The Antitumor Activity of s-Triazine **Derivatives**

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1,3,5-triazine derivatives, also called s-triazines, are a series of containing-nitrogen heterocyclic compounds that play an important role in anticancer drug design and development. Three s-triazine derivatives, including altretamine, gedatolisib, and enasidenib, have already been approved for refractory ovarian cancer, metastatic breast cancer, and leukemia therapy, respectively, demonstrating that the s-triazine core is a useful scaffold for the discovery of novel anticancer drugs.

s-triazines antitumor drug target

1. Topoisomerases Inhibition

In recent years, a number of DNA topoisomerase inhibitors have been reported in both basic research and clinical applications. Topoisomerase was first discovered in E. coli by Wang et al. in 1971. Since then, the study of topoisomerase's structure, function, and mechanism has been initiated. Generally, human DNA topoisomerases are classified into two large groups: type I and type II topoisomerases. Type I topoisomerases could generate transient single-strand breaks (SSBs) in the DNA molecule by forming a covalent phosphotyrosyl linkage without ATP. In contrast, type II topoisomerases are large homodimeric proteins that require ATP for overall catalytic activity, which could generate transient DNA double-strand breaks (DSBs) [1]. Thus, topoisomerases play important roles in several cell processes, such as replication, transcription, chromosome separation, and segregation. Therefore, topoisomerases are ideal targets for the development of antitumor drugs.

Human DNA topoisomerase inhibitors could be classified into two categories based on their inhibition modes: topoisomerase poisons and topoisomerase catalytic inhibitors [2]. Topoisomerase poisons cover the majority of clinical antitumor agents (e.g., etoposide, doxorubicin, mitoxantrone, salvicine, and teniposide), which can kill cancer cells by stabilizing the covalent topo-DNA complexes and transforming this enzyme into a cellular toxin. Topoisomerase catalytic inhibitors can kill tumor cells by inhibiting the essential enzymatic activity of topoisomerase. A number of structurally diverse compounds have been identified as catalytic inhibitors: preventing DNA cleavage (e.g., merbarone), blocking the enzyme on the ATP-binding site (e.g., purine analogues), or inhibiting ATP hydrolysis (e.g., bisdioxipiperazine analogues). Because of the ubiquitous role of topoisomerase in correlation with various carcinomas, intensive efforts attempting to explore effective tumor therapeutics have led to a panel of chemical agents with diversified structures. In this research, the researchers focused on s-triazine derivatives as topoisomerase inhibitors.

The Novartis research group reported a kind of ATP-competitive catalytic topoisomerase II inhibitor 1 with the core 9H-purine scaffold in 2009 [3], as shown in **Figure 1**. Then the 9H-purine motif of the compounds was monocyclically substituted by 1,3,5-triazines, which was first reported [4] by Perdih et al. using a virtual screening protocol. In the hit selection, 4-amino-6-(phenylamino)-1,3,5-triazines 2 displayed potent topoisomerase IIa inhibitory activity with IC_{50} values in the micromolar range ($IC_{50} = 229 \mu M$). Moreover, compound 2 exhibited cytotoxicity against the HepG2 cell lines (IC₅₀ = 20.53 μ M), MCF-7 (IC₅₀ = 129.0 μ M), and normal HUVEC cells $(IC_{50} = 122 \mu M)$. Furthermore, Perdih et al. designed a series of 4,6-disubstituted-1,3,5-triazin-2(1H)-one analogs [5]. This research provided valuable structure-activity relationship (SAR) data regarding the role of substituents introduced at position 6 of the 1,3,5-triazin-2(1H)-one core. Among which, 6-(benzylthio)-4-((3chlorobenzyl)thio)-1,3,5-triazin-2(1H)-one 3 was identified as a topoisomerase II inhibitor with an improved IC₅₀ of 57.6 µM. However, these compounds showed low cytotoxicity in cancer cells. Perdih et al. further optimize the substituents at position 4 of the 1,3,5-triazin-2(1H)-one scaffold to improve the inhibition potency that would display activity on the cellular level. After virtual screening and experimental evaluation for human topo IIa inhibition, the most potent compounds were 4 and 5, with IC₅₀ values of 8.1 μM and 11.1 μM, respectively, as well as being superior to the positive drug etoposide ($IC_{50} = 28.6 \mu M$) [6]. Additionally, compound 5 showed better HepG2 cell inhibitory activity with EC₅₀ values of 38.7 μ M.

Figure 1. Chemical structures of topoisomerase II inhibitors.

2. Tyrosine Kinases Inhibition

Tyrosine kinases are implicated in tumorigenesis and progression and have emerged as major targets for drug discovery. Tyrosine kinase inhibitors (TKIs) inhibit corresponding kinases from phosphorylating tyrosine residues of their substrates and then block the activation of downstream signaling pathways. Over the past several decades,

multiple robust and well-tolerated TKIs with single or multiple targets, including BTK, EGFR, FAK, VEGFR, ALK, ROS1, HER2, MET, MEK, FGFR, and PDGFR, have been developed.

2.1. BTK-TKIs

Bruton's tyrosine kinase (BTK) is the first Tec family tyrosine kinase and also a key member of the B-cell receptor signalling pathway [Z]. Since BTK was confirmed to play a crucial role in B-cell maturation as well as in mast cell activation through the high-affinity IgE receptor, studies of BTK as a target have attracted substantial attention from drug researchers, resulting in the development of a diverse array of BTK inhibitors. BTK inhibitors can be divided into reversible and irreversible inhibitors. Most published BTK inhibitors are irreversible inhibitors, including imidazopyrimidine, 2,4-diaminopyrimidines, imidazoquinoxaline, 1,3,5-trazine, and other groups. There are five covalent irreversible BTK inhibitors (Ibrutinib, Acalabrutinib, Zanubrutinib, Tirabrutinib, Orelabrutinib) that have been marketed so far. Irreversible BTK inhibitors usually react with the key amino acid residue Cys481 of BTK via covalent bond modification. However, they may also inhibit other kinases with structurally related cysteines (such as EGFR, BMX, ITK, and JAK3), resulting in off-target effects and potential toxicity after prolonged administration. Meanwhile, several potent noncovalent reversible inhibitors have also been reported, including ARQ-531, Fenebrutinib, Vecabrutinib, and BIIB068, but none of these reversible BTK inhibitors has yet been approved to date.

Xiang et al. designed a kind of novel covalent BTK inhibitor by substituting the 1H-pyrazolo[3,4-d] pyrimidine core (Ibrutinib) with a 1,3,5-trazines scaffold containing a key acrylamide warhead, as shown in **Figure 2**. As reported, compound **6** showed potent BTK inhibitory activity (BTK IC₅₀ = 21 nM), validating the potential of the 1,3,5-trazine scaffold as a new skeleton for developing effective BTK inhibitors $^{[8]}$. However, **6** also potently inhibited EGFR with an IC₅₀ value of 31 nM. In order to improve its selectivity for BTK over EGFR, the 1,3,5-triazine scaffold modified by more hydrophobic side chains, which would additionally occupy the H3 pocket of BTK, was designed. Structure-activity relationship (SAR) and extensive pharmacological screening led to the discovery of a potent irreversible BTK inhibitor **7** $^{[9]}$. Compound **7** possessed desirable BTK selectivity (IC₅₀ = 17.0 nM) over EGFR (IC₅₀ > 1000 nM) and JAK3 (IC₅₀ = 104.6 nM), which shared a high degree of homology with BTK. However, it is worth noting that broad kinase panel screening is further needed to clarify the selectivity profile of **7**.

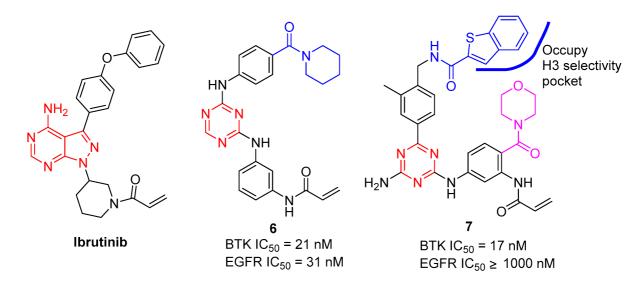


Figure 2. Chemical structures of BTK inhibitors.

2.2. EGFR-TKIS

The epidermal growth factor receptor (EGFR) is a trans-membrane protein belonging to the erbB/HER family of tyrosine kinase (TK) receptors. EGFR signalling in tumour cells, as opposed to normal cells, is changed and often becomes dysregulated. Accordingly, interrupting EGFR communicating signals is considered one of the targets for curing tumors. Multiple EGFR suppressors have been developed, including 1,3,5-triazine derivatives.

Based on virtual screening, Bai et al. identified a number of 1,3,5-triazine derivatives as potent dual-effective inhibitors against both wild-type and mutant EGFR in 2012 $^{[10]}$. As shown in **Figure 3**, compound **8** exhibited the most potent activity against wild-type EGFR (IC₅₀ = 25.9 μ M) and mutant EGFR T790M/L858R (IC₅₀ = 6.5 μ M). Structure–activity relationships (SAR) were analyzed among the triazine analogs. Compound **9** was obtained by removing the fluorine group in the phenyl part along with the inclusion of the para-hydroxy group on another phenyl, with a low inhibitory potency for wild-type EGFR (IC₅₀ > 100 μ M) and reduced about fivefold for mutant EGFR T790M/L858R (IC₅₀ = 30.7 μ M). Singh groups reported a novel hybrid analogue of monastrol-1,3,5-triazine **10** as an effective anti-breast cancer agent $^{[11]}$ in 2017, which showed excellent EGFR-TK enzyme inhibitory activity (96.3% at 10 μ M). Additionally, their group further modified 1,3,5-triazine to obtain 4-aminoquinoline-1,3,5-triazine derivatives $^{[12]}$. Compound **11** showed the most EGFR-TK enzyme inhibitory activity (96.4% at 10 μ M) and excellent anticancer activities against the entire tested cell lines. In 2021, hybrid quinazoline-1,3,5-triazine derivatives as EGFR inhibitors were reported by Pathak $^{[13]}$. Through pharmacological evaluation, they found that the substitution of the 1,3,5-triazine ring by morpholine and aniline rings could increase the ligand's potency in EGFR inhibition. Among them, the most active compound **12** showed an IC₅₀ value of 36.8 nM against the EGFR enzyme.

Figure 3. Chemical structures of EGFR inhibitors.

In 2018, He et al. investigated four previously reported 1,3,5-triazine compounds as anti-breast cancer agents via modulation of EGFR-TK [14]. Among the compounds, $\mathbf{13}$ (8.45 ± 0.65 μ M) and $\mathbf{14}$ (2.54 ± 0.22 μ M) significantly inhibited EGFR-TK and exhibited potent anticancer activity in all the tested cell lines in a dose-dependent manner. In 2021, 1,3,5-triazine-based pyrazole derivatives were synthesized by Osman et al. with anticancer activity targeting EGFR [15]. Compounds $\mathbf{15}$, $\mathbf{16}$, and $\mathbf{17}$ displayed excellent activity with an IC₅₀ value of 305.1, 286.9, and 229.4 nM, respectively, in comparison with reference erlotinib.

In 2022, El-Faham et al. synthesized a new series of mono- and bis(dimethylpyrazolyl)-s-triazine derivatives $^{[16]}$. Of these compounds, **18** showed excellent potency against the HCT116 cell lines (IC₅₀ = 500 ± 80 nM) through

apoptosis induction and exhibited potent EGFR inhibitory activity with an IC₅₀ value of 61 nM (tamoxifen, IC₅₀ = 69 nM). For the downstream pathway of PI3K/AKT/mTOR, **18** showed remarkable inhibitory activity with a 0.18-, 0.27-, and 0.39-fold decrease in their concentration. Additionally, pyrazole and fused pyrazole-s-triazine derivatives **19** and **20** were obtained by a one-pot synthesis method, which displayed EGFR inhibitory activity with IC₅₀ values of 70.3 and 59.24 nM, respectively, and similar PI3K/AKT/mTOR inhibitory activity compared with **18** $^{[17]}$. Therefore, compounds **18**, **19**, and **20** possessed EGFR/PI3K/AKT/mTOR signaling cascade inhibitors.

EGFR signaling dysregulation in tumors could result in EGFR overexpression or a gain-of-function mutation. According to the reports, acquired drug resistance to the first-generation EGFR-TKIs was revealed due to the T790M and L858R mutations in EGFR. Although a drug related to EGFR^{T790M} or EGFR^{T790M/L858R} mutation has been discovered, it is still needed to discover new EGFR inhibitors with high selectivity for EGFR^{T790M} and EGFR^{T790M/L858R} kinases. Azmy et al. reported a series of 1,3,5-triazine-based derivatives as potential EGFR^{WT} and EGFR^{T790M} inhibitors [18]. Among the derivatives, compound 21 as the most potent antiproliferative agent showed inhibitory activity with EGFR^{WT} (0.22 \pm 0.05 μ M) and EGFR^{T790M} (0.18 \pm 0.11 μ M), compared to the reference drugs erlotinib (0.09 \pm 0.05 μ M) and AZD9291 (0.55 \pm 0.10 μ M).

2.3. FAK-TKIS

Focal Adhesion Kinase (FAK) is an intracellular non-receptor tyrosine kinase that plays a crucial role in reproduction, early embryonic development, and tumorigenesis through its kinase activity and scaffold function. So far, s-triazines have been proven to be effective inhibitors targeting FAK [19].

In 2013, Chen and co-workers reported a series of diarylamino-1,3,5-triazine derivatives as FAK inhibitors $^{[20]}$. As shown in **Figure 4**, compound **22** showed poor activity against FAK ($^{[1]}$ C₅₀ = 41.9 $^{[1]}$ M) owing to the introduction of a 3,4,5-trimethoxyanilino group and a methanesulfonamide phenyl group on the 1,3,5-triazine ring. Then, they removed the chlorine atom from **22**, and compound **23** with good inhibitory activity against FAK ($^{[1]}$ C₅₀ = 5.1 $^{[1]}$ M) was obtained. Moreover, they found that when the methanesulfonamide phenyl group was displaced with an amide moiety, compound **24** showed significantly increased inhibitory activity against FAK ($^{[1]}$ C₅₀ = 0.4 $^{[1]}$ M). While changes in the 3,4,5-trimethoxyanilino group of compounds **24** and **25** exhibited similar inhibitory activity against FAK ($^{[1]}$ C₅₀ = 0.31 $^{[1]}$ M), in order to improve the FAK kinase activity, they continued to design imidazo[1,2-a][1,3,5] triazines as novel FAK inhibitors in 2015 $^{[21]}$. The new compounds displayed 10–100 nM IC₅₀ values against FAK, and compound **26** ($^{[1]}$ C₅₀ = 50 nM) was identified as the best inhibitor.

Figure 4. Chemical structures of FAK inhibitors.

3. Phosphoinositide 3-Kinase Inhibition

Phosphatidylinositol 3-kinase (PI3K) is a lipid kinase, that is, a central component in the PI3K/Akt/mTOR signaling pathway [22]. The most widely studied type I PI3Ks are further subdivided into class IA (PI3K α , β , and δ) and class IB (PI3K γ) based on the type of catalytic structural domain. Blocking the PI3K/AKT/mTOR pathway has been widely recognized as an attractive cancer therapeutic strategy owing to its crucial role in cell growth and survival. A number of PI3K inhibitors are currently under research, including pan-PI3K-mTOR inhibitors, pan-class I PI3K inhibitors, and isoformspecific PI3K inhibitors [23]. Dimorpholino-substituted s-triazine derivatives have shown great potential in PI3K-related cancer therapy. Several promising candidates, such as ZSTK474, gedatolisib, and bimiralisib, are currently in clinical trials.

ZSTK474 is an s-triazine derivative that was selected by Zenyaku Kogyo together with more than 1500 other analogues [24]. ZSTK474 potently inhibits the four isoforms of PI3K, with IC₅₀ values of 16, 44, 5, and 49 nM for PI3K α , β , δ , and γ , respectively, suggesting that it is a pan-PI3K inhibitor [25]. Unfortunately, ZSTK-474 has been withdrawn from clinical trials due to its resistance and on-target/off-tumor side effects [26]. Therefore, the structure of ZSTK-474 needs to be optimized to improve its selective targeting of PI3K.

As shown in **Figure 5**, D. Ross et al., using ZSTK474 as a lead compound, explored compound **27** by displacing a single morpholine group in ZSTK474 with piperazine. Compound **27** showed a 36-fold reduction in its PI3K α (IC₅₀ = 180 nM) and PI3K δ (IC₅₀ = 142 nM) inhibition and a >70-fold reduction in PI3K β (IC₅₀ = 1093 nM) and PI3K γ (IC₅₀ = 1873 nM) inhibition, indicating the sensitivity of this region towards oxygen replacement [27]. However, N-acetylation of **27** to provide **28** showed PI3K isoform inhibition (IC₅₀ = 2.9–21 nM) similar to ZSTK474 (IC₅₀ = 3.9–20.8 nM). To further investigate the role of the morpholine oxygen in these compounds on PI3K inhibition, they

found that when the 2-aminoethyl functional groups were displaced with a single morpholine group of ZSTK474 [28], bifunctional inhibitors **29** and **30** displayed nanomolar inhibition towards PI3K α (IC₅₀ = 130 nM and 107 nM, respectively) and PI3K δ (IC₅₀ = 236 nM and 137 nM, respectively) and low micromolar inhibition for PI3K β and PI3K γ (IC₅₀ = 1.5–3.9 μ M) in enzymatic inhibition assays. Cell viability assays demonstrated that compound **29** showed superior anti-proliferative activities over compound **30** in three tumor-derived cell lines (A375, D54, and SET-2), owing to **29** being able to inhibit downstream AKT and ERK1/2 phosphorylation.

$$F_{2}HC \longrightarrow F_{2}HC \longrightarrow F_{2$$

Figure 5. Chemical structures of PI3K inhibitors of ZSTK474 and its derivatives.

In 2011, Shepherd and co-workers found that the introduction of a methoxy group (31) at the 4'-position of the 2-(difluoromethyl)-1H-benzo[d]imidazol of ZST474 could enhance interaction with the PI3K α protein [29]. Based on this, Hou et al. designed a new 1,3,5-triazine derivative 32 containing semicarbazones as new potential PI3K α inhibitors, 32 also further by replacing a single morpholine group of 31 with piperazine [30]. Further research indicated that compound 32 displayed excellent inhibitory activity with an IC50 value of 0.32 nM against PI3K α . Meanwhile, in the U87-MG human glioblastoma xenograft model assay by intragastric administration, 32 exhibited similar antitumor activity at 20 mg/kg/day compared with ZSTK-474 at 40 mg/kg/day in vivo antitumor efficacy. However, the pharmacokinetic properties of 32 are not ideal, so the compound needs to be further modified structurally to improve its physicochemical properties.

Gedatolisib (PKI-587), another dimorpholino-substituted s-triazine derivative developed by Pfizer, is in a phase III clinical trial as a potential agent for the treatment of acute myeloid leukemia, hormone receptor-positive breast cancer, and HER2-negative breast cancer. Gedatolisib exhibits potent in an in vitro inhibitory activity against both PI3K and mTOR and displays significant antitumor effects in vivo xenograft model. However, it suffers from low selectivity over different PI3K isozymes, leading to possible off-target effects and reducing its therapeutic utility. In addition, gedatolisib must be administered by injection, which may not be suitable as a practical drug for cancer patients. Hence, structural modification of gedatolisib is desirable to develop PI3Ks and mTOR inhibitors with enhanced isozyme selectivity and metabolic stability.

The Venkatesan group has reported the bis-morpholino s-triazine-based compounds as potent dual PI3K/mTOR inhibitors [31]. Gedatolisib was found to have poor plasma levels when administered orally, which may owe to its poor permeability, low clogP (1.24), and high molecular weight (Mw: 615). In order to increase the clogP value and reduce molecular weight, Mansour et al. reported a series of mono-morpholino 1,3,5-triazine derivatives by a single morpholine group in gedatolisib that were replaced with 3-oxa-8-azabicyclo[3.2.1]octane [32]. As shown in **Figure 6**, **33** showed significantly increased potency against PI3K α (IC₅₀ = 8 nM) and mTOR (IC₅₀ = 0.42 nM) and excellent cellular potency against both MDA-361 (IC₅₀ = 22 nM) and PC3mm2 (IC₅₀ = 29 nM) cell lines. Pharmacokinetic studies of **33** showed good oral bioavailability in nude mice after oral administration (10 mg/kg) and a high half-life (>60 min). Metabolite evaluation studies found that the ethylene bridge on the bridged-morpholine group was the main site of metabolism, and its metabolite structure was determined to be **34** [33]. Therefore, compound **33** is an orally effective dual PI3K/mTOR inhibitor.

Figure 6. Chemical structures of PI3K inhibitors of gedatolisib and its derivatives.

Zhang et al. designed and synthesized a series of novel substituted s-triazines bearing a benzimidazole group as PI3K/mTOR inhibitors using the ring closing and scaffold hopping methods [34]. Compound 35 showed remarkable inhibitory activity against PI3K δ with an IC50 value of 2.3 nM and moderate activity against other class I PI3K

isoforms and mTOR (with IC₅₀ values of 14.6, 34.0, 849.0, and 15.4 nM for PI3K α , β , γ , and mTOR, respectively). Therefore, compound **35** could significantly suppress the PI3K/Akt/mTOR signaling pathway.

BKM120 is one of the clinically most advanced pan-PI3K inhibitors, but it interferes with an off-target effect on microtubule polymerization. As shown in **Figure 7**, Wymann et al. have identified that it differs from BKM120 by having only one atom that could be separated into discrete PI3K and tubulin activities [35]. Initially inspired by ZSTK474 and BKM120, they replaced the BKM120 core with s-triazine to yield bimiralisib (36), aiming to maximize compound solubility and bioavailability and avoid microtubule interactions. As expected, 36 displayed potent inhibition against pan-PI3K and showed no microtubule-destabilizing agent activity [36]. Above all, 36 is a highly selective pan-PI3K inhibitor with balanced targeting of mTOR kinase, which passed phase I studies and is now in phase II studies in relapsed and refractory lymphoma and advanced solid tumors [37][38].

Figure 7. Chemical structures of PI3K inhibitors of BKM120 and its derivatives.

Subsequently, Wymann and co-workers mainly focused on the 4-(difluoromethyl)-pyrimidin-2-amine group as an optimized residue for PI3K binding; compound **37** was obtained by substituting 4-(trifluoromethyl)-pyridine-2-amine of **36** with the group. Compound **37** showed low nanomolar affinity PI3K α (IC₅₀ = 2.2 nM) and demonstrated

significant antitumor activity in a mice xenograft model at a concentration almost eight times lower than the parental phase-II inhibitor **35** [39]. Recently, starting from the lead compound **37**, they designed various potent covalent PI3K α inhibitors by targeting the solvent-exposed cysteines at a distance > 10 Å from an ATP-site-directed core group [40]. A number of compounds bearing an acrylamide warhead and different linker modules were synthesized to assess the required warhead reactivity and the spatial trajectory of the Michael acceptor. Among these compounds, compound **38** showed the most PI3K α enzyme inhibitory activity (IC₅₀ = 1 nM) and excellent stability in rat liver microsomes.

Based on the structures of PI3K inhibitor BKM120 and Hh inhibitor vismodegib, the researchers' group reported a novel series of unsymmetrical diaryl ureas as potent inhibitors simultaneously inhibiting PI3K/Akt/mTOR and Hh signalings [41]. After biological activities were examined, compound **39** showed excellent antiproliferative activity against MDA-MB-231, T47D, and MCF-7 cells. The enzymatic activity of **39** (IC₅₀ = 180 nM) against mTOR was of nanomolecular value. Furthermore, based on the structures of the PI3K inhibitor ZSTK474 and the CRBN ligand pomalidomide, the researchers have designed a series of new small-molecule PROTACs for the degradation of PI3K [42]. Compound **40** indicated great potency on PI3K α with an IC₅₀ of 24 nM and could inhibit tumor cell proliferation by inducing autophagy instead of apoptosis or cell cycle arrest.

In 2012, Baselga and Wulf's groups independently found that PI3K inhibition could promote HR deficiency by downregulating BRCA1/2 and could sensitize BRCA-proficient tumors to PARP inhibition, which provides an effective therapy for the combined administration of PI3K and PARP inhibitors to expand the application of PARP inhibitors [43][44]. Based on these works, Xu et al. reported a series of dual PARP/PI3K inhibitors by merging the pharmacophores of PARP and PI3K inhibitors [45]. As shown in Figure 8, structure-activity relationships (SARs) indicate that the R1 group, which serves as a hydrogen-bond donor, could be modified to change the compounds' PI3K inhibitory activity. Several compounds were synthesized and evaluated for PARP-1 and PI3Kα inhibitory activities. As expected, all the remaining compounds showed excellent PARP-1 inhibitory activity in the low nanomolar range. The first round of structural optimization provided compound 41, which displayed appropriate PARP-1 and PI3Kα inhibitory activities with IC₅₀ values of 9.08 and 7.04 nM, respectively. Subsequently, to further optimize the 1,3,5-triazine core by a bicyclic system of tetrahydropyrido[3,4-d]pyrimidine, compound 42 displayed the most promising candidate with IC₅₀ values of 8.22 and 8.25 nM against PARP-1 and PI3Kα. Next, they replaced the phthalazine moiety with a benzofuran-7-carboxamide moiety to obtain compound 43, which exhibited good PARP-1 and PI3Ka inhibitory activity with an inhibition ratio of more than 50% at 10 nM and 100 nM, respectively. After SAR screening and experimental evaluation, the most potent compound was 44 against PARP-1 and PI3K α with IC₅₀ values of 13.8 and 64.0 nM, respectively [46].

Figure 8. Chemical structures of s-triazine simultaneous targeting PI3K and PARP.

The Ras/MEK/ERK and PI3K/Akt/mTOR pathways play important biological functions. MEK inhibition could promote a compensatory activation of PI3K/Akt kinase activity. As shown in **Figure 9**, Ross et al. reported a striazine analog **45** by combining the PI3K inhibitor ZSTK474 with the Raf/MEK inhibitor RO5126766. Bifunctional inhibitor **45** showed good inhibitory activity against MEK1 (IC $_{50}$ = 473 nM) and PI3K (IC $_{50}$ = 172 nM) [47]. However, the corresponding inhibition of pERK1/2 activity was low (35–40%) in two representative human cancer cell lines (A549, PANC-1) at the 5 μ M concentration level. Recently, they continued to report a series of bifunctional MEK1/PI3K inhibitors by covalent linking the PI3K inhibitor ZSTK474 and the MEK inhibitor PD0325901 [48]. Among them, compound **46** showed excellent inhibition against MEK1 (IC $_{50}$ = 0.015 nM) and PI3K (IC $_{50}$ = 191 nM) and displayed a 95% and 67% inhibition of tumor ERK1/2 and Akt phosphorylation, respectively, at 2 h postadministration.

Figure 9. Chemical structures of PI3K and MEK bifunctional inhibitors.

Zhu and co-workers reported 2-(thiophen-2-yl)-1,3,5-triazine derivatives 47 as PI3K and mTOR inhibitors in 2020 $^{[49]}$. By experimental screening, as shown in **Figure 10**, compound 47 showed excellent inhibition activity of cell proliferation against A549, MCF-7, and Hela cancer cell lines with IC₅₀ values of 0.20 \pm 0.05 μ M, 1.25 \pm 0.11 μ M, and 1.03 \pm 0.24 μ M, and potently inhibited PI3K and mTOR kinases with IC₅₀ values of 7.0 and 48 nM, respectively. Further, they also reported the 2-arylurea-1,3,5-triazine derivative 48 as a PI3K and mTOR inhibitor $^{[50]}$. Compound 48 exhibited potent inhibition against PI3K and mTOR kinases with IC₅₀ values of 23.8 and 10.9 nM, respectively. Compound 48 was further evaluated in MCF-7 cells and MCF-7 xenograft models, which showed significant in vitro and in vivo anticancer efficacies. Besides, in order to enhance the mTOR kinase activity, they introduced aryl urea units with various substituents into the triazine core and modified it with arylurea by pyridine to obtain a series of novel thiophene-triazine derivatives bearing arylurea $^{[51]}$. Among them, the inhibitory activity of 49 against PI3K α and mTOR kinase was excellent, with IC₅₀ values of 177.41 and 12.24 nM, respectively, indicating that it was a potential dual PI3K α /mTOR inhibitor.

$$O = S - N$$

PI3K $IC_{50} = 7.0 \text{ nM}$

mTOR $IC_{50} = 48 \text{ nM}$

PI3K $IC_{50} = 23.8 \text{ nM}$

mTOR $IC_{50} = 10.9 \text{ nM}$

PI3K $IC_{50} = 177.41 \text{ nM}$

mTOR $IC_{50} = 12.24 \text{ nM}$

Figure 10. Chemical structures of PI3K and mTOR bifunctional inhibitors.

4. NADP⁺-Dependent Isocitrate Dehydrogenases Inhibition

Metabolic reprogramming is a hallmark of cancer, promoting the initiation and maintenance of tumors. The NADP⁺-dependent isocitrate dehydrogenases (IDH) are critical metabolic enzymes, converting isocitrate to α-ketoglutarate (αKG) in the tricarboxylic acid cycle. IDH mutations occur in multiple tumors, mainly in IDH1 (R132) and IDH2 (R140 and R172). In recent years, several IDH inhibitors have been approved by the FDA. Among them, AG-221 (enasidenib) is a first-in-class, orally available, small-molecule s-triazine IDH2 inhibitor ^[52]. Through high-throughput screening, as shown in **Figure 11**, Su et al. ^[53] reported several s-triazine compounds as effective inhibitors against IDH2^{R140Q}, and the initial hit compound **50** was obtained. Compound **50** showed micromolar inhibitory potency for IDH2^{R140Q} with IC₅₀ values of 1.9 μM. SAR analysis among the s-triazine analogs obtained compound **51**, the first nanomolar inhibitor of IDH2^{R140Q} (IC₅₀ = 7 nM at 16 h). Although compound **51** showed potency in enzymatic and cellular assays, **51** displayed high lipophilicity, resulting in solubility-limited absorption in vivo. Additionally, it emerged poor in vitro liver microsomal stability, which translated to high clearance in vivo. Thus, they continue optimizing the substituents around the s-triazine core. Through the addition of mildly polar substituents with trifluoromethyl pyridine and 2-methyl-2-propanol to provide **52**, AG-221(**52**) showed excellent

potency for 2HG inhibition, improved solubility, low clearance, and good oral bioavailability in vivo in rats, supporting the clinical trials of AG-221 in patients with IDH2 mutation.

Figure 11. Chemical structures of IDH inhibitors of AG-221 and its derivatives.

5. Cyclin-Dependent Kinases Inhibiton

Cyclin-dependent kinases (CDKs) are members of a complex family of heterodimeric serine/threonine protein kinases that play an important role in regulating cell cycle machinery. Depending on their biological functions, CDKs can be divided into cell cycle regulatory CDKs (CDK1, CDK2, CDK4, and CDK6 are included) and transcription-associated CDKs (CDK7-9, CDK11-13, and CDK19 are included) [54]. Murray et al. have identified [1,3,5]triazine-pyridine as a new series of potent CDK inhibitors [55]. Among them, compound **53** (**Figure 12**) displayed excellent inhibitory potency at CDK1 (IC₅₀ = 0.021 μ M), CDK2 (IC₅₀ = 0.007 μ M), and CDK5 (IC₅₀ = 0.003 μ M) and submicromolar potency at CDK4 (IC₅₀ = 0.308 μ M), CDK6 (IC₅₀ = 0.356 μ M), and CDK7 (IC₅₀ = 0.126 μ M). The broad spectrum of potent CDK inhibitory activities and the in vitro and in vivo antitumor efficacy of **53** may render it as a valuable pharmacological tool in elucidating the complex roles of CDK signaling pathways.

Figure 12. Chemical structures of s-triazine targeting CDK.

Positive transcription elongation factor b (PTEFb) is a hetero-dimer of CDK9 and one of four cyclin partners: cyclin T1, cyclin K, cyclin T2a, or cyclin T2b. Deregulated kinase activity of CDK9 of the PTEFb hetero-dimer is associated with cancer. Starting from lead compound **54**, Lucking et al. have optimized structurally characterized by an unuaual benzyl sulfoximine group to obtain **55** [56]. Through kinase selectivity, physicochemical properties, DMPK properties, in vitro and in vivo efficacy, **55** is the first potent and highly selective PTEFb/CDK9 (IC₅₀ CDK9/CycT1: 13 nm, ratio of IC₅₀ values CDK2/CDK9: 100) inhibitor to enter clinical trials for the treatment of cancer.

6. Others

Human adenosine receptors (hARs) can be classified into four subtypes: hA1, hA2A, hA2B, and hA3. All four belong to the G protein-coupled receptor (GPCR) family, and each has a different pharmacological profile, tissue distribution, and function. Langmead et al., in an effort to find a novel hit by virtual screening, reported several compounds containing 1,3,5-triazine that bind to hA1 and hA2A. Subsequently, 1,3,5-triazine analogs were identified as potent human adenosine receptor antagonists for Parkinson's disease $^{[57][58]}$. To modify s-triazine to produce selective ligands for subtypes other than hA2A. Yu et al. designed and synthesized novel 1,3,5-triazine derivatives targeting the hA1 and hA3 adenosine receptors for cancer therapy $^{[59]}$. As shown in **Figure 13**, compounds **56** and **57** showed good binding affinity to both hA1 (IC₅₀ = 139.3 nM, 78.1 nM, respectively) and hA3 AR (IC₅₀ = 55.5 nM, 13.3 nM, respectively) and could inhibit cell viability, leading to cell death in lung cancer cell lines.

Figure 13. Chemical structures of hAR inhibitors **56**, **57**, Hsp90 inhibitors **58**, PD-L1. Inhibitors **59**, and MMP and VEGF inhibitors **60**.

Heat shock protein 90 (Hsp90), a kind of molecular chaperone, is widely expressed and highly conserved in cells. DCZ5248 (**58**, **Figure 13**), a triazine derivative by the Lou group, is a novel Hsp90 inhibitor that directly binds to the Hsp90 protein and does not induce a heat shock response [60], and identified **58** was a dual inhibitor of both Hsp90 and late-autophagy with potent antitumor activity against colon cancer cells in vitro and in vivo.

Programmed cell death protein 1 (PD-1) and its ligand PD-L1 comprise immune checkpoints located on T-cells. The PD-1/PD-L1 axis is hijacked by tumor cells to suppress immune surveillance. At present, inhibition of the PD-

1/PD-L1 axis by monoclonal antibodies has achieved remarkable success and is approved for cancer. However, a number of small molecules targeting PD-L1 have been reported [61]. Marinelli and co-workers have identified 2,4,6-tri- and 2,4-disubstituted 1,3,5-triazines as PD-L1 inhibitors [62]. Among them, compound **59** demonstrated to strongly bind with the PD-L1 protein with a nanomolar IC₅₀ of 0.115 μ M and not to PD-1, as shown in **Figure 13**.

Matrix metalloproteinases (MMPs) are a characteristic family of zinc-dependent endopeptidases that could promote extracellular matrix turnover, tumor growth, angiogenesis, and metastasis. Considering MMP-9 could promote cell migration and trigger the angiogenic switch during carcinogenesis via expression of vascular endothelial growth factor (VEGF), therefore, Khattab et al. developed a series of star-shaped triazine-based dendrimers as MMP and VEGF inhibitors [63]. As shown in **Figure 13**, the most active anticancer dendrimer **60** showed potent MMP-9, -10, and -13 inhibitors with IC_{50} values of 156, 145, and 124 nM, respectively. Furthermore, **60** suppressed the correlated oncogenic mediators, VEGF expression, induced apoptosis (>75%), and inhibited tumor cell migration (~84%) in Caco-2 cells.

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