

Ganoderma Triterpenoids and Their Bioactivities

Subjects: [Biology](#) | [Medicine, Research & Experimental](#)

Contributor: Mahesh C. A. Galappaththi , Nimesha M. Patabendige , Bhagya M. Premarathne , Kalani K. Hapuarachchi , Saowaluck Tibpromma , Dong-Qin Dai , Nakarin Suwannarach , Sylvie Rapior , Samantha C. Karunarathna

Ganoderma has been used as a traditional medicine in Asian countries to prevent and treat various diseases. Numerous publications are stating that *Ganoderma* species have a variety of beneficial medicinal properties, and investigations on different metabolic regulations of *Ganoderma* species, extracts or isolated compounds have been performed both in vitro and in vivo.

[bioactivity](#)
[bioactive molecules](#)
[drug](#)
[Ganoderma](#)
[Triterpenoids](#)

1. Introduction

Ganoderma P. Karst. 1881 ^[1] is an old polypore genus typified by *Ganoderma lucidum* (Curtis) P. Karst. belonging to the family Ganodermataceae (basidiomycete) ^[2], which has been synonymized with the family Polyporaceae ^[3], with the members normally growing on woody plants and logs ^{[2][4][5]}. The genus was originally reported in the UK ^[6] and is considered to have a worldwide distribution ^{[7][8][9][10][11][12][13][14][15][16][17][18]}. *Ganoderma* species are used for medicinal purposes in China ^{[19][20][21][22]}, Japan, South Korea ^[22] and the French West Indies ^[12]. The common names for *Ganoderma* include Lingzhi, Munnertake, Sachitake, Reishi, and Youngzhi. *Ganoderma* was first recorded in Shennong's Classic of Materia Medica and classified as an upper-grade medicine in medical books ^[23]. Index Fungorum (2022) (<http://www.indexfungorum.org/>, accessed on 6 September 2022) lists 488 records of *Ganoderma* while MycoBank lists 529 records ^[18]. Approximately 80 species of *Ganoderma* are recorded in Chinese Fungi ^[24], of which *G. lucidum* and *G. sinense* are described as medicinally beneficial macrofungi in Chinese medicine ^[25]. However, other species, such as *G. capense*, *G. cochlear* and *G. tsugae* also play an important role in traditional folk medicine. In addition, pharmacological studies have also used the extract and chemical constituents of other *Ganoderma* species ^{[22][26][27][28][29][30]}.

The chemical constituents and the biological activities of 25 species of *Ganoderma*, namely *G. amboinense*, *G. annulare*, *G. applanatum* (synonym *G. lipsiense*), *G. australe*, *G. boninense*, *G. capense*, *G. carnosum*, *G. casuarinicola*, *G. cochlear*, *G. colossus*, *G. concinnum*, *G. ellipsoideum*, *G. fornicatum*, *G. hainanense*, *G. lucidum*, *G. mastoporum*, *G. neo-japonicum*, *G. orbiforme*, *G. pfeifferi*, *G. resinaceum*, *G. sinense*, *G. theaecola* (as *theaecolum*), *G. tropicum*, *G. tsugae* and *G. weberianum*. Several chemical constituents such as ganoderic acids, lucidenic acids, 12-hydroxyganoderic acid, ganorbiformin, lucidimines ^[31] and other compounds have been reported from various *Ganoderma* species during recent decades ^{[29][32][33][34][35]}.

Triterpenes, polysaccharides and peptidoglycans are one of the major types of bioactive substances [36][37] responsible for the various biological activities of several species in *Ganoderma* (e.g., *Ganoderma lucidum*). Triterpenoids and ganoderic acids that play a critical role in pharmacological activities are also present in *G. applanatum* and *G. orbiforme* [33][36][38][39][40][41][42]. Besides *G. applanatum* [43][44], *G. colossus* [45] and *G. pfeifferi* [46], *G. resinaceum* [45] have also been identified as potential *Ganoderma* natural antioxidants.

G. amboinense [47], *G. annulare* [48], *G. applanatum* [49][50][51], *G. boninense* [52], *G. calidophilum* [53], *G. capense* [54], *G. carnosum* [55], *G. cochlear* [56][57][58], *G. colossus* [59], *G. concinnum*, *G. neo-japonicum* [60], *G. fornicatum* [61], *G. hainanense* [62], *G. leucocontextum* [63], *G. lucidum* [64][65][66][67], *G. mastoporum* [68], *G. mbrekobenum* [69], *G. resinaceum* [70], *G. sinense* [71][72][73], *G. theaecola* (as *theaecolum*) [74], *G. tropicum* [75], *G. tsugae* [76], *G. weberianum* [77] were reported to contain bioactive compounds of great interest.

The members of the genus *Ganoderma* are rich in novel “mycochemicals”, including polysaccharides [78], steroids, fatty acids, phenolic compounds, vitamins, amino acids and triterpenoids [69][79][80][81]. *Ganoderma* polysaccharides are a hot topic in the medicinal mushroom research field [78][82][83][84]. Although polysaccharides are found to be one of the main bioactive constituents, their high molecular weight and complex structure limit their use in the drug market [85]. The *Ganoderma* polysaccharides are well known for their diverse bioactivities such as antitumor, immunomodulatory, anti-hypertensive, antidyslipidemic and hepatoprotective activities. *Ganoderma* polysaccharides have been developed into medicines, dietary supplements, healthy foods, treat and prevent diseases, and are continuing to serve as important leads in modern drug discovery [78][86][87][88].

Taxonomic Studies of *Ganoderma*

The word *Ganoderma* is derived from the Greek words “Gano”, meaning “shiny”, and “derma”, meaning “skin” [22][89]. Originating from the tropics and recently extending its range into temperate zones [90], *Ganoderma* is an old genus with a taxonomic history dating back to 1881, and the Finnish mycologist P. A. Karsten erected the genus to place a single species, *Polyporus lucidus* [= *Ganoderma lucidum* (Curtis: Fr.) P. Karst.] [1][91]. *Ganoderma lucidum* is the type species of the genus *Ganoderma*, which was originally described as from the UK and later found to have a worldwide distribution [6]. A number of *Ganoderma* species morphologically closely related and belonging to *G. lucidum* complex viz. *G. multipileum* Ding Hou 1950 [92], *G. sichuanense* J.D. Zhao & X.Q. Zhang 1983 [93] and *G. lingzhi* Sheng H. [94][95] from China, *G. resinaceum* Boud. 1889 [96] from Europe, and *G. oregonense* Murrill 1908, *G. sessile* Murrill 1902, *G. tsugae* Murrill 1902 and *G. zonatum* Murrill 1902 [97][98] from the USA, have been described as from all over the world, and are mainly characterized by laccate pileus. Zhou et al. [99] considered 32 collections belonging to the *G. lucidum* complex from Asia, Europe and North America in terms of their morphology and phylogeny as derived from analyses of four protein-coding genes viz. Internal transcribed spacer (ITS), translational elongation factor 1- α (*tef1- α*) and retinol-binding protein 1 and 2 (*rpb1*, *rpb2*). Different molecular techniques have been used to study the genetic diversity in *Ganoderma*, such as amplified fragment length polymorphism, isozyme analysis, inter simple sequence repeat, random amplified polymorphic DNA, single-nucleotide polymorphism, restricted fragment length polymorphisms, sequence-related amplified polymorphism, single-strand conformational polymorphism and sequence characterized amplified region [100]. These different

molecular identifications used in different taxonomic classifications of *Ganoderma* have caused great improvements and provide information for the further research of *Ganoderma* [18][90][101].

Ganoderma has long been treated as one of the most important medicinal fungi worldwide [39], and laccate species of *Ganoderma* have been considered for centuries [102], i.e., *G. lucidum* complex have been used as medicinal mushrooms in traditional Chinese medicine [103]. Anyhow, the species concept of the *G. lucidum* complex lacks agreement in morphology, and the taxonomy of this species complex is thus problematic, and this ultimately limits both further research on this complex and their medicinal usefulness. For example, the widely used medicinal species in biochemical and pharmaceutical studies have been assumed to be *G. lucidum*, but evidence has emerged that this medicinal species is, in fact, a different species [104] and was described as *G. lingzhi* [94].

The taxonomic position of the *G. lucidum* complex has long been subjected to debate [9][18][90][101][105] and different opinions have been expressed regarding the members and their validity in the complex. Haddow [106] treated *G. sessile* as a synonym of *G. resinaceum*, while Overholts [107] pointed out that *G. lucidum* should be the correct name of the specimens classified as *G. sessile*. *Ganoderma lucidum* has also been considered as the correct name over its later synonym *G. tsugae* [106][108]. However, with the support of mating tests, Nobles [109] pointed out that the specimens classified as *G. lucidum* in the USA represented *G. sessile*. All names of the *G. lucidum* complex in East Africa were parsimoniously treated as the “*G. lucidum* group”, because of the lack of morpho-taxonomic solution to the problem in this complex [7]. Asian specimens classified as *G. lucidum* were divided into two clades; both were separated from European *G. lucidum* [110], with one clade being composed of tropical collections represented by *G. multipileum*, while the other clade was unknown [110], yet later recognized as *G. sichuanense* [111]. It was found that the holotype of *G. sichuanense* was not conspecific with the unknown clade, and the unknown clade was identified as a new species, *G. lingzhi*, which also is the most widely cultivated species in China [94]. Meanwhile, the distribution of genuine *G. lucidum* in China was also confirmed [94][112]. The taxonomy of the *G. lucidum* complex remains problematic even after several decades of debates [99]. Most of the studies previously conducted were focused on the species in a continent or specific region [94][111], or the phylogeny was described with low resolution to certain clades [104][112]. On the other hand, a strong phylogeny together with species originally described as from the USA is greatly needed, as most of these species are old and never referred to in any phylogenetic analyses.

2. Triterpenoids

2.1. Biosynthesis of Triterpenoids

Triterpenoids belong to a large and structurally diverse class of natural products [113]. Basidiomycetes are considered a main source for triterpenoids. Compared to plants, very few types of triterpenoid skeletons have been reported in basidiomycetes, thus more research is needed [114]. Triterpenoids extracted from *Ganoderma* spp. are named as *Ganoderma* triterpenoids (GTs) [113].

Ganoderma triterpenoids (GTs) isolated from the fruiting bodies, cultured mycelia and basidiospores of *Ganoderma* [115][116][117] belong to the lanostane-type triterpenoids and are one of the major chemical constituents in *Ganoderma*. Studies have confirmed that GTs are biosynthesized using isoprenoid pathways [118][119]. This pathway was considered to start from acetyl-coenzyme A and termed as a Mevalonate pathway (MVA) [120]. The MVA pathway (**Figure 1**) is one of the important metabolic pathways that can be divided into four main processes: conversion, construction, condensation, and postmodification [35]. The initial step is the transformation of acetyl-coenzyme A to isopentenyl pyrophosphate (IPP). Then the activities of different prenyltransferases produce farnesyl pyrophosphate (FPP), geranyl pyrophosphate (GPP) and geranylgeranyl pyrophosphate (GGPP), which are higher-order terpenoid building blocks from this IPP precursor. Then, these junction mediators can self-condense, and are also used in alkylation processes to produce prenyl side chains (for a range of nonterpenoids) or create a ring (to build the basic skeletons of triterpenoids). Ultimately, oxidation and reduction reactions, conjugation, isomerization or other secondary reactions magnify the distinctive characteristics of the triterpenoids [35][121][122].

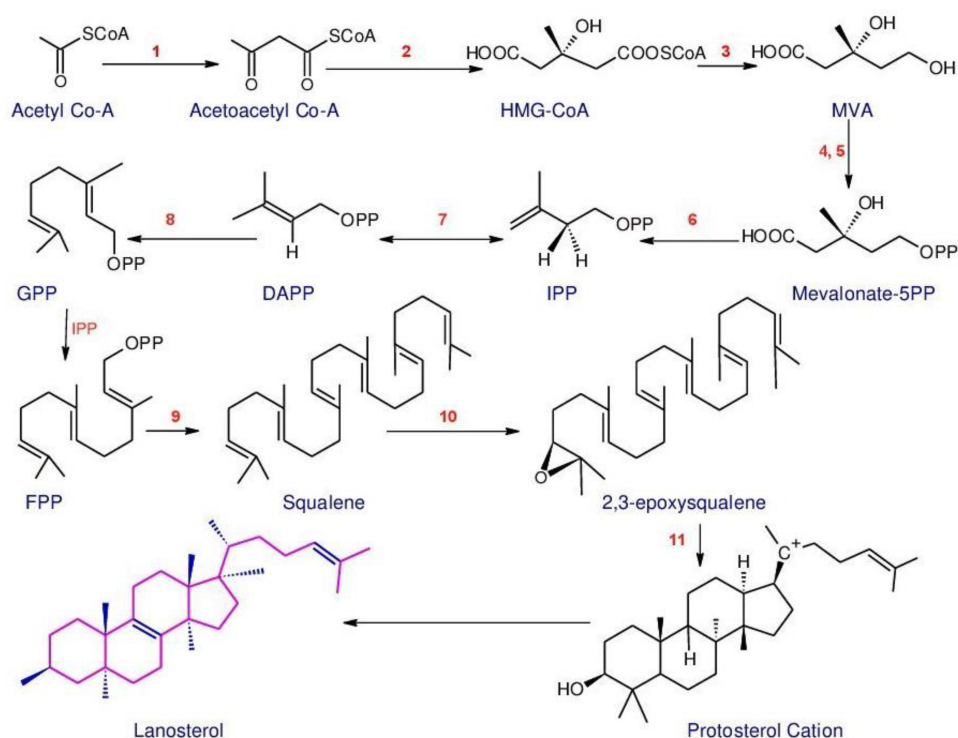


Figure 1. Outline of the MVA and lanostane-type triterpenoids biosynthesis. Enzymes involved in the pathway are: 1. Acetyl-CoA acetyltransferase, AACT; 2. 3-Hydroxy-3-methylglutaryl-CoA synthase, HMGS; 3. 3-Hydroxy-3-methylglutaryl-CoA reductase, HMGR; 4. Mevalonate kinase, MK; 5. Phosphomevalonate kinase, MPK; 6. Phosphomevalonate decarboxylase, MVD; 7. Isopentenylidiphosphate isomerase, IDI; 8. Farnesyl diphosphate synthase, FPPs; 9. Squalene synthase, SQS; 10. Squalene monooxygenase, SE; 11. 2,3-Oxidosqualene-lanosterol cyclase, OSC (The structures were redrawn in ACD/ChemSketch: Freeware: 2012).

The relationship between the content of *Ganoderma* triterpenoids and the expression levels of major genes has been evaluated in several studies. These kinds of studies may provide valuable data for studying the role of key

genes and, finally, for raising the production of triterpenoids. According to Wang et al. [114], the most comprehensive studies on *Ganoderma* triterpenoids were concentrated on *G. lucidum*. A considerable number of essential enzyme genes are involved in the production of *G. lucidum* triterpenoids during the biosynthesis. Liu et al. [123] inspected the *G. lucidum* genes in the “terpenoid backbone biosynthesis (map00900)” pathway and discovered that the genes are solely spread in the MVA pathway, not the MEP/DOXP (methylerythritol 4-phosphate/deoxyxylulose 5-phosphate) pathway. Meanwhile, several genes in this pathway have been cloned in *G. lucidum*, including 3-hydroxy-3-methylglutaryl-CoA reductase (HMGR) [124], Farnesyl diphosphate synthase (FPPs) [125], squalene synthase (SQS) [126], and lanosterol synthase (also namely 2,3-oxidosqualene lanosterol cyclase, OSC) [123]. These observations demonstrated that triterpenoid backbone biosynthesis could only be accomplished via the MVA pathway at the genome level in fungi.

Scientists have performed a large number of studies to find out key enzyme genes involved in increasing the yield of ganoderic acid (GA). Ye et al. [127] examined the effect of salicylic acid (SA) and calcium ions on the biosynthesis of triterpenoids in *G. lucidum* through spraying during the fruiting stage. Calcium ions had no effect on the production of triterpenoids because the six key triterpenoid biosynthetic genes did not respond. However, SA increased triterpenoid content by 23.32% compared to the control, and the combined induction of both increased triterpenoid content by 13.61% compared to the control since in the case of SA and the combined induction of both on the six-triterpenoid biosynthetic genes were up-regulated [127]. Fei et al. [128] have utilized the homologous farnesyl diphosphate synthase gene to overexpress GA (which actually increases the generation of GA) to determine the function of MVD in the biosynthesis of GA. However, it also increases expression of squalene synthase (SQS) and lanosterol synthase (LS).

According to the results of Shi et al. [129], it was revealed that MVD plays a key role in the biosynthesis of GA. The SE gene was cloned from *G. lucidum* and overexpressed to examine the impact on the biosynthetic pathway of GA. The results indicated that the SE gene promotes the biosynthesis of GA. In addition, SE and HMGR genes were simultaneously expressed, with the co-expressed strain having a higher acid content than the single expressed strain, which demonstrated that the co-expression of the two genes stimulated biosynthesis of GA [101][130][131]. LS is a key enzyme of the MVA synthesis pathway and is at the second branch point, [101][114][130][131] whilst it was revealed that the overexpression of LS increases the content of GA [101][130][131]. As a summary, the fundamental enzyme gene in the biosynthesis pathway of GA is intensely involved in the amount of GA.

2.2. Structures and Bioactivities of Triterpenoids

Triterpenes are a major class of widely dispersed secondary metabolites in nature [132]. Triterpenoids structures with a carbon skeleton are considered to be derived from the acyclic precursor squalene [133]. More than 30,000 structures of triterpenes [134] such as dammarane, lanostane, lupine, oleanane and ursane types have been isolated and identified [135].

The structures of triterpenoids isolated from *Ganoderma* spp. are complicated. These compounds consist with lanostane carbon skeleton and pentacyclic triterpenoids (**Figure 2**). According to the number of carbon atoms in

their skeleton, GTs can be divided into three types viz. C30, C27 and C24 [136]. On the basis of the substituent groups, they are classified into different groups such as triterpenoid acids, triterpenoid alcohols and triterpenoid lactones [136].

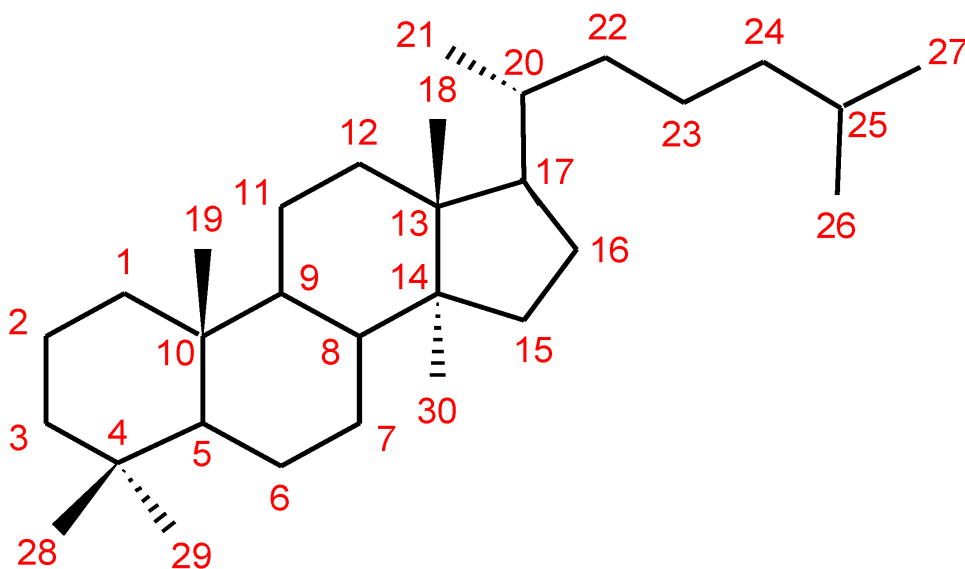


Figure 2. Chemical structure of lanostane triterpene.

With the development of separation techniques and extraction methods, structurally diverse triterpenoids were isolated [137][138], which were formed through oxidation, reduction, cleavage, rearrangement and cyclization within MVA pathway [139]. It is known that the bioactivity of GTs has long been the research focus and that new changes of structures can affect their bioactivities [29][140]. Thus, researchers summarize the triterpenoids of 25 species of *Ganoderma* and classify them into seven types on the basis of the number and type of skeleton carbons (i.e., C31, C30, C29, C27, C25, C24, and rearranged novel skeleton).

2.2.1. C30 Triterpenoids

The majority of GTs was C30 triterpenoids on the basis of the biosynthetic pathway of GTs. Because of the oxidation and reduction processes, their structures should be divided into six groups: 8,9-ene, 8,9-dihydro, 8,9-epoxy, 7(8),9(11)-diene, triterpenoid saponins and rearranged novel skeleton triterpenoids.

8,9-ene-triterpenoids

This type of GT is the lanostane-type triterpenoid with a double bond between C-8 and C-9. In addition, C-3, C-7, C-11 and C-15 were generally substituted by the hydroxyl or carbonyl groups. The minority of compounds possessed hydroxyl or carbonyl groups at C-12. Different oxidation and reduction can happen in the side chain. Except the above structural features, compounds ganodermacetal (**44**) and methyl ganoderate A acetonide (**45**) possessed an uncommon acetonide unit. Compound **44** was isolated from the basidiomycete *G. amboinense*, and was a natural product, but not an artefact, which resulted from the acetalization of native ganoderic acid C (**3**) and with acetone not being used during the isolation procedure [141]. However, compound **45** isolated from the fruiting

bodies of *G. lucidum* was reported to be most likely not of natural origin due to the utilization of acetone during the isolation procedure [142].

Several studies on the bioactivity of triterpenoids showed that ganoderic acids had significant biological activities [95][143][144][145][146][147] (Table 1). Ganoderic acid DM (50) displayed stronger 5 α -reductase inhibitory activity (IC₅₀ value of 10.6 μ M) than the positive control (α -linolenic acid, 116 μ M). Meanwhile, compared to its methyl derivative, the inhibitory activities of 5 α -reductase at 20 μ M were 55% and 3% for 50 and its derivative, suggesting that the carboxyl group of the side chain for 50 is essential to elicit the inhibitory activity [143]. Liu et al. [144] evaluated the structure–activity relationship for inhibition of 5 α -reductase using ganoderic acid A (1), B (2), C (3), D (19), I (5) and DM (50). The results showed that the presence of the carbonyl group at C-3, and of the α,β -unsaturated carbonyl group at C-26, was characteristic of almost all inhibitors.

Table 1. 8,9-double-bond triterpenoids and bioactivities from *Ganoderma*.

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
1.	Ganoderic acid A	Promising anticancer agent (via potent inhibitory effect on JAK/STAT3 pathway)	<i>G. lucidum</i> , <i>G. tsugae</i>	[148][149] [150]
2.	Ganoderic acid B	Moderately active inhibitor against HIV-1 PR (0.17 mM)	<i>G. lucidum</i> , <i>G. tsugae</i>	[148][149] [151]
3.	Ganoderic acid C	Suppressed LPS-induced TNF- α (IC ₅₀ = 24.5 μ g/mL) production through down-regulating MAPK, NF-kappa B and AP-1 signaling pathways in macrophages	<i>G. lucidum</i> , <i>G. tsugae</i>	[148][152]
4.	Ganoderic acid G	Antinociceptive effect	<i>G. lucidum</i>	[153][154]
5.	Ganoderic acid I	Cytotoxicity against Hep G2 cells (IC ₅₀ = 0.26 mg/mL), HeLa cells (IC ₅₀ = 0.33 mg/mL), Caco-2 cells (IC ₅₀ = 0.39 mg/mL)	<i>G. lucidum</i>	[153][155]
6.	Ganolucidic acid A	-	<i>G. lucidum</i>	[156]
7.	Ganolucidic acid B	-	<i>G. lucidum</i>	[157]
8.	Methyl ganoderate M	-	<i>G. lucidum</i>	[158]
9.	Methyl ganoderate N	-	<i>G. lucidum</i>	[158]
10.	Methyl ganoderate O	-	<i>G. lucidum</i>	[158]
11.	Methyl ganoderate K	-	<i>G. lucidum</i>	[158]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
12.	Compound B9	-	<i>G. lucidum</i>	[158]
13.	Methyl ganoderate H	-	<i>G. lucidum</i>	[158]
14.	Ganoderic acid α	Anti-HIV protease (0.19 mM)	<i>G. lucidum</i>	[151]
15.	3-O-acetyl ganoderic acid B	-	<i>G. lucidum</i>	[159]
16.	Ethyl 3-O-acetyl ganoderate B	-	<i>G. lucidum</i>	[159]
17.	3-O-Acetyl ganoderic acid K	-	<i>G. lucidum</i>	[159]
18.	Ethyl ganoderate J	-	<i>G. lucidum</i>	[159]
19.	Ganoderic acid D	Cytotoxicity against HeLa cells (17.3 μ M), 5 α -reductase inhibition—NE	<i>G. lucidum</i> , <i>G. applanatum</i> , <i>G. tsugae</i>	[144][148] [160][161] [162]
20.	Ganoderic acid F	Cytotoxicity against HeLa cells (19.5 μ M)	<i>G. lucidum</i>	[160][162]
21.	Ganoderic acid E	Cytotoxicity against tumor cell lines [Hep G2 (1.44 $\times 10^{-4}$ μ M), HepG2,2,15 (1.05 $\times 10^{-4}$ μ M), κ B—NE, CCM2 (31.25 μ M), p388 (5.02 μ M)]	<i>G. lucidum</i> , <i>G. tsugae</i>	[148][160] [163]
22.	Ganoderic acid Df	Human aldose reductase inhibitory activity (22.8 μ M/mL)	<i>G. lucidum</i>	[164]
23.	Ganosporeric acid A	-	<i>G. lucidum</i>	[165]
24.	Ganohainanic acid A	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
25.	Acetyl ganohainanic acid A	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
26.	Ganohainanic acid B	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
27.	Ganohainanic acid C	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
28.	Ganohainanic acid D	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
29.	Acetyl ganohainanic acid D	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
30.	Methyl ganoderate D	-	<i>G. lucidum</i>	[166][167]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources Ganoderma Species	References
31.	Methyl ganoderate E	-	<i>G. lucidum</i>	[168]
32.	Methyl ganoderate F	Inhibitory effects on EBV-EA induction (289 mol ratio/32 pmol TPA)	<i>G. lucidum</i>	[168][169]
33.	12 β -Acetoxy-3 β ,7 β - dihydroxy-11,15,23- trioxolanost-8-en-26-oic acid butyl ester	Antimicrobial [<i>Staphylococcus aureus</i> ATCC 6538 (68.5 μ M) and <i>Bacillus subtilis</i> ATCC6633 (123.8 μ M)]	<i>G. lucidum</i>	[170]
34.	12 β -acetoxy- 3,7,11,15,23- pentaioxolanost-8-en-26- oic acid butyl ester	Antimicrobial—NE	<i>G. lucidum</i>	[170]
35.	n-Butyl ganoderate H	Selective cholinesterase inhibition	<i>G. lucidum</i>	[142]
36.	Butyl ganoderate A	Cytotoxicity against 3T3-L1 cells —NE	<i>G. lucidum</i>	[167]
37.	Butyl ganoderate B	Cytotoxicity against 3T3-L1 cells —NE	<i>G. lucidum</i>	[167]
38.	3 β ,7 β ,15 β -Trihydroxy- 11,23-dioxo-lanost- 8,16-dien-26-oic acid	Anti-AChE—NE	<i>G. tropicum</i>	[171]
39.	3 β ,7 β ,15 β -Trihydroxy- 11,23-dioxo-lanost-8,16- dien-26-oic acid methyl ester	Anti-AChE (15.72%)	<i>G. tropicum</i>	[171]
40.	3 β ,15 β -Dihydroxy- 7,11,23-trioxo-lanost- 8,16-dien-26-oic acid methyl ester	Anti-AChE—NE	<i>G. tropicum</i>	[171]
41.	Ganoderenic acid G	-	<i>G. applanatum</i>	[172]
42.	Ganoderenic acid F	-	<i>G. applanatum</i>	[172]
43.	Methyl ganoderate I	-	<i>G. applanatum</i>	[172]
44.	Ganodermacetal	Toxic activity against brine shrimp larvae	<i>G. amboinense</i>	[141]
45.	Methyl ganoderate A acetanide	Anti-AChE (18.35 μ M), anti-BChE—NE	<i>G. lucidum</i>	[142]
46.	Ganoderic acid Z	Cytotoxicity	<i>G. lucidum</i>	[173]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources Ganoderma Species	References
47.	Ganoderic acid W	Cytotoxicity	<i>G. lucidum</i>	[173]
48.	Ganoderic acid V	Cytotoxicity	<i>G. lucidum</i>	[173]
49.	Ganoderic acid U	-	<i>G. lucidum</i>	[174]
50.	Ganoderic acid DM	5 α -Reductase inhibition (10.6 μ M), anti-androgen and anti-proliferative activities, osteoclastogenesis inhibitor, inhibits prostate cancer cell growth, inhibits breast cancer cell growth	<i>G. lucidum</i> , <i>G. sinense</i>	[95][145] [146][175] [176][177]
51.	7-Oxo-ganoderic acid Z ₂	-	<i>G. resinaceum</i>	[178]
52.	7-Oxo-ganoderic acid Z ₃	-	<i>G. resinaceum</i>	[178]
53.	Ganoderic acid GS-1	Anti-HIV protease (58 μ M)	<i>G. sinense</i>	[177]
54.	Ganoderic acid GS-2	Anti-HIV protease (30 μ M)	<i>G. sinense</i>	[177]
55.	Ganoderic acid GS-3	Anti-HIV protease—NE	<i>G. sinense</i>	[177]
56.	Ganoderic acid Ma	-	<i>G. lucidum</i>	[179]
57.	Ganoderic acid Mb	-	<i>G. lucidum</i>	[179]
58.	Ganoderic acid Mc	-	<i>G. lucidum</i>	[179]
59.	Ganoderic acid Md	-	<i>G. lucidum</i>	[179]
60.	Ganoderic acid Mg	-	<i>G. lucidum</i>	[174]
61.	Ganoderic acid Mh	-	<i>G. lucidum</i>	[174]
62.	Ganoderic acid Mi	-	<i>G. lucidum</i>	[174]
63.	Ganoderic acid Mj	-	<i>G. lucidum</i>	[174]
64.	3 α ,22 β -Diacetoxy-7 α -hydroxyl-5 α -lanost-8,24 E -dien-26-oic acid	Cytotoxicity against 95D (IC ₅₀ = 23 μ M/mL), HeLa human tumor cell lines (IC ₅₀ = 14.7 μ M/mL)	<i>G. lucidum</i> (mycelia)	[180]
65.	7-O-Ethyl ganoderic acid O	Cytotoxicity against 95D (46.7 μ M), HeLa cells (59.1 μ M)	<i>G. lucidum</i>	[181]
66.	Ganorbiformin B	-	<i>G. orbiforme</i>	[36]
67.	Ganorbiformin C	-	<i>G. orbiforme</i>	[36]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources Ganoderma Species	References
68.	Ganorbiformin D	Cytotoxicity (against NCIH187, MCF-7, and κB—NE), nonmalignant Vero cells, antimalarial, antitubercular—NE	<i>G. orbiforme</i>	[36]
69.	Ganorbiformin E	Cytotoxicity against NCIH187 (70 μM), MCF-7, κB—NE, nonmalignant Vero cells, antimalarial, antitubercular—NE	<i>G. orbiforme</i>	[36]
70.	Ganorbiformin F	Cytotoxicity against NCIH187 (44 μM), MCF-7—NE and κB (63 μM), nonmalignant Vero cells (36 μM), antimalarial, antitubercular—NE	<i>G. orbiforme</i>	[36]
71.	7β,23ξ-Dihydroxy-3,11,15-trioxolanosta-8,20E(22)-dien-26-oic acid	-	<i>G. applanatum</i>	[38]
72.	Methyl ganoderenate D	-	<i>G. applanatum</i>	[38]
73.	3β,7β,20,23ξ-Tetrahydroxy-11,15-dioxolanosta-8-en-26-oic acid	-	<i>G. applanatum</i>	[38]
74.	7β,20,23ξ-Trihydroxy-3,11,15-trioxolanosta-8-en-26-oic acid	-	<i>G. applanatum</i>	[38]
75.	Ganoderic acid L	-	<i>G. lucidum</i>	[182]
76.	Methyl ganoderate L	-	<i>G. lucidum</i>	[182]
77.	Ganolucidic acid ya	PXR-mediated CYP3A4 expression—NE	<i>G. sinense</i>	[183]
78.	Ganolucide F	PXR-mediated CYP3A4 expression	<i>G. sinense</i>	[183]
79.	Ganolucidic acid D	Cytotoxicity on tumor growth cells—NE	<i>G. lucidum</i>	[182][184]
80.	Methyl ganolucide D	-	<i>G. lucidum</i>	[182]
81.	Hainanic acid A	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
82.	Hainanic acid B	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
83.	Ganoderic acid γ	Cytotoxicity against tumor cell growth Meth-A (ED ₅₀ = 15.6 μg/mL), LLC—NE	<i>G. lucidum</i>	[184]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources Ganoderma Species	References
84.	Ganoderic acid δ	Cytotoxicity against tumor cell growth Meth-A and LLC—NE	<i>G. lucidum</i>	[184]
85.	Ganoderic acid ε	Cytotoxicity against tumor cell growth Meth-A (ED ₅₀ = 12.2 µg/mL), LLC—NE	<i>G. lucidum</i>	[184]
86.	Ganoderic acid ζ	Cytotoxicity against tumor cell growth Meth-A and LLC—NE	<i>G. lucidum</i>	[184]
87.	Ganoderic acid η	Cytotoxicity against tumor cell growth Meth-A and LLC—NE	<i>G. lucidum</i>	[184]
88.	Ganoderic acid θ	Cytotoxicity against tumor cell growth Meth-A (ED ₅₀ = 5.7 µg/mL), LLC (ED ₅₀ = 15.2 µg/mL)	<i>G. lucidum</i>	[184]
89.	Ganoderiol G	-	<i>G. lucidum</i>	[185]
90.	Ganoderiol H	-	<i>G. lucidum</i>	[185]
91.	Ganoderiol I	-	<i>G. lucidum</i>	[185]
92.	24 <i>S</i> ,25 <i>R</i> -Dihydroxy-3,7-dioxo-8-en-5 <i>α</i> -lanost-26-ol	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
93.	Ganoderone A	Antiviral: influenza A—NE, HSV (0.3 µg/mL)	<i>G. pfeifferi</i>	[46]
94.	Ganoderone C	Antiviral—NE	<i>G. pfeifferi</i>	[46]
95.	3,7,24-Trioxo-5 <i>α</i> -lanost-8,25-dien-26-ol	Cytotoxicity against HL-60 (15.70 µM), SMMC-7721 (15.52 µM), A-549 (15.81 µM), MCF-7 (20.08 µM), SW480—NE	<i>G. hainanense</i>	[62]
96.	Hainanaldehyde A	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
97.	21-Hydroxy-3,7-dioxo-5 <i>α</i> -lanost-8,24 <i>E</i> -dien-26-ol	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
98.	3 <i>β</i> ,11 <i>α</i> -Dihydroxy-7-oxo-5 <i>α</i> -lanost-8,24 <i>E</i> -dien-26-ol	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
99.	Lucialdehyde D	-	<i>G. lucidum</i> , <i>G. pfeifferi</i>	[46][186] [187]
100.	Ganoderiol J	-	<i>G. sinense</i>	[183]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
101.	16 α ,26-Dihydroxylanosta-8,24-dien-3-one	Cytotoxicity against K-562 cells (13.3 μ g/mL)	<i>G. hainanense</i>	[75]
102.	Lucidadiol	Antiviral: influenza virus type A (ED ₅₀ = 0.22 mmol/L), HSV—NE	<i>G. lucidum</i> , <i>G. pfeifferi</i>	[188][189]
103.	Sinensoic acid	-	<i>G. sinense</i>	[190]
104.	Tsugaric acid C	Cytotoxicity—NE	<i>G. tsugae</i>	[191]
105.	Colossolactone A	Moderate cytotoxicity against L-929, K-562, HeLa cells—NE, anti-inflammatory properties	<i>G. colossum</i>	[192]
106.	Ganoderenicfy A	Promoting angiogenesis activities	<i>G. applanatum</i>	[193]
107.	Colossolactone I	Moderate cytotoxicity against HCT-116 colorectal cancer cells, Antimalarial: <i>Plasmodium falciparum</i> —NE	<i>G. colossum</i>	[194][195] [196]
108.	Colossolactone II	Low cytotoxicity against HCT-116 colorectal cancer cells	<i>G. colossum</i>	[194][195]
109.	Ganodermalactone E	Antimalarial: <i>Plasmodium falciparum</i> —NE	<i>G. colossum</i>	[196]
110.	Colossolactone B	Moderate cytotoxicity against L-929, K-562, and HeLa cells, antimicrobial—NE, antibacterial—NE	<i>G. colossum</i>	[192][196] [197]
111.	Methyl ganolucidate A	-	<i>G. lucidum</i>	[153][157]
112.	Methyl ganolucidate B	-	<i>G. lucidum</i>	[153][157]
113.	Methyl ganoderate A	-	<i>G. lucidum</i>	[166]
114.	Methyl ganoderate B	-	<i>G. lucidum</i>	[166]
115.	Methyl ganoderate C	-	<i>G. lucidum</i>	[166]
116.	Methyl ganoderate C ₂	-	<i>G. lucidum</i> (dried fruit bodies)	[198]
117.	Compound B ₈	-	<i>G. lucidum</i> (dried fruit	[198]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species bodies)	References
118.	3β-Oxo-formyl-7β,12β-dihydroxy-5α-lanost-11,15,23-trioxo-8-en(<i>E</i>)-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[199]
119.	Ganoderic acid B ₈	Cytotoxicity against LLC—NE, T47-D—NE, S-180—NE, Meth-A—NE	<i>G. lucidum</i> (fruit bodies)	[200]
120.	Ganoderic acid C ₁	Inhibitory activity against HIV-PR (0.18 mM)	<i>G. lucidum</i> (fruit bodies)	[151][200]
121.	12β-Acetoxy-3,7,11,15,23-pentaoxo-5α-lanosta-8-en-26-oic acid ethyl ester	Cytotoxicity against human HeLa cervical cancer cell lines (63 μM)	<i>G. lucidum</i>	[201]
122.	3β,7β-Dihydroxy-12β-acetoxy-11,15,23-trioxo-5α-lanosta-8-en-26-oic acid methyl ester	-	<i>G. lucidum</i>	[32]
123.	3β-Hydroxy-7,11,12,15,23-pentaoxolanost-8-en-26-oic acid	Cytotoxic against p388 cell (9.85 μM), HeLa cell (17.10 μM), BEL-7402 cell (51.00 μM), SGC-7901 cells (42.00 μM)	<i>G. lucidum</i> (fruit bodies)	[202]
124.	Ganoderic acid H	Inhibitory activity against HIV-PR (0.20 mM)	<i>G. lucidum</i> (fruit bodies)	[160][203]
125.	Ganoderic acid K	Cytotoxicity against p388 cell (13. 8 μM), HeLa cell (8.23 μM); BEL-7402 cell (16.5 μM), SGC-7901cell (21.0 μM)	<i>G. lucidum</i> (fruit bodies)	[204]
126.	Ganoderic acid AM ₁	Cytotoxicity against p388 cell (13. 2 μM), HeLa cell (9.75 μM), BEL-7402 cell (20.9 μM), SGC-7901 cell (23.0 μM)	<i>G. lucidum</i> (fruit bodies)	[204]
127.	Ganoderic acid J	Cytotoxicity against p388 cell (15. 8 μM), HeLa cell (12.2 μM), BEL-7402 cell (25.2 μM), SGC-7901 cell (20.2 μM)	<i>G. lucidum</i> (fruit bodies)	[204]
128.	Ganoderic acid AP ₂	-	<i>G. applanatum</i> (fruit bodies)	[161]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
129.	23S-Hydroxy-3,7,11,15-tetraoxolanost-8,24E-diene-26-oic acid	Cytotoxicity against p388 cell (15.7 µM), HeLa cell (9.72 µM), BEL-7402 cell (25.6 µM), SGC-7901 cell (23.1 µM)	<i>G. lucidum</i> (fruit bodies)	[204]
130.	7-Oxoganoderic acid Z	Inhibitory activities against the HMG-CoA reductase (22.3 µM), acyl CoA acyltransferase (5.5 µM)	<i>G. lucidum</i> (fruit bodies)	[205]
131.	Ganoderic acid LM ₂	Potent enhancement of ConA-induced mice splenocytes proliferation in vitro	<i>G. lucidum</i> (fruit bodies)	[206]
132.	Lucialdehyde B	Cytotoxic effect on tested tumor cells	<i>G. lucidum</i> (fruit bodies)	[200]
133.	Lucialdehyde C	Cytotoxicity against LLC, T-47D (10.7 µg/mL), Sarcoma 180 (4.7 µg/mL), Meth-A tumor cells (3.8 µg/mL)	<i>G. lucidum</i> (fruit bodies)	[200]
134.	Ganoderic acid β	HIV-I protease inhibitory activity (20 µM)	<i>G. lucidum</i> (spores)	[207]
135.	Ganolucidic acid E	-	<i>G. lucidum</i> (fruit bodies)	[185]
136.	Ganoderal B	-	<i>G. lucidum</i>	[208]
137.	11α-Hydroxy-3,7-dioxo-5α-lanosta-8,24(E)-dien-26-oic acid	Cytotoxicity against human HeLa cervical cancer cell lines (123 µM)	<i>G. lucidum</i>	[201]
138.	11β-Hydroxy-3,7-dioxo-5α-lanosta-8,24(E)-dien-26-oic acid	Cytotoxicity against human HeLa cervical cancer cell lines (51 µM)	<i>G. lucidum</i>	[201]
139.	Lucidal	-	<i>G. lucidum</i> (cultured fruit bodies)	[188]
140.	Lucialdehyde E	Cytotoxic activity against esophageal tumor EC109 cell line (18.7 mg/mL)	<i>G. lucidum</i> (spores)	[187]
141.	3α,22β-Diacetoxy-7α-hydroxyl-5α-lanost-8,24E-dien-26-oic acid	Cytotoxicity against HeLa cell lines (14.7 µM), 95D cell lines (23.01 µM)	<i>G. lucidum</i> (mycelial mat)	[180]
142.	Ganoderic acid O	-	<i>G. lucidum</i> (cultured	[209]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
			mycelium)	
143.	7-O-Methylganoderic acid O	-	<i>G. lucidum</i> (cultured mycelium)	[209]
	12 β -Acetoxy-3 β -hydroxy-7,11,15,23-tetraoxo-lanost-8,20E-diene-26-oic acid	Cytotoxicity against human cancer cell p388 (12.7 μ M), HeLa cell (8.72 μ M), BEL-7402 (24.2 μ M), SGC-7901 (18.7 μ M)	<i>G. lucidum</i> (fruit bodies)	[204]
145.	23-Dihydroganoderenic acid D	-	<i>G. applanatum</i> (fruit bodies)	[38]
146.	Ganoderenic acid A	-	<i>G. lucidum</i> (dried fruit bodies)	[160]
147.	Ganoderenic acid B	-	<i>G. lucidum</i> (dried fruit bodies)	[160]
148.	Ganoderenic acid C	-	<i>G. lucidum</i> (dried fruit bodies)	[160]
149.	Ganoderenic acid D	-	<i>G. lucidum</i> (dried fruit bodies)	[160]
150.	12 β -Acetoxy-7 β -hydroxy-3,11,15,23-tetraoxo-5 α -lanosta-8,20-dien-26-oic acid	Cytotoxicity against human HeLa cervical cancer cell lines—NE	<i>G. lucidum</i>	[201]
151.	Methy ganoderenate H	-	<i>G. applanatum</i> (fruit bodies)	[172]
152.	Methyl ganoderenate I	-	<i>G. applanatum</i> (fruit bodies)	[172]
153.	12 β -Acetoxy-3 β ,7 β -dihydroxy-11,15,23-trioxo-5 α -lanosta-8,20-dien-26-oic acid	-	<i>G. lucidum</i>	[32]
154.	Methyl ganoderate G	-	<i>G. lucidum</i>	[153]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
155.	Compound C5	-	<i>G. lucidum</i> (fruit bodies)	[203]
156.	Compound C6	-	<i>G. lucidum</i> (fruit bodies)	[203]
157.	Ganoderic acid AP ₃	-	<i>G. applanatum</i> (fruit bodies)	[161]
158.	23-Dihydroganoderic acid I	-	<i>G. applanatum</i> (fruit bodies)	[38]
159.	23-Dihydroganoderic acid N	-	<i>G. applanatum</i> (fruit bodies)	[38]
160.	20-Hydroxyganoderic acid G	-	<i>G. lucidum</i> (fruit bodies)	[210]
161.	Lucidumol A	HIV-I protease inhibitory activity—NE	<i>G. lucidum</i> (spores)	[207]
162.	Ganoderiol C	-	<i>G. lucidum</i> (fruit bodies)	[185]
163.	Ganoderiol D	-	<i>G. lucidum</i> (fruit bodies)	[185]
164.	Ganoderitriol M	-	<i>G. lucidum</i> (fruit bodies)	[211]
165.	Tsugaric acid A	-	<i>G. tsugae</i>	[212]
166.	Tsugarioside A	Cytotoxicity against PLC/PRF/5 (ED ₅₀ = 6.5 µg/mL), T-24 (ED ₅₀ = 8.6 µg/mL), HT-3 (ED ₅₀ = 7.2 µg/mL), SiHa (ED ₅₀ = 9.5 µg/mL)	<i>G. tsugae</i> (fruit bodies)	[191]
167.	3-Oxo-5 α -lanosta-8,24-dien-21-oic acid	Cytotoxicity—NE	<i>G. resinaceum</i> (fruit bodies)	[213]
168.	3 β -Hydroxy-5 α -lanosta-8,24-dien-21-oic acid	Cytotoxicity against T-24 (ED ₅₀ = 4.4 µg/mL), HT-3 (ED ₅₀ = 3.5 µg/mL), SiHa (ED ₅₀ = 5.5 µg/mL), CaSKi (ED ₅₀ = 6.2 µg/mL)	<i>G. tsugae</i> (fruit bodies)	[191]
169.	3 β ,7 β -Dihydroxy-11,15,23-trioxolanost-8,16-dien-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[188]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
170.	3 β ,7 β -Dihydroxy-11,15,23-trioxolanost-8,16-dien-26-oic acid methyl ester	-	<i>G. lucidum</i> (fruit bodies)	[188]
171.	12 β -Acetoxy-3 β ,7 β -dihydroxy-11,15,23-trioxolanost-8,16-dien-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[214]
172.	Methyl ganoderate AP	-	<i>G. applanatum</i> (fruit bodies)	[172]
173.	Ganoderiol E	-	<i>G. lucidum</i> (fruit bodies)	[185]
174.	Epoxyganoderiol A	-	<i>G. lucidum</i>	[208]
175.	3 α -Carboxyacetoxy-24-methylene-23-oxolanost-8-en-26-oic acid	Cytotoxicity—NE	<i>G. applanatum</i> (fruit bodies)	[215]
176.	3 α -Carboxyacetoxy-24-methyl-23-oxolanost-8-en-26-oic acid	Cytotoxicity—NE	<i>G. applanatum</i> (fruit bodies)	[215]
177.	3-Epipachymic acid	-	<i>G. resinaceum</i> (fruit bodies)	[213]
178.	3 β ,15 α -Diacetoxylanosta-8,24-dien-26-oic acid	-	<i>G. lucidum</i> (mycelia)	[216]
179.	Ganoderic acid V ₁	-	<i>G. lucidum</i>	[217]
180.	Tsugaric acid B	-	<i>G. tsugae</i>	[212]
181.	Methyl ganoderenate E	-	<i>G. lucidum</i> (fruit bodies)	[158]
182.	Lucidumol D	Selective anti-proliferative and cytotoxic effects	<i>G. lingzhi</i>	[218]
183.	Lucidumol C	Selective anti-proliferative and cytotoxic effects	<i>G. lingzhi</i>	[218]
184.	Leucocontextin A	-	<i>G. leucocontextum</i>	[219]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
185.	Leucocontextin B	-	G. <i>leucocontextum</i>	[219]
186.	Leucocontextin C	-	G. <i>leucocontextum</i>	[219]
187.	Leucocontextin D	-	G. <i>leucocontextum</i>	[219]
188.	Leucocontextin E	-	G. <i>leucocontextum</i>	[219]
189.	Leucocontextin F	-	G. <i>leucocontextum</i>	[219]
190.	Leucocontextin G	-	G. <i>leucocontextum</i>	[219]
191.	Leucocontextin H	-	G. <i>leucocontextum</i>	[219]
192.	Leucocontextin I	-	G. <i>leucocontextum</i>	[219]
193.	Leucocontextin R	Cytotoxicity against K562 and MCF-7 cell lines (IC ₅₀ = 20–30 μM)	G. <i>leucocontextum</i>	[219]
194.	Ganoleuconin A	Cytotoxicity against K562 (17.8 μM), PC-3 cell lines—NE	G. <i>leucocontextum</i>	[34]
195.	Ganoleuconin B	Cytotoxicity against K562 (19.7 μM), PC-3 cell lines—NE	G. <i>leucocontextum</i>	[34]
196.	Ganoleuconin E	Cytotoxicity against K562 and PC-3 cell lines—NE	G. <i>leucocontextum</i>	[34]
197.	Ganoleuconin G	Cytotoxicity against K562 (11.4 μM), PC-3 cell lines (132.4 μM)	G. <i>leucocontextum</i>	[34]
198.	Ganoleuconin H	Cytotoxicity against K562 (115.4 μM), PC-3 cell lines (24.2 μM)	G. <i>leucocontextum</i>	[34]
199.	Ganoleuconin I	Cytotoxicity against K562 and PC-3 cell lines—NE	G. <i>leucocontextum</i>	[34]
200.	Ganoderenicfy B	Promoting angiogenesis activities	<i>G. applanatum</i>	[193]
201.	(24 <i>E</i>)-15 <i>α</i> ,26-Dihydroxy-	Antimycobacteria (50 μg/mL),	<i>G. casuarinicola</i>	[220]

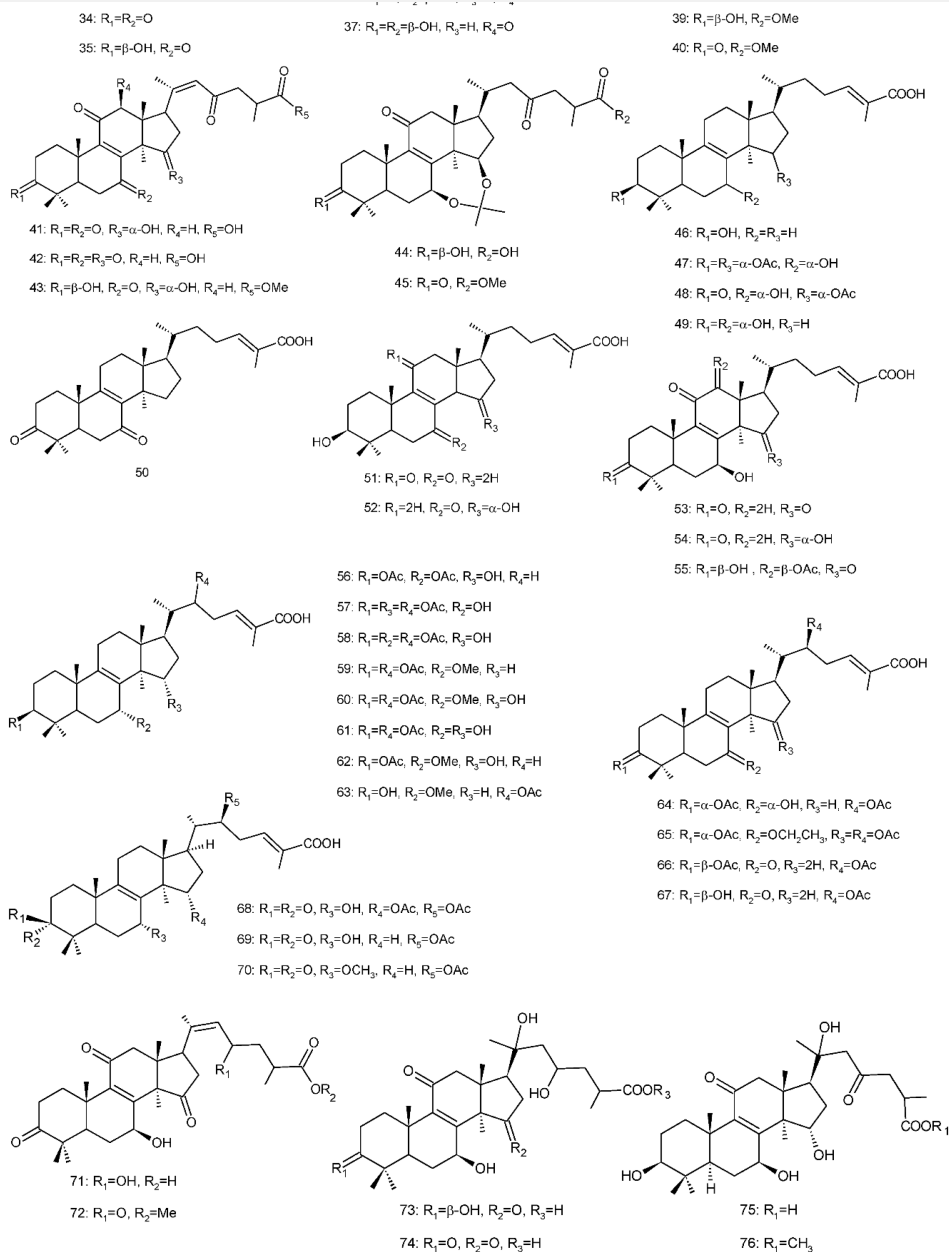
No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources Ganoderma Species	References
	3-oxo-lanosta-8,24-diene	cytotoxicity (5.9 µg/mL)		
202.	(24E)-7α,26-Dihydroxy-3-oxo-lanosta-8,24-diene.	Antimalarial activity (IC ₅₀ = 9.7 µg/mL)	<i>G. casuarinicola</i>	[220]
203.	(24E)-3-Oxo-7α,15α,26-trihydroxylanosta-8,24-diene	Antimalarial activity (IC ₅₀ = 9.2 µg/mL)	<i>G. casuarinicola</i>	[220]
204.	(24E)-3β-Acetoxy-15α,26-dihydroxylanosta-8,24-diene	Antimycobacteria (25 µg/mL), cytotoxicity (6 µg/mL)	<i>G. casuarinicola</i>	[220]
205.	(24E)-3β-Acetoxy-7α,15α,26-trihydroxylanosta-8,24-diene	Antimycobacteria (25 µg/mL), cytotoxicity (9 µg/mL)	<i>G. casuarinicola</i>	[220]
206.	(24E)-3β,7α,15α,26-Tetra-hydroxylanosta-8,24-diene	-	<i>G. casuarinicola</i>	[220]
207.	7β,15α,20-Trihydroxy-3,11,23-trioxo-5α-lanosta-8-en-26-oic acid	-	<i>G. lucidum</i>	[221]
208.	Ganoderic acid XL ₃	-	<i>G. theaecolum</i>	[222]
209.	Ganoderic acid XL ₄	-	<i>G. theaecolum</i>	[222]
210.	Ganodecalone A	Cytotoxicity against K562 (17.22 µM)	<i>G. calidophilum</i>	[53]
211.	Ganoderic acid C ₆ [145][146]	- [147]	<i>G. lucidum</i> (fruit bodies)	[65]
212.	Ganoderic acid D ₁	-	<i>G. lucidum</i> (fruit bodies)	[65]
[146] 213.	Ganoderic acid M	-	<i>G. lucidum</i> (fruit bodies)	[65]
214.	Ganoderic acid N	-	<i>G. lucidum</i> (fruit bodies)	[65]
215.	12-Hydroxylganoderic acid C ₂	- [95]	<i>G. lucidum</i> (fruit bodies)	[65]
216.	3-Acetylganoderic acid H	-	<i>G. lucidum</i> (fruit bodies)	[65]

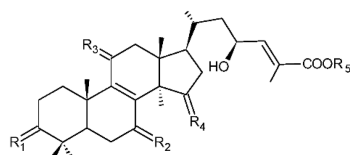
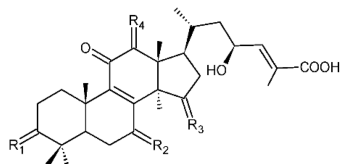
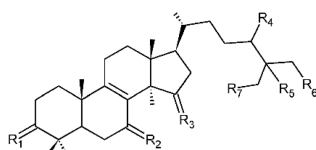
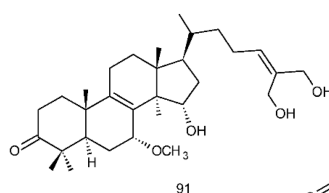
st genesis
c-Fos and
dritic cell-
et al. [145]
ancer cell
et al. [175]
androgen-
e, G1 cell
et protein
[226].

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
217.	12-Acetoxyganoderic acid D	-	<i>G. lucidum</i> (fruit bodies)	[65]
218.	12-Hydroxyganoderic acid D	-	<i>G. lucidum</i> (fruit bodies)	[65]
219.	12-Acetoxyganoderic acid F	-	<i>G. lucidum</i> (fruit bodies)	[65]
220.	12 β -Hydroxy-3,7,11,15,23-pentaoxo-5 α -lanosta-8-en-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[65]
221.	12-Hydroxy-3,7,11,15,23-pentaoxo-lanost-8-en-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[65]
222.	12,15-Bis(acetyloxy)-3-hydroxy-7,11,23-trioxo-lanost-8-en-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[65]
223.	Methyl ganoderate J	-	<i>G. lucidum</i> (fruit bodies)	[65]
224.	Methyl-O-acetylganoderate C	-	<i>G. lucidum</i> (fruit bodies)	[65]
225.	Methyl ganoderate C1 ₅₀	-	<i>G. lucidum</i> (fruit bodies)	[65]
226. [227]	Methyl ganoderate AM	-	[164] <i>G. lucidum</i> (fruit bodies)	[65]
227.	Ganoderic aldehyde A	-	<i>G. lucidum</i> (fruit bodies)	[65]
228.	Ganoderenic acid K	-	<i>G. lucidum</i> (fruit bodies)	[65]
229.	Ganoderenic acid [228]	-	<i>G. lucidum</i> (fruit bodies)	[65]
230.	Elfvingic acid A	-	<i>G. lucidum</i> (fruit bodies)	[65]
231.	12 β -Acetoxy-3 β ,7 β -dihydroxy-11,15,23-	-	<i>G. lucidum</i> (fruit bodies)	[65]

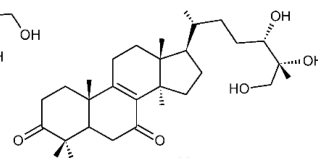
No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
	trioxo-5 α -lanosta-8,20-dien-26-oic acid			
232.	Methyl ganolucidate C	-	<i>G. lucidum</i> (fruit bodies)	[65]
233.	Ganolucidic acid C	-	<i>G. lucidum</i> (fruit bodies)	[65]
234.	Ganoderic acid C ₂	-	<i>G. lucidum</i> (fruit bodies /spore)	[65]
235.	Ganodrol A	Moderately inhibits FAAH (Inhibition rate in between 50–60%)	<i>G. lucidum</i>	[116]
236.	Ganodrol C	Moderately inhibits FAAH (Inhibition rate in between 50–60%)	<i>G. lucidum</i>	[116]
237.	Ganodrol D	Moderately inhibits FAAH (Inhibition rate in between 30–40%)	<i>G. lucidum</i>	[116]
238.	Ganoderic acid XL ₅	Cytotoxicity against human tumor cell lines—NE	<i>G. theaeacolum</i>	[222]
239.	Methyl gibbosate M	Anti-adipogenesis activity—NE	<i>G. applanatum</i>	[51]
240.	Methyl ganoapplate E	Anti-adipogenesis activity—NE	<i>G. applanatum</i>	[51]
241.	Applandiketone A	-	<i>G. applanatum</i>	[223]
242.	Applandiketone B	Significant inhibitory effect against NO production in LPS-induced RAW264.7 cells (IC ₅₀ = 20.65 μ M)	<i>G. applanatum</i>	[223]
243.	15 α -Acetoxy-3 α -hydroxy-lanosta-8,24-dien-26-oic	-	<i>G. capense</i>	[224]
244.	Ganoderterpene A	Strongly suppressed NO generation in BV-2 microglial cells treated with lipopolysaccharide (LPS) (IC ₅₀ = 7.15 μ M), significantly suppressed the activation of MAPK and TLR-4/NF- κ B signaling pathways, effectively improved the LPS-induced mitochondrial membrane potential and apoptosis	<i>G. lucidum</i>	[225]

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
245.	Ganodeweberiol G	Significant α -glucosidase inhibitory activity (IC ₅₀ = 165.9 μ M)	<i>G. weberianum</i>	[77]

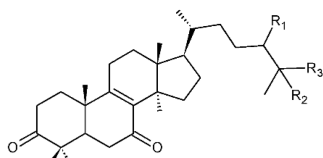
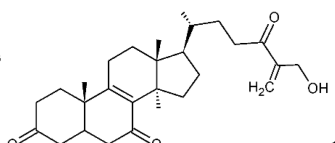


77: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{O}$, $R_4=\alpha\text{-OH}$, $R_5=\text{H}$ 78: $R_1=\beta\text{-OH}$, $R_2=\text{H}$, $R_3=\text{O}$, $R_4=\alpha\text{-OH}$, $R_5=\text{H}$ 79: $R_1=\text{O}$, $R_2=\text{H}_2$, $R_3=\text{O}$, $R_4=\alpha\text{-OH}$, $R_5=\text{H}$ 80: $R_1=\text{O}$, $R_2=\text{H}_2$, $R_3=\text{O}$, $R_4=\alpha\text{-OH}$, $R_5=\text{Me}$ 81: $R_1=R_2=\text{O}$, $R_3=R_4=R_5=\text{H}$ 82: $R_1=R_2=R_3=\text{O}$, $R_4=\beta\text{-OH}$, $R_5=\text{H}$ 83: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\alpha\text{-OH}$, $R_4=2\text{H}$ 84: $R_1=\text{O}$, $R_2=\alpha\text{-OH}$, $R_3=\alpha\text{-OH}$, $R_4=2\text{H}$ 85: $R_1=R_2=\beta\text{-OH}$, $R_3=\text{O}$, $R_4=2\text{H}$ 86: $R_1=\beta\text{-OH}$, $R_2=R_3=\text{O}$, $R_4=2\text{H}$ 87: $R_1=R_2=R_4=\beta\text{-OH}$, $R_3=\text{O}$ 88: $R_1=R_4=\beta\text{-OH}$, $R_2=R_3=\text{O}$ 89: $R_1=\text{O}$, $R_2=\text{OMe}$, $R_3=2\text{H}$, $R_4=R_5=R_6=\text{OH}$, $R_7=\text{H}$ 90: $R_1=\text{OH}$, $R_2=\text{O}$, $R_3=2\text{H}$, $R_4=R_5=R_6=\text{OH}$, $R_7=\text{H}$ 

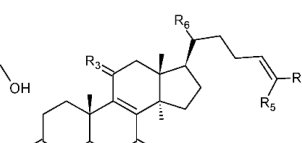
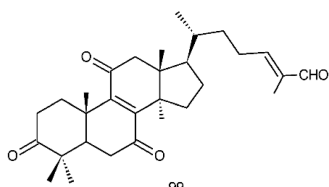
91



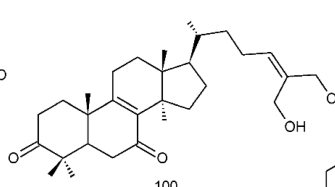
92

93: $R_1, R_2=\Delta^{24(25)}$, $R_3=\text{CH}_2\text{OH}$ 94: $R_1, R_2=\text{O}$, $R_3=\text{CH}_2\text{OH}$ 

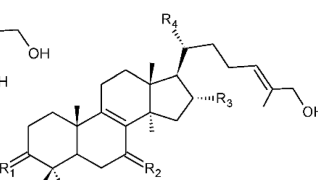
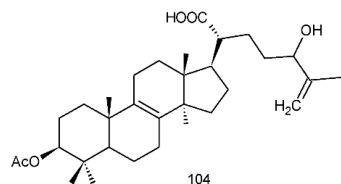
95

96: $R_1=R_2=\text{O}$, $R_3=\alpha\text{-OH}$, $R_4=\text{CHO}$, $R_5=R_6=\text{CH}_3$ 97: $R_1=R_2=\text{O}$, $R_3=2\text{H}$, $R_4=\text{CH}_2\text{OH}$, $R_5=\text{CH}_3$, $R_6=\text{CH}_2\text{OH}$ 98: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\alpha\text{-OH}$, $R_4=\text{CH}_2\text{OH}$, $R_5=R_6=\text{CH}_3$ 

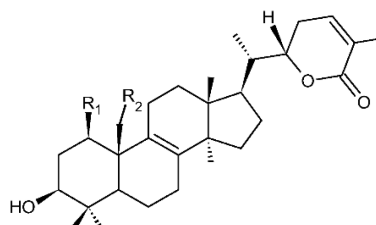
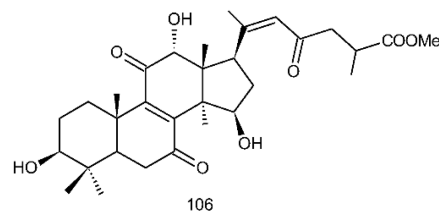
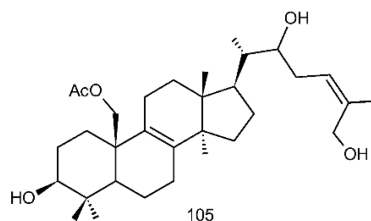
99



100

101: $R_1=\text{O}$, $R_2=\text{H}_2$, $R_3=\text{OH}$, $R_4=\text{Me}$ 102: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{Me}$ 103: $R_1=\beta\text{-OH}$, $R_2=\text{H}_2$, $R_3=\text{H}$, $R_4=\text{COOH}$ 

104

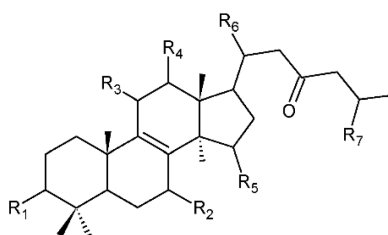


107: $R_1=H$, $R_2=H$

108: $R_1=\beta-OH$, $R_2=H$

109: $R_1=H$, $R_2=OH$

110: $R_1=H$, $R_2=OAc$



111: $R_1=O$, $R_2=H$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\beta-CH_3$, $R_7=COOCH_3$

112: $R_1=\beta-OH$, $R_2=H$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\beta-CH_3$, $R_7=COOCH_3$

113: $R_1=O$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\beta-CH_3$, $R_7=COOCH_3$

114: $R_1=\beta-OH$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\beta-CH_3$, $R_7=COOCH_3$

115: $R_1=\beta-OH$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\beta-CH_3$, $R_7=COOCH_3$

116: $R_1=\beta-OH$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\alpha-CH_3$, $R_7=COOCH_3$

117: $R_1=O$, $R_2=\alpha-OH$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\alpha-CH_3$, $R_7=COOCH_3$

118: $R_1=O-CHO$, $R_2=\beta-OH$, $R_3=O$, $R_4=\beta-OH$, $R_5=O$, $R_6=\beta-CH_3$, $R_7=COOH$

119: $R_1=O$, $R_2=\alpha-OH$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\alpha-CH_3$, $R_7=COOH$

120: $R_1=O$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=COOH$

121: $R_1=O$, $R_2=O$, $R_3=O$, $R_4=\beta-O-COCH_3$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=COOEt$

122: $R_1=\beta-OH$, $R_2=\beta-OH$, $R_3=O$, $R_4=\beta-O-COCH_3$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=COOCH_3$

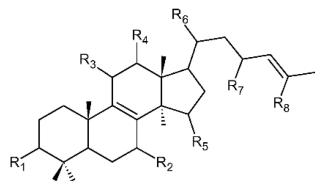
123: $R_1=\beta-OH$, $R_2=O$, $R_3=O$, $R_4=O$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=COOH$

124: $R_1=\beta-OH$, $R_2=O$, $R_3=O$, $R_4=\beta-OAc$, $R_5=O$, $R_6=\beta-CH_3$, $R_7=COOH$

125: $R_1=\beta-OH$, $R_2=\beta-OH$, $R_3=O$, $R_4=\beta-OAc$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=COOH$

126: $R_1=\beta-OH$, $R_2=O$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=COOH$

127: $R_1=O$, $R_2=O$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\alpha-CH_3$, $R_7=COOH$



128: $R_1=\beta\text{-OH}$, $R_2=\text{H}$, $R_3=\text{O}$, $R_4=\beta\text{-OAc}$, $R_5=\alpha\text{-OAc}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{COOH}$

129: $R_1=O$, $R_2=O$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\alpha\text{-CH}_3$, $R_7=\beta\text{-OH}$, $R_8=\text{COOH}$

130: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\beta\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{COOH}$

131: $R_1=O$, $R_2=OH$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=OH$, $R_8=COOH$

132: $R_1=O$, $R_2=O$, $R_3=H$, $R_4=H$, $R_5=H$, $R_6=\alpha\text{-CH}_3$, $R_7=H$, $R_8=\text{CHO}$

133: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{CHO}$

134: R₁=β-OH, R₂=β-OH, R₃=O, R₄=H, R₅=O, R₆=α-CH₂, R₇=H, R₈=COOH

135: $R_1=O$, $R_2=H$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\beta-CH_3$, $R_7=H$, $R_8=COOH$

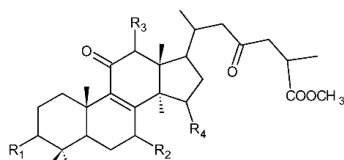
136: $R_1=O, R_2=\alpha-OH, R_3=H, R_4=H, R_5=H, R_6=\beta-CH_2, R_7=H, R_8=CHO$

137: $R_1=O$, $R_2=O$, $R_3=\alpha-OH$, $R_4=H$, $R_5=H$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOH$

138: $R_1=O, R_2=O, R_3=\beta-OH, R_4=H, R_5=H, R_6=\alpha-CH_3, R_7=H, R_8=COOH$

139: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{CHO}$

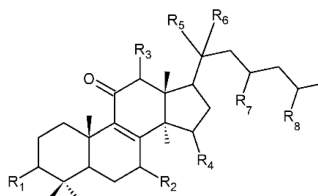
140: $R_1=O$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=CHO$



154: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OH}$, $R_4=\text{O}$

155: $R_1=O$, $R_2=\beta-OH$, $R_3=OH$, $R_4=O$

156: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{OH}$, $R_4=\text{O}$

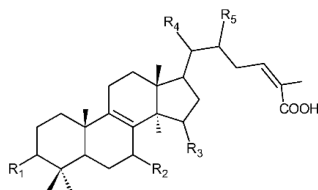


157: $R_1=O$, $R_2=O$, $R_3=H$, $R_4=\alpha-OH$, $R_5=\beta-CH_3$, $R_6=\xi-OH$, $R_7=O$, $R_8=COOH$

158: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\alpha\text{-CH}_3$, $R_6=\beta\text{-OH}$, $R_7=\xi\text{-OH}$, $R_8=\xi\text{-COOH}$

159: R₁=O, R₂=β-OH, R₃=H, R₄=O, R₅=α-CH₃, R₆=β-OH, R₇=ξ-OH, R₈=ξ-COOH

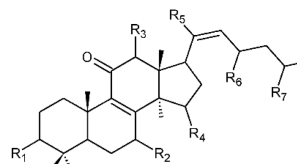
160: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OH}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_2$, $R_6=\beta\text{-OH}$, $R_7=\text{O}$, $R_8=\text{COOH}$



141: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OH}$, $R_3=\text{H}$, $R_4=\alpha\text{-CH}_3$, $R_5=\beta\text{-OAc}$

142: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OH}$, $R_3=\alpha\text{-OAc}$, $R_4=\alpha\text{-CH}_3$, $R_5=\beta\text{-OAc}$

143: R₁=α-OAc, R₂=α-OCH₃, R₃=α-OAc, R₄=α-CH₃, R₅=β-OAc



144: R₁=β-OH, R₂=O, R₃=β-OAc, R₄=O, R₅=β-CH₃, R₆=O, R₇=COOH

145: $R_1=O$, $R_2=\beta-OH$, $R_3=H$, $R_4=O$, $R_5=\beta-CH_3$, $R_6=\xi-OH$, $R_7=\xi-COOH$

146: $R_1=O$, $R_2=\beta-OH$, $R_3=H$, $R_4=\alpha-OH$, $R_5=\beta-CH_3$, $R_6=O$, $R_7=COOH$

147: R₁=β-OH, R₂=β-OH, R₃=H, R₄=O, R₅=β-CH₃, R₆=O, R₇=COOH

148: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\alpha\text{-OH}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COOH}$

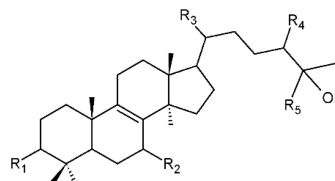
149: R₁=O, R₂=β-OH, R₃=H, R₄=O, R₅=β-CH₃, R₆=O, R₇=COOH

150: $R_1=O$, $R_2=\beta-OH$, $R_3=\beta-OAc$, $R_4=O$, $R_5=\beta-CH_3$, $R_6=O$, $R_7=COOH$

151: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COCH}_3$

152: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\alpha\text{-OH}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COCH}_3$

153: R₁=β-OH, R₂=β-OH, R₃=β-OAc, R₄=O, R₅=α-CH₃, R₆=O, R₇=COOH

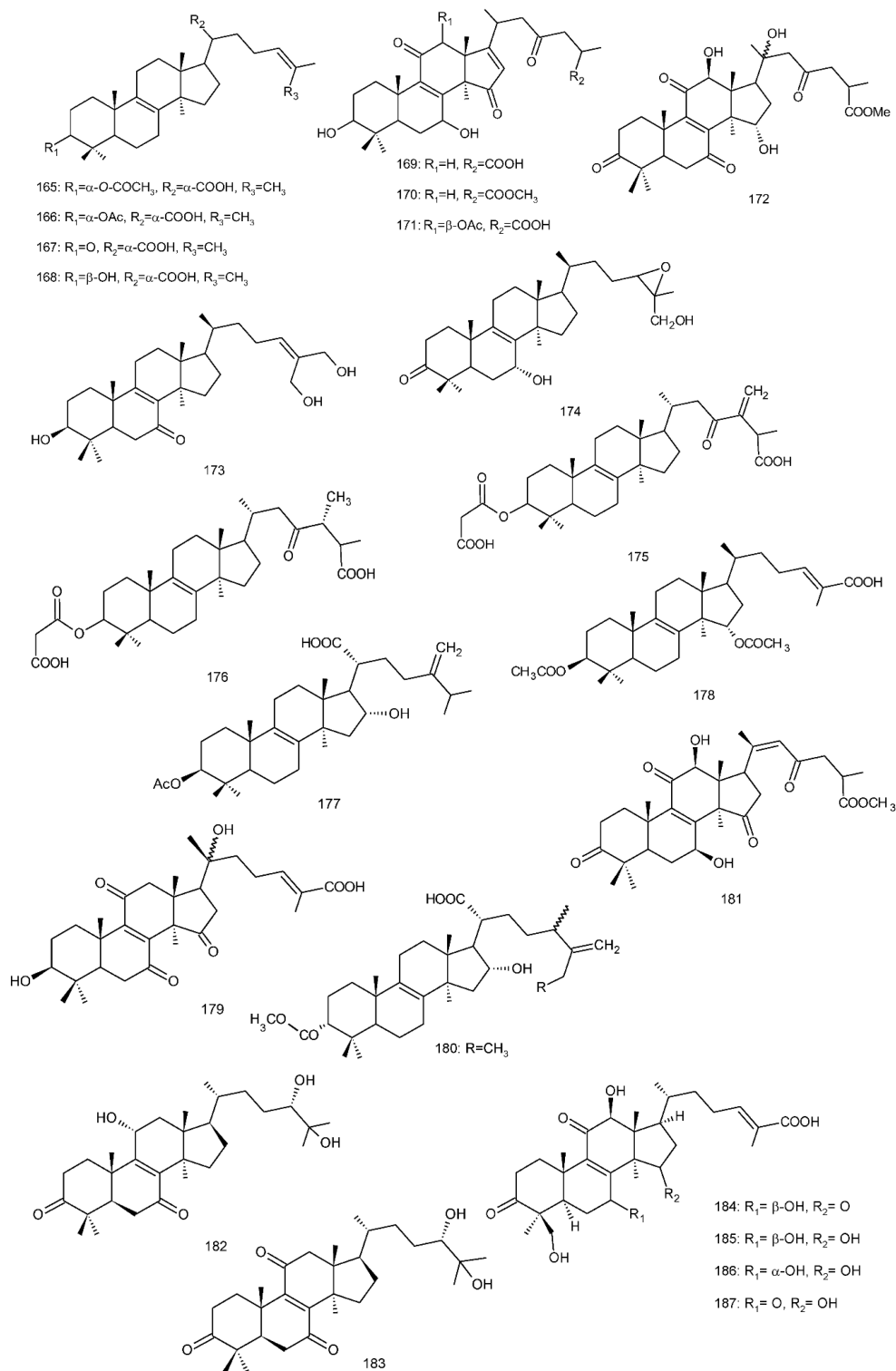


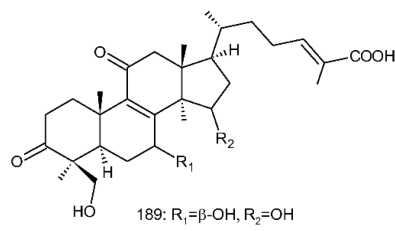
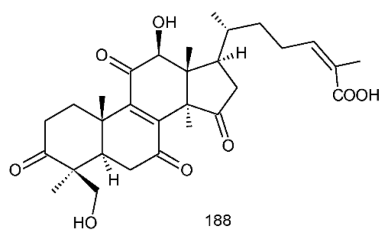
161: $R_1=O$, $R_2=O$, $R_3=\alpha\text{-CH}_3$, $R_4=\alpha\text{-OH}$, $R_5=\text{CH}_3$

162: $R_1=O$, $R_2=\alpha\text{-OEt}$, $R_3=\beta\text{-CH}_3$, $R_4=\xi\text{-OH}$, $R_5=\text{CH}_2\text{OH}$

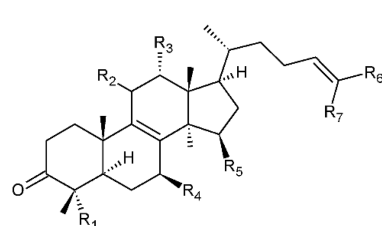
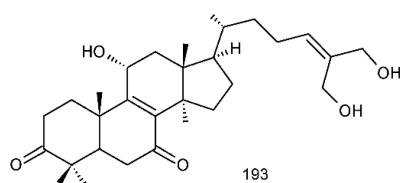
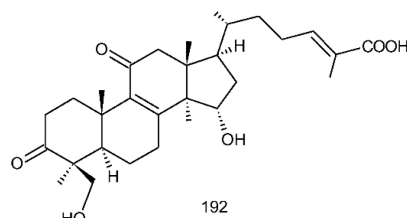
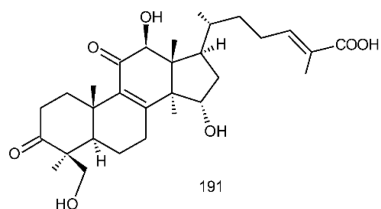
163: $R_1=O$, $R_2=O$, $R_3=\beta\text{-CH}_3$, $R_4=\xi\text{-OH}$, $R_5=\text{CH}_2\text{OH}$

164: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\alpha\text{-CH}_3$, $R_4=\alpha\text{-OH}$, $R_5=\text{CH}_3$.





190: $R_1 = \text{O}$, $R_2 = \text{O}$



194: $R_1 = \text{CH}_2\text{OH}$, $R_2 = \text{O}$, $R_3 = \text{H}$, $R_4 = \text{OH}$, $R_5 = \text{H}$, $R_6 = \text{COOH}$, $R_7 = \text{CH}_3$

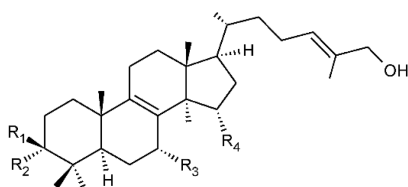
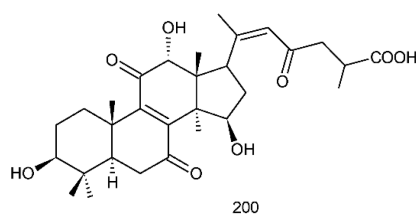
195: $R_1 = \text{CH}_2\text{OH}$, $R_2 = \text{O}$, $R_3 = \text{OH}$, $R_4 = \text{OH}$, $R_5 = \text{H}$, $R_6 = \text{COOH}$, $R_7 = \text{CH}_3$

196: $R_1 = \text{CH}_2\text{OH}$, $R_2 = \text{O}$, $R_3 = \text{OH}$, $R_4 = \text{OH}$, $R_5 = \text{O}$, $R_6 = \text{COOH}$, $R_7 = \text{CH}_3$

197: $R_1 = \text{CH}_3$, $R_2 = \text{O}$, $R_3 = \text{H}$, $R_4 = \text{OH}$, $R_5 = \text{H}$, $R_6 = \text{CH}_2\text{OH}$, $R_7 = \text{CH}_2\text{OH}$

198: $R_1 = \text{CH}_3$, $R_2 = \text{O}$, $R_3 = \text{H}$, $R_4 = \text{O}$, $R_5 = \text{H}$, $R_6 = \text{CH}_2\text{OH}$, $R_7 = \text{CH}_2\text{OH}$

199: $R_1 = \text{CH}_3$, $R_2 = \text{O}$, $R_3 = \text{H}$, $R_4 = \text{H}$, $R_5 = \text{OH}$, $R_6 = \text{CH}_2\text{OH}$, $R_7 = \text{CH}_2\text{OH}$



201: $R_1, R_2 = \text{O}$, $R_3 = \text{H}$, $R_4 = \text{OH}$

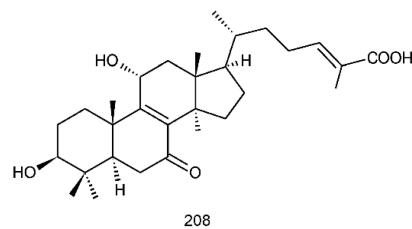
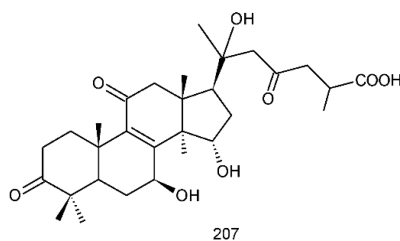
202: $R_1, R_2 = \text{O}$, $R_3 = \text{OH}$, $R_4 = \text{H}$

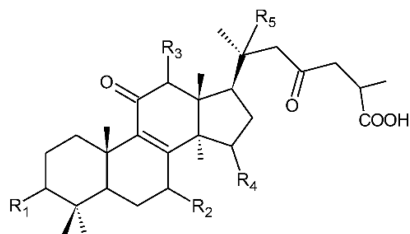
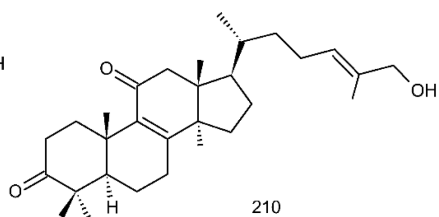
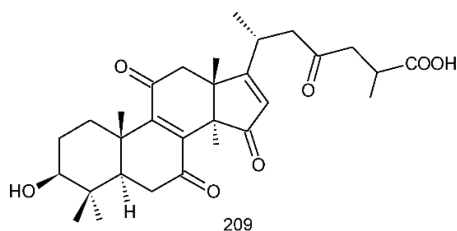
203: $R_1, R_2 = \text{O}$, $R_3 = \text{OH}$, $R_4 = \text{OH}$

204: $R_1 = \text{OAc}$, $R_2 = \text{H}$, $R_3 = \text{H}$, $R_4 = \text{OH}$

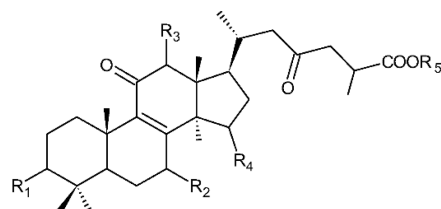
205: $R_1 = \text{OAc}$, $R_2 = \text{H}$, $R_3 = \text{OH}$, $R_4 = \text{OH}$

206: $R_1 = \text{OH}$, $R_2 = \text{H}$, $R_3 = \text{OH}$, $R_4 = \text{OH}$

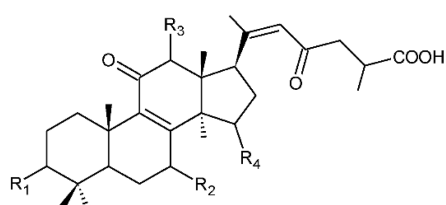
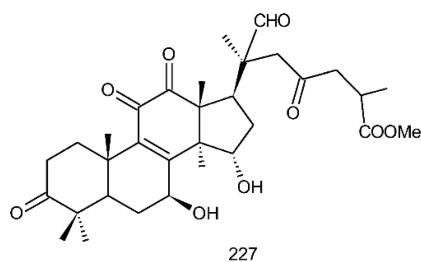




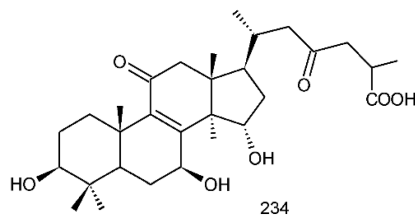
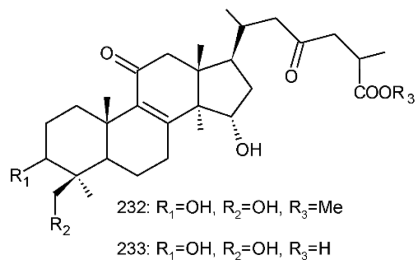
- 211: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\beta\text{-OH}$, $R_4=\text{O}$, $R_5=\text{H}$
 212: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OH}$, $R_4=\text{O}$, $R_5=\text{H}$
 213: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\alpha\text{-OH}$, $R_4=\text{O}$, $R_5=\text{H}$
 214: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\xi\text{-OH}$
 215: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{OH}$, $R_4=\alpha\text{-OH}$, $R_5=\text{H}$
 216: $R_1=\beta\text{-OAc}$, $R_2=\text{O}$, $R_3=\beta\text{-OAc}$, $R_4=\text{O}$, $R_5=\text{H}$
 217: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{OAc}$, $R_4=\text{O}$, $R_5=\text{H}$
 218: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{OH}$, $R_4=\text{O}$, $R_5=\text{H}$
 219: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\beta\text{-OAc}$, $R_4=\text{O}$, $R_5=\text{H}$
 220: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\beta\text{-OH}$, $R_4=\text{O}$, $R_5=\text{H}$
 221: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\text{OH}$, $R_4=\text{O}$, $R_5=\text{H}$
 222: $R_1=\text{OH}$, $R_2=\text{O}$, $R_3=\text{OAc}$, $R_4=\text{OAc}$, $R_5=\text{H}$



- 223: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\alpha\text{-OH}$, $R_5=\text{Me}$
 224: $R_1=\beta\text{-OAc}$, $R_2=\text{O}$, $R_3=\beta\text{-OAc}$, $R_4=\text{O}$, $R_5=\text{Me}$
 225: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\text{CH}_2$
 226: $R_1=\text{O}$, $R_2=\text{H}$, $R_3=\text{H}$, $R_4=\beta\text{-OH}$, $R_5=\text{H}$



- 228: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OAc}$, $R_4=\text{O}$
 229: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OH}$, $R_4=\text{O}$
 230: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\alpha\text{-OH}$, $R_4=\beta\text{-OH}$
 231: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OAc}$, $R_4=\text{O}$



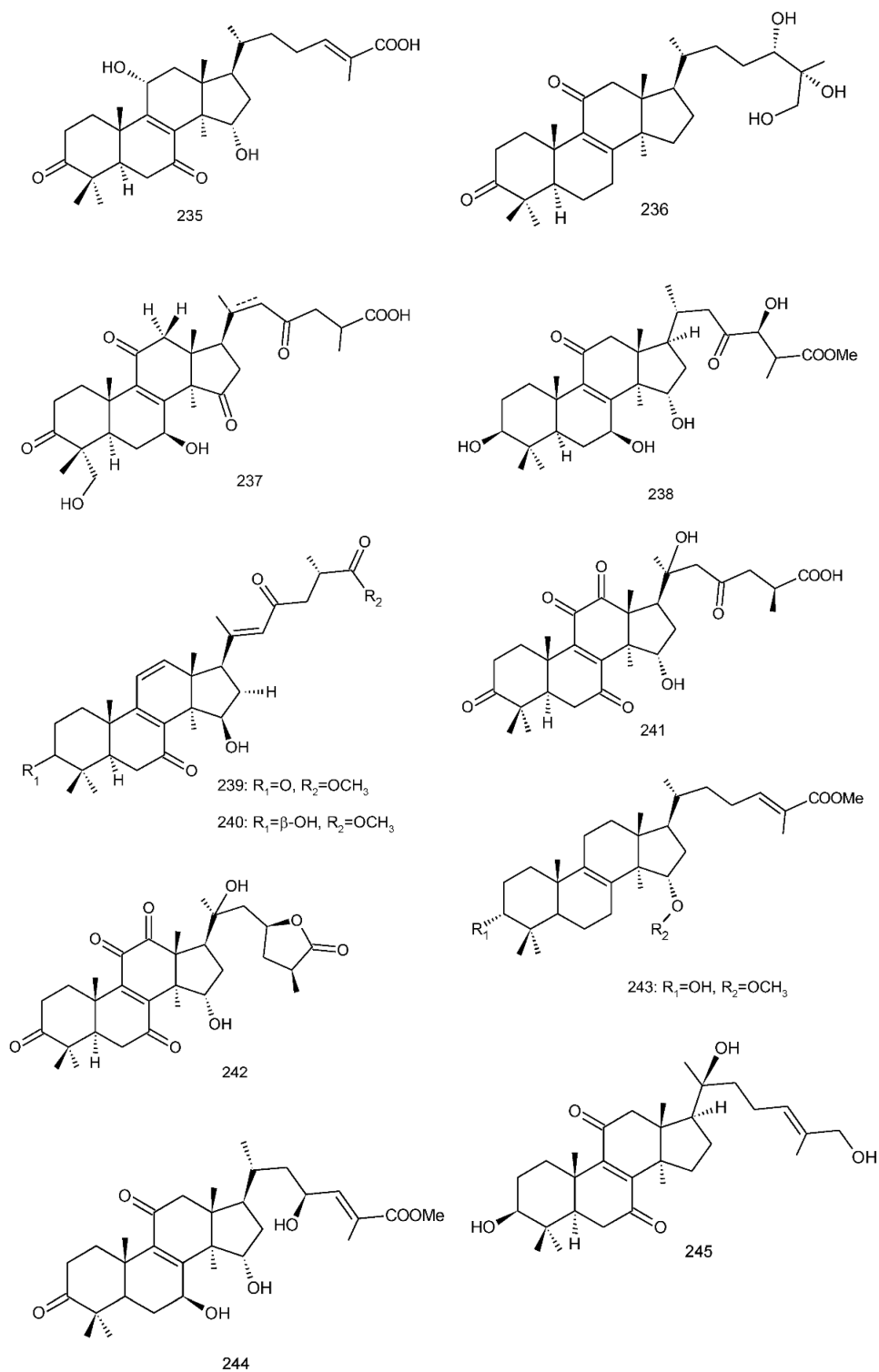


Figure 4. Chemical structures of several 8,9-double-bond triterpenoids (1–245).

References

1. Karsten, P.A. Enumeratio boletinearum et poly-porearum fennicarum. Systemate novo dispositarum. Rev. Mycol. 1881, 3, 16–19.
2. Upadhyay, M.; Shrivastava, B.; Jain, A.; Kidwai, M.; Kumar, S.; Gomes, J.; Goswami, D.G.; Panda, A.K.; Kuhad, R.C. Production of ganoderic acid by *Ganoderma lucidum* RCKB-2010 and its therapeutic potential. Ann. Microbiol. 2014, 64, 839–846.
3. He, M.-Q.; Zhao, R.-L.; Hyde, K.D.; Begerow, D.; Kemler, M.; Yurkov, A.; McKenzie, E.H.; Raspé, O.; Kakishima, M.; Sánchez-Ramírez, S.; et al. Notes, outline and divergence times of basidiomycota. Fungal Divers. 2019, 99, 105–367.
4. Badalyan, S.M.; Gharibyan, N.G.; Iotti, M.; Zambonelli, A. Morphological and ecological screening of different collections of medicinal white-rot bracket fungus *Ganoderma adspersum* (Schulzer) Donk (Agaricomycetes, Polyporales). Ital. J. Mycol. 2019, 48, 1–15.
5. Gryzenhout, M.; Ghosh, S.; Tchotet Tchoumi, J.M.; Vermeulen, M.; Kinge, T.R. *Ganoderma*: Diversity, ecological significances, and potential applications in industry and allied sectors. In Industrially Important Fungi for Sustainable Development, Fungal Biology; Abdel-Azeem, A.M., Yadav, A.N., Yadav, N., Usmani, Z., Eds.; Springer Nature: Cham, Switzerland, 2021; Volume 1, pp. 295–334.
6. Moncalvo, J.M.; Ryvarden, L. A Nomenclatural Study of the Ganodermataceae Donk, Synopsis Fungorum 11; Fungiflora: Oslo, Norway, 1997; pp. 1–114.
7. Ryvarden, L.; Johansen, I. A Preliminary Polypore Flora of East Africa; Fungiflora: Oslo, Norway, 1980; pp. 1–636.
8. Gilbertson, R.L.; Ryvarden, L. North American polypores. Abortiporus-Lindtneria; Fungiflora A/S: Oslo, Norway, 1986; Volume 1, pp. 1–433.
9. Ryvarden, L.; Gilbertson, R.L. European Polypores 1, Synop Fungorum 6; Fungiflora A/S: Oslo, Norway, 1993; pp. 1–387.
10. Quanten, E. The Polypores (Polyporaceae s.l.) of Papua New Guinea: A Preliminary Conspectus, Opera Botanica Belgica 11; National Botanic Garden of Belgium: Meise, Belgium, 1997; pp. 1–352.
11. Nunez, M.; Ryvarden, L. East Asian Polypores 1. Ganodermataceae and Hymenochaetaceae, Synop Fungorum 13; Fungiflora A/S: Oslo, Norway, 2000; pp. 1–168.
12. Welti, S.; Courtecuisse, R. The Ganodermataceae in the French West Indies (Guadeloupe and Martinique). Fungal Divers. 2010, 43, 103–126.
13. Arenas, M.C.; Tadiosa, E.R.; Reyes, R.G. Taxonomic inventory based on physical distribution of macrofungi in Mt. Maculot, Cuenca, Batangas, Philippines. Int. J. Biol. Pharm. Allied Sci. 2018, 7, 672–687.

14. Luangharn, T.; Karunarathna, S.C.; Dutta, A.K.; Paloi, S.; Promputtha, I.; Hyde, K.D.; Xu, J.; Mortimer, P.E. *Ganoderma* (Ganodermataceae, Basidiomycota) species from the Greater Mekong Subregion. *J. Fungi* 2021, 7, 819.
15. Morera, G.; Lupo, S.; Alaniz, S.; Robledo, G. Diversity of the *Ganoderma* species in Uruguay. *Neotrop. Biodivers.* 2021, 7, 570–585.
16. Runnel, K.; Miettinen, O.; Lõhmus, A. Polypore fungi as a flagship group to indicate changes in biodiversity—A test case from Estonia. *IMA Fungus* 2021, 12, 2.
17. Ueitele, I.S.E.; Horn, L.N.; Kadhila, N.P. *Ganoderma* research activities and development in Namibia. *AJOM* 2021, 4, 29–39.
18. He, J.; Han, X.; Luo, Z.-L.; Li, E.-X.; Tang, S.-M.; Luo, H.-M.; Niu, K.-Y.; Su, X.-J.; Li, S.-H. Species diversity of *Ganoderma* (Ganodermataceae, Polyporales) with three new species and a key to *Ganoderma* in Yunnan province, China. *Front. Microbiol.* 2022, 13, 1035434.
19. Ying, C.-C.; Wang, Y.-C.; Tang, H. *Icones of Medicinal Fungi from China*; Science Press: Beijing, China, 1987; p. 575.
20. Dai, Y.C.; Yang, Z.L.; Cui, B.K.; Yu, C.J.; Zhou, L.W. Species diversity and utilization of medicinal mushrooms and fungi in China (review). *Int. J. Med. Mushrooms* 2009, 3, 287–302.
21. Wu, F.; Zhou, L.-W.; Yang, Z.-L.; Bau, T.; Li, T.-H.; Dai, Y.-C. Resource diversity of Chinese macrofungi: Edible, medicinal and poisonous species. *Fungal Divers.* 2019, 98, 1–76.
22. El Sheikha, A.F.E. Nutritional profile and health benefits of *Ganoderma lucidum* “Lingzhi, Reishi, or Mannentake” as functional foods: Current scenario and future perspectives. *Foods* 2022, 11, 1030.
23. Li, Y.; Zhu, Z.; Yao, W.; Chen, R. Research status and progress of the triterpenoids in *Ganoderma lucidum*. *Med. Plant* 2012, 3, 75–81.
24. Liu, H.; Guo, L.-J.; Li, S.-L.; Fan, L. *Ganoderma shanxiense*, a new species from northern China based on morphological and molecular evidence. *Phytotaxa* 2019, 406, 129–136.
25. Li, L.F.; Liu, H.B.; Zhang, Q.W.; Li, Z.P.; Wong, T.L.; Fung, H.Y.; Zhang, J.X.; Bai, S.P.; Lu, A.P.; Han, Q.B. Comprehensive comparison of polysaccharides from *Ganoderma lucidum* and *G. sinense*: Chemical, anti-tumor, immunomodulating and gut-microbiota modulatory properties. *Sci. Rep.* 2018, 8, 6172.
26. Ngai, P.H.K.; Ng, T.B. A mushroom (*Ganoderma capense*) lectin with spectacular thermostability, potent mitogenic activity on splenocytes, and antiproliferative activity toward tumor cells. *Biochem. Bioph. Res. Commun.* 2004, 314, 988–993.
27. Lee, S.; Shim, S.H.; Kim, J.S.; Shin, K.H.; Kang, S.S. Aldose reductase inhibitors from the fruiting bodies of *Ganoderma applanatum*. *Biol. Pharm. Bull.* 2005, 28, 1103–1105.

28. Gurunathan, S.; Raman, J.; Malek, S.N.A.; John, P.A.; Vikineswary, S. Green synthesis of silver nanoparticles using *Ganoderma neo-japonicum* Imazeki: A potential cytotoxic agent against breast cancer cells. *Int. J. Nanomed.* 2013, 8, 4399–4413.
29. Wang, L.; Li, J.-Q.; Zhang, J.; Li, Z.-M.; Liu, H.-G.; Wang, Y.-Z. Traditional uses, chemical components and pharmacological activities of the genus *Ganoderma* P. Karst: A review. *RSC Adv.* 2020, 10, 42084–42097.
30. Shankar, A.; Sharma, K.K. Fungal secondary metabolites in food and pharmaceuticals in the era of multiomics. *Appl. Microbiol. Biotechnol.* 2022, 106, 3465–3488.
31. Chen, Y.; Lan, P. Total syntheses and biological evaluation of the *Ganoderma lucidum* alkaloids lucidimines B and C. *ACS Omega* 2018, 3, 3471–3481.
32. Xia, Q.; Zhang, H.; Sun, X.; Zhao, H.; Wu, L.; Zhu, D.; Yang, G.; Shao, Y.; Zhang, X.; Mao, X.; et al. A comprehensive review of the structure elucidation and biological activity of triterpenoids from *Ganoderma* spp. *Molecules* 2014, 19, 17478–17535.
33. Baby, S.; Johnson, A.J.; Govindan, B. Secondary metabolites from *Ganoderma*. *Phytochemistry* 2015, 114, 66–101.
34. Wang, K.; Bao, L.; Xiong, W.; Ma, K.; Han, J.; Wang, W.; Yin, W.; Liu, H. Lanostane triterpenes from the Tibetan medicinal mushroom *Ganoderma leucocontextum* and their inhibitory effects on HMG-CoA reductase and α -glucosidase. *J. Nat. Prod.* 2015, 78, 1977–1989.
35. Angulo-Sanchez, L.T.; López-Peña, D.; Torres-Moreno, H.; Gutiérrez, A.; Gaitán-Hernández, R.; Esquedaa, M. Biosynthesis, gene expression, and pharmacological properties of triterpenoids of *Ganoderma* species (Agaricomycetes): A review. *Int. J. Med. Mushrooms* 2022, 24, 1–17.
36. Isaka, M.; Chinthanom, P.; Kongthong, S.; Srichomthong, K.; Choeyklin, R. Lanostane triterpenes from cultures of the basidiomycete *Ganoderma orbiforme* BCC 22324. *Phytochemistry* 2013, 87, 133–139.
37. Martínez-Montemayor, M.M.; Ling, T.; Suárez-Arroyo, I.J.; Ortiz-Soto, G.; Santiago-Negrón, C.L.; Lacourt-Ventura, M.Y.; Valentín-Acevedo, A.; Lang, W.H.; Rivas, F. Identification of biologically active *Ganoderma lucidum* compounds and synthesis of improved derivatives that confer anti-cancer activities in vitro. *Front. Pharmacol.* 2019, 10, 115.
38. Shim, S.H.; Ryu, J.; Kim, J.S.; Kang, S.S.; Xu, Y.; Jung, S.H.; Lee, Y.S.; Lee, S.; Shin, K.H. New lanostane-type triterpenoids from *Ganoderma applanatum*. *J. Nat. Prod.* 2004, 67, 1110–1113.
39. Paterson, R.R. *Ganoderma*—A therapeutic fungal biofactory. *Phytochemistry* 2006, 67, 1985–2001.
40. Cilerdzic, J.; Vukojevic, J.; Stajic, M.; Stanojkovic, T.; Glamoclija, J. Biological activity of *Ganoderma lucidum* basidiocarps cultivated on alternative and commercial substrate. *J.*

- Ethnopharmacol. 2014, 155, 312–329.
41. Peng, X.R.; Li, L.; Dong, J.R.; Lu, S.Y.; Lu, J.; Li, X.N.; Zhou, L.; Qiu, M.H. Lanostane-type triterpenoids from the fruiting bodies of *Ganoderma applanatum*. *Phytochemistry* 2019, 157, 103–110.
 42. Shi, J.-X.; Chen, G.-Y.; Sun, Q.; Meng, S.-Y.; Chi, W.-Q. Antimicrobial lanostane triterpenoids from the fruiting bodies of *Ganoderma applanatum*. *J. Asian Nat. Prod. Res.* 2021, 157, 1001–1007.
 43. Muhsin, T.M.; Al-Duboon, A.-H.A.; Khalaf, K.T. Bioactive compounds from a polypore fungus *Ganoderma applanatum* (Pers. ex Wallr.) Pat. *Jordan J. Biol. Sci.* 2011, 4, 205–212.
 44. Kozarski, M.; Klaus, A.; Nikšić, M.; Vrvic, M.M.; Todorović, N.; Jakovljević, D.; Van Griensven, L.J.L.D. Antioxidative activities and chemical characterization of polysaccharide extracts from the widely used mushrooms *Ganoderma applanatum*, *Ganoderma lucidum*, *Lentinus edodes* and *Trametes versicolor*. *J. Food Compost. Anal.* 2012, 26, 144–153.
 45. Al-Fatimi, M.; Wurster, M.; Kreisel, H.; Lindequist, U. Antimicrobial, cytotoxic and antioxidant activity of selected basidiomycetes from Yemen. *Pharmazie* 2005, 60, 776–780.
 46. Niedermeyer, T.H.; Lindequist, U.; Mentel, R.; Gordes, D.; Schmidt, E.; Thurow, K.; Lalk, M. Antiviral terpenoid constituents of *Ganoderma pfeifferi*. *J. Nat. Prod.* 2005, 68, 1728–1731.
 47. Hsu, C.L.; Yu, Y.S.; Yen, G.C. Lucidenic acid B induces apoptosis in human leukemia cells via a mitochondria-mediated pathway. *J. Agric. Food Chem.* 2008, 56, 3973–3980.
 48. Smania, E.F.A.; Delle Monache, F.; Smania, A.; Yunes, R.A.; Cuneo, R.S. Antifungal activity of sterols and triterpenes isolated from *Ganoderma annulare*. *Fitoterapia* 2003, 74, 375–377.
 49. Rosecke, J.; König, W.A. Constituents of various wood-rotting basidiomycetes. *Phytochemistry* 2000, 54, 603–610.
 50. Li, L.; Peng, X.R.; Dong, J.R.; Lu, S.Y.; Li, X.N.; Zhou, L.; Qiu, M.H. Rearranged lanostane-type triterpenoids with anti-hepatic fibrosis activities from *Ganoderma applanatum*. *RSC Adv.* 2018, 8, 31287–31295.
 51. Peng, X.-R.; Wang, Q.; Su, H.-G.; Zhou, L.; Xiong, W.-Y.; Qiu, M.-H. Anti-adipogenic lanostane-type triterpenoids from the edible and medicinal mushroom *Ganoderma applanatum*. *J. Fungi* 2022, 8, 331.
 52. Ma, K.; Ren, J.; Han, J.; Bao, L.; Li, L.; Yao, Y.; Sun, C.; Zhou, B.; Liu, H. Ganoboninketals A–C, antiplasmodial 3,4-seco-27-norlanostane triterpenes from *Ganoderma boninense* Pat. *J. Nat. Prod.* 2014, 77, 1847–1852.
 53. Huang, S.-Z.; Ma, Q.-Y.; Kong, F.-D.; Guo, Z.-K.; Cai, C.-H.; Hu, L.-L.; Zhou, L.-M.; Wang, Q.; Dai, H.-F.; Mei, W.-L.; et al. Lanostane-type triterpenoids from the fruiting body of *Ganoderma calidophilum*. *Phytochemistry* 2017, 143, 104–110.

54. Li, N.; Yan, C.; Hua, D.; Zhang, D. Isolation, purification, and structural characterization of a novel polysaccharide from *Ganoderma capense*. *Int. J. Biol. Macromol.* 2013, 57, 285–290.
55. Keller, A.C.; Keller, J.; Maillard, M.P.; Hostettmann, K. A lanostane-type steroid from the fungus *Ganoderma carnosum*. *Phytochemistry* 1997, 46, 963–965.
56. Peng, X.-R.; Liu, J.-Q.; Wan, L.-S.; Li, X.-N.; Yan, Y.-X.; Qiu, M.-H. Four new polycyclic meroterpenoids from *Ganoderma cochlear*. *Org. Lett.* 2014, 16, 5262–5265.
57. Peng, X.-R.; Liu, J.-Q.; Wang, C.-F.; Li, X.-Y.; Shu, Y.; Zhou, L.; Qiu, M.-H. Hepatoprotective effects of triterpenoids from *Ganoderma cochlear*. *J. Nat. Prod.* 2014, 77, 737–743.
58. Yu, C.; Cao, C.Y.; Shi, P.D.; Yang, A.A.; Yang, Y.X.; Huang, D.S.; Chen, X.; Chen, Z.M.; Gao, J.M.; Yin, X. Highly oxygenated chemical constitutes and rearranged derivatives with neurotrophic activity from *Ganoderma cochlear*. *J. Ethnopharmacol.* 2022, 295, 115393.
59. Chen, S.-Y.; Chang, C.-L.; Chen, T.-H.; Chang, Y.-W.; Lin, S.-B. Colossolactone H, a new *Ganoderma* triterpenoid exhibits cytotoxicity and potentiates drug efficacy of gefitinib in lung cancer. *Fitoterapia* 2016, 114, 81–91.
60. Jo, W.-S.; Park, H.-N.; Cho, D.-H.; Yoo, Y.-B.; Park, S.-C. Detection of extracellular enzyme activities in *Ganoderma neo-japonicum*. *Mycobiology* 2011, 39, 118–120.
61. Qiao, Y.; Zhang, X.-M.; Dong, X.-C.; Qiu, M.-H. A new 18(13 → 12 β)-abeo-lanostadiene triterpenoid from *Ganoderma fornicatum*. *Helv. Chim. Acta* 2006, 89, 1038–1041.
62. Peng, X.; Liu, J.; Xia, J.; Wang, C.; Li, X.; Deng, Y.; Bao, N.; Zhang, Z.; Qiu, M. Lanostane triterpenoids from *Ganoderma hainanense* J. D. Zhao. *Phytochemistry* 2015, 114, 137–145.
63. Li, T.-H.; Hu, H.-P.; Deng, W.-Q.; Wu, S.-H.; Wang, D.-M.; Tsering, T. *Ganoderma leucocontextum*, a new member of the *G. lucidum* complex from southwestern China. *Mycoscience* 2015, 56, 81–85.
64. Wei, J.-C.; Wang, Y.-X.; Dai, R.; Tian, X.-G.; Sun, C.-P.; Ma, X.-C.; Jia, J.-M.; Zhang, B.-J.; Huo, X.-K.; Wang, C. C27-Nor lanostane triterpenoids of the fungus *Ganoderma lucidum* and their inhibitory effects on acetylcholinesterase. *Phytochem. Lett.* 2017, 20, 263–268.
65. Yang, Y.; Zhang, H.; Zuo, J.; Gong, X.; Yi, F.; Zhu, W.; Li, L. Advances in research on the active constituents and physiological effects of *Ganoderma lucidum*. *Biomed. Dermatol.* 2019, 3, 6.
66. Li, D.; Leng, Y.; Liao, Z.; Hu, J.; Sun, Y.; Deng, S.; Wang, C.; Tian, X.; Zhou, J.; Wang, R. Nor-triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their inhibitory activity against FAAH. *Nat. Prod. Res.* 2022, 158, 105161.
67. Su, H.; Peng, X.; Shi, Q.; Huang, Y.; Zhou, L.; Qiu, M. Lanostane triterpenoids with anti-inflammatory activities from *Ganoderma lucidum*. *Phytochemistry* 2020, 173, 112256.

68. Hirotani, M.; Ino, C.; Hatano, A.; Takayanagi, H.; Furuya, T. Ganomastenols A, B, C and D, cadinene sesquiterpenes, from *Ganoderma mastoporum*. *Phytochemistry* 1995, 40, 161–165.
69. Yangchum, A.; Fujii, R.; Choowong, W.; Rachtawee, P.; Pobkwamsuk, M.; Boonpratuang, T.; Mori, S.; Isaka, M. Lanostane triterpenoids from cultivated fruiting bodies of basidiomycete *Ganoderma mbrekobenum*. *Phytochemistry* 2022, 196, 113075.
70. Chen, X.Q.; Chen, L.X.; Zhao, J.; Tang, Y.P.; Li, S.P. Nortriterpenoids from the fruiting bodies of the mushroom *Ganoderma resinaceum*. *Molecules* 2017, 22, 1073.
71. Liu, C.; Zhao, F.; Chen, R. A novel alkaloid from the fruiting bodies of *Ganoderma sinense* Zhao, Xu et Zhang. *Chin. Chem. Lett.* 2010, 21, 197–199.
72. Liu, J.-Q.; Wang, C.-F.; Peng, X.-R.; Qiu, M.-H. New alkaloids from the fruiting bodies of *Ganoderma sinense*. *Nat. Prod. Bioprospect.* 2011, 1, 93–96.
73. Da, J.; Wu, W.-Y.; Hou, J.-J.; Long, H.-L.; Yao, S.; Yang, Z.; Cai, L.-Y.; Yang, M.; Jiang, B.-H.; Liu, X.; et al. Comparison of two officinal Chinese pharmacopoeia species of *Ganoderma* based on chemical research with multiple technologies and chemometrics analysis. *J. Chromatogr. A* 2012, 1222, 59–70.
74. Liu, L.-Y.; Chen, H.; Liu, C.; Wang, H.-Q.; Kang, J.; Li, Y.; Chen, R.-Y. Triterpenoids of *Ganoderma theaecolum* and their hepatoprotective activities. *Fitoterapia* 2014, 98, 254–259.
75. Ma, Q.-Y.; Luo, Y.; Huang, S.-Z.; Guo, Z.-K.; Dai, H.-F.; Zhao, Y.-X. Lanostane triterpenoids with cytotoxic activities from the fruiting bodies of *Ganoderma hainanense*. *J. Asian Nat. Prod. Res.* 2013, 15, 1214–1219.
76. Chien, R.-C.; Tsai, S.-Y.; Lai, E.Y.-C.; Mau, J.-L. Antiproliferative activities of hot water extracts from culinary-medicinal mushrooms, *Ganoderma tsugae* and *Agrocybe cylindracea* (Higher Basidiomycetes) on cancer cells. *Int. J. Med. Mushrooms* 2015, 17, 453–462.
77. Yang, L.; Kong, D.-X.; Xiao, N.; Ma, Q.-Y.; Xie, Q.-Y.; Guo, J.-C.; Ying Deng, C.; Ma, H.-X.; Hua, Y.; Dai, H.-F.; et al. Antidiabetic lanostane triterpenoids from the fruiting bodies of *Ganoderma weberianum*. *Bioorg. Chem.* 2022, 127, 106025.
78. Zhang, J.; Liu, Y.; Tang, Q.; Zhou, S.; Feng, J.; Chen, H. Polysaccharide of *Ganoderma* and its bioactivities. In *Ganoderma and Health: Advances in Experimental Medicine and Biology*; Lin, Z., Yang, B., Eds.; Springer: Singapore, 2019; Volume 1181, pp. 107–134.
79. Chan, J.S.; Asatiani, M.D.; Sharvit, L.E.; Trabelcy, B.; Barseghyan, G.S.; Wasser, S.P. Chemical composition and medicinal value of the new *Ganoderma tsugae* var. *jannieae* CBS-120304 medicinal higher basidiomycete mushroom. *Int. J. Med. Mushrooms* 2015, 17, 735–747.
80. Ahmad, R.; Riaz, M.; Khan, A.; Aljamea, A.; Algheryafi, M.; Sewaket, D.; Alqathama, A. *Ganoderma lucidum* (Reishi) an edible mushroom; a comprehensive and critical review of its

- nutritional, cosmeceutical, mycochemical, pharmacological, clinical, and toxicological properties. *Phytother. Res.* 2021, 35, 6030–6062.
81. Kolniak-Ostek, J.; Oszmianski, J.; Szyjka, A.; Moreira, H.; Barg, E. Anticancer and antioxidant activities in *Ganoderma lucidum* wild mushrooms in Poland, as well as their phenolic and triterpenoid compounds. *Int. J. Mol. Sci.* 2022, 23, 9359.
 82. Bhat, Z.A.; Wani, A.H.; War, J.M.; Bhat, M.Y. Major bioactive properties of *Ganoderma* polysaccharides: A review. *Asian J. Pharm. Clin. Res.* 2021, 14, 11–24.
 83. Gallo, A.L.; Soler, F.; Pellizas, C.; Vélez, M.L. Polysaccharide extracts from mycelia of *Ganoderma australe*: Effect on dendritic cell immunomodulation and antioxidant activity. *J. Food Meas. Charact.* 2022, 16, 3251–3262.
 84. Veena, R.K.; Janardhanan, K.K. Polysaccharide-protein complex isolated from fruiting bodies and cultured mycelia of Lingzhi or reishi medicinal mushroom, *Ganoderma lucidum* (agaricomycetes), attenuates doxorubicin-induced oxidative stress and myocardial injury in rats. *Int. J. Med. Mushrooms* 2022, 24, 31–40.
 85. Cragg, G.M.; Kingston, D.G.I.; Newman, D.J. *Anticancer Agents from Natural Products*, 1st ed.; CRC Press: Boca Raton, FL, USA, 2005; pp. 1–600.
 86. Mfopa, A.; Mediesse, F.K.; Mvongo, C.; Nkoubatchoundjwen, S.; Lum, A.A.; Sobngwi, E.; Kamgang, R.; Boudjeko, T. Antidyslipidemic potential of water-soluble polysaccharides of *Ganoderma applanatum* in MACAPOS-2-induced obese rats. *Evid. Based Complement. Altern. Med.* 2021, 2021, 2452057.
 87. Seweryn, E.; Ziała, A.; Gamian, A. Health-promoting of polysaccharides extracted from *Ganoderma lucidum*. *Nutrients* 2021, 13, 2725.
 88. Bleha, R.; Třešnáková, L.; Sushytskyi, L.; Capek, P.; Čopíková, J.; Klouček, P.; Jablonský, I.; Synytsya, A. Polysaccharides from basidiocarps of the polypore fungus *Ganoderma resinaceum*: Isolation and structure. *Polymers* 2022, 14, 255.
 89. Loyd, A.L.; Barnes, C.W.; Held, B.W.; Schink, M.J.; Smith, M.E.; Smith, J.A.; Blanchette, R.A. Elucidating “*lucidum*”: Distinguishing the diverse laccate *Ganoderma* species of the United States. *PLoS ONE* 2018, 13, e0199738.
 90. Jargalmaa, S.; Eimes, J.A.; Park, M.S.; Park, J.Y.; Oh, S.-Y.; Lim, Y.W. Taxonomic evaluation of selected *Ganoderma* species and database sequence validation. *PeerJ* 2017, 5, e3596.
 91. Tchet Tchoumi, J.M.; Coetzee, M.P.; Rajchenberg, M.; Roux, J. Taxonomy and species diversity of *Ganoderma* species in the Garden Route National Park of South Africa inferred from morphology and multilocus phylogenies. *Mycologia* 2019, 111, 730–747.
 92. Hou, D. A new species of *Ganoderma* from Taiwan. *Quat. J. Taiwan Mus.* 1950, 3, 101–105.

93. Zhao, J.D.; Xu, L.W.; Zhang, X.Q. Taxonomic studies on the family Ganodermataceae of China II. *Acta Mycol. Sin.* 1983, 2, 159–167.
94. Cao, Y.; Wu, S.H.; Dai, Y.C. Species clarification of the prize medicinal Ganoderma mushroom “Lingzhi”. *Fungal Divers.* 2012, 56, 49–62.
95. Wu, G.-S.; Lu, J.J.; Guo, J.-J.; Li, Y.-B.; Tan, W.; Dang, Y.-Y.; Zhong, Z.-F.; Xu, Z.-T.; Chen, X.-P.; Wang, Y.-T. Ganoderic acid DM, a natural triterpenoid, induces DNA damage, G1 cell cycle arrest and apoptosis in human breast cancer cells. *Fitoterapia* 2012, 83, 408–414.
96. Patouillard, N.T. Le genre *Ganoderma*. *Bull. Soc. Mycol. Fr.* 1889, 5, 64–80.
97. Murrill, W.A. The polyporaceae of North America. I. The genus *Ganoderma*. *Bull. Torrey Bot. Club* 1902, 29, 599–608.
98. Murrill, W.A. Polyporaceae. *North American Flora; The New York Botanical Garden: New York, NY, USA*, 1908; Volume 9, pp. 73–131.
99. Zhou, L.-W.; Cao, Y.; Wu, S.-H.; Vlasák, J.; Li, D.-W.; Li, M.-J.; Dai, Y.-C. Global diversity of the *Ganoderma lucidum* complex (Ganodermataceae, Polyporales) inferred from morphology and multilocus phylogeny. *Phytochemistry* 2015, 114, 7–15.
100. Du, Z.; Dong, C.-H.; Wang, K.; Yao, Y.-J. Classification, biological characteristics and cultivations of *Ganoderma*. In *Ganoderma and Health: Advances in Experimental Medicine and Biology*; Lin, Z., Yang, B., Eds.; Springer: Singapore, 2019; Volume 1181, pp. 15–58.
101. Zhang, X.; Xu, Z.; Pei, H.; Chen, Z.; Tan, X.; Hu, J.; Yang, B.; Sun, J. Intraspecific variation and phylogenetic relationships are revealed by ITS1 secondary structure analysis and single-nucleotide polymorphism in *Ganoderma lucidum*. *PLoS ONE* 2017, 12, e0169042.
102. Loyd, A.L.; Smith, J.A.; Richter, B.S.; Blanchette, R.A.; Smith, M.E. The laccate *Ganoderma* of the South-Eastern United States: A cosmopolitan and important genus of wood decay fungi. *EDIS* 2017, 2017, 6.
103. Anonymous. *Shen Nong Materia Medica*; Beijing People's Hygiene Press: Beijing, China, 1955.
104. Moncalvo, J.M.; Wang, H.F.; Hseu, R.S. Gene phylogeny of the *Ganoderma lucidum* complex based on ribosomal DNA sequences. Comparison with traditional taxonomic characters. *Mycol. Res.* 1995, 99, 1489–1499.
105. Hapuarachchi, K.K.; Wen, T.C.; Deng, C.Y.; Kang, J.C.; Hyde, K.D. *Mycosphere Essays 1: Taxonomic confusion in the Ganoderma lucidum species complex*. *Mycosphere* 2015, 6, 542–559.
106. Haddow, W.R. Studies in *Ganoderma*. *J. Arnold Arbor.* 1931, 12, 25–46.

107. Overholts, L.O. The Polyporaceae of the United States, Alaska and Canada; University of Michigan Press: Ann Arbor, MI, USA, 1953; pp. 1–466.
108. Steyaert, R.L. Basidiospores of two *Ganoderma* Species and others of two related genera under the scanning electron microscope. *Kew Bull.* 1977, 31, 437–442.
109. Nobles, M.K. Identification of cultures of wood-inhabiting Hymenomycetes. *Can. J. Bot.* 1965, 43, 1097–1139.
110. Wang, D.M.; Wu, S.H.; Su, C.H.; Peng, J.T.; Shih, Y.H. *Ganoderma multipileum*, the correct name for “*G. lucidum*” in tropical Asia. *Bot. Stud.* 2009, 50, 451–458.
111. Wang, X.-C.; Xi, R.-J.; Li, Y.; Wang, D.-M.; Yao, Y.-J. The species identity of the widely cultivated *Ganoderma*, ‘*G. lucidum*’ (Ling-zhi), in China. *PLoS ONE* 2012, 7, e40857.
112. Yang, L.Z.; Feng, B. What is the Chinese “Lingzhi”?—A taxonomic mini-review. *Mycology* 2013, 4, 1–4.
113. Talapatra, S.K.; Talapatra, B. Triterpenes (C30). In *Chemistry of Plant Natural Products: Stereochemistry, Conformation, Synthesis, Biology, and Medicine*, 1st ed.; Talapatra, S.K., Talapatra, B., Eds.; Springer: Berlin/Heidelberg, Germany, 2014; pp. 517–552.
114. Wang, Q.; Cao, R.; Zhang, Y. Biosynthesis and regulation of terpenoids from basidiomycetes: Exploration of new research. *AMB Expr.* 2021, 11, 150.
115. Yang, M.; Wang, X.; Guan, S.; Xia, J.; Sun, J.; Guo, H.; Guo, D.A. Analysis of triterpenoids in *Ganoderma lucidum* using liquid chromatography coupled with electrospray ionization mass spectrometry. *J. Am. Soc. Mass Spectr.* 2007, 18, 927–939.
116. Lin, Y.-X.; Sun, J.-T.; Liao, Z.-Z.; Sun, Y.; Tian, X.-G.; Jin, L.-L.; Wang, C.; Leng, A.-J.; Zhou, J.; Li, D.-W. Triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their inhibitory activity against FAAH. *Fitoterapia* 2022, 158, 105161.
117. Sappan, M.; Rachtawee, P.; Srichomthong, K.; Boonpratuang, T.; Choeyklin, R.; Feng, T.; Liu, J.-K.; Isaka, M. Ganoellipsic acids A–C, lanostane triterpenoids from artificially cultivated fruiting bodies of *Ganoderma ellipsoideum*. *Phytochem. Lett.* 2022, 49, 27–31.
118. Hirotani, M.; Asaka, I.; Furuya, T. Investigation of the biosynthesis of 3 α -hydroxy triterpenoids, ganoderic acids T and S, by application of a feeding experiment using acetate. *J. Chem. Soc. Perkin Trans.* 1990, 1, 2751–2754.
119. Noushahi, H.A.; Khan, A.H.; Noushahi, U.F.; Hussain, M.; Javed, T.; Zafar, M.; Batool, M.; Ahmed, U.; Liu, K.; Harrison, M.T.; et al. Biosynthetic pathways of triterpenoids and strategies to improve their biosynthetic efficiency. *Plant Growth Regul.* 2022, 97, 439–454.
120. Vranová, E.; Coman, D.; Gruissem, W. Network analysis of the MVA and MEP pathways for isoprenoid synthesis. *Annu. Rev. Plant Biol.* 2013, 64, 665–700.

121. McGarvey, D.J.; Croteau, R. Terpenoid metabolism. *Plant Cell* 1995, 7, 1015–1026.
122. Shi, L.; Ren, A.; Mu, D.; Zhao, M. Current progress in the study on biosynthesis and regulation of ganoderic acids. *Appl. Microbiol. Biot.* 2010, 88, 1243–1251.
123. Liu, D.; Gong, J.; Dai, W.; Kang, X.; Huang, Z.; Zhang, H.M.; Liu, W.; Liu, L.; Ma, J.; Xia, Z.; et al. The genome of *Ganoderma lucidum* provides insights into triterpene biosynthesis and wood degradation. *PLoS ONE* 2012, 7, e36146.
124. Shang, C.-H.; Zhu, F.; Li, N.; Ou-Yang, X.; Shi, L.; Zhao, M.-W.; Li, Y.-X. Cloning and characterization of a gene encoding HMG-CoA reductase from *Ganoderma lucidum* and its functional identification in yeast. *Biosci. Biotechnol. Biochem.* 2008, 72, 1333–1339.
125. Ding, Y.-X.; Ou-Yang, X.; Shang, C.-H.; Ren, A.; Shi, L.; Li, Y.-X.; Zhao, M.-W. Molecular cloning, characterization, and differential expression of a farnesyl-diphosphate synthase gene from the basidiomycetous fungus *Ganoderma lucidum*. *Biosci. Biotechnol. Biochem.* 2008, 72, 1571–1579.
126. Zhao, M.-W.; Liang, W.-Q.; Zhang, D.-B.; Wang, N.; Wang, C.-G.; Pan, Y.-J. Cloning and characterization of Squalene Synthase (SQS) gene from *Ganoderma lucidum*. *J. Microbiol. Biotechnol.* 2007, 17, 1106–1112.
127. Ye, L.; Liu, S.; Xie, F.; Zhao, L.; Wu, X. Enhanced production of polysaccharides and triterpenoids in *Ganoderma lucidum* fruit bodies on induction with signal transduction during the fruiting stage. *PLoS ONE* 2018, 13, e0196287.
128. Fei, Y.; Li, N.; Zhang, D.H.; Xu, J.W. Increased production of ganoderic acids by overexpression of homologous farnesyl diphosphate synthase and kinetic modeling of ganoderic acid production in *Ganoderma lucidum*. *Microb. Cell Fact.* 2019, 18, 115.
129. Shi, L.; Qin, L.; Xu, Y.; Ren, A.; Fang, X.; Mu, D.; Tan, Q.; Zhao, M. Molecular cloning, characterization, and function analysis of a mevalonate pyrophosphate decarboxylase gene from *Ganoderma lucidum*. *Mol. Biol. Rep.* 2012, 39, 6149–6159.
130. Zhang, D.H.; Jiang, L.X.; Li, N.; Yu, X.; Zhao, P.; Li, T.; Xu, J.W. Overexpression of the squalene epoxidase gene alone and in combination with the 3-hydroxy-3-methylglutaryl coenzyme A gene increases ganoderic acid production in *Ganoderma lingzhi*. *J. Agric. Food Chem.* 2017, 65, 4683–4690.
131. Zhang, D.H.; Li, N.; Yu, X.Y.; Zhao, P.; Li, T.; Xu, J.W. Overexpression of the homologous lanosterol synthase gene in ganoderic acid biosynthesis in *Ganoderma lingzhi*. *Phytochemistry* 2017, 134, 46–53.
132. Howes, M.R. Phytochemicals as anti-inflammatory nutraceuticals and phytopharmaceuticals. In *Immunity and Inflammation in Health and Disease: Emerging Roles of Nutraceuticals and Functional Foods in Immune Support*; Chatterjee, S., Jungraithmayr, W., Bagchi, D., Eds.; Academic Press: Cambridge, MA, USA, 2018; pp. 363–388.

133. Ludwiczuk, A.; Skalicka-Woźniak, K.; Georgiev, M. Terpenoids. In *Pharmacognosy: Fundamentals, Applications and Strategies*, 1st ed.; Badal, S., Delgoda, R., Eds.; Academic Press: London, UK, 2017; pp. 233–266.
134. Muffler, K.; Leipold, D.; Scheller, M.C.; Haas, C.; Steingroewer, J.; Bley, T.; Neuhaus, H.E.; Mirata, M.A.; Schrader, J.; Ulber, R. Biotransformation of triterpenes. *Process Biochem.* 2011, 46, 1–15.
135. Blundell, R.; Azzopardi, J.; Briffa, J.; Rasul, A.; Vargas-de la Cruz, C.; Shah, M.A. Analysis of pentaterpenoids. In *Recent Advances in Natural Products Analysis*, 1st ed.; Nabavi, S.M., Saeedi, M., Nabavi, S.F., Silva, A.S., Eds.; Elsevier: Amsterdam, The Netherlands, 2020; pp. 457–475.
136. Gong, T.; Yan, R.; Kang, J.; Chen, R. Chemical components of Ganoderma. In *Ganoderma and Health: Advances in Experimental Medicine and Biology*; Lin, Z., Yang, B., Eds.; Springer: Singapore, 2019; Volume 1181, pp. 59–106.
137. Chen, R.; Kang, J. Quantitative analysis of components in Ganoderma. In *Ganoderma and Health: Advances in Experimental Medicine and Biology*; Lin, Z., Yang, B., Eds.; Springer: Singapore, 2019; Volume 1181, pp. 135–155.
138. Du, Q.; Cao, Y.; Liu, C. Lingzhi; an overview. In *The Lingzhi Mushroom Genome. Compendium of Plant Genomes*, 1st ed.; Liu, C., Ed.; Springer: Cham, Switzerland, 2021; pp. 1–25.
139. Ma, B.; Ren, W.; Zhou, Y.; Ma, J.; Ruan, Y.; Wen, C.N. Triterpenoids from the spores of *Ganoderma lucidum*. *N. Am. J. Med. Sci.* 2011, 3, 495–498.
140. Liu, R.-M.; Li, Y.-B.; Liang, X.-F.; Liu, H.-Z.; Xiao, J.-H.; Zhong, J.-J. Structurally related ganoderic acids induce apoptosis in human cervical cancer HeLa cells: Involvement of oxidative stress and antioxidant protective system. *Chem. Biol. Interact.* 2015, 240, 134–144.
141. Yang, S.-X.; Yu, Z.-C.; Lu, Q.-Q.; Shi, W.-Q.; Laatsch, H.; Gao, J.-M. Toxic lanostane triterpenes from the basidiomycete *Ganoderma amboinense*. *Phytochem. Lett.* 2012, 5, 576–580.
142. Lee, I.S.; Ahn, B.R.; Choi, J.S.; Hattori, M.; Min, B.S.; Bae, K.H. Selective cholinesterase inhibition by lanostane triterpenes from fruiting bodies of *Ganoderma lucidum*. *Bioorg. Med. Chem. Lett.* 2011, 21, 6603–6607.
143. Liu, J.; Kurashiki, K.; Shimizu, K.; Kondo, R. 5 α -Reductase inhibitory effect of triterpenoids isolated from *Ganoderma lucidum*. *Biol. Pharm. Bull.* 2006, 29, 392–395.
144. Liu, J.; Kurashiki, K.; Shimizu, K.; Kondo, R. Structure-activity relationship for inhibition of alpha-reductase by triterpenoids isolated from *Ganoderma lucidum*. *Bioorg. Med. Chem.* 2006, 14, 8654–8660.
145. Liu, J.; Shiono, J.; Shimizu, K.; Kukita, A.; Kukita, T.; Kondo, R. Ganoderic acid DM: Anti-androgenic osteoclastogenesis inhibitor. *Bioorg. Med. Chem. Lett.* 2009, 19, 2154–2157.

146. Liu, J.; Shiono, J.; Tsuji, Y.; Shimizu, K.; Kondo, R. Methyl ganoderic acid DM: A selective potent osteoclastogenesis inhibitor. *Open Bioact. Compd. J.* 2009, 2, 37–42.
147. Miyamoto, I.; Liu, J.; Shimizu, K.; Sato, M.; Kukita, A.; Kukita, T.; Kondo, R. Regulation of osteoclastogenesis by ganoderic acid DM isolated from *Ganoderma lucidum*. *Eur. J. Pharmacol.* 2009, 602, 1–7.
148. Chen, D.-H.; Chen, W.K.-D. Determination of ganoderic acids in triterpenoid constituents of *Ganoderma tsugae*. *J. Food Drug Anal.* 2003, 11, 195–201.
149. Kubota, T.; Asaka, Y.; Miura, I.; Mori, H. Structures of ganoderic acid A and B, two new lanostane type bitter triterpenes from *Ganoderma lucidum* (FR.) Karst. *Helv. Chim. Acta* 1982, 65, 611–619.
150. Yao, X.; Li, G.; Xu, H.; Lu, C. Inhibition of the JAK-STAT3 signaling pathway by ganoderic acid A enhances chemosensitivity of HepG2 cells to cisplatin. *Planta Med.* 2012, 78, 1740–1748.
151. El-Mekkawy, S.; Meselhy, M.R.; Nakamura, N.; Tezuka, Y.; Hattori, M.; Kakiuchi, N.; Shimotohno, K.; Kawahata, T.; Otake, T. Anti-HIV-1 and anti-HIV-1-protease substances from *Ganoderma lucidum*. *Phytochemistry* 1998, 49, 1651–1657.
152. Liu, C.; Yang, N.; Folder, W.; Cohn, J.; Wang, R.; Li, X. Ganoderic acid C isolated from *Ganoderma lucidum* suppress Lps-induced macrophage Tnf- α production by down-regulating Mapk, Nf-kappab and Ap-1 signaling pathways. *J. Allergy. Clin. Immunol.* 2012, 129, AB127.
153. Kikuchi, T.; Matsuda, S.; Kadota, S.; Ogita, Z. Ganoderic acid G and I and ganolucidic acid A and B, new triterpenoids from *Ganoderma lucidum*. *Chem. Pharm. Bull.* 1985, 33, 2628–2631.
154. Koyama, K.; Imaizumi, T.; Akiba, M.; Kinoshita, K.; Takahashi, K.; Suzuki, A.; Yano, S.; Horie, S.; Watanabe, K.; Naoi, Y. Antinociceptive components of *Ganoderma lucidum*. *Planta Med.* 1997, 63, 224–227.
155. Ruan, W.; Lim, A.H.H.; Huang, L.G.; Popovich, D.G. Extraction optimisation and isolation of triterpenoids from *Ganoderma lucidum* and their effect on human carcinoma cell growth. *Nat. Prod. Res.* 2014, 28, 2264–2272.
156. Hennicke, F.; Cheikh-Ali, Z.; Liebisch, T.; Maciá-Vicente, J.G.; Bode, H.B.; Piepenbring, M. Distinguishing commercially grown *Ganoderma lucidum* from *Ganoderma lingzhi* from Europe and East Asia on the basis of morphology, molecular phylogeny, and triterpenic acid profiles. *Phytochemistry* 2016, 127, 29–37.
157. Kikuchi, T.; Kanomi, S.; Murai, Y.; Kadota, S.; Tsubono, K.; Ogita, Z.I. Constituents of the fungus *Ganoderma lucidum* (FR.) Karst. III. Structures of ganolucidic acids A and B, new lanostane-type triterpenoids. *Chem. Pharm. Bull.* 1986, 34, 4030–4036.
158. Nishitoba, T.; Sato, H.; Sakamura, S. Triterpenoids from the fungus *Ganoderma lucidum*. *Phytochemistry* 1987, 26, 1777–1784.

159. Li, Y.-Y.; Mi, Z.-Y.; Tang, Y.; Wang, G.; Li, D.-S.; Tang, Y.-J. Lanostanoids isolated from *Ganoderma lucidum* mycelium cultured by submerged fermentation. *Helv. Chim. Acta* 2009, 92, 1586–1593.
160. Komoda, Y.; Nakamura, H.; Ishihara, S.; Uchida, M.; Kohda, H.; Yamasaki, K. Structures of new terpenoid constituents of *Ganoderma lucidum* (Fr.) Karst (Polyporaceae). *Chem. Pharm. Bull.* 1985, 33, 4829–4835.
161. Wang, F.; Liu, J.K. Highly oxygenated lanostane triterpenoids from the fungus *Ganoderma applanatum*. *Chem. Pharm. Bull.* 2008, 56, 1035–1037.
162. Yue, Q.-X.; Song, X.-Y.; Ma, C.; Feng, L.-X.; Guan, S.-H.; Wu, W.-Y.; Yang, M.; Jiang, B.-H.; Liu, X.; Cui, Y.-J.; et al. Effects of triterpenes from *Ganoderma lucidum* on protein expression profile of HeLa cells. *Phytomedicine* 2010, 17, 606–613.
163. Wu, T.-S.; Shi, L.-S.; Kuo, S.-C. Cytotoxicity of *Ganoderma lucidum* triterpenes. *J. Nat. Prod.* 2001, 64, 1121–1122.
164. Fatmawati, S.; Shimizu, K.; Kondo, R. Ganoderic acid Df, a new triterpenoid with aldose reductase inhibitory activity from the fruiting body of *Ganoderma lucidum*. *Fitoterapia* 2010, 81, 1033–1036.
165. Chen, R.-Y.; Yu, D.-Q. Studies on the triterpenoid constituents of the spores of *Ganoderma lucidum* (Curt.: Fr.) P. Karst. (Aphylllophoromycetideae). *Int. J. Med. Mushrooms* 1999, 1, 147–152.
166. Kohda, H.; Tokumoto, W.; Sakamoto, K.; Fujii, M.; Hirai, Y.; Yamasaki, K.; Komoda, Y.; Nakamura, H.; Ishihara, S. The biologically active constituents of *Ganoderma lucidum* (FR.) Karst. Histamine release-inhibitory triterpenes. *Chem. Pharm. Bull.* 1985, 33, 1367–1374.
167. Lee, I.S.; Seo, J.J.; Kim, J.P.; Kim, H.J.; Youn, U.J.; Lee, J.S.; Jung, H.J.; Na, M.K.; Hattori, M.; Min, B.S.; et al. Lanostane triterpenes from the fruiting bodies of *Ganoderma lucidum* and their inhibitory effects on adipocyte differentiation in 3T3-L1 cells. *J. Nat. Prod.* 2010, 73, 172–176.
168. Kikuchi, T.; Matsuda, S.; Kadota, S.; Murai, Y.; Ogita, Z. Ganoderic acid D, E, F, and H and lucidenic acid D, E, and F, new triterpenoids from *Ganoderma lucidum*. *Chem. Pharm. Bull.* 1985, 33, 2624–2627.
169. Iwatsuki, K.; Akihisa, T.; Tokuda, H.; Ukiya, M.; Oshikubo, M.; Kimura, Y.; Asano, T.; Nomura, A.; Nishino, H. Lucidenic acids P and Q, methyl lucidenate P, and other triterpenoids from the fungus *Ganoderma lucidum* and their inhibitory effects on epstein-barr virus activation. *J. Nat. Prod.* 2003, 66, 1582–1585.
170. Liu, D.-Z.; Zhu, Y.-Q.; Li, X.-F.; Shan, W.-G.; Gao, P.-F. New triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their bioactivities. *Chem. Biodivers.* 2014, 11, 982–986.

171. Hu, L.-L.; Ma, Q.-Y.; Huang, S.-Z.; Guo, Z.-K.; Ma, H.-X.; Guo, J.-C.; Dai, H.-F.; Zhao, Y.-X. Three new lanostanoid triterpenes from the fruiting bodies of *Ganoderma tropicum*. *J. Asian Nat. Prod. Res.* 2013, 15, 357–362.
172. Nishitoba, T.; Goto, S.; Sato, H.; Sakamura, S. Bitter triterpenoids from the fungus *Ganoderma applanatum*. *Phytochemistry* 1989, 28, 193–197.
173. Toth, J.O.; Luu, B.; Ourisson, G. Les acides ganoderiques t  : Triterpenes cytotoxiques de *Ganoderma lucidum* (Polyporac  e). *Tetrahedron Lett.* 1983, 24, 1081–1084.
174. Nishitoba, T.; Sato, H.; Sakamura, S. Novel mycelial components, ganoderic acid Mg, Mh, Mi, Mj, and Mk, from the fungus *Ganoderma lucidum*. *Agric. Biol. Chem.* 1987, 51, 1149–1153.
175. Johnson, B.M.; Doonan, B.P.; Radwan, F.F.; Haque, A. Ganoderic acid DM: An alternative agent for the treatment of advanced prostate cancer. *Open Prostate Cancer J.* 2010, 3, 78–85.
176. Cai, H.; Wang, F.S.; Yang, J.S.; Zhang, Y.M. Structure of ganoderic acid DM, a new triterpenoid from the fruiting body of *Ganoderma lucidum*. *Chin. Chem. Lett.* 1995, 6, 1051–1052.
177. Sato, N.; Zhang, Q.; Ma, C.-M.; Hattori, M. Anti-human immunodeficiency virus-1 protease activity of new lanostane-type triterpenoids from *Ganoderma sinense*. *Chem. Pharm. Bull.* 2009, 57, 1076–1080.
178. Peng, X.-R.; Liu, J.-Q.; Han, Z.-H.; Yuan, X.-X.; Luo, H.-R.; Qiu, M.-H. Protective effects of triterpenoids from *Ganoderma resinaceum* on H₂O₂-induced toxicity in HepG2 cells. *Food Chem.* 2013, 141, 920–926.
179. Nishitoba, T.; Sato, H.; Shirasu, S.; Sakamura, S. Novel triterpenoids from the mycelial mat at the previous stage of fruiting of *Ganoderma lucidum*. *Agric. Biol. Chem.* 1987, 51, 619–622.
180. Li, Y.-B.; Liu, R.-M.; Zhong, J.-J. A new ganoderic acid from *Ganoderma lucidum* mycelia and its stability. *Fitoterapia* 2013, 84, 115–122.
181. Wang, J.-L.; Li, Y.-B.; Liu, R.-M.; Zhong, J.-J. A new ganoderic acid from *Ganoderma lucidum* mycelia. *J. Asian Nat. Prod. Res.* 2010, 12, 727–730.
182. Nishitoba, T.; Sato, H.; Sakamura, S. New terpenoids, ganolucidic acid D, ganoderic acid L, lucidone C and lucidenic acid G, from the fungus *Ganoderma lucidum*. *Agric. Biol. Chem.* 1986, 50, 809–811.
183. Liu, J.-Q.; Wang, C.-F.; Li, Y.; Luo, H.-R.; Qiu, M.-H. Isolation and bioactivity evaluation of terpenoids from the medicinal fungus *Ganoderma sinense*. *Planta Med.* 2012, 78, 368–376.
184. Min, B.-S.; Gao, J.-J.; Nakamura, N.; Hattori, M. Triterpenes from the spores of *Ganoderma lucidum* and their cytotoxicity against Meth-A and LLC tumor cells. *Chem. Pharm. Bull.* 2000, 48, 1026–1033.

185. Nishitoba, T.; Oda, K.; Sato, H.; Sakamura, S. Novel triterpenoids from the fungus *Ganoderma lucidum*. *Agric. Biol. Chem.* 1988, 52, 367–372.
186. Hapuarachchi, K.K.; Cheng, C.R.; Wen, T.C.; Jeewon, R.; Kakumyan, P. *Mycosphere Essays 20: Therapeutic potential of Ganoderma species: Insights into its use as traditional medicine.* *Mycosphere* 2017, 8, 1653–1694.
187. Ma, B.-J.; Zhou, Y.; Ruan, Y.; Ma, J.-C.; Ren, W.; Wen, C.-N. Lanostane-type triterpenes from the sporoderm-broken spores of *Ganoderma lucidum*. *J. Antibiot.* 2012, 65, 165–167.
188. Gonzalez, A.G.; Leon, F.; Rivera, A.; Munoz, C.M.; Bermejo, J. Lanostanoid triterpenes from *Ganoderma lucidum*. *J. Nat. Prod.* 1999, 62, 1700–1701.
189. Mothana, R.A.A.; Awadh Ali, N.A.; Jansen, R.; Wegner, U.; Mentel, R.; Lindequist, U. Antiviral lanostanoid triterpenes from the fungus *Ganoderma pfeifferi*. *Fitoterapia* 2003, 74, 177–180.
190. Liu, C.; Chen, R. A new triterpene from fungal fruiting bodies of *Ganoderma sinense*. *Zhongcaoyao* 2010, 41, 8–11.
191. Su, H.-J.; Fann, Y.-F.; Chung, M.-I.; Won, S.-J.; Lin, C.-N. New lanostanoids of *Ganoderma tsugae*. *J. Nat. Prod.* 2000, 63, 514–516.
192. Kleinwaechter, P.; Anh, N.; Kiet, T.T.; Schlegel, B.; Dahse, H.-M.; Haertl, A.; Graefe, U. Colossolactones, new triterpenoid metabolites from a Vietnamese mushroom *Ganoderma colossum*. *J. Nat. Prod.* 2001, 64, 236–239.
193. Jiang, C.; Ji, J.; Li, P.; Liu, W.; Yu, H.; Yang, X.; Xu, L.; Guo, L.; Fan, Y. New lanostane-type triterpenoids with proangiogenic activity from the fruiting body of *Ganoderma applanatum*. *Nat. Prod. Res.* 2022, 36, 1529–1535.
194. Algehani, R.A.; Abou Khouzam, R.; Hegazy, G.A.; Alamoudi, A.A.; El-Halawany, A.M.; El Dine, R.S.; Ajabnoor, G.A.; Al-Abbasi, F.A.; Baghdadi, M.A.; Elsayed, I.; et al. Colossolactone-G synergizes the anticancer properties of 5-fluorouracil and gemcitabine against colorectal cancer cells. *Biomed. Pharmacother.* 2021, 140, 111730.
195. El Dine, R.S.; El Halawany, A.M.; Nakamura, N.; Ma, C.-M.; Hattori, M. New lanostane triterpene lactones from the Vietnamese mushroom *Ganoderma colossum* (FR.) C. F. Baker. *Chem. Pharm. Bull.* 2008, 56, 642–646.
196. Lakornwong, W.; Kanokmedhakul, K.; Kanokmedhakul, S.; Kongsaree, P.; Prabpai, S.; Sibounnavong, P.; Soyong, K. Triterpene lactones from cultures of *Ganoderma* sp. KM01. *J. Nat. Prod.* 2014, 77, 1545–1553.
197. Ofodile, L.N.; Uma, N.; Grayer, R.J.; Ogundipe, O.T.; Simmonds, M.S.J. Antibacterial compounds from the mushroom *Ganoderma colossum* from Nigeria. *Phytother. Res.* 2012, 26, 748–751.

198. Kikuchi, T.; Matsuda, S.; Kadota, S.; Murai, Y.; Tsubono, K.; Ogita, Z. Constituents of the fungus *Ganoderma lucidum* (FR.) Karst. I. Structure of ganoderic acids of C2, E, I, and K, lucidenic acid F and related compounds. *Chem. Pharm. Bull.* 1986, 34, 3695–3712.
199. Luo, J.; Lin, Z.B. Structure identification of triterpenes from fruiting bodies of *Ganoderma lucidum* by NMR spectra and X-ray diffraction analysis. *Chin. Tradit. Herb. Drugs* 2002, 33, 197–200.
200. Gao, J.-J.; Min, B.-S.; Ahn, E.-M.; Nakamura, N.; Lee, H.-K.; Hattori, M. New triterpene aldehydes, lucialdehydes A-C, from *Ganoderma lucidum* and their cytotoxicity against murine and human tumor cells. *Chem. Pharm. Bull.* 2002, 50, 837–840.
201. Cheng, C.-R.; Yue, Q.-X.; Wu, Z.-Y.; Song, X.-Y.; Tao, S.-J.; Wu, X.-H.; Xu, P.-P.; Liu, X.; Guan, S.-H.; Guo, D.-A. Cytotoxic triterpenoids from *Ganoderma lucidum*. *Phytochemistry* 2010, 71, 1579–1585.
202. Guan, S.H.; Yang, M.; Liu, X.; Xia, J.M.; Wang, X.M.; Jin, H.; Guo, D.A. Two new lanostanoid triterpenes from fruit body of *Ganoderma lucidum*-The major component of SunRecome®. *Nat. Prod. Commun.* 2006, 1, 177–181.
203. Kikuchi, T.; Kanomi, S.; Murai, Y.; Kadota, S.; Tsubono, K.; Ogita, Z.I. Constituents of the fungus *Ganoderma lucidum* (FR.) Karst. II. Structure of ganoderic acids F, G, and H, lucidenic acids D2 and E2, and related compounds. *Chem. Pharm. Bull.* 1986, 34, 4018–4029.
204. Guan, S.H.; Xia, J.M.; Yang, M.; Wang, X.M.; Liu, X.; Guo, D.A. Cytotoxic lanostanoid triterpenes from *Ganoderma lucidum*. *J. Asian Nat. Prod. Res.* 2008, 10, 695–700.
205. Li, C.J.; Li, Y.M.; Sun, H.H. New ganoderic acids, bioactive triterpenoid metabolites from the mushroom *Ganoderma lucidum*. *Nat. Prod. Res.* 2006, 20, 985–991.
206. Luo, J.; Zhao, Y.Y.; Lin, Z.B. A new lanostane-type triterpene from the fruiting bodies of *Ganoderma lucidum*. *J. Asian Nat. Prod. Res.* 2002, 4, 129–134.
207. Min, B.S.; Nakamura, N.; Miyashiro, H.; Bae, K.W.; Hattori, M. Triterpenes from the spores of *Ganoderma lucidum* and their inhibitory activity against HIV-1 protease. *Chem. Pharm. Bull.* 1998, 46, 1607–1612.
208. Nishitoba, T.; Sato, H.; Oda, K.; Sakamura, S. Novel triterpenoids and a steroid from the fungus *Ganoderma lucidum*. *Agric. Biol. Chem.* 1988, 52, 211–216.
209. Hirotsu, M.; Asaka, I.; Ino, C.; Furuya, T.; Shiro, M. Ganoderic acid derivatives and ergosta-4,7,22-triene-3,6-dione from *Ganoderma lucidum*. *Phytochemistry* 1987, 26, 2797–2803.
210. Ma, J.; Ye, Q.; Hua, Y.; Zhang, D.; Cooper, R.; Chang, M.N.; Chang, J.Y.; Sun, H.H. New lanostanoids from the mushroom *Ganoderma lucidum*. *J. Nat. Prod.* 2002, 65, 72–75.
211. Chen, M.; Zhang, M.; Sun, S.; Xia, B.; Zhang, H.Q. A new triterpene from the fruiting bodies of *Ganoderma lucidum*. *Acta Pharm. Sin.* 2009, 44, 768–770.

212. Lin, C.N.; Fann, Y.F.; Chung, M.I. Steroids of formosan *Ganoderma tsugae*. *Phytochemistry* 1997, 46, 1143–1146.
213. Niu, X.-M.; Li, S.-H.; Xiao, W.-L.; Sun, H.-D.; Che, C.-T. Two new lanostanoids from *Ganoderma resinaceum*. *J. Asian Nat. Prod. Res.* 2007, 9, 659–664.
214. Guan, S.H.; Yang, M.; Wang, X.M.; Xia, J.M.; Zhang, Z.M.; Liu, X.; Guo, D.A. Structure elucidation and complete NMR spectral assignments of three new lanostanoid triterpenes with unprecedented $\Delta^{16,17}$ double bond from *Ganoderma lucidum*. *Magn. Reson. Chem.* 2007, 45, 789–791.
215. De Silva, E.D.; van der Sar, S.A.; Santha, R.G.L.; Wijesundera, R.L.C.; Cole, A.L.J.; Blunt, J.W.; Munro, M.H.G. Lanostane triterpenoids from the Sri Lankan basidiomycete *Ganoderma applanatum*. *J. Nat. Prod.* 2006, 69, 1245–1248.
216. Lin, L.J.; Shiao, M.S.; Yeh, S.F. Seven new triterpenes from *Ganoderma lucidum*. *J. Nat. Prod.* 1988, 51, 918–924.
217. Hirotani, M.; Ino, C.; Furuya, T. Comparative study on the strain-specific triterpenoid components of *Ganoderma lucidum*. *Phytochemistry* 1993, 33, 379–382.
218. Satria, D.; Amen, Y.; Niwa, Y.; Ashour, A.; Allam, A.E.; Shimizu, K. Lucidumol D, a new lanostane-type triterpene from fruiting bodies of Reishi (*Ganoderma lingzhi*). *Nat. Prod. Res.* 2019, 33, 189–195.
219. Zhao, Z.-Z.; Chen, H.-P.; Li, Z.-H.; Dong, Z.-J.; Bai, X.; Zhou, Z.-Y.; Feng, T.; Liu, J.-K. Leucocontextins A–R, lanostane-type triterpenoids from *Ganoderma leucocontextum*. *Fitoterapia* 2016, 109, 91–98.
220. Isaka, M.; Chinthanom, P.; Rachtawee, P.; Choowong, W.; Choeyklin, R.; Thummarukcharoen, T. Lanostane triterpenoids from cultivated fruiting bodies of the wood-rot basidiomycete *Ganoderma casuarinicola*. *Phytochemistry* 2020, 170, 112225.
221. Wu, Y.; Han, F.; Luan, S.; Ai, R.; Zhang, P.; Li, H.; Chen, L. Triterpenoids from *Ganoderma lucidum* and their potential anti-inflammatory effects. *J. Agric. Food Chem.* 2019, 67, 5147–5158.
222. Liu, L.; Yan, Z.; Kang, J.; Chen, R.; Yu, D. Three new triterpenoids from *Ganoderma theaeecolum*. *J. Asian Nat. Prod. Res.* 2017, 19, 847–853.
223. Gao, J.; Chen, Y.; Liu, W.; Liu, Y.; Li, M.; Chen, G.; Yuan, T. Applanhydrides A and B, lanostane triterpenoids with unprecedented seven-membered cyclo-anhydride in ring C from *Ganoderma applanatum*. *Tetrahedron* 2021, 79, 131839.
224. Tan, Z.; Zhao, J.-L.; Liu, J.-M.; Zhang, M.; Chen, R.-D.; Xie, K.-B.; Chen, D.-W.; Dai, J.-G. Lanostane triterpenoids and ergostane-type steroids from the cultured mycelia of *Ganoderma capense*. *J. Asian Nat. Prod. Res.* 2017, 20, 844–851.

225. Kou, R.-W.; Gao, Y.-Q.; Xia, B.; Wang, J.-Y.; Liu, X.-N.; Tang, J.-J.; Yin, X.; Gao, J.-M. Ganoderterpene A, a new triterpenoid from *Ganoderma lucidum*, attenuates LPS-induced inflammation and apoptosis via suppressing MAPK and TLR-4/NF-KB pathways in BV-2 cells. *J. Agric. Food Chem.* 2021, 69, 12730–12740.
226. Liu, L.; Shimizu, K.; Tanaka, A.; Shinobu, W.; Ohnuki, K.; Nakamura, T.; Kondo, R. Target proteins of ganoderic acid DM provides clues to various pharmacological mechanisms. *Sci. Rep.* 2012, 2, 905.
227. Fatmawati, S.; Shimizu, K.; Kondo, R. Inhibition of aldose reductase in vitro by constituents of *Ganoderma lucidum*. *Planta Med.* 2010, 76, 1691–1693.
228. Fatmawati, S.; Kondo, R.; Shimizu, K. Structure–activity relationships of lanostane-type triterpenoids from *Ganoderma lingzhi* as α -glucosidase inhibitors. *Bioorg. Med. Chem. Lett.* 2013, 23, 5900–5903.

Retrieved from <https://encyclopedia.pub/entry/history/show/89828>