

# Resistant Starches and Non-Communicable Disease

Subjects: Nutrition & Dietetics

Contributor: Erika Cione

Resistant starch (RS) is the starch fraction that eludes digestion in the small intestine. RS is classified into five subtypes (RS1–RS5), some of which occur naturally in plant-derived foods, whereas the others may be produced by several processing conditions. The different RS subtypes are widely found in processed foods, but their physiological effects depend on their structural characteristics.

Keywords: resistant starches ; glycemic control ; cancer ; inflammation ; microbiome

---

## 1. Introduction

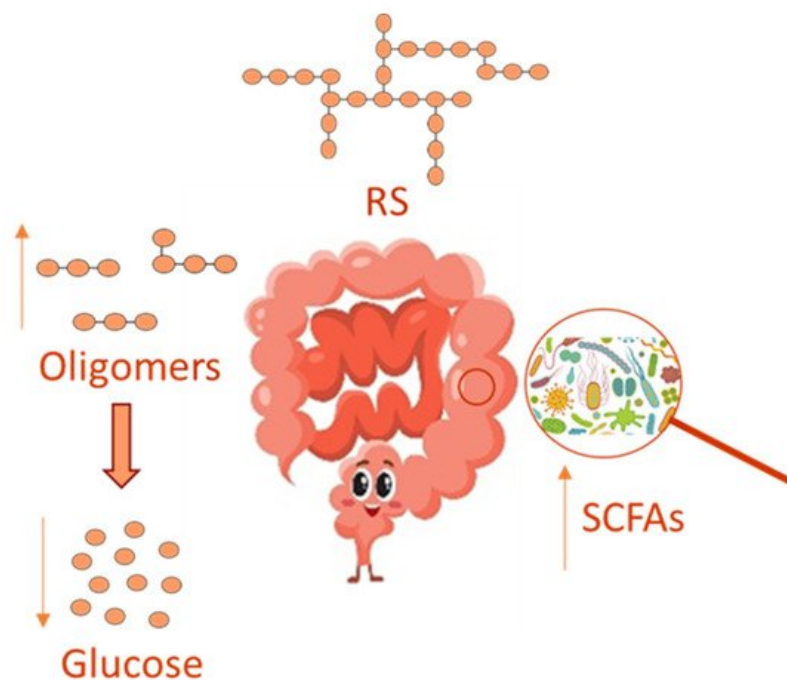
Non-communicable diseases (NCDs) are chronic disorders, which tend to be of long duration. NCDs are due to a combination of genetic, physiological, environmental and behavioral factors, including unhealthy diets, physical inactivity, tobacco smoke or alcohol use. NCDs can be preventable by nutrition and the adoption of an active lifestyle <sup>[1][2]</sup>. Nutritional habits and physical activity represent a winning combination to counteract the rising burden of NCDs. Both unhealthy diets and a lack of physical activity are determinants to develop obesity, which in turn is linked to: (i) rise of blood pressure, (ii) increase of blood glucose and (iii) elevation in blood lipids , i.e., all conditions leading to the development of metabolic diseases. Therefore, obesity is strongly associated with chronic inflammation, which in turn leads to metabolic and cardiovascular diseases and even cancer <sup>[3]</sup>. On this basis, understanding the mechanisms leading to obesity is essential to develop preventive strategies and new treatments <sup>[4][5]</sup>. Food intake plays a critical role in the development of metabolic diseases. Recently, an investigation on European children from six different nations has highlighted that the consumption of fiber-rich foods does not meet present recommended daily intake guidelines <sup>[6]</sup>. The authors have found out a weak but significant correlation between an increase in BMI and a decrease in consumption frequency of wholegrain cereals and wholemeal products (bread, cereals, biscuits, pasta, rice). Furthermore, adopting the consumption of high-fiber foods in early childhood can delay the initiation of impaired condition <sup>[6][7]</sup>. These results are mostly driven by Italy (lower cereals consumption frequency). Angel Keys was the first to describe the Mediterranean Diet (MeD) as such and to present it to the popular thoughts as it is today. He was a biologist and physiologist who focused his studies on the dietary habits of people living in the South of Italy. Following the joint candidacy of Italy, Spain, Greece and Morocco, succeeded by Cyprus, Croatia and Portugal, the MeD was recognized by UNESCO, WHO and FAO. Such a diet varies by country and region; therefore, it has a range of definitions. It usually includes a low intake of meat and eggs, a moderate intake of dairy products and milk, and a large consumption of vegetables, fruits rich in phenols, fish, olive oil and seeds, rich in unsaturated fats <sup>[8][9]</sup>. Furthermore, legumes, whole cereals, potato, bread and rice are indicated in the Mediterranean food pie chart ( **Figure 1** ).



**Figure 1.** Mediterranean food pie chart.

## 2. Resistant Starch in Human Nutritional Intervention Studies: GI and Impact on Inflammation and Gut Microbiome

In recent years, the gut microbiota has been widely investigated, and its imbalance, due to antibiotics use <sup>[10]</sup>, has been related to many disorders, including inflammation and oxidative stress, which underlie several chronic diseases, such as obesity, type 2 diabetes and chronic kidney disease <sup>[11]</sup>. Several reports have evidenced that a prebiotic supplemented diet can healthily regulate the gut microbiota, thus relieving disorders due to its imbalance. Prebiotics comprises non-digestible dietary soluble fiber, which can be used by the gut microbiota for fermentation. In this context, RS can act as a substrate for microbial fermentation in the large intestine <sup>[11]</sup> by supplying an energy source and fermentative products, such as short-chain fatty acids (SCFAs) in their anionic forms, in such a way RS is able to modulate microbial growth and could influence colonic health ( **Figure 2** ). On this basis, RS is believed to be a prebiotic and to influence the GI of foods favoring a lower intestinal absorption of glucose, even in the presence of polyphenols that in turn inhibits enterocytic starch digestion enzymes <sup>[12]</sup>



**Figure 2.** Resistant starch effect on the GI and microbiome SCFAs synthesis.

When compared to food containing only readily digestible starch, the rate of digestion of RS-containing foods in the small intestine is substantially slower. As a result, consumption of such food leads to a sustained and lower level of glucose release [13]. This effect is reflected by the GI, a ranking system that organizes different food products based on the glycemic response to food consumption [13]. Researchers discovered a decrease in starch digestibility in treated food compared to untreated food after producing retrogradation in test meals [14]. They also observed a slower rise in blood glucose levels in human subjects upon consuming treated food when compared to those consuming untreated food [14][15]. Several studies have reported that potatoes generally have medium to high GI, which has often negatively impacted their consumption, but such studies have overlooked the many nutritional and health benefits of potatoes [16][17]. Interestingly, the GI varies depending on the potato variety, origin, maturity and processing methods, which can alter the starch digestibility of consumed foods. Hence, the concept of glycemic load (GL) was developed to simultaneously describe the quality (i.e., GI) and quantity of carbohydrates in a meal or diet [18][19]. Potato starch consists of 70–80% amylopectin, which is a highly branched, high molecular weight biopolymer. Amylose represents approximately 20–30% of starch, and it is a relatively long, linear,  $\alpha$ -glucan with only a few branches. The relative proportion of amylose and amylopectin is important, given that amylose acts as a restraint to swelling and, upon cooling, it forms retrograded starch more readily [20][21][22]. Other than total starch, structure and moisture content [23], as stated above, other factors can also impact the glycemic response, including the growing conditions, maturity of the potato variety and cooking methods [24]. Potato varieties, maturity level, starch structure, methods of food processing and composition of the meal affect the GI of potatoes. Boiling, baking, microwave or oven cooking, extrusion and frying result in diverse degrees of gelatinization and starch crystallinity in potatoes. Cooling or storage after processing potatoes significantly reduces their GI because of the retrogradation of starch molecules. Although beneficial effects of resistant starch consumption have been observed, the current results of studies on the underlying molecular mechanisms in both animals and humans are not yet conclusive. Additional research efforts are necessary in order to reach a better understanding of the effects of habitual RS consumption on glycemic control.

The mammalian gastrointestinal microbiota makes important contributions to the health of the host, including immune system development, nutrient metabolism and absorption, drug metabolism as well as protection against infection [25]. An altered microbiota (dysbiosis) has been associated with human diseases, such as diabetes, obesity, inflammatory bowel diseases, fecal occult blood and colorectal cancer [25][26][27]. By the way, diet is also considered a key modulator of the composition and function of the gut microbiota. Over the past few decades, the dietary intake of RS has been investigated. RS is a type of fermentable fiber considered a prebiotic since it can reach the large intestine, in which gut bacteria ferment it. RS fermentation leads to SCFAs production and pH reduction in the proximal large intestine. It has long been known that diet influences the microbial communities of the gastrointestinal tract. Although studies to understand how different classes of RS can affect microbiota are limited, it is clear that high-fiber diets greatly modulate the composition of mammalian microbiota [28]. Among the different classes of resistant starches, RS1–RS5 ( **Table 1** ), type 3 (RS3) is endowed with the strongest prebiotic properties [29]. Recent studies have explored the comparative physiological effects of diverse types of RS, investigating RS-induced changes in the microbiome that might be substantial in health and disease. Bacteria in the large intestine can be exposed to as much as 20 g of RS per day in humans [30]. RS may exert protective effects through broader mechanisms associated with fermentation. Acetate, propionate and butyrate are the most abundant anionic form of SCFAs generated from colonic microbial metabolism. SCFAs play a key role in the regulation of the inflammation process; contextually, they induce protective effects by stimulating or lessening inflammatory cytokines production, as well as by inhibiting or facilitating immune cells recruitment [31]. Furthermore, the fermentation of RS in the colon results in the production of gases (methane, hydrogen and carbon dioxide), small amounts of organic acid in its anionic form (lactate, succinate and formate), branched SCFAs either in anionic form such as valerate and butyrate. This latter is mainly obtained from the fermentation of RS2 than that of other RSs; notably, it has displayed interesting anti-inflammatory properties [30][32][33]. In particular, RS2 can promote a greater growth of bacteria belonging to the families of Bifidobacteriaceae and Lactobacillaceae , which are known to reduce inflammation [32]. Additionally, RS2 supplementation led to an increase in the gut level of Faecalibacterium ; this prompted the authors to hypothesize a bacterial involvement in the anti-inflammatory effect exerted by the prebiotic fiber [33]. A future research direction to better assess gut/fecal microbial composition and serum concentration of anionic form SCFAs before and after RS2 intake could be represented by rodent models [34][35].

**Table 1.** Classification of RS and example of foods rich in RS.

Classification	Description	Example
RS1	Physically inaccessible starch	Whole grains

Classification	Description	Example
RS2	Starch with B- or C-polymorph	Uncooked potato, high-amylose maize starch
RS3	Retrograded starch	Cooked and cooled potato starch
RS4	Chemically modified starch	Cross-linked starch in thickeners
RS5	Amylose–lipid complex	Palmitic acid-amylose complex

The beneficial effect of a long-term (12 months) dietary intervention with increased fiber intake, including RS on humans, was assessed [36][37]. In the study, two dietary groups were investigated, RS group subjects ingested higher amounts of food rich in RS (especially cereals and legumes) in order to consume about 15 g/day of RS, while subjects in the fiber group received general advice to ingest vegetables rich in fiber, without specific advice on the intake of RS-rich foods. At the end of the study, in the RS group, anthropometric parameters, such as body weight, body mass index (BMI) and waist circumference, were slightly more decreased, and a negative correlation was found between RS intake and adiponectin level, along with a negative correlation between RS intake and blood level of resistin, a possible pro-inflammatory mediator of insulin resistance. Furthermore, leptin and apelin levels were significantly decreased only in the RS group [38][39]. As already discussed above, RS exists in different types, having different chemical structures. In a double-blind controlled, crossover intervention study, a diet enriched with RS4, a chemically modified starch, was found to lower blood cholesterol and improve body composition measured by dual-energy, X-ray absorptiometry (DXA) [40]. The RS4 enriched diet was used for 26 weeks in the management of metabolic syndrome (MetS) using RS4 flour at 30% (v / v). Regular flour was used as a control (CF). Similar results on human lipid profiles were highlighted with RS3 (40 g/d for 21 days) in overweight and obese females [40] and RS2 (>25 g/d for 12 months) in overweight and obese subjects [40][41]. High-RS potato starch (with low protein) was used as a source of RS in patients with early type 2 diabetic nephropathy (DN) for 12 weeks [42]. In the study, the control group subjects consumed each day protein restriction diet with an everyday staple, whereas subjects of the intervention group ingested 50 g of high-RS, low-protein flour instead of an everyday staple of equal quality at lunch and dinner every day. A significant reduction in the levels of hemoglobin A1c (HbA1c) and lipid profile was found in the intervention group, along with a marked decrease of serum uric acid and urinary  $\beta$ 2-microglobulin levels [41], thus delaying the progression of early type 2 DN. Several short-term intervention studies employing whole grain (WG) foods, including WG barley or rye derivatives rich in intact kernels, dietary fiber and RS, exhibited anti-obesogenic and anti-diabetic effects in healthy subjects [43][44][42]. More recently, a short-term, crossover, randomized study investigated the effect of rye-based bread preparation on healthy middle-aged subjects by employing white wheat flour bread (WWB) as a reference in order to evaluate possible effects on cardiometabolic risk markers, cognitive functions and mood [44]. Rye-based bread was prepared using a WG rye kernel/flour mixture (1:1 ratio) supplemented with RS2 (RB + RS2). Such a dietary treatment significantly increased insulin sensitivity, fasting concentrations of plasma butyrate, acetate and total SCFAs anionic form, which in turn have beneficial systemic effects [44]. Moreover, the dietary treatment significantly increased the fasting levels of plasma gut hormones, such as the peptide YY (PYY) and the glucagon-like peptide (GLP)-2. Furthermore, fasting levels of the inflammatory marker interleukin (IL)-1 $\beta$  were significantly decreased. Remarkably, the fasting concentrations of butyrate and acetate and the breath hydrogen excretion were significantly increased when preceded by the RB + RS2 intervention, suggesting that increased gut microbial fermentation of dietary fiber could mediate the observed good effects. On the other hand, blood levels of lipids, and of other inflammatory markers, including C-reactive protein, brain-derived neurotrophic factor, IL-6 and IL-18, did not significantly differ between the RB + RS2 and WWB group, as well as no significant differences in appetite sensations were observed (satiety, desire to eat or hunger), and in cognitive performance. However, insulin sensitivity was found to be positively correlated with working memory test performance. Furthermore, subjects receiving the RB + RS2 intervention felt glad more, pleased, happy, active, awake and peppy when compared to those of the WWB group. Overall, studies in humans have highlighted that the dietary intake of RS seems to not directly and significantly affect body weight and composition. In the same way, the effects of RS on reducing energy intake, increasing satiety and improving lipid profiles are controversial. On the other hand, RS supplementation can healthily affect glucose homeostasis by decreasing fasting or postprandial glucose levels and improving insulin sensitivity. Additionally, RS exerts beneficial effects on the gut microbiota, and it positively modulates gut hormones, such as GLP-1 and PYY. Nevertheless, such statements are still not definitive since further research is required, especially regarding the need for a larger sample size and longer intervention times [45]. Additionally, Chang et al. showed that the anti-inflammatory properties, as well as the inhibition of pro-inflammatory responses by intestinal macrophages, performed by butyrate, were dependent on its histone deacetylase (HDAC) inhibitory activity. Such inhibition, in turn, increased histone H3 acetylation within genetic loci required for regulatory T cells (also called Tregs) induction [46]. Moreover, it was evidenced that butyrate increased the release of the anti-inflammatory cytokine IL-10 and decreased the production of pro-inflammatory cytokines, such as TNF- $\alpha$ , IL-6 and nitric oxide (NO) [47]. Recent studies highlighted that a diet supplemented with buckwheat RS induced a significant decrease in TNF- $\alpha$  and interleukin-6

(IL-6) levels in high-fat diet (HFD)-fed mice; therefore, buckwheat RS supplementation could improve inflammatory response in plasma [48]. Instead, controversial results emerged from human nutritional clinical studies regarding the RS2 effect on inflammatory mediators. Indeed, a study performed on hemodialysis patients, who had taken RS2 for 4 weeks, evidenced a decreased IL-6 concentration [49]. In agreement, in patients with systemic inflammation associated with chronic kidney disease, a supplemented diet containing high-amylose maize RS2 elicited a significant decrease in serum levels of IL-6 and TNF $\alpha$  with respect to the placebo [50]. On the contrary, in women with type 2 diabetes, RS2 dietary intake did not induce any significant change in IL-6 concentration compared to the placebo [51]. RS2 significantly decreased TNF- $\alpha$  level and ameliorated both glycemic and lipid profiles in women with T2DM after 8 weeks of treatment [52]. Conversely, after 12 weeks of supplementation with RS, prediabetes adults displayed reduced concentrations of plasmatic TNF- $\alpha$  but no significant improvement in insulin resistance [53]. These controversial results were recently reported in a systematic review with meta-analysis. The authors concluded that RS2 could not reduce inflammatory mediators, but they also recognized the need for more randomized controlled trials with longer intervention in terms of duration, use of higher dose and studies in different countries [54].

## References

1. Budreviciute, A.; Damiati, S.; Sabir, D.K.; Onder, K.; Schuller-Goetzburg, P.; Plakys, G.; Katileviciute, A.; Khoja, S.; Kodzius, R. Management and Prevention Strategies for Non-communicable Diseases (NCDs) and Their Risk Factors. *Front. Public Health* 2020, 8.
2. Caprara, G. Mediterranean-Type Dietary Pattern and Physical Activity: The Winning Combination to Counteract the Rising Burden of Non-Communicable Diseases (NCDs). *Nutrients* 2021, 13, 429.
3. Bhattacharya, S.; Juyal, R.; Hossain, M.M.; Singh, A. Non-communicable diseases viewed as “collateral damage” of our decisions: Fixing accountabilities and finding solutions in primary care settings. *J. Fam. Med. Prim. Care* 2020, 9, 2176–2179.
4. Gortmaker, S.L.; Swinburn, B.A.; Levy, D.; Carter, R.; Mabry, P.L.; Finegood, D.; Huang, T.; Moodie, M. Changing the future of obesity: Science, policy, and action. *Lancet* 2011, 378, 838–847.
5. González-Muniesa, P.; Martínez-González, M.A.; Hu, F.; Depres, J.P.; Matsuzawa, Y.; Loos, R.J.F.; Moreno, L.A.; Bray, G.A.; Martínez, J.A. Obesity. *Nat. Rev. Dis. Primers* 2017, 3, 17034.
6. Hörmann-Wallner, M.; Krause, R.; Alfaro, B.; Jilani, H.; Laureati, M.; Almi, V.L.; Sandell, M.; Sandvik, P.; Zeinstra, G.Z.; Methven, L. Intake of Fibre-Associated Foods and Texture Preferences in Relation to Weight Status Among 9-12 Years Old Children in 6 European Countries. *Front Nutr.* 2021, 8, 633807.
7. Cannataro, R.; Fazio, A.; La Torre, C.; Caroleo, M.C.; Cione, E. Polyphenols in the Mediterranean Diet: From Dietary Sources to microRNA Modulation. *Antioxidants* 2021, 10, 328.
8. Trichopoulou, A.; Martínez-González, M.A.; Tong, T.Y.; Forouhi, N.G.; Khandelwal, S.; Prabhakaran, D.; Mozaffarian, D.; de Longeril, M. Definitions and potential health benefits of the Mediterranean diet: Views from experts around the world. *BMC Med.* 2014, 12, 112.
9. Ghosh, T.S.; Rampelli, S.; Jeffery, I.B.; Santoro, A.; Neto, M.; Capri, M.; Giampieri, E.; Jennings, A.; Candela, M.; Turroni, S.; et al. Mediterranean diet intervention alters the gut microbiome in older people reducing frailty and improving health status: The NU-AGE 1-year dietary intervention across five European countries. *Gut* 2020, 69, 1218–1228.
10. Ramirez, J.; Guarner, F.; Bustos Fernandez, L.; Maruy, A.; Sdepanian, V.L.; Cohen, H. Antibiotics as Major Disruptors of Gut Microbiota. *Front. Cell. Infect. Microbiol.* 2020, 10, 572912.
11. Dolan, K.T.; Chang, E.B. Diet, gut microbes, and the pathogenesis of inflammatory bowel diseases. *Mol. Nutr. Food Res.* 2017, 61, 1600129.
12. Ayua, E.O.; Nkhata, S.G.; Namaumbo, S.J.; Kamau, E.H.; Ngoma, T.N.; Aduol, K.O. Polyphenolic inhibition of enterocytic starch digestion enzymes and glucose transporters for managing type 2 diabetes may be reduced in food systems. *Heliyon* 2021, 12, e06245.
13. Burton, P.; Lightowler, H. The impact of freezing and toasting on the glycaemic response of white bread. *Eur. J. Clin. Nutr.* 2008, 62, 594–599.
14. Frei, M.; Siddhuraju, P.; Becker, K. Studies on the in vitro starch digestibility and the glycemic index of six different indigenous rice cultivars from the Philippines. *Food Chem.* 2003, 83, 395–402.
15. Foster-Powell, K.; Holt, S.H.; Brand-Miller, J.C. International Tables of Glycemic Index and Glycemic Load Values: 2002. *Am. J. Clin. Nutr.* 2002, 76, 5–56.

16. Atkinson, F.S.; Foster-Powell, K.; Brand-Miller, J.C. International Tables of Glycemic Index and Glycemic Load Values. *Diabet. Care* 2008, 3, 2281–2283.
17. Salmeron, J.; Monson, J.E.; Stampfer, M.S.; Colditz, G.A.; Wing, A.L.; Willett, W.C. Dietary Fiber, Glycemic Load and Risk of Non-Insulin Dependent Diabetes Mellitus on Woman. *JAMA* 1997, 277, 472–477.
18. Brennan, C.S. Dietary Fiber, Glycemic Response and Diabetes. *Mol. Nutr. Food Res.* 2005, 49, 560–570.
19. Hoover, R. Composition, Molecular Structure, and Physico-Chemical Properties of Tuber and Root Starches: A Review. *Carbohydr. Polym.* 2001, 45, 253–267.
20. Bertoft, E.; Blennow, A. Structure of Potato Starch. In *Advances in Potato Chemistry and Technology*, 2nd ed.; Singh, J., Kaur, L., Eds.; Elsevier: Copenhagen, Denmark, 2008; pp. 83–98.
21. Pérez, S.; Bertoft, E. The Molecular Structures of Starch Components and Their Contribution to the Architecture of Starch Granules: A Comprehensive Review. *Starch* 2010, 62, 389–420.
22. Lynch, D.R.; Liu, Q.; Tarn, T.R.; Bizimungu, B.; Chen, Q.; Harris, P.; Chik, C.L.; Skjodt, N.M. Glycemic Index—A Review and Implications for the Potato Industry. *Am. J. Potato Res.* 2007, 84, 179–190.
23. Fernandes, G.; Velangi, A.; Wolever, T.M.S. Glycemic Index of Potatoes Commonly Consumed in North America. *J. Acad. Nutr. Diet.* 2005, 105, 557–562.
24. Dodevska, M.S.; Sobajic, S.S.; Djordjevic, P.B.; Dimitrijevic-Sreckovic, V.S.; Spasojevic-Kalimanovska, V.S.; Djordjevic, B.I. Effects of total fibre or resistant starch-rich diets within lifestyle intervention in obese prediabetic adults. *Eur. J. Nutr.* 2016, 55, 127–137.
25. Cafiero, C.; Re, A.; Pisconti, S.; Trombetti, M.; Perri, M.; Colosimo, M.; D'Amato, G.; Gallelli, L.; Cannataro, R.; Molinari, C.; et al. Dysbiosis in intestinal microbiome linked to fecal blood determined by direct hybridization. *Biotech* 2020, 10, 358.
26. Wong, S.H.; Yu, J. Gut microbiota in colorectal cancer: Mechanisms of action and clinical applications. *Nat. Rev. Gastroenterol. Hepatol.* 2019, 16, 690–704.
27. Wu, G.D.; Chen, Y.Y.; Hoffmann, C.; Bittinger, K.; Chen, Y.; Keilbaugh, S.A.; Bewtra, M.; Knights, D.; Walters, W.A.; Knight, R.; et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science* 2011, 334, 105–108.
28. Asp, N.G.; van Amelsvoort, J.M.M.; Hautvast, J.G.A.J. Nutritional implications of resistant starch. *Nutr. Res. Rev.* 1996, 9, 1–31.
29. Topping, D.L.; Clifton, P.M. Short-chain fatty acids and human colonic function: Roles of resistant starch and non-starch polysaccharides. *Physiol Rev.* 2001, 81, 1031–1064.
30. Corrêa-Oliveira, R.; Fachi, J.L.; Vieira, A.; Sato, F.T.; Vinolo, M.A. Regulation of immune cell function by short-chain fatty acids. *Clin. Transl. Immunol.* 2016, 5, e73.
31. Le Leu, R.K.; Hu, Y.; Brown, I.L.; Young, G.P. Effect of high amylose maize starches on colonic fermentation and apoptotic response to DNA-damage in the colon of rats. *J. Nutr. Metab.* 2009, 6, 11.
32. Arpaia, N.; Rudensky, A.Y. Microbial metabolites control gut inflammatory responses. *Proc. Natl. Acad. Sci. USA* 2014, 111, 2058–2059.
33. Chang, P.V.; Hao, L.; Offermanns, S.; Medzhitov, R. The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition. *Proc. Natl. Acad. Sci. USA* 2014, 111, 2247–2252.
34. Birt, D.F.; Phillips, G.J. Diet, Genes, and Microbes: Complexities of Colon Cancer Prevention. *Toxicol Pathol.* 2014, 42, 182–188.
35. Sekirov, I.; Russell, S.L.; Antunes, C.M.; Finlay, B.B. Gut microbiota in health and disease. *Physiol. Rev.* 2010, 90, 859–904.
36. Bush, J.R.; Alfa, M.J. Increasing levels of *Parasutterella* in the gut microbiome correlate with improving low-density lipoprotein levels in healthy adults consuming resistant potato starch during a randomised trial. *BMC Nutr.* 2020, 11, 72.
37. Alfa, M.J.; Strang, D.; Tappia, P.S.; Olson, N.; DeGagne, P.; Bray, D.; Murray, B.L.; Hiebert, B. A Randomized Placebo Controlled Clinical Trial to Determine the Impact of Digestion Resistant Starch MSPrebiotic® on Glucose, Insulin, and Insulin Resistance in Elderly and Mid-Age Adults. *Front. Med.* 2018, 4, 260.
38. Oliver, A.; Chase, A.B.; Weihe, C.; Orchanian, S.B.; Riedel, S.F.; Hendrickson, C.L.; Lay, M.; Sewall, J.M.; Martiny, J.B.H.; Whiteson, K. High-Fiber, Whole-Food Dietary Intervention Alters the Human Gut Microbiome but Not Fecal Short-Chain Fatty Acids. *mSystems* 2021, 6, e00115–e00121.
39. Luo, R.; Li, X.; Jiang, R.; Gao, X.; Lu, Z.; Hua, W. Serum Concentrations of Resistin and Adiponectin and Their Relationship to Insulin Resistance in Subjects with Impaired Glucose Tolerance. *J. Int. Med. Res.* 2012, 40, 621–630.

40. Nichenametla, S.N.; Lee, A.; Weidauer, L.A.; Wey, H.E.; Beare, T.M.; Specker, B.L.; Dey, M. Resistant starch type 4-enriched diet lowered blood cholesterol and improved body composition in a double-blind controlled crossover intervention. *Mol. Nutr. Food Res.* 2014, 58, 1365–1369.
41. Park, O.J.; Ekang, N.; Chang, M.J.; Kim, W.K. Resistant starch supplementation influences blood lipid concentrations and glucose control in overweight subjects. *J. Nutr. Sci. Vitaminol.* 2004, 50, 93–99.
42. Meng, Y.; Bai, H.; Yu, Q.; Yan, J.; Zhao, L.; Wang, S.; Zhaoping, L.; Wang, Q.; Chen, L. High-resistant starch, low-protein flour intervention on patients with early type 2 diabetic nephropathy: A randomized trial. *J. Ren Nutr.* 2019, 29, 386–393.
43. Johansson, E.V.; Nilsson, A.C.; Östman, E.M.; Björck, I.M.E. Effects of indigestible carbohydrates in barley on glucose metabolism, appetite and voluntary food intake over 16 h in healthy adults. *Nutr. J.* 2013, 12, 46.
44. Nilsson, A.C.; Johansson-Boll, E.V.; Björck, I.M. Increased gut hormones and insulin sensitivity index following a 3-d intervention with a barley kernel-based product: A randomised cross-over study in healthy middle-aged subjects. *Br. J. Nutr.* 2015, 114, 899–907.
45. Sandberg, J.C.; Björck, I.M.E.; Nilsson, A.C. Impact of rye-based evening meals on cognitive functions, mood and cardiometabolic risk factors: A randomized controlled study in healthy middle-aged subjects. *Nutr. J.* 2018, 17, 102.
46. Guo, J.; Tan, L.; Kong, L. Impact of dietary intake of resistant starch on obesity and associated metabolic profiles in human: A systematic review of the literature. *Crit. Rev. Food* 2021, 61, 889–905.
47. Vinolo, M.A.; Rodrigues, H.G.; Nachbar, R.T.; Curi, R. Regulation of inflammation by short chain fatty acids. *Nutrients* 2011, 3, 858–876.
48. Zhou, Y.; Zhao, S.; Jiang, Y.; Wei, Y.; Zhou, X. Regulatory function of buckwheat-resistant starch supplementation on lipid profile and gut microbiota in mice fed with a high-fat diet. *J. Food. Sci* 2019, 84, 2674–2681.
49. Gargari, B.P.; Namazi, N.; Khalili, M.; Sarmadi, B.; Jafarabadi, M.A.; Dehghan, P. Is there any place for resistant starch, as alimentary prebiotic, for patients with type 2 diabetes? *Complement. Ther. Med.* 2015, 23, 810–815.
50. Dobranowski, P.A.; Stintzi, A. Resistant starch, microbiome, and precision modulation. *Gut Microbes.* 2021, 13, 1926842.
51. Esgalhado, M.; Kemp, J.A.; Azevedo, R.; Laffin, L.J.; Madsen, K.; Kafil, H.S.; Abedi, B.; Shiralizadeh, S.; Vaziri, N.D. Could resistant starch supplementation improve inflammatory and oxidative stress biomarkers and uremic toxins levels in hemodialysis patients? A pilot randomized controlled trial. *Food Funct.* 2018, 9, 6508–6516.
52. Deehan, E.C.; Yang, C.; Perez-Muñoz, M.E.; Nguyen, N.K.; Cheng, C.C.; Triador, L.; Zhang, Z.; Bakal, J.A.; Walter, J. Precision Microbiome Modulation with Discrete Dietary Fiber Structures Directs Short-Chain Fatty Acid Production. *Cell Host Microbe* 2020, 27, 389–404.e6.
53. Peterson, C.M.; Beyl, R.A.; Marlatt, K.L.; Martin, C.K.; Aryana, K.J.; Marco, M.L.; Martin, R.J.; Keenan, M.J.; Ravussin, E. Effect of 12 wk of resistant starch supplementation on cardiometabolic risk factors in adults with prediabetes: A randomized controlled trial. *Am. J. Clin. Nutr.* 2018, 108, 492–501.
54. Haghighatdoost, F.; Gholami, A.; Hariri, M. Effect of resistant starch type 2 on inflammatory mediators: A systematic review and meta-analysis of randomized controlled trials. *Complement. Ther. Med.* 2021, 56, 102597.