

# Secondary Marine Metabolites

Subjects: [Oncology](#)

Contributor: Arijit Mondal , Sankhadip Bose , Sabyasachi Banerjee , Jayanta Kumar Patra , Jai Malik , Sudip Kumar Mandal , Kaitlyn L. Kilpatrick , Gitishree Das , Rout George Kerry , Carmela Fimognari , Anupam Bishayee

**Secondary marine metabolites** are defined as molecules with a molecular weight ranging between 100 to 1000 Da and, unlike primary marine metabolites, are often found to be unique to an organism or a specific taxonomic group of the marine source. Other than marine macroorganisms such as algae, sponges, or corals, specifically marine fungi and bacteria have shown to produce novel secondary metabolites with unique as well as diverse chemical structures that may hold the key for developing novel drugs or drug leads.

Secondary marine metabolites have been found in both prokaryotic and eukaryotic microorganisms, with unicellular bacteria (e.g., *Bacillus sp.*, *Pseudomonas sp.*), eukaryotic fungi (e.g., *Penicillium sp.*, *Aspergillus sp.*), filamentous actinomyces (e.g., *Streptomyces sp.*), and terrestrial plants being the most frequently studied and versatile producers. A potential clarification why organisms produce a high assortment of bioactive secondary marine metabolites is that these molecules furnish producers with a specific advantage against competing organisms and, furthermore, act as an adaptation to ecological conditions.

Secondary marine metabolites

Secondary metabolites

Marine metabolites

Marine bacteria

Marine fungi

Algae

## 1. Introduction

The ocean holds a broad reservoir of marine species full of natural pharmaceutical components of potential significance. Marine bioprospecting is a recent phenomenon; thus, aquatic life is a largely unexplored field of research [\[1\]\[2\]\[3\]\[4\]](#). Terrestrial life is the source of most pharmaceutically effective natural products [\[5\]](#). For centuries, marine flora has been exploited for their potential medicinal applications throughout the world [\[6\]](#). Among the marine organisms, bacteria, fungi, actinobacteria, seaweeds, and sponges have been utilized for cancer treatment [\[7\]\[8\]\[9\]\[10\]\[11\]](#). The most effective anti-cancer drugs are natural products. However, the natural product research and innovation phases are expensive, tedious, and time-consuming [\[12\]\[13\]](#).

Few marine pharmaceutical products for marketing have been licensed and some molecules are being clinically trialed within Phases I and III, along with preclinical studies. It is notable that among the scientifically developed marine compounds, four compounds are used in cancer treatment, such as cytarabine (Cytosar), trabectedin (Yondelis), eribulin mesylate (Halaven) and the conjugated antibody brentuximab vedotin (Acentris).

The dynamic and extremely sensitive global marine ecosystem covers about 70.8% of Earth [14]. This extreme biodiversity encompasses a heterogeneous array of micro- and macro-organisms. Among them, microorganisms like marine bacteria, fungi, and micro-algae orchestrate a pivotal role in restoring the balance within the aquatic environment by being both a producer and decomposer [15]. Marine microalgae are essential ecologically which are used as food and medicinal products from ancient times. Marine microalgae are eukaryotic plants that contribute to drug discovery by their metabolic plasticity, which can trigger the production of several compounds with possible applications in combating various diseases, such as cancer [16]. Presently, these aquatic species have gained increasing acclaim for their bioactive metabolites, which provides an unparalleled potential for a range of pharmaceutical activity. The distribution of marine species differs depending on the type of open sea ecosystem, such as pelagic environment, epipelagic environment, mesopelagic zone, bathypelagic zone, abyssopelagic zone, and benthic environment. These diversified divisions of the aquatic world harbors heterogeneous species of marine microflora and microalgae. Therefore, summing all the types of microflora and microalgae, and their extracted bioactive chemical metabolites, would be too tedious to elaborate in a single article. Thus, only certain specific microflora including bacteria, fungus, and microalgae (cyanobacteria), along with their potent bioactive metabolites displaying anticancer activity are emphasized. The most common bioactive compounds having anticancer activity from these marine sources are alkaloids (staurosporine, ambigols, amycolactam, and marinoquinoline A), polyketides (chromone, engyodontiumones H, pestalpolyol I, hytidchromone A, B, C, and E), terpenes (meroterpenes, diterpene, and scopararane I), peptides (beauvericin, polymyxin B and other non-ribosomal peptides), nucleosides (cytarabine, gemcitabine and other nucleoside analogues) and carbohydrates (laminaran, alginic acid, and other sulfated polysaccharides) [17][18][19].

Microflora and, up to a certain extent, microalgae could be regarded as chief drivers of nutrient transformations in a marine ecosystem. Shift in abiotic factors of marine ecosystem, such as temperature, salinity, nutrients, oxygen, solar energy, water clarity, tides, waves, aerial exposure and current has immensely influenced the production and secretion of marine metabolites/ bioactive chemical compounds from these organisms. To survive under these harsh abiotic factors, the marine microflora tends to form a symbiotic relationship with other marine microorganisms. These synergetic relationships enable them to endure and secrete a number of exotic secondary metabolites/bioactive compounds which they would not do under natural circumstances [20]. Over the last two decades, natural product research has observed a paradigm shift from terrestrial areas to oceans. Approximately 15,000 marine-origin metabolites, which have demonstrated cytotoxicity in cancer models, have been identified during the past three decades and from these, 28 agents are under clinical trials for their anticancer properties [21]. There is myriad of structural motifs undiscovered from the marine ecosystem and these metabolites can serve as new potent anticancer agents.

## 2. Marine Cyanobacteria Metabolites with Anticancer Property

### 2.1. Anthracyclines

Komodoquinone A (**1**) (Figure 1), produced by *Streptomyces* sp. KS3. This is a new anthracycline that caused neurogenesis (process of forming new neurites) in the neuro 2A neuroblastoma cell line [22].

**Figure 1.** Isolated marine cyanobacteria-derived anthracyclines, phenoxazin-3-one compounds, polyketides and peptides (1–8).

## 2.2. Phenoxazin-3-One Compounds

The novel anticancer antibiotics chandrananimycins A (2), B (3), and C (4) with phenoxazin-3-one moiety have been isolated from the marine *Actinomadura* sp. [23]. These compounds exhibited anticancer activity by inhibiting the proliferation of cancer cell lines such as CCL HT29 (colon cancer cell), MEXF 514L (melanoma cells), LXFA 526L, LXFL 529L (lung cancer cells), CNCL SF268, LCL H460, MACL MCF-7 (breast cancer cells), and PRCL PC3M, RXF 631L (kidney tumor) [23].

## 2.3. Polyketides

Ankaraholide A (5) is a glycosylated swinholide compound, which is procured from *Geitlerinema* sp. [24]. It inhibited the proliferation of NCI-H460, Neuro-2a, and MDAMB-435 cells [25]. Swinholide A (6) was initially obtained from the

marine sponge *Theonella swinhoei*. Later, it was also reported to be the metabolites of the marine cyanobacterium *Symploca sp.* It exhibited its antitumor activity by disruption of actin [24].

#### 2.4. Peptides

Symplostatin 1 (7) (Figure 1) is an analogue of dolastatin 10, isolated from *Symploca hydroides*, a marine bacterium. The antimitotic activity of symplostatin was shown against a panel of cancer cell lines, such as MDA-MB-435, SK-OV-3, NCI/ADR, A-10, and HUVEC [26]. It also showed profound antitumor activity against murine colon 38 and murine mammary 16/C carcinoma cells. This metabolite triggered the phosphorylation of Bcl-2, micronuclei formation, caspase-3 activation, and induced apoptosis that led to cell cycle arrest at the G2/M phase. It also prevented the accumulation of tubulin [26]. Grassypeptolide is a macrocyclic depsipeptide formed by the cyanobacteria *Lyngbya confervoides* [27]. It showed cytotoxic antiproliferative activity against various cell lines, such as human osteosarcoma (U2OS), cervical carcinoma (HeLa), colorectal adenocarcinoma (HT-29), and neuroblastoma (IMR-32) cell lines [27]. Grassypeptolide A (8) (Figure 1), B (9) and C (10) (Figure 2), significantly inhibited the proliferation of colorectal adenocarcinoma (HT-29) cell lines and cervical carcinoma (HeLa) cell lines in a concentration-dependent manner by inducing cell cycle arrest at either G1 the phase or G2/M phase [28].

**Figure 2.** Isolated marine cyanobacteria-derived peptides (9–16).

Curacin A (**11**) ([Figure 2](#)), a linear and complex hybridketopeptide, was the first curacin that was isolated from extracts of the Caribbean cyanobacterium *Lyngbya majuscula* [\[29\]](#). After curacin A, other curacin compounds, namely curacin B, C, and D, were also identified as the constituents of *L. majuscula* [\[30\]](#). Amongst these, curacin A was the most active anticancer compound that inhibited the proliferation of non-small cell lung cancer cells (A549) by triggering apoptosis and causing cell cycle arrests in the G2/M phase [\[31\]](#). It acts by binding to tubulin at colchicine binding site and acting as a competitive antagonist, and tubulin polymerization inhibitor [\[32\]](#). Its structural activity relationship (SAR) studies indicated that the existence of four double bonds, a conjugated diene, a readily oxidized thiazoline heterocycle, and high lipophilicity are the factors that govern the pharmacological function of curacin A [\[33\]](#). Various analogues were prepared, but they exhibit activity, lower than that the parent compound [\[34\]](#).

### 3. Conclusions, Current Challenges and Future Perspectives

Marine resources certainly have valuable and undiscovered biochemical versatility and demonstrate a greater opportunity for the development of new anticancer agents. While a variety of compounds have been identified to suppress cell growth in a broad spectrum of cancer cell types, the mechanism of action still remains unclear. A handful of marine molecules have demonstrated possible cytotoxic actions toward specific cancer by inhibition of cell proliferation, its antimitotic behavior (anti-tubulin impacts), induced apoptosis and inhibition of movement, invasion, or metastatic potential of cancer cells.

While these metabolites have shown potential for cancer treatment, there are several challenges associated with the development of these drugs that need consideration. Oceans certainly provide a large supply of valuable species, but the researchers still cannot access any of these regions. For years, the selection of entities in readily accessible places was preferred. Therefore, the problem is that marine research is not always easily accessible where researchers and the oceanographer need to have strong working ties. Genetic engineering is undergoing development in order to enhance drug production through the conversion of genetic data from the target compound into the host cells. It is an important field for regulation of the isolation and expression of aquatic genes, helping us produce lead compounds from the aquatic ecosystem in a more controlled way. Secondary metabolites are hard to produce independently from cultures, since their development is directly or indirectly dependent on the host. Therefore, several of the main genes stay silent while these things are attempted to evolve *In vitro*. Another big concern is that the development of a specific molecule requires sufficient resources. There is still little research available on the toxicity studies of these marine metabolites in normal cells; this has to be addressed.

Marine cyanobacteria and microalgae tend to be an effective source of anticancer drugs. Nevertheless, more studies are required to understand the basic targets and pathways behind the cytotoxicity of these compounds in cancer cells.

---

### References

1. Grothaus, P.G.; Cragg, G.M.; Newman, D.J. Plant natural products in anticancer drug discovery. *Curr. Org. Chem.* 2010, 14, 1781–1791.
2. Demain, A.L.; Vaishnav, P. Natural products for cancer chemotherapy. *Microb. Biotechnol.* 2011, 4, 687–699.
3. Cragg, G.M.; Kingston, D.G.I.; Newman, D.J. *Anticancer Agents from Natural Products*, 2nd ed.; CRC/Taylor & Francis: Boca Raton, FL, USA, 2012.
4. Basmadjian, C.; Zhao, Q.; Djehal, A.; Bentouhami, E.; Nebigil, C.G.; Johnson, R.A.; Serova, M.; De Gramont, A.; Faivre, S.; Raymond, E.; et al. Cancer wars: Natural products strike back. *Front. Chem.* 2014, 2, 20.

5. Cragg, G.M.; Newman, D.J. Plants as a source of anti-cancer agents. *J. Ethnopharmacol.* 2005, 100, 72–79.
6. Wang, W.; Wang, S.X.; Guan, H.S. The antiviral activities and mechanisms of marine polysaccharides: An overview. *Mar. Drugs* 2012, 10, 2795–2816.
7. Khalifa, S.A.M.; Elias, N.; Farag, M.A.; Chen, L.; Saeed, A.; Hegazy, M.F.; Moustafa, M.S.; Abd El-Wahed, A.; Al-Mousawi, S.M.; Musharraf, S.G.; et al. Marine natural products: A source of novel anticancer drugs. *Mar. Drugs* 2019, 17, 491.
8. Faulkner, D.J. Marine pharmacology. *Antonie Leeuwenhoek* 2000, 77, 135–145.
9. Schwartzmann, G.; Brondani, A.; Berlinck, R.G.S.; Jimeno, J. Marine organisms and other novel natural sources of new cancer drugs. *Ann. Oncol.* 2000, 11, 235–243. [Google Scholar] [CrossRef] [PubMed]
10. White, M.C.; Holman, D.M.; Boehm, J.E.; Peipins, L.A.; Grossman, M.; Henley, S.J. Age and cancer risk: A potentially modifiable relationship. *Am. J. Prev. Med.* 2014, 46, S7–S15.
11. Newman, D.; Cragg, G. Marine-sourced anti-cancer and cancer pain control agents in clinical and late preclinical development. *Mar. Drugs* 2014, 12, 255–278.
12. Gerwick, W.H.; Moore, B.S. Lessons from the past and charting the future of marine natural products drug discovery and chemical biology. *Chem. Biol.* 2012, 19, 85–98.
13. Montaser, R.; Luesch, H. Marine natural products: A new wave of drugs? *Future Med. Chem.* 2011, 3, 1475–1489.
14. Townsend, M.; Davies, K.; Hanley, N.; Hewitt, J.E.; Lundquist, C.J.; Lohrer, A.M. The challenge of implementing the marine ecosystem services concept. *Front. Mar. Sci.* 2018, 5, 359.
15. Snelgrove, P.V.R. Getting to the bottom of marine biodiversity: Sedimentary habitats: Ocean bottom are the most widespread habitats on earth and support high biodiversity and key ecosystem services. *BioScience* 1999, 49, 129–138.
16. Lauritano, C.; Andersen, J.H.; Hansen, E.; Albrigtsen, M.; Escalera, L.; Esposito, F.; Helland, K.; Hanssen, K.Ø.; Romano, G.; Ianora, A. Bioactivity screening of microalgae for antioxidant, anti-inflammatory, anticancer, anti-diabetes and antibacterial activities. *Front. Mar. Sci.* 2016, 3, 1–12.
17. Ruiz-Torres, V.; Encinar, J.A.; Herranz-López, M.; Pérez-Sánchez, A.; Galiano, V.; Barrajión-Catalán, E.; Micol, V. An Updated Review on Marine Anticancer Compounds: The Use of Virtual Screening for the Discovery of Small-Molecule Cancer Drugs. *Molecules* 2017, 22, 1037.
18. Deshmukh, S.K.; Prakash, V.; Ranjan, N. Marine Fungi: A Source of Potential Anticancer Compounds. *Front. Microbiol.* 2018, 8, 2536.

19. Vasconcelos, A.A.; Pomin, V.H. Marine Carbohydrate-Based Compounds with Medicinal Properties. *Mar. Drugs* 2018, 16, 233.
20. Mahapatra, G.P.; Raman, S.; Nayak, S.; Gouda, S.; Das, G.; Patra, J.K. Metagenomics approaches in discovery and development of new bioactive compounds from marine actinomycetes. *Curr. Microbiol.* 2019.
21. Malve, H. Exploring the ocean for new drug developments: Marine pharmacology. *J. Pharm. Bioallied Sci.* 2016, 8, 83–91.
22. Itoh, T.; Kinoshita, M.; Aoki, S.; Kobayashi, M. Komodoquinone A, a novel neuritogenic anthracycline, from marine *Streptomyces* sp. KS3. *J. Nat. Prod.* 2003, 66, 1373–1377.
23. Maskey, R.P.; Li, F.; Qin, S.; Fiebig, H.H.; Laatsch, H. Chandrananimycins A approximately C: Production of novel anticancer antibiotics from a marine *Actinomadura* sp. isolate M048 by variation of medium composition and growth conditions. *J. Antibiot. (Tokyo)* 2003, 56, 622–629.
24. Andrianasolo, E.H.; Gross, H.; Goeger, D.; Musafija-Girt, M.; McPhail, K.; Leal, R.M.; Mooberry, S.L.; Gerwick, W.H. Isolation of swinholide A and related glycosylated derivatives from two field collections of marine cyanobacteria. *Org. Lett.* 2005, 7, 1375–1378.
25. Williams, P.G.; Yoshida, W.Y.; Moore, R.E.; Paul, V.J. The isolation and structure elucidation of Tasiamide B, a 4-amino-3-hydroxy-5-phenylpentanoic acid containing peptide from the marine cyanobacterium *Symploca* sp. *J. Nat. Prod.* 2003, 66, 1006–1009.
26. Mooberry, S.L.; Leal, R.M.; Tinley, T.L.; Luesch, H.; Moore, R.E.; Corbett, T.H. The molecular pharmacology of symprostatin 1: A new antimitotic dolastatin 10 analog. *Int. J. Cancer* 2003, 104, 512–521.
27. Kwan, J.C.; Rocca, J.R.; Abboud, K.A.; Paul, V.J.; Luesch, H. Total Structure Determination of Grassypeptolide, a New Marine Cyanobacterial Cytotoxin. *Org. Lett.* 2008, 10, 789–792.
28. Kwan, J.C.; Ratnayake, R.; Abboud, K.A.; Paul, V.J.; Luesch, H. Grassypeptolides A-C, cytotoxic bis-thiazoline containing marine cyclodepsipeptides. *J. Org. Chem.* 2010, 75, 8012–8023.
29. Gerwick, W.H.; Proteau, P.J.; Nagle, D.G.; Hamel, E.; Blokhin, A.; Slate, D. Structure of curacin A, a novel antimitotic, antiproliferative, and brine shrimp toxic natural product from the marine cyanobacterium *Lyngbya majuscula*. *J. Org. Chem.* 1994, 59, 1243–1245.
30. Yoo, H.D.; Nagle, D.G.; Gerald, R.S.; Gerwick, W.H.; Kim, T.S.; Nambu, M.; White, J.D. Absolute configuration of curacin A, a novel antimitotic agent from the tropical marine cyanobacterium *Lyngbyamajuscula*. *Tetrahedron Lett.* 1995, 36, 1189–1192.
31. Catassi, A.; Cesario, A.; Arzani, D.; Menichini, P.; Alama, A.; Bruzzo, C.; Imperatori, A.; Rotolo, N.; Granone, P.; Russo, P. Characterization of apoptosis induced by marine natural products in non small cell lung cancer A549 cells. *Cell Mol. Life Sci.* 2006, 63, 2377–2386.



32. Blokhin, A.V.; Yoo, H.D.; Gerald, R.S.; Nagle, D.G.; Gerwick, W.H.; Hamel, E. Characterization of the interaction of the marine cyanobacterial natural product curacin A with the colchicine site of tubulin and initial structure-activity studies with analogues. *Mol. Pharmacol.* 1995, 48, 523–531.
33. Verdier-Pinard, P.; Lai, J.Y.; Yoo, H.D.; Yu, J.; Marquez, B.; Nagle, D.G.; Nambu, M.; White, J.D.; Falck, J.R.; Gerwick, W.H.; et al. Structure-activity analysis of the interaction of curacin A, the potent colchicine site antimitotic agent, with tubulin and effects of analogs on the growth of MCF-7 breast cancer cells. *Mol. Pharmacol.* 1998, 53, 62–76.
34. Wipf, P.; Reeves, J.; Day, B. Chemistry and Biology of Curacin A. *Curr. Pharm. Des.* 2004, 10, 1417–1437.

---

Retrieved from <https://encyclopedia.pub/entry/history/show/5459>