

Fast Prototyping Microfluidics

Subjects: Automation & Control Systems | Biotechnology & Applied Microbiology | Biochemistry & Molecular Biology

Contributor: Pedro Baptista

Microfluidic (MF) advancements have been leveraged toward the development of state-of-the-art platforms for molecular diagnostics, where isothermal amplification schemes allow for further simplification of DNA detection and quantification protocols. The MF integration with loop-mediated isothermal amplification (LAMP) is today the focus of a new generation of chip-based devices for molecular detection, aiming at fast and automated nucleic acid analysis. Here, we combined MF with droplet digital LAMP (ddLAMP) on an all-in-one device that allows for droplet generation, target amplification, and absolute quantification. This multilayer 3D chip was developed in less than 30 minutes by using a low-cost and extremely adaptable production process that exploits direct laser writing technology in “Shrinky-dinks” polystyrene sheets. ddLAMP and target quantification were performed directly on-chip, showing a high correlation between target concentration and positive droplet score. We validated this integrated chip via the amplification of targets ranging from five to 500,000 copies/reaction. Furthermore, on-chip amplification was performed in a 10 μ L volume, attaining a limit of detection of five copies/ μ L under 60 min. This technology was applied to quantify a cancer biomarker, *c-MYC*, but it can be further extended to any other disease biomarker.

Keywords: digital amplification ; integrated microfluidics device ; loop-mediated isothermal amplification ; oncogenes ; lab-on-a-chip

1. Introduction

The increasing demand for faster and inexpensive diagnostic tools toward integrated devices has been pushed forward by combining microfluidics with molecular detection approaches, both considered key features of lab-on-a-chip (LOC) technology ^[1]. These integrated LOC platforms bring numerous advantages, mostly due to the scale effect such as enhanced sensitivity, throughput, and portability, while reducing the costs and sample volume ^[1].

Nucleic acid amplification technologies (NAATs) benefit from microfluidics integration, which allows for faster and decentralized analysis of molecular disease biomarkers, especially those requiring the detection and quantification of DNA/RNA targets. From these, digital amplification systems are today changing the field of molecular sensing, particularly those that rely on dividing a solution containing the nucleic acid molecules and the reaction components into numerous discrete reaction vessels: droplets ^{[2][3]}. Considering that the number of target molecules inside each droplet follows a Poisson distribution, the absolute quantification of initial target molecules is estimated after endpoint amplification by modeling the fraction of positive droplets ^[4].

So far, polymerase chain reaction (PCR) has been the most widely exploited nucleic acid amplification system, be it on its conventional, quantitative, or digital arrangements ^[5]. Nevertheless, PCR requires thermal cycling with temperatures as high as 95 °C, resulting in complex and cumbersome challenges in the design and operation of chip-based systems. Conversely, isothermal amplification schemes remove the need for temperature control instrumentation, while reducing the amplification time, thus making them more suitable for screening scenarios ^[4]. From these, LAMP is the most commonly used method as it provides an outstanding specificity when compared with other isothermal amplification architectures due to the requirement of four to six primers ^[6]. The potential of LAMP-on-a-chip relies on the simplicity of its thermal requirements, easy integration with sample preparation steps, multiple detection methods and tolerance to biological components present in clinical samples. Although paper-based and digital variants of LAMP-on-a-chip have been proposed ^{[7][8]}, the feasibility of false positive phenomena is a shortcoming concerning the LAMP mechanism, which is outmatched by adding an extra step of sequence recognition, often accomplished with specific fluorescence probes or by restriction enzyme profiling ^[9].

Chip-based microfluidic systems have played an important role in droplet generation technology for biological, biomedical, and diagnostic applications ^{[10][11][12][13]}. Droplet-based microfluidics uses microchannels and immiscible fluids to generate discrete fractions of aqueous solutions. One of the most widely used geometries is the flow-focusing ^{[10][14]}, in which the dispersed phase is symmetrically sheared by the continuous phase in the cross-junction ^{[10][15]}. This architecture yields

highly monodisperse droplets ^{[16][17]} that can be used in a diverse range of applications ^[12]. Devices exploiting this microchannel geometry are also commercially available (e.g., Bio-Rad QX200, Stilla Naica, Raindance Raindrop Plus). Nevertheless, these devices are costly and, for screening more than one target at a time, require multiple apparatus that are not easily integrable. Additionally, the requirement for specific proprietary materials and reagents have been limiting the adoption of this technology into a larger scale.

Microfluidic chip fabrication and optimization have not been easily accessible to everyone. Standard production processes have been based in lithographic techniques ^{[18][19][20]}. These methods pose several limitations in a R&D environment, since they mostly rely on cumbersome, lengthy, and expensive fabrication protocols; additionally, they also present limited design flexibility ^[24]. Recently, several methods have been developed toward fast prototyping of multilayer microfluidic chips ^{[22][23][24][25]}. From these, direct patterning of complex three-dimensional, stacked polystyrene (PS) microfluidic chips has been accomplished ^[26]. By leveraging the inherent shrinkage properties of biaxially pre-stressed thermoplastic sheets, microfluidic channels become thinner and deeper upon heating ^[26]. These advantages make PS a very promising material to fabricate microfluidic devices, although there is a need for inexpensive and fast processes that allow for polystyrene micro-structure fabrication ^[25]. On this, laser direct-writing micromachining has become a promising alternative to lithography, because of its low cost, fast speed, scalability, and non-contact characteristics ^{[27][28]}. CO₂ laser systems have been widely used for rapid production of microfluidic systems with several different polymers (e.g., PS, poly(methyl methacrylate) (PMMA), polydimethylsiloxane (PDMS), polycarbonate (PC), and polytetrafluoroethylene (PTFE)) ^{[29][30][31][32][33]}. While several of these fabrication concepts have already been reported, no research has been pursued toward their application in integrated molecular detection approaches.

Despite all the recent breakthroughs in the molecular mechanisms of cancer, there is still a pressing demand to develop platforms, especially for earlier cancer diagnosis, that combine the specificity and accuracy of molecular methods with portability, user friendly, and cost-effective features such as those presented by LoC technologies ^[34]. Cancer is one of the leading causes of death in the developed world, as estimated by the World Health Organization (WHO) ^[35]. This disease is generally defined as a consequence of multiple genetic events that can exert two broad effects: gain of function mutations leading to oncogenes (e.g., *c-MYC*, *KRAS*, *HER2*, and *SRC*) ^[36] and loss of function mutations resulting in the inactivation of tumor-suppressor genes (e.g., *Tp53*, *RB1*, and *CDKN2A*) ^[37]. Constitutive expression of the proto-oncogene *c-MYC* plays an important role in tumor progression and has been associated with a variety of hematopoietic tumors, leukemias, and lymphomas including Burkitt lymphoma ^[38]. The protein encoded by this gene controls the regulation of the cell cycle and cell growth, activating genomic instability, stimulating angiogenesis, cell transformation, and apoptosis ^[38]. As proof-of-concept, the *c-MYC* proto-oncogene was used in this research.

Herein, we demonstrate a novel approach for multilayered PS-based microfluidic device production that integrates an inexpensive and fast fabrication process based on the shrinkage properties of PS. This integration significantly increases the design flexibility while reducing the prototyping time and cost of the device. As such, we demonstrate the use of this approach for a fully functional droplet-based microfluidic chip and its application with the aim of an integrated ddLAMP assay for absolute target quantification. Additionally, the innovative incorporation of multilayered channels with fully transparent chambers allows for in situ detection of the amplification. The principle here demonstrated can also be applied to absolutely quantify other disease biomarkers and further gene expression analysis.

2. Results and Discussion

2.1. Chip Design and Fabrication Process

Most of the microfluidic devices require complicated fabrication processes such as photolithography, hot embossing, and injection molding. In R&D settings, there is a continuous need to change/optimize chip design, so the improvement of prototyping time and costs is of much value. This microfluidic chip was produced with a laser ablation mechanism in shrinkable PS sheets, followed by thermal bonding to assemble the 3D multilayered chip ^[39]. The reported approach allows for fast (under 30 min) production of functional chips, while eliminating cumbersome fabrication steps. Additionally, it also enables the creation of integrated structures with high design flexibility, thus lowering material expenditure and reducing the costs of chip development (under 50 cents per chip for the proposed design). Even though the fabrication process here describes the results from combining two previously reported techniques: CO₂ laser engraving ^[29] and “Shrinky-Dinks” PS sheets ^[25], what is truly compelling is the advantage given by the multilayered design, which can not only be adapted for several applications, but also enables the creation of fully transparent chambers suitable for on-chip and real-time optical data acquisition. Furthermore, this is the first time that these techniques have been translated into a functional biodetection platform.

We used shrinkable PS sheets because it allowed us to design the channels in a “large” scale version that were then heat-shrunk (~66% of size reduction) (Figure 1A), contributing to an increase in the X and Y resolution while achieving precise control over the channel height. The 3D chips were designed with the aim of an all-in-one integrated platform (Figure 1B) for droplet generation, amplification reaction, and detection. For this, channels on both the top and bottom sections, and a full depth incubation chamber were engraved (Figure 1C). The incubation chamber is enclosed (top and bottom) by a single sheet of PS, increasing the transparency, and reducing the optical defects that could interfere with fluorescence acquisition. Additionally, the bottom overflow channel allowed for a continuous operation without the loss of droplets (droplets float in the oil phase).

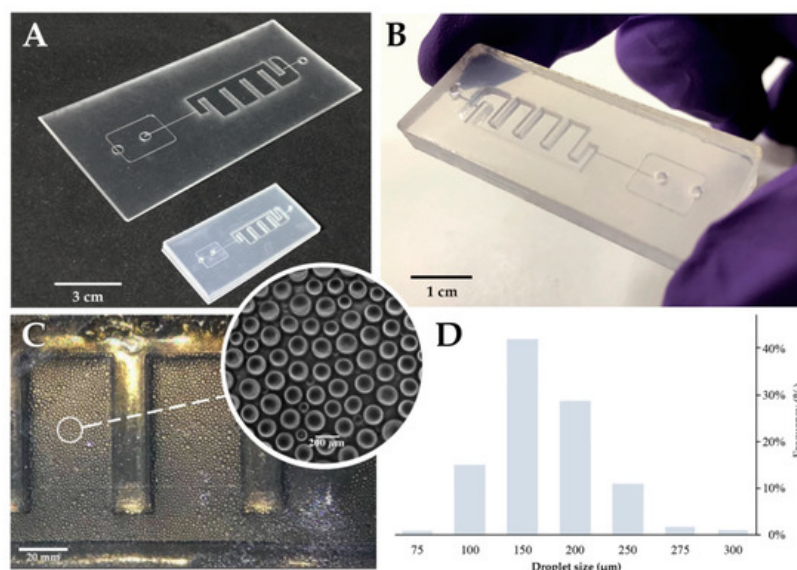


Figure 1. Multilayer Chip production and performance. (A) Multilayer chip before and after the shrinking process; the shrink was isotopically in plane and reduced around 66% of the original size. Additionally, this occurred in an increase in the height over 500%. Scale bar represents 3 cm. (B) Final chip appearance after frame removal. This step also helps seal the edges. Scale bar represents 1 cm. (C) On-chip droplets. Scale bar represents 20 mm; (inset) bright-field microscope image of the produced droplets inside the incubation chamber. Scale bar represents 200 μm. (D) Frequency distribution of the droplet sizes. Droplets present a weighted average size of 170 μm, a coefficient of variation equal to 3% and a standard error of mean (SEM) of 0.27. Plot of the droplet size distribution for all the experiments (n = 6000 droplets). The mean volume of each droplets is ~2.6 nL.

The profile of the microchannel depends on the intensity distribution of the laser beam given by laser power, cutting speed, and number of beam paths on the same channel. Single line designs show a cross section with an extremely sharp Gaussian distribution. To circumvent this effect, a parallel vector design was used and optimized for a 1:1 depth/width channel ratio design (see Figures C1 and C2) (available online at <https://www.mdpi.com/1424-8220/20/6/1624/s1>). The engraved channels present a 200 μm width and height, that are suitable to generate droplets of 150–200 μm in diameter (Figure 1D). Flow rates were optimized to generate droplets with an average size of approximately 170 μm in diameter (corresponding to ~2.6 nL). This size range correlated to those obtained with the standard commercially available devices for digital droplet PCR (dd PCR) [40]. Additionally, the droplets showed a narrow polydispersity and a low coefficient of variation (CV = 3%; defined as CV = standard deviation/mean), which were among the lowest polydispersity obtained with microchannel droplet generation schemes (usually 1–3%) [41].

2.2. Chip-Based Droplet Digital Lamp

Following the design and fabrication of the device, this platform was applied to a digital LAMP approach. Digital amplification techniques are based on the partitioning of the amplification reaction into many small reaction vessels, where amplification occurs. The absolute quantification of the initial sample concentration was achieved through a Poisson statistical analysis of the positive droplets (droplets holding at least one target molecule) versus negative droplets (droplets with zero target molecules) [42], which requires the proper distinction of these two populations.

For this, a threshold line was set to allow scoring the positive from negative droplets based on their fluorescence amplitude. Positive droplets inherently exhibit higher fluorescence than negative droplets due to the presence of Evagreen, a dsDNA binding dye in the amplification reaction. However, the fluorescence amplitude of Evagreen fluctuates depending on the droplet size, amplicon size, amplification efficiency, and primer-dimer formation [43]. Furthermore, larger droplets show an increased probability of encompassing more Evagreen molecules, yielding higher basal fluorescence

[40]. Therefore, to define the threshold for droplet scoring, several reactions without template molecules were assessed (Figure 2A). Data showed a clear correlation between the fluorescence and droplet size, indicating that the threshold line must have a two-dimensional equation with a positive slope value [44].

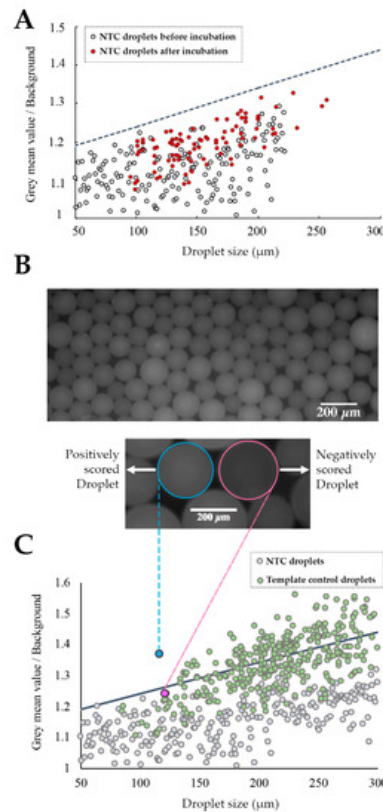


Figure 2. Chip-based ddLAMP. **(A)** Threshold definition; fluorescence measurements of droplets without a template before and after end-point reaction. (○) Represents the NTC droplets before incubation at 65 °C and (●) Represents the NTC droplets after incubation at 65 °C. The threshold (---) equation is $y = 0.001x + 1.14$. **(B, top)** Fluorescence image of on-chip ddLAMP droplets from a template sample after end-point amplification. **(Bottom)** Representative fluorescence image of a (○) negative droplet vs. a (○) positive droplet. Scale bar of 200 μm in length; **(C)** Assessment of the threshold differentiation capability. Negative/positive droplet score. Only droplets with fluorescence above the threshold were scored as positive. (●) Represents droplets from the NTC sample after end-point reaction. (●) Represents droplets from the template sample after end-point LAMP reaction. (●) Represents a negatively scored droplet from the template sample (fluorescence image shown in Figure 2B and (●) represents a positively scored from the template sample (fluorescence image shown in Figure 2B).

Figure 2A demonstrates that droplets from the NTC reaction did not present a significantly increased basal fluorescence at end-point reaction. Thus, the threshold line was iteratively defined taking into account all the attained baseline measurements to distinctively distinguish positive from negative droplets. To confirm if the defined threshold was suitable to discern between negative and positive droplets (Figure 2B), this was applied to the output of both (NTC and template) reactions (Figure 2C). These results show that the defined threshold allows for the differentiation of positive from negative droplets, since only droplets from the template control sample have fluorescence values above the threshold, thus being scored as positive while all of the droplets from the NTC have a fluorescence below the threshold (negatively scored droplets). Nevertheless, some droplets from template control also present fluorescence values below the threshold, which may be attributed to the Poisson distribution of target molecules (i.e., some of these droplets do not hold any target molecules, while some of the positive droplets may contain more than one).

2.3. Device Application on Target Quantification

To evaluate the effectiveness of this chip on a ddLAMP approach for target quantification, serial dilutions of a selected target DNA were analyzed (logarithm of dilution factor ranging from −6 to −12). As a model, *c-MYC* oncogene was used, since this is a valuable biomarker of malignant transformation leading to cancer development [38], however, any other genetic marker could be applied. End-point amplification reactions were conducted for the above-mentioned template dilutions, followed by fluorescence imaging and data processing. Figure 3A shows the increase of the fraction of positive droplets (E_{pos}) as a function of the sample dilution factor. These results were obtained by modeling the fraction of positive droplets to Poisson partitioning statistics (see Figure D1) (available online at <https://www.mdpi.com/1424-8220/20/6/1624/s1>). The increase of E_{pos} occurs due to a rise in the number of target molecules, and consequently, the

probability of a droplet containing at least one target molecule also increases, which in turn leads to an increase in the number of fluorescence droplets after the end-point reaction. The algorithm for scoring positive droplets correlates perfectly to the defined threshold, yielding a very robust correlation to target dilution ($R^2 = 0.98$). This indicates that the designed chip is capable of discriminating the target concentrations for a dynamic range of five orders of magnitude. Nevertheless, the attained dynamic range is only constrained by the number of droplets, which can be easily adjusted by scaling the incubation chamber.

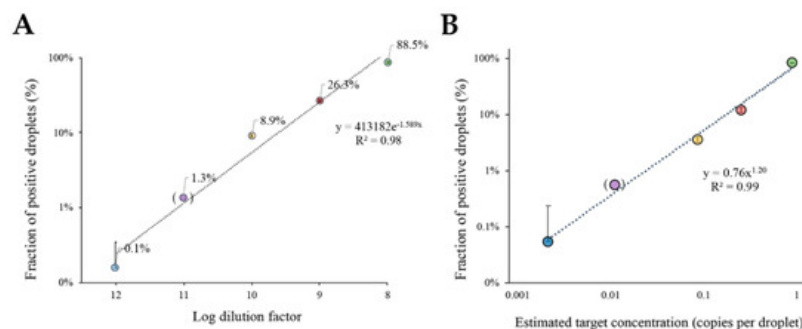


Figure 3. Target DNA quantification with ddLAMP. (A) Fraction of positive events attained with Poisson-based correction for different target dilutions. The fraction of positive events corrected to the Poisson statistics are represented in a logarithmic scale. The trendline presents an exponential equation: $y = 413182e^{-1.589x}$ and a $R^2 = 0.98$. Data were obtained through the measurement of the area of the droplets and corresponding mean grey value with ImageJ software. (B) The plot of the positive droplet fraction against the expected DNA concentration in copies per droplet shows an exponential relationship predicted by Poisson distribution. The trendline for the power adjustment has a $R^2 = 0.99$ and an equation: $y = 0.76x^{1.20}$. Copy number for each dilution was calculated with the equation $C = -\ln(1 - E_{pos})$. For both panels: The error bars correspond to the standard deviation of two independent experiments ($n = 2$) with 400 measured droplets for each reaction; dilution factor 11 (between brackets) was only performed once ($n = 1$). (●) Represents the 10^{-12} dilution, (●) Represents the 10^{-11} dilution (●) Represents the 10^{-10} dilution, (●) Represents the 10^{-9} dilution, and (●) Represents the 10^{-8} dilution.

A similar approach was then used for absolute quantification of *c-MYC* molecules in each sample. The percentage of positive droplets as a function of estimated target DNA concentration provides a robust calibration curve (Figure 3B). The plot shows a high linear correlation factor between the fraction of positive droplets and the predicted number of molecules (per droplet) given by the Poisson distribution. As a result, not only is the chip design suitable for direct ddLAMP, but the developed algorithm also provides a simple and straightforward quantification of the target molecules in the samples. It should be noted that the plot in Figure 3A,B only shows the target dilutions for five orders of magnitude. Taking this into consideration, for each additional order of magnitude, the number of droplets should be increased 10-fold.

Using this chip approach, it was possible to determine the concentration of template target DNA by assessing only a total of 400 droplets. This is an improvement compared to the commercially available devices that require the measurement of thousands of droplets/events for the same dynamic range [45]. As such, the device herein proposed is capable of delivering similar results, but without the need for complex and expensive detection units. In fact, our approach was capable of quantifying the target concentration down to 0.001 copies/droplet corresponding to 5 copies/ μ L.

3. Conclusions

Herein, an integrated chip for digital nucleic acid detection system relying on isothermal LAMP reaction in a digital readout strategy was presented. The fabrication strategy exploits two previously reported techniques. Their combination allows for fast chip development and testing, which is particularly beneficial in R&D settings. In fact, one of the major advantages of this approach is the fast-interactive development of multilayered chips under 30 minutes. Moreover, this is the first time that this production scheme is applied into a working biodetection platform. The proposed design allows for uniform droplet generation, amplification, and measurement of the resulting fluorescence in a single device. This innovative chip-based ddLAMP exhibited the capability of distinguishing between different target concentrations, paving the way for comparative determinations of target DNA/RNA such as those relating to gene expression analysis, which is an essential feature for the quantification of nucleic acid biomarkers in cancer diagnostics.

The laser ablation technique, together with the use of biocompatible shrink PS sheets, allowed for the production of a flow focusing droplet generator, which attained droplets with a coefficient of variation of 3% and average size of $\sim 170 \mu$ m, corresponding to a volume of ~ 2.6 nL. The included fully transparent chamber allows for clear fluorescence image acquisition, contributing to correct droplet score. The current setting allows for the incubation of roughly 6000 droplets,

providing for target quantification in a working range of five decades. Nevertheless, this range is only limited by the number of droplets produced, which in turn is only restricted by the size of the incubation chamber. Further optimizations (chamber size and design) are extremely easy to implement and fabricate, making this an attractive alternative to the traditional fabrication processes for the R&D scenario.

Still, some aspects should be improved in order to enhance the score assessment such as the optimization of droplet generation to remove size dispersity, optimization of the LAMP reaction conditions (i.e., primer concentration), lowering the basal fluorescence of the droplets, and using an external reference dye to normalize the initial fluorescence of the droplets. Moreover, the current setting requires a fluorescence microscope for image acquisition. Nonetheless, in the future, we envision its integration with commercially available portable measurement systems, aiming at a truly standalone biodetection platform. This approach can be further advanced into a multiplex platform, allowing for seamless processing of multiple samples on a single device, thus achieving gene expression analysis capability. Despite this 3-layered design, previous tests showed the capability of fabricating functional devices with up to five layers, increasing the design freedom for any forceable application.

References

1. Francesko, A.; Cardoso, V.F.; Lanceros-Méndez, S. *Lab-On-a-Chip Technology and Microfluidics*; Elsevier: Amsterdam, The Nederland, 2019.
2. Sanders, R.; Huggett, J.F.; Bushell, C.A.; Cowen, S.; Scott, D.J.; Foy, C.A. Evaluation of digital PCR for absolute DNA quantification. *Anal. Chem.* 2011, 83, 6474–6484.
3. Zanolli, L.M.; Spoto, G. Isothermal amplification methods for the detection of nucleic acids in microfluidic devices. *Biosensors* 2013, 3, 18–43.
4. Gansen, A.; Herrick, A.M.; Dimov, I.K.; Lee, L.P.; Chiu, D.T. Digital LAMP in a sample self-digitization (SD) chip. *Lab Chip* 2012, 12, 2247–2254.
5. Kopp, M.U.; De Mello, A.J.; Manz, A. Chemical amplification: Continuous-flow PCR on a chip. *Science* 1998, 280, 1046–1048.
6. Notomi, T.; Okayama, H.; Masubuchi, H.; Yonekawa, T.; Watanabe, K.; Amino, N.; Hase, T. Loop-mediated isothermal amplification of DNA. *Nucleic Acids Res.* 2000, 28, e63.
7. Zhang, H.; Xu, Y.; Fohlerova, Z.; Chang, H.; Iliescu, C.; Neuzil, P. LAMP-on-a-chip: Revising microfluidic platforms for loop-mediated DNA amplification. *TrAC Trends Anal. Chem.* 2019, 113, 44–53.
8. Magro, L.; Escadafal, C.; Garneret, P.; Jacquelin, B.; Kwasiborski, A.; Manuguerra, J.C.; Monti, F.; Sakuntabhai, A.; Vanhomwegen, J.; Lafaye, P.; et al. Paper microfluidics for nucleic acid amplification testing (NAAT) of infectious diseases. *Lab Chip* 2017, 17, 2347–2371.
9. Francois, P.; Tangomo, M.; Hibbs, J.; Bonetti, E.J.; Boehme, C.C.; Notomi, T.; Perkins, M.D.; Schrenzel, J. Robustness of a loop-mediated isothermal amplification reaction for diagnostic applications. *FEMS Immunol. Med. Microbiol.* 2011, 62, 41–48.
10. Sontti, S.G.; Atta, A. Numerical Insights on Controlled Droplet Formation in a Microfluidic Flow-Focusing Device. *Ind. Eng. Chem. Res.* 2019, 59, 3702–3716.
11. Haeberle, S.; Zengerle, R. Microfluidic platforms for lab-on-a-chip applications. *Lab Chip* 2007, 7, 1094–1110.
12. Zhu, P.; Wang, L. Passive and active droplet generation with microfluidics: A review. *Lab Chip* 2017, 17, 34–75.
13. Liu, P.; Mathies, R.A. Integrated microfluidic systems for high-performance genetic analysis. *Trends Biotechnol.* 2009, 27, 572–581.
14. Teh, S.Y.; Lin, R.; Hung, L.H.; Lee, A.P. Droplet microfluidics. *Lab Chip* 2008, 8, 198–220.
15. Liu, H.; Zhang, Y. Droplet formation in microfluidic cross-junctions. *Phys. Fluids* 2011, 23, 82101.
16. Anna, S.L.; Bontoux, N.; Stone, H.A. Formation of dispersions using “flow focusing” in microchannels. *Appl. Phys. Lett.* 2003, 82, 364–366.
17. Krutkramelis, K.; Xia, B.; Oakey, J. Monodisperse polyethylene glycol diacrylate hydrogel microsphere formation by oxygen-controlled photopolymerization in a microfluidic device. *Lab Chip* 2016, 16, 1457–1465.
18. Ren, K.; Zhou, J.; Wu, H. Materials for microfluidic chip fabrication. *Acc. Chem. Res.* 2013, 46, 2396–2406.

19. Kim, P.; Kwon, K.W.; Park, M.C.; Lee, S.H.; Kim, S.M.; Suh, K.Y. Soft lithography for microfluidics: A Review. *Biochip J.* 2008, 2, 1–11.
20. Gale, B.K.; Jafek, A.R.; Lambert, C.J.; Goenner, B.L.; Moghimifam, H.; Nze, U.C.; Kamarapu, S.K. A review of current methods in microfluidic device fabrication and future commercialization prospects. *Inventions* 2018, 3, 60.
21. LaFratta, C.N.; Simoska, O.; Pelse, I.; Weng, S.; Ingram, M. A convenient direct laser writing system for the creation of microfluidic masters. *Microfluid. Nanofluidics* 2015, 19, 419–426.
22. Bartholomeusz, D.A.; Boutté, R.W.; Andrade, J.D. Xurography: Rapid prototyping of microstructures using a cutting plotter. *J. Microelectromechanical Syst.* 2005, 14, 1364–1374.
23. Koppa, V.L.; Crews, N.D. Microfab in a microwave oven: Simultaneous patterning and bonding of glass microfluidic devices. *J. Microelectromechanical Syst.* 2018, 27, 434–439.
24. Cosson, S.; Aeberli, L.G.; Brandenburg, N.; Lutolf, M.P. Ultra-rapid prototyping of flexible, multi-layered microfluidic devices via razor writing. *Lab Chip* 2015, 15, 72–76.
25. Chen, C.S.; Breslauer, D.N.; Luna, J.I.; Grimes, A.; Chin, W.C.; Lee, L.P.; Khine, M. Shrinky-Dink microfluidics: 3D polystyrene chips. *Lab Chip* 2008, 8, 622–624.
26. Grimes, A.; Breslauer, D.N.; Long, M.; Pegan, J.; Lee, L.P.; Khine, M. Shrinky-Dink microfluidics: Rapid generation of deep and rounded patterns. *Lab Chip* 2007, 8, 170–172.
27. Nayak, N.C.; Lam, Y.C.; Yue, C.Y.; Sinha, A.T. CO₂-laser micromachining of PMMA: The effect of polymer molecular weight. *J. Micromechanics Microengineering* 2008, 18, 095020.
28. Urech, L.; Lippert, T. Photoablation of polymer materials. In *Photochemistry and Photophysics of Polymer Materials*; Wiley: Hoboken, NJ, USA, 2010; pp. 541–568.
29. Li, H.; Fan, Y.; Kodzius, R.; Foulds, I.G. Fabrication of polystyrene microfluidic devices using a pulsed CO₂ laser system. *Microsyst. Technol.* 2012, 18, 373–379.
30. Snakenborg, D.; Klank, H.; Kutter, J.P. Microstructure fabrication with a CO₂ laser system. *J. Micromechanics Microengineering* 2004, 14, 182–189.
31. Huft, J.; Da Costa, D.J.; Walker, D.; Hansen, C.L. Three-dimensional large-scale microfluidic integration by laser ablation of interlayer connections. *Lab Chip* 2010, 10, 2358–2365.
32. Qi, H.; Chen, T.; Yao, L.; Zuo, T. Hydrophilicity modification of poly(methyl methacrylate) by excimer laser ablation and irradiation. *Microfluid. Nanofluidics* 2008, 5, 139–143.
33. Tolstopyatov, E.M. Ablation of polytetrafluoroethylene using a continuous CO₂ laser beam. *J. Phys. D. Appl. Phys.* 2005, 38, 1993–1999.
34. Sandbhor Gaikwad, P.; Banerjee, R. Advances in point-of-care diagnostic devices in cancers. *Analyst* 2018, 143, 1326–1348.
35. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA. Cancer J. Clin.* 2018, 68, 394–424.
36. Klein, G. Oncogenes and tumor suppressor genes. *Acta Oncol.* 1988, 27, 427–437.
37. Paige, A.J.W. Redefining tumour suppressor genes: Exceptions to the two-hit hypothesis. *Cell. Mol. Life Sci.* 2003, 60, 2147–2163.
38. Gartel, A.L.; Shchors, K. Mechanisms of c-myc-mediated transcriptional repression of growth arrest genes. *Exp. Cell Res.* 2003, 283, 17–21.
39. Cao, Y.; Bontrager-Singer, J.; Zhu, L. A 3D microfluidic device fabrication method using thermopress bonding with multiple layers of polystyrene film. *J. Micromechanics Microengineering* 2015, 25, 65005.
40. Bio-Rad. Droplet Digital TM PCR Applications Guide; Bio-Rad: Hercules, CA, USA, 2018; pp. 1–10.
41. Baroud, C.N.; Gallaire, F.; Dangla, R. Dynamics of microfluidic droplets. *Lab Chip* 2010, 10, 2032–2045.
42. Vogelstein, B.; Kinzler, K.W. Digital PCR. *Proc. Natl. Acad. Sci. USA* 1999, 96, 9236–9241.
43. Košir, A.B.; Divieto, C.; Pavšič, J.; Pavarelli, S.; Dobnik, D.; Dreo, T.; Bellotti, R.; Sassi, M.P.; Žel, J. Droplet volume variability as a critical factor for accuracy of absolute quantification using droplet digital PCR. *Anal. Bioanal. Chem.* 2017, 409, 6689–6697.
44. Corbisier, P.; Pinheiro, L.; Mazoua, S.; Kortekaas, A.M.; Chung, P.Y.J.; Gerganova, T.; Roebben, G.; Emons, H.; Emslie, K. DNA copy number concentration measured by digital and droplet digital quantitative PCR using certified reference materials. *Anal. Bioanal. Chem.* 2015, 407, 1831–1840.

45. Pinheiro, L.B.; Coleman, V.A.; Hindson, C.M.; Herrmann, J.; Hindson, B.J.; Bhat, S.; Emslie, K.R. Evaluation of a droplet digital polymerase chain reaction format for DNA copy number quantification. *Anal. Chem.* 2012, 84, 1003–1011.
-

Retrieved from <https://encyclopedia.pub/entry/history/show/15849>