248.
- 38. Amaro-Ortiz, A.; Yan, B.; D'Orazio, J.A. Ultraviolet radiation, aging and the skin: Prevention of damage by topical cAMP Protect against Rat [119]
- 39. Azam, M.S.; Choi, J.; Lee, M.S.; Kim, H.R. Hypopigmenting effects of brown algae-derived phytochemicals: A review on molecular mechanisms. Mar. Drugs 2017, 15, 297.
- Reduced the 40. Hwang, H.; Chen, T.; Nines, R.G.; Shin, H.C.; Stoner, G.D. Photochemoprevention of UVB-induced skin carcinogenesis production of L-In SKH-1 mice by brown algae polyphenols. Int. J. Cancer 2006, <u>119, 2742</u>, 2749 In SKH-1 mice by brown algae polyphenols. Int. J. Cancer 2006, 119, 2742, 2749
- Anti-41. Mahendra, C.K.; Tahçī كالمجاب المجاب المجاب المحالي المحالي
- 42. Shibata, T.; Fujimoto, K.; Nagayama, K.; Yamaguchi, K.; Nakagura, T. Inhibitory activity of brown algal phlorotannins against hyaluronidase. Int. J. Food Sci. Technol. 2002, 37, 703–709.
- 43. Joe, M.J.; Kim, S.N.; Choi, H.Y.; Shin, W.S.; Park, G.M.; Kang, D.W.; Kim, Y.K. The inhibitory effects of eckol and dieckol from Ecklonia stolonifera on the expression of matrix metalloproteinase-1 in human dermal fibroblasts. Biol. Pharm. Bull. 2006, 29, 1735–1739.
- Yoon, N.Y.; Eom, T.K.; Kim, M.M.; Kim, S.K. Inhibitory effect of phlorotannins isolated from Ecklonia cava on mushroom tyrosinase activity and melanin formation in mouse B16F10 melanoma cells. J. Agric. Food Chem. 2009, 57, 4124– 4129.
- 45. Anastyuk, S.; Shervchenko, N.; Ermakova, S.; Vishchuk, O.; Nazarenko, E.; Dmitrenok, P.; Zvyagintseva, T. Anticancer activity in vitro of a fucoidan from the brown algae Fucus evanescens and its low-molecular fragments, structurally characterized by tandem mass-spectrometry. Carbohydr. Polym. 2012, 87, 186–194.
- 46. Wang, Z.-J.; Xu, W.; Liang, J.-W.; Wang, C.-S.; Kang, Y. Effect of fucoidan on B16 murine melanoma cell melanin formation and apoptosis. Afr. J. Tradit. Complement. Altern. Med. 2017, 14, 149–155.

- 47. Fernando, I.S.; Sanjeewa, K.A.; Samarakoon, K<u>Benefictar</u>, W.W. Kim, H.S.; Kang, N.; Banasinghe, P.; Lee, H.S.; Jeon, Alge Alge Algebra fraction duffer work of the second state of th
- 48. Shimoda, H.; Tanaka, J.; Shan, S.J.; Maoka, T. Anti-pigmentary activity of fucox antibia and its influence on skin mRNA expression of melanogenic molecules. J. Pharman flagrage 2010, 197–1145. human dermal expression
- 49 Mise T. Ueda, M.; Yasumoto, T. Production of fucoxanthin-rich powder from Cladosiphon okamuranus. Adv. J. Food provinals chool. 2011, Astaxanthin (carotenoid)
- 50. Urikura, I.; Sugawara, T.; Hirata, T. Protective effect of fucoxanthin against UVB-induced skin photoaging in hairless Anticancer scavenging Mice Mice potential
- 51. Sakai, S.; Komura, Y.; Nishimura, Y.; Sugawara, T.; Hirata, T. Inhibition of mast cell degranulation by phycoerythrin and its pigment moiety phycoerythrobilin, prepared from Porphyra yezoensis. Food Sci. Technol. Res. 2011, 17, 171–177. *Nannochloropsis*
- 52. Suh, S.S.; Hwang, 영森中和和帕纳.; Seo, H.; Kim, H独tiqueenu.; M的尺对;iuebeitique. Anti-immammation activities of granulata mycosporine-like amino acids (MAAs) in response to UV radiation suggest potential anti-skin aging activity. Mar. Drugs 2014, 12, 5174–5187.
- 53 Name Bar Sostner, J.; Fuchs, J.E.; Chaita, EntAligiannis, Nhiskaltsounis, L.; Ganzera, M. Inhibition of collagenase obview and compared to the com
- 54. Abu, R.; Jiang, Z.; Ueno, M.; Isaka, S.; Nakazono, S.; Okimura, T.; Cho, K.; Yamaguchi, K.; Kim, D.; Oda, T metastatic effects of the sulfated polysaccharide ascophyllan isolated from Ascophyllum nodosum on B16 melanoma. Biochem, Biophys. Res. Commun. 2015, 458, 727–732 Antioxidant Reduced Human [125] oxidative stress Cliama Calla
- 55. Silva, J.; Alves, C.; Freitas, R.; Martins, A.; Pinteus, S.; Ribeiro, J.; Gaspar, H.; Alfonso, A.; Pedrosa, R. Antioxidant and <u>Neuroprotective Potential of the Brown Seaweed Bifurcaria bifurcata in an in vitro Parkinson's Disease Mod</u>el. Mar. Drugs 2019, 17, 85. ROS
- 56. Muñoz-Ochoa, M.; Murillo-Álvarez, J.I.; Zermeño-Cervantes, L.A.; Martínez-Díaz, S.; Rodríguez-Riosmena, R. Screening of extracts of algae from Baja California Sur, Mexico as reversers of the antibiotic resistance of some pathogenic bacteria. Eur. Rev. Med. Pharmacol. Sci. 2010, 14, 739–747.
- Enduce 57**9Pante/kaeva**ta/.; M岳尼的hanova, A.; Dobreva, D.A.,过2xidamov, M.: Makedonski, L. 图的ids of black sea algae: Unveiling their potential for pharmaceutical and cosmetic applications. J. IMAB–Ann. Proc. Sci. Pap. 2017, 23, 1747–1751.
- 58. Messina, C.M.; Renda, G.; Laudicella, V.A.; Trepos, R.; Fauchon, M.; Hellio, C.; Santyllip Arerom ecology to phinkechnology study of the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the phinkechnology in the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus nore and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus and the defense strategies of algae and habaphytas (from Tranani Saltyrorks, NW Sicily) with a focus and the defense strategies (from Tranani Saltyrorks, NW Salt
- 59. Airanthi, M.W.A.; Hosokawa, M.; Miyashita, K. Comparative antioxidant activity of edible Japanese brown seaweeds. J. Food Sci. 2011, 76, C104–C111.
- 60. Kosanić, M.; Ranković, B.; Stanojković, T. Brown macroalgae holhi^tthe Adriatic Sea as a promising source of bioactive proinflammatory modulator Unknown
- 61**Porphyrivitikm size**o, SSWfa**ted** polysactlearide.M.; Park, P.M.; Byun, H.G.; Choi, Y.H.; Park, S.G.; Choi, T.W. Inhib tory effects and molecular mechanism of dieckol isolated from the probability of COX-2 and NOS in microglial cells. J. Agric. oxidative damage
- 62. Kang, K.A.; Lee, K.H.; Chae, S.; Koh, Y.S.; Yoo, B.S.; Kim, J.H.; Ham, Y.M.; Baik, J.S.; Lee, N.H.; Hyun, J.W. Triphlorethol-A from Ecklonia cava protects V79-4 lung fibroblast against hydrogen peroxide induced cell damage. Free Ros Sulfated polysaccharide Antioxidant scavenging In vitro
- 63. Lee, M.S.; Kwon, M.S.; Choi, J.W.; Shin, T.; No, H.K.; Choi, J. Soter Millin, D.S.; Kim, J.I.; Kim, H.R. Anti-inflammatory activities of an ethanol extract of Ecklonia stolonifera in lipopolysaccharide-stimulated RAW 264.7 murine m cells. J. Agric. Food Chem. 2012, 60, 9120–9129. Skeletonema Polyunsaturated aldehyde Inhibit cell
- SkeletonemaPolyunsaturated aldehydeInhibit cell64. Sugiura, Y.; Takeuchi, Y.; Kakinuma, M.; Amano, Mic Mike intory effects of seaweeds and fatty acidInhibit cellmarmoiand fatty acidproliferationbasophile leukemia cells (RBL-2H3). Fish. Sci. 2006, 72, 1286–1291.(A2058)
- 65. Teas, J.; Irhimeh, M.R. Melanoma and brown seaweed: An integrative hypothesis. J. Appl. Phycol. 2017, 29, 941–948.
- 66. Zaragozá, M.C.; López, D.P.; Sáiz, M.; Poquet, M.; Pérez, J.; Puig-Parellada, P.; Marmol, F.; Simonetti, P.; Gardana, C.; Lerat, Y.; et al. Toxicity and antioxidant activity in vitro and in vivo of two Fucus vesiculosus extracts. J. Agric. Food Chem. 2008, 56, 7773–7780.
- 67. Campos, A.M.; Matos, J.; Afonso, C.; Gomes, R.; Bandarra, N.M.; Cardoso, C. Azorean macroalgae (Petalonia binghamiae, Halopteris scoparia and Osmundea pinnatifida) bioprospection: A study of fatty acid profiles and bioactivity. Int. J. Food Sci. Technol. 2018, 54, 880–890.

- 68. Plaza, M.; Santoyo, S.; Jaime, L.; Reina, G.G.B.; Herrero, M.; Señoráns, F.J.; Ibáñez, F. Screening for bioactive Algor Bolinias from abjae: المجادة المجادة المحافة ال
- 69. Choi, E.O.; Kim, H.S.; Han, M.H.; Choi, Y.H.; Park, C.; Kim, B.W.; Hwang, H.J. Effects of Hizikia fusiforme fractions on melanin synthesis in B16F10 melanoma cells. J. Life Sci. 2013;1231 1495–1500.

Mouse

- 70. Yan, X.; Chuda, Y.; Suzuki, M.; Nagata, T. Fuco Altiminas the Mayouration in Hijikia fusiformis, a common edible seaweed. Biosci. Biotachool Biochomoclaga, 605–607. Human dermal [130]
- 71. Kim, K.N.; Yang, H.M.; Kang, S.M.; Kim, D.; Ahiŋ^{fl}@ŋŋŋetion, Y.ArOdetaphlorethol A isolated from Ishige foliacea inhibits α-(CCD-986sk) MSH-stimulated induced melanogenesis via ERK pathway in **B**វាøFstionmelanoma cells. Food Chem. Toxicol. 2013, 59, sjārlattīfa²6.
- 72. PRIMINS N.; Yang, H.M.; Kang, S.M.; Ahn, G.N.; Roh, S.W.; Lee, W.; Kim, D.K.; Jeon, Y.J. Whitening effect of octaphlorethol A isolated from Ishige foliacea in an in vivo zebrafish model. J. Microhigh, Biggechnol. 2015, 25, 448–451.
- 73. Heo, S.J.; Hwang, LY: Choi, J.L; Lee, S.H.; Park H.J.; Kang, BAHE, OH, CM, MARGE, MARGE, MARGE, CARL, Lee, S.H.; Park H.J.; et al. Protective effect of diphlorethohydroxycarmalol isolated from ishibit Okamurae agailist filght ignoce-induced-oxidative stress in human umbilical vein endothelial cells. Food Chem. Toxicol. 2010, 48, 92448-94994.
- 74. Del Olmo, A.; Picon, A.; Nuñez, M. High pressure processing for the extension of Laminaria ochroleuca (kombu) shelflife: A comparative study with seaweed salting and freezing. Innov. Food Sci. Emerg. Technol. 2019, 52, 420–428.
- 75. Special Chem The Anjverse and S. [68] 75. Special Chem — The Anjverse and S. [68] Special Chem — The Anjverse and Spe
- 76. Antony, T.; Chakraborty, K. Xenicanes attenuate pro-inflammatory 5-lipoxygenase: Prospective natural antiinflammatory leads from intertidal brown seaweed Padina tetrastromatica. Med. Chem. Res. 2019, 28, 591–607.
- 77. Mohsin, S.; Kurup, G.M. Mechanism underlying the anti-inflammatory effect of sulphated polysaccharide from Padina tetrastromatica against carrageenan induced paw edema in rats. Biomed. Prev. Nutr. 2011, 1, 294–301.
- Yoon, H.S.; Koh, W.B.; Oh, Y.S.; Kim, I.J. The Anti-Melanogenic Effects of Petalonia binghamiae extarcts in αmelanocyte stimulating hormone-induced B16/F10 murine melanoma cells. J. Korean Soc. Appl. Biol. Chem. 2009, 52, 564–567.
- Sappati, P.K.; Nayak, B.; VanWalsum, G.P.; Mulrey, O.T. Combined effects of seasonal variation and drying methods on the physicochemical properties and antioxidant activity of sugar kelp (Saccharina latissima). J. Appl. Phycol. 2019, 31, 1311–1332.
- Vasconcelos, J.B.; de Vasconcelos, E.R.; Urrea-Victoria, V.; Bezerra, P.S.; Reis, T.N.; Cocentino, A.L.; Navarro, D.M.; Chow, F.; Areces, A.J.; Fujii, M.T. Antioxidant activity of three seaweeds from tropical reefs of Brazil: Potential sources for bioprospecting. J. Appl. Phycol. 2019, 31, 835–846.
- Hwang, P.A.; Chien, S.Y.; Chan, Y.L.; Lu, M.K.; Wu, C.H.; Kong, Z.L.; Wu, C.J. Inhibition of lipopolysaccharide (LPS)induced inflammatory responses by Sargassum hemiphyllum sulfated polysaccharide extract in RAW 264.7 macrophage cells. J. Agric. Food Chem. 2011, 59, 2062–2068.
- Ale, M.T.; Maruyama, H.; Tamauchi, H.; Mikkelsen, J.D.; Meyer, A.S. Fucose-containing sulfated polysaccharides from brown seaweeds inhibit proliferation of melanoma cells and induce apoptosis by activation of caspase-3 in vitro. Mar. Drugs 2011, 9, 2605–2621.
- 83. Kim, J.A.; Ahn, B.N.; Kong, C.S.; Kim, S.K. The chromene sargachromanol E inhibits ultraviolet A-induced ageing of skin in human dermal fibroblasts. Br. J. Dermatol. 2013, 168, 968–976.
- Fernando, I.S.; Jayawardena, T.U.; Sanjeewa, K.A.; Wang, L.; Jeon, Y.J.; Lee, W.W. Anti-inflammatory potential of alginic acid from Sargassum horneri against urban aerosol-induced inflammatory responses in keratinocytes and macrophages. Ecotoxicol. Environ. Saf. 2018, 160, 24–31.
- Balboa, E.M.; Li, Y.X.; Ahn, B.N.; Eom, S.H.; Domínguez, H.; Jiménez, C.; Rodríguez, J. Photodamage attenuation effect by a tetraprenyltoluquinol chromane meroterpenoid isolated from Sargassum muticum. J. Photochem. Photobiol. B Biol. 2015, 148, 51–58.
- 86. Heo, S.-J.; Jeon, Y.-J. Protective effect of fucoxanthin isolated from Sargassum siliquastrum on UV-B induced cell damage. J. Photochem. Photobiol. B 2009, 95, 101–107.
- 87. Seo, Y.; Park, K.E.; Kim, Y.A.; Lee, H.J.; Yoo, J.S.; Ahn, J.W.; Lee, B.J. Isolation of tetraprenyltoluquinols from the brown alga Sargassum thunbergii. Chem. Pharm. Bull. 2006, 54, 1730–1733.
- Santos, J.P.; Torres, P.B.; dos Santos, D.Y.; Motta, L.B.; Chow, F. Seasonal effects on antioxidant and anti-HIV activities of Brazilian seaweeds. J. Appl. Phycol. 2018, 31, 1333–1341.

- Velatooru, L.R.; Baggu, C.B.; Janapala, V.R. Spatane diterpinoid from the brown algae, Stoechospermum marginatum induces apoptosis via ROS induced mitochondrial mediated caspase dependent pathway in murine B16F10 melanoma cells. Mol. Carcinog. 2016, 55, 2222–2235.
- 90. Kelman, D.; Posner, E.K.; McDermid, K.J.; Tabandera, N.K.; Wright, P.R.; Wright, A.D. Antioxidant activity of Hawaiian marine algae. Mar. Drugs 2012, 10, 403–416.
- Kim, M.; Kim, D.; Yoon, H.; Lee, W.; Lee, N.; Hyun, C. Melanogenesis inhibitory activity of Korean Undaria pinnatifida in mouse B16 melanoma cells. Interdiscip. Toxicol. 2014, 7, 89–92.
- 92. Khan, M.N.A.; Yoon, S.J.; Choi, J.S.; Park, N.G.; Lee, H.H.; Cho, J.Y.; Hong, Y.K. Anti-edema effects of brown seaweed (Undaria pinnatifida) extract on phorbol 12-myristate 13-acetate-induced mouse ear inflammation. Am. J. Chin. Med. 2009, 37, 373–381.
- 93. Rhimou, B.; Hassane, R.; José, M.; Nathalie, B. The antibacterial potential of the seaweeds (Rhodophyceae) of the Strait of Gibraltar and the Mediterranean Coast of Morocco. Afr. J. Biotechnol. 2010, 9, 6365–6372.
- 94. Wang, W.J.; Li, X.L.; Zhu, J.Y.; Liang, Z.R.; Liu, F.L.; Sun, X.T.; Wang, F.J.; Shen, Z.G. Antioxidant response to salinity stress in freshwater and marine Bangia (Bangiales, Rhodophyta). Aquat. Bot. 2019, 154, 35–41.
- 95. Jaballi, I.; Sallem, I.; Feki, A.; Cherif, B.; Kallel, C.; Boudawara, O.; Jamoussi, K.; Mellouli, L.; Nasri, M.; Amara, I.B. Polysaccharide from a Tunisian red seaweed Chondrus canaliculatus: Structural characteristics, antioxidant activity and in vivo hemato-nephroprotective properties on maneb induced toxicity. Int. J. Biol. Macromol. 2019, 123, 1267–1277.
- 96. Kulshreshtha, G.; Borza, T.; Rathgeber, B.; Stratton, G.S.; Thomas, N.A.; Critchley, A.; Hafting, J.; Prithiviraj, B. Red seaweeds Sarcodiotheca gaudichaudii and Chondrus crispus down regulate virulence factors of Salmonella enteritidis and induce immune responses in Caenorhabditis elegans. Front. Microbiol. 2016, 7, 421.
- Ryu, B.; Qian, Z.J.; Kim, M.M.; Nam, K.W.; Kim, S.K. Anti-photoaging activity and inhibition of matrix metalloproteinase (MMP) by marine red alga, Corallina pilulifera methanol extract. Radiat. Phys. Chem. 2009, 78, 98–105.
- Francavilla, M.; Franchi, M.; Monteleone, M.; Caroppo, C. The red seaweed Gracilaria gracilis as a multi products source. Mar. Drugs 2013, 11, 3754–3776.
- Wang, X.; Zhang, Z.; Wu, Y.; Sun, X.; Xu, N. Synthesized sulfated and acetylated derivatives of polysaccharide extracted from Gracilariopsis lemaneiformis and their potential antioxidant and immunological activity. Int. J. Boil. Macromol. 2019, 124, 568–572.
- 100. Antony, T.; Chakraborty, K. First report of antioxidative 2H-chromenyl derivatives from the intertidal red seaweed Gracilaria salicornia as potential anti-inflammatory agents. Nat. Prod. Res. 2019.
- 101. Moshfegh, A.; Salehzadeh, A.; Shandiz, S.A.S.; Shafaghi, M.; Naeemi, A.S.; Salehi, S. Phytochemical analysis, antioxidant, anticancer and antibacterial properties of the Caspian Sea red macroalgae, Laurencia caspica. Iran. J. Sci. Technol. Trans. A Sci. 2019, 43, 49–56.
- 102. Lajili, S.; Ammar, H.H.; Mzoughi, Z.; Amor, H.B.H.; Muller, C.D.; Majdoub, H.; Bouraoui, A. Characterization of sulfated polysaccharide from Laurencia obtusa and its apoptotic, gastroprotective and antioxidant activities. Int. J. Biol. Macromol. 2019, 126, 326–336.
- 103. De Jesus Raposo, M.; de Morais, A.; de Morais, R. Marine polysaccharides from algae with potential biomedical applications. Mar. Drugs 2015, 13, 2967–3028.
- 104. Pimentel, F.B.; Alves, R.C.; Rodrigues, F.; PP Oliveira, M.B. Macroalgae-derived ingredients for cosmetic industry—An Update. Cosmetics 2017, 5, 2.
- 105. Cian, R.E.; Bacchetta, C.; Rossi, A.; Cazenave, J.; Drago, S.R. Red seaweed Pyropia columbina as antioxidant supplement in feed for cultured juvenile Pacú (Piaractus mesopotamicus). J. Appl. Phycol. 2019, 31, 1455–1465.
- 106. Kim, C.R.; Kim, Y.M.; Lee, M.K.; Kim, I.H.; Choi, Y.H.; Nam, T.J. Pyropia yezoensis peptide promotes collagen synthesis by activating the TGF-β/Smad signaling pathway in the human dermal fibroblast cell line Hs27. Int. J. Mol. Med. 2017, 39, 31–38.
- 107. Li, K.; Li, X.M.; Gloer, J.B.; Wang, B.G. New nitrogen-containing bromophenols from the marine red alga Rhodomela confervoides and their radical scavenging activity. Food chem. 2012, 135, 868–872.
- 108. Zhang, Z.; Wang, F.; Wang, X.; Liu, X.; Hou, Y.; Zhang, Q. Extraction of the polysaccharides from five algae and their potential antioxidant activity in vitro. Carbohydr. Polym. 2010, 82, 118–121.
- 109. Premalatha, M.; Dhasarathan, P.; Theriappan, P. Phytochemical characterization and antimicrobial efficiency of seaweed samples, Ulva fasciata and Chaetomorpha antennina. Int. J. Pharm. Biol. Sci. 2011, 2, 288–293.
- 110. Lee, C.; Park, G.H.; Ahn, E.M.; Kim, B.A.; Park, C.I.; Jang, J.H. Protective effect of Codium fragile against UVBinduced pro-inflammatory and oxidative damages in HaCaT cells and BALB/c mice. Fitoterapia 2013, 86, 54–63.

- 111. Pezeshk, F.; Babaei, S.; Abedian Kenari, A.; Hedayati, M.; Naseri, M. The effect of supplementing diets with extracts derived from three different species of macroalgae on growth, thermal stress resistance, antioxidant enzyme activities and skin colour of electric yellow cichlid (Labidochromis caeruleus). Aquac. Nutr. 2019, 25, 436–443.
- 112. Farasat, M.; Khavari-Nejad, R.A.; Nabavi, S.M.B.; Namjooyan, F. Antioxidant properties of two edible green seaweeds from northern coasts of the Persian Gulf. Jundishapur. J. Nat. Pharm. Prod. 2013, 8, 47.
- 113. Fernandes, H.; Salgado, J.M.; Martins, N.; Peres, H.; Oliva-Teles, A.; Belo, I. Sequential bioprocessing of Ulva rigida to produce lignocellulolytic enzymes and to improve its nutritional value as aquaculture feed. Bioresour. Technol. 2019, 281, 277–285.
- 114. Adrien, A.; Bonnet, A.; Dufour, D.; Baudouin, S.; Maugard, T.; Bridiau, N. Pilot production of ulvans from Ulva sp. and their effects on hyaluronan and collagen production in cultured dermal fibroblasts. Carbohydr. Polym. 2017, 157, 1306–1314.
- 115. Mourelle, M.L.; Gómez, C.P.; Legido, J.L. The potential use of marine microalgae and cyanobacteria in cosmetics and thalassotherapy. Cosmetics 2017, 4, 46.
- 116. José de Andrade, C.; Maria de Andrade, L. An overview on the application of genus Chlorella in biotechnological processes. J. Adv. Res. Biotechnol. 2017, 2, 1–9.
- 117. Makpol, S.; Yeoh, T.W.; Ruslam, F.A.C.; Arifin, K.T.; Yusof, Y.A.M. Comparative effect of Piper betle, Chlorella vulgaris and tocotrienol-rich fraction on antioxidant enzymes activity in cellular ageing of human diploid fibroblasts. BMC Complement. Altern. Med. 2013, 13, 210.
- 118. Kang, H.; Lee, C.H.; Kim, J.R.; Kwon, J.Y.; Seo, S.G.; Han, J.G.; Kim, B.; Kim, J.; Lee, K.W. Chlorella vulgaris attenuates dermatophagoides farinae-induced atopic dermatitis-like symptoms in NC/Nga mice. Int. J. Mol. Sci. 2015, 16, 21021–21034.
- 119. Murthy, K.; Vanitha, A.; Rajesha, J.; Swamy, M.; Sowmya, P.; Ravishankar, G. In vivo antioxidant activity of carotenoids from Dunaliella salina—A green microalga. Life Sci. 2005, 76, 1381–1390.
- 120. Yang, D.J.; Lin, J.T.; Chen, Y.C.; Liu, S.C.; Lu, F.J.; Chang, T.J.; Wang, M.; Lin, H.W.; Chang, Y.Y. Suppressive effect of carotenoid extract of Dunaliella salina alga on production of LPS-stimulated pro-inflammatory mediators in RAW264. 7 cells via NF-κB and JNK inactivation. J. Funct. Foods 2013, 5, 607–615.
- 121. Shin, J.; Kim, J.E.; Pak, K.J.; Kang, J.I.; Kim, T.S.; Lee, S.Y.; Yeo, I.H.; Park, J.H.Y.; Kim, J.H.; Kang, N.J.; et al. A Combination of soybean and Haematococcus extract alleviates ultraviolet B-induced photoaging. Int. J. Mol. Sci. 2017, 18, 682.
- 122. Rao, A.R.; Sindhuja, H.N.; Dharmesh, S.M.; Sankar, K.U.; Sarada, R.; Ravishankar, G.A. Effective inhibition of skin cancer, tyrosinase, and antioxidative properties by astaxanthin and astaxanthin esters from the green alga Haematococcus pluvialis. J. Agric. Food Chem. 2013, 61, 3842–3851.
- 123. Banskota, A.H.; Sperker, S.; Stefanova, R.; McGinn, P.J.; O'Leary, S.J. Antioxidant properties and lipid composition of selected microalgae. J. Appl. Phycol. 2019, 31, 309–318.
- 124. Shen, C.T.; Chen, P.Y.; Wu, J.J.; Lee, T.M.; Hsu, S.L.; Chang, C.M.J.; Young, C.C.; Shieh, C.J. Purification of algal antityrosinase zeaxanthin from Nannochloropsis oculate using supercritical anti-solvent precipitation. J. Supercrit. Fluids 2011, 55, 955–962.
- 125. Wu, H.L.; Fu, X.Y.; Cao, W.Q.; Xiang, W.Z.; Hou, Y.J.; Ma, J.K.; Wang, Y.; Fan, C.D. Induction of apoptosis in human glioma cells by fucoxanthin via triggering of ROS-mediated oxidative damage and regulation of MAPKs and PI3K-AKT pathways. J. Agric. Food Chem. 2019, 67, 2212.
- 126. Rastogi, R.P.; Sonani, R.R.; Madamwar, D.; Incharoensakdi, A. Characterization and antioxidant functions of mycosporine-like amino acids in the cyanobacterium Nostoc sp. R76DM. Algal Res. 2016, 16, 110–118.
- 127. Haimeur, A.; Ulmann, L.; Mimouni, V.; Guéno, F.; Pineau-Vincent, F.; Meskini, N.; Tremblin, G. The role of Odontella aurita, a marine diatom rich in EPA, as a dietary supplement in dyslipidemia, platelet function and oxidative stress in high-fat fed rats. Lipids Health Dis. 2012, 11, 147.
- 128. Shannon, E.; Abu-Ghannam, N. Antibacterial derivatives of marine algae: An overview of pharmacological mechanisms and applications. Mar. Drugs 2016, 14, 81.
- 129. Lauritano, C.; Andersen, J.H.; Hansen, E.; Albrigtsen, M.; Escalera, L.; Esposito, F.; Helland, K.; Hanssen, K.Ø.; Romano, G.; Ianora, A. Bioactivity screening of microalgae for antioxidant, anti-inflammatory, anticancer, anti-diabetes, and antibacterial activities. Front. Mar. Sci. 2016, 3, 68.
- 130. Wu, Q.; Liu, L.; Miron, A.; Klímová, B.; Wan, D.; Kuča, K. The antioxidant, immunomodulatory, and anti-inflammatory activities of Spirulina: An overview. Arch. Toxicol. 2016, 90, 1817–1840.

131. El-Sheekh, M.M.; Daboor, S.M.; Swelim, M.A.; Mohamed, S. Production and characterization of antimicrobial active substance from Spirulina platensis. Iran. J. Microbiol. 2014, 6, 112–119.

Retrieved from https://encyclopedia.pub/entry/history/show/44038