Insulin in Health and Disease

Subjects: Biochemistry & Molecular Biology Contributor: Md-Saidur Rahman

Insulin is a polypeptide hormone mainly secreted by β cells in the islets of Langerhans of the pancreas. The hormone potentially coordinates with glucagon to modulate blood glucose levels; insulin acts via an anabolic pathway, while glucagon performs catabolic functions. Insulin regulates glucose levels in the bloodstream and induces glucose storage in the liver, muscles, and adipose tissue, resulting in overall weight gain. The modulation of a wide range of physiological processes by insulin makes its synthesis and levels critical in the onset and progression of several chronic diseases. Although clinical and basic research has made significant progress in understanding the role of insulin in several pathophysiological processes, many aspects of these functions have yet to be elucidated. This review provides an update on insulin secretion and regulation, and its physiological roles and functions in different organs and cells, and implications to overall health. We cast light on recent advances in insulin-signaling targeted therapies, the protective effects of insulin signaling activators against disease, and recommendations and directions for future research.

Keywords: insulin ; health ; disease ; homeostasis ; regulation ; glucose

1. Introduction

Although the biosynthesis and secretion of insulin are controlled by circulating glucose levels, the concentrations required to initiate the two processes differ ^{[1][2]}. While glucose concentrations above 5 mM are required to initiate insulin secretion, fluctuations within 2 mM to 4 mM stimulate its biosynthesis ^[1]. The metabolism of glucose is triggered by food intake, leading to simultaneously increased β cell insulin production and decreased α cell glucagon secretion to bring serum glucose levels back to normal ^[2]. Skeletal muscle cells and adipocytes, the other major targets of circulating insulin, also take up glucose, thereby reducing the blood glucose concentration to baseline ^[3].

To date, the main focus of research has been to investigate the role of insulin in the onset and progression of pathological conditions and chronic diseases, such as diabetes. The literature shows that insulin deficiency makes it impossible for cells to use glucose as an energy source ^[3]; consequently, high-glucose concentrations in the bloodstream lead to a condition known as hyperglycemia ^[3]. Prolonged hyperglycemia leads to diabetes mellitus and can cause health complications such as nervous system damage and dysfunction of the eyes and kidneys. Similarly, a cell's inability to use glucose as an energy source that results from the lack of insulin can trigger reliance on fat stores as the only energy supply.

Recent advances in insulin research have led to insulin-signaling targeted therapies and insulin-signaling activators being used as protective measures against several diseases. Similarly, sulfonylurea, through its activities on enhanced insulin secretion via their actions on pancreatic β cells, augmented insulin secretion ^[1]. Currently available forms of insulin include insulin mixtures, concentrated insulins, and insulins with alternate routes of administration, providing several options for people living with diabetes. This article provides an updated review of insulin secretion and regulation, its physiological roles in body organs, the health consequences of insulin deficiency, and recent advances in insulin-signaling targeted therapies.

2. Regulation of Insulin Secretion

The physiology of insulin-producing cells is fundamental to understanding the regulation of insulin secretion. The human pancreas contains one to two million pancreatic islets ^[4] housing different endocrine cells, primarily insulin-secreting β cells, glucagon-producing α cells, and somatostatin-secreting δ cells ^[5]. Generally, insulin is released after ingesting glucose in a process named glucose-induced insulin stimulation. In human β cells, glucose transporter 1 (GLUT1, encoded by SLC2A1) and GLUT3 (encoded by SLC2A3) are the prominent glucose transporters, whereas GLUT2 (encoded by SLC2A2) has been reported as a major glucose transporter in rodent ^{[6][Z]}.

The phosphorylation of glucose by the enzyme glucokinase (GCK) is the first step in glucose metabolism. Glucose phosphorylation by GCK is related to insulin secretion; therefore, GCK gene dysfunction or aberration leads to decreased glucose-mediated insulin release and glucose intolerance or diabetes [B]. A major understanding of insulin secretion is derived from the research using rodent models, whereas few studies have been described in humans [9].

In nondiabetic donors, an increase in glucose concentration from 1 mM to 6 mM raises the rate of glucose oxidation threefold (as determined by measuring the levels of C14O2produced from uniformly C14-labeled glucose), and an approximately 25% acceleration takes place when the concentration of glucose is increased to >12 mM $^{[10]}$. About one-tenth of the ingested glucose enters into glycolysis, which occurs through mitochondrial oxidation in human islets $^{[10]}$, but the ultimate destination of the rest of the ingested glucose needs to be further elucidated.

largely promote nutrient-induced insulin secretion, and they are highly crucial in the overall physiology of insulin secretion $[\mathfrak{Y}]$. Research has indicated that incretins are capable of binding to G-protein-coupled receptors on β cell membranes and increase cellular 3',5'-cyclic adenosine monophosphate (cAMP) levels and glucose-stimulated insulin secretion (GSIS) in the presence of higher glucose levels. As a consequence, cAMP induces an upsurge in the size/amount of readily releasable pools in a glucose concentration-dependent manner within insulin granule dynamics. It is worth noting that incretin peaks the activity of the β cells in the presence of active glucose concentrations in a Ca2+-independent manner, even in Ca2+

3. Insulin Signaling Pathways

After being secreted from pancreatic β cells and circulating through the body, insulin binds to insulin receptors (IRs) on target cell membranes. This results in the phosphorylation of insulin receptor substrate (IRS) and the subsequent activation of two primary signaling pathways, viz. the phosphoinositide3-kinase (PI3K)/protein kinase B (Akt) pathway and the mitogen-activated protein kinase (MAPK) pathway (**Figure 1**).

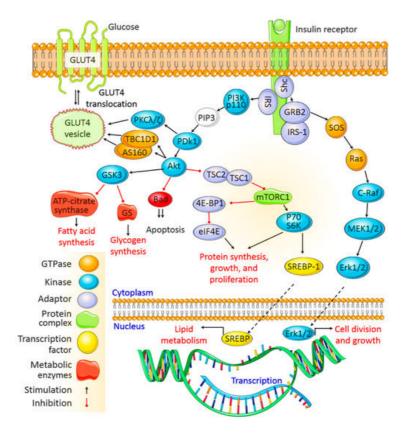


Figure 1. Classic insulin signaling pathway. Insulin regulates cellular functions and metabolic activity by binding to the insulin receptor. Core cellular processes downstream of the insulin signaling system include the PI3K/Akt and MAPK signaling pathways; details on these pathways are given. IRS, insulin receptor substrate; IR, insulin receptor; PI3K, phosphoinositide3-kinase; MAPK, mitogen-activated protein kinase; PIP3, phosphatidylinositol 3,4,5-triphosphate; PDK1, 3-phosphoinositide dependent protein kinase-1; Grb2, growth factor receptor-bound protein 2; GSK, glycogen synthase kinase; GS, glycogen synthase; mTORC1, mammalian target of rapamycin complex 1; SREBP, sterol regulatory element-binding protein; glucose transporter 4, GLUT4; MEK, MAPK/Erk kinase; ERK, extracellular signal-regulated kinase.

PDK1 then activates Akt, which mediates multiple cellular functions. Activated Akt phosphorylates glycogen synthase kinase to deactivate it and inhibits glycogen synthase and ATP-citrate lyase activity, thereby inhibiting glycogen and fatty acid synthesis, respectively. Akt also inactivates the mammalian target of rapamycin complex 1 to promote protein synthesis. In addition, Akt mediates cell survival by inhibiting the proapoptotic pathway, and it activates sterol regulatory binding proteins (SREBPs), which translocate to the nucleus to transcribe genes associated with fatty acid and cholesterol synthesis.

The MAPK pathway is activated when IRS-1 binds to growth factor receptor-bound protein 2 (Grb2). SOS binds to Grb2 and then to Ras, causing GDP –GTP exchange and the activation of Ras.

Activated Ras recruits c-Raf, which phosphorylates and activates MAPK/Erk kinase (MEK). MEK then phosphorylates extracellular signal-regulated kinase (Erk). Once activated, Erk is translocated to the nucleus, where its subsequent phosphorylation and transcriptional activation by transcription factors, such as ELK1, ultimately promote cell division, protein synthesis, and cell growth ^{[12][13]} (**Figure 1**).

4. Physiological Roles of Insulin

The major purpose of insulin is to regulate the body's energy supply by balancing micronutrient levels during the fed state ^[14]. Insulin is critical for transporting intracellular glucose to insulin-dependent cells/tissues, such as liver, muscle, and adipose tissue. Any imbalance in exogenous energy supplies results in the breakdown of fats stored in adipose tissue and eventually accelerates insulin secretion. In the following sections, we discuss the major role of insulin in regulating several insulin-dependent tissue/organ functions.

Among the many important functions of insulin, nutrient homeostasis, i.e., synthesizing glycogen from glucose and the conversion of excess glucose into fatty acids and precursor triglyceride (TAG), are the most crucial ^[15]. A search of the literature indicates that insulin upregulates glucose-utilizing activity via accelerated hepatic glucose utilization, glycolysis, and glycogenesis, and it downregulates glucose production by suppressing net glucose production, gluconeogenesis, and glycogenolysis ^[16]. Directly, it can bind with hepatic insulin receptors and subsequently activates insulin signaling pathways in the liver, which has been demonstrated both in vitro and in vivo experimental models ^{[12][18][19][20]}. On the other hand, indirect insulin action is mostly regulated by the reduction in pancreatic glucagon secretion ^[21], the inhibition of fat lipolysis ^[22], and the influence of overall hypothalamic insulin signaling ^[23], which subsequently affects hepatic glucose production.

Insulin performs several important functions in all body parts, and skeletal muscle is no exception. Approximately 70% of the glucose is used by skeletal muscle from the whole-body glucose uptake ^[24], and the rest is used by the liver through an insulin-dependent mechanism. Postprandial hyperglycemia encourages the pancreas to secrete insulin, and an increase in plasma insulin concentration triggers the uptake and use of glucose by skeletal muscle ^{[25][26][27]}. Given that skeletal muscle is a major site for glucose disposal, quantitative declines in muscle volume in patients with type 2 diabetes might adversely affect overall glucose metabolism; thus, insulin therapy could improve optimal glucose targets ^[28].

Like other parts of the body, the muscle needs a continuous source of energy to work efficiently, and carbohydrates and fats are the main energy sources used by muscle cells to produce ATP $^{[29]}$. At the same time, insulin stimulates skeletal muscle to take up glucose by activating several enzymes $^{[27]}$ In skeletal muscle control, glucose uptake and energy metabolism increase GLUT4 Insulin also controls the amount of branched-chain amino acids, non-esterified fatty acids, plasma glucose, and muscle mitochondrial ATP production $^{[30]}$.

In some organisms, specialized cells store extra energy in the form of lipids, and in multicellular organisms, adipose tissue is responsible for the lipid storage ^[31]. Therefore, the biological features of adipose tissue from different sites could play a significant role in the onset and progression of metabolic derangements, particularly in obese individuals. It has been reported that insulin regulates several aspects of adipose cells' functional development and differentiation ^[32]. However, further studies are needed to understand the molecular basis of adipose tissue response to insulin as it is highly relevant to the pharmacotherapy of diabetes and weight gain.

The endothelium, which forms the inner layer of blood and lymph vessels, is considered a highly active part of the body involved in several pathophysiological processes, such as leukocyte adhesion, the control of vasomotor tone, inflammation, and barrier function ^[33]. Insulin is thought to be important for several functions relating to the endothelium, among which endothelium dysfunction is the most discussed ^[34]. Generally, endothelial dysfunction indicates events such as reduced nitric oxide (NO) bioavailability, increased oxidative stress mediated by elevated reactive oxygen species

(ROS) production, the expression of pro-inflammatory and pro-thrombotic factors, and abnormal vasoreactivity ^[34]. Indeed, an in-depth understanding of insulin's role in both the physiology and physiology of endothelial cells is essential for developing novel approaches for treating and preventing cardiovascular complications ^[35].

Initially, the brain was considered insulin insensitive because glucose uptake in this organ is unaffected by insulin $^{[36][37]}$. However, growing evidence suggested that insulin can increase glucose uptake in the spinal cord tissues and some brain regions, such as the choroid plexus, the pineal gland, and the pituitary $^{[38][39][40]}$. In addition to glucose metabolism and the energy balance in the brain, insulin is reported to control other vital physiological functions, such as neuronal plasticity, memory processing, and cognition $^{[41][42][43]}$. Therefore, depending on the evidence described above, insulin plays a very important role in brain function and might be an excellent therapeutic compound for treating both obesity and Alzheimer's disease, as suggested by other contemporaries $^{[44][45][46]}$.

The initial evidence of insulin actions in the kidney has been suggested in the 1950s [47]. Insulin involves various metabolic processes, including kidney homeostasis [48]. Hyperglycemia is responsible for the development of diabetic kidney diseases (DKDs). Therefore, the progression of kidney diseases, including DKD, could be slowed or reversed by controlling hyperglycemia with insulin therapy [49].

Another important physiological role of insulin is that it promotes bone development ^[50]. Bone formation may be regulated by insulin signaling through osteoblast development and bone resorption by osteoclasts ^[50]. In another in vitro study, it has been reported that insulin is an anabolic agent capable of increasing the rate of osteoblast proliferation, collagen synthesis, and alkaline phosphatase production, and it facilitates glucose uptake and subsequently inhibits the activities of osteoclasts ^[51]. Additionally, insulin-deficient models displayed a decline in mineralized and osteoid surface areas due to altered mineral apposition rates and osteoblast activity ^[51], and epidemiological investigations have further reinforced this finding.

Therefore, chronic decreases in oxygen and nutrient supplies due to hyperglycemic conditions may lead to follicular damage, which results in the alteration of regular hair growth (e.g., thinning, fragility, sparseness of hair, and decline in hair growth) The corresponding relationship between insulin action and the hair follicle is not fully understood. However, certain skin conditions, such as acrochordon and acne, have been linked with insulin action ^{[52][53]}. Another skin disease, psoriasis, is connected to other metabolic diseases, including obesity and metabolic syndrome, and there is a link between psoriasis and IR through the common factor of adipose tissue ^{[54][55]}.

References

- Alarcon, C.; Lincoln, B.; Rhodes, C.J. The biosynthesis of the subtilisin-related proprotein convertase PC3, but no that of the PC2 convertase, is regulated by glucose in parallel to proinsulin biosynthesis in rat pancreatic islets. J. Biol. Chem. 1993, 268, 4276–4280.
- Kaufman, B.A.; Li, C.; Soleimanpour, S.A. Mitochondrial regulation of beta-cell function: Maintaining the momentum for insulin release. Mol. Asp. Med. 2015, 42, 91–104.
- 3. Vasiljevic, J.; Torkko, J.M.; Knoch, K.P.; Solimena, M. The making of insulin in health and disease. Diabetologia 2020, 63, 1981–1989.
- 4. Wendt, A.; Eliasson, L. Pancreatic alpha-cells—The unsung heroes in islet function. Semin. Cell Dev. Biol 2020, 103, 41–50.
- Cabrera, O.; Berman, D.M.; Kenyon, N.S.; Ricordi, C.; Berggren, P.O.; Caicedo, A. The unique cytoarchitecture of human pancreatic islets has implications for islet cell function. Proc. Natl. Acad. Sci. USA 2006, 103, 2334–2339.
- De Vos, A.; Heimberg, H.; Quartier, E.; Huypens, P.; Bouwens, L.; Pipeleers, D.; Schuit, F. Human and rat beta cells differ in glucose transporter but not in glucokinase gene expression. J. Clin. Investig. 1995, 96, 2489–2495.
- McCulloch, L.J.; van de Bunt, M.; Braun, M.; Frayn, K.N.; Clark, A.; Gloyn, A.L. GLUT2 (SLC2A2) is not the principal glucose transporter in human pancreatic beta cells: Implications for understanding genetic association signals at this locus. Mol. Genet. Metab. 2011, 104, 648–653.
- Gloyn, A.L.; Odili, S.; Zelent, D.; Buettger, C.; Castleden, H.A.; Steele, A.M.; Stride, A.; Shiota, C.; Magnuson, M.A.; Lorini, R.; et al. Insights into the structure and regulation of glucokinase from a novel mutation (V62M), which causes maturity-onset diabetes of the young. J. Biol. Chem. 2005, 280, 14105–14113.
- 9. Rorsman, P.; Braun, M. Regulation of insulin secretion in human pancreatic islets. Annu. Rev. Physiol. 2013, 75, 155– 179.

- Doliba, N.M.; Qin, W.; Najafi, H.; Liu, C.; Buettger, C.W.; Sotiris, J.; Collins, H.W.; Li, C.; Stanley, C.A.; Wilson, D.F.; et al. Glucokinase activation repairs defective bioenergetics of islets of Langerhans isolated from type 2 diabetics. Am. J. Physiol. Endocrinol. Metab. 2012, 302, E87–E102.
- 11. Yajima, H.; Komatsu, M.; Schermerhorn, T.; Aizawa, T.; Kaneko, T.; Nagai, M.; Sharp, G.W.; Hashizume, K. cAMP enhances insulin secretion by an action on the ATP-sensitive K+ channel-independent pathway of glucose signaling in rat pancreatic islets. Diabetes 1999, 48, 1006–1012.
- 12. Haeusler, R.A.; McGraw, T.E.; Accili, D. Biochemical and cellular properties of insulin receptor signalling. Nat. Rev. Mol. Cell Biol. 2018, 19, 31–44.
- 13. Saltiel, A.R. Insulin signaling in health and disease. J. Clin. Investig. 2021, 131, e142241.
- 14. Burks, D.J.; White, M.F. IRS proteins and beta-cell function. Diabetes 2001, 50 (Suppl. 1), S140–S145.
- 15. Alves-Bezerra, M.; Cohen, D.E. Triglyceride Metabolism in the Liver. Compr. Physiol. 2017, 8, 1-8.
- Hatting, M.; Tavares, C.D.J.; Sharabi, K.; Rines, A.K.; Puigserver, P. Insulin regulation of gluconeogenesis. Ann. NY Acad. Sci. 2018, 1411, 21–35.
- 17. Claus, T.H.; Pilkis, S.J. Regulation by insulin of gluconeogenesis in isolated rat hepatocytes. Biochim. Biophys. Acta 1976, 421, 246–262.
- 18. Marks, J.S.; Botelho, L.H. Synergistic inhibition of glucagon-induced effects on hepatic glucose metabolism in the presence of insulin and a cAMP antagonist. J. Biol. Chem. 1986, 261, 15895–15899.
- Sindelar, D.K.; Balcom, J.H.; Chu, C.A.; Neal, D.W.; Cherrington, A.D. A comparison of the effects of selective increases in peripheral or portal insulin on hepatic glucose production in the conscious dog. Diabetes 1996, 45, 1594– 1604.
- Sindelar, D.K.; Chu, C.A.; Venson, P.; Donahue, E.P.; Neal, D.W.; Cherrington, A.D. Basal hepatic glucose production is regulated by the portal vein insulin concentration. Diabetes 1998, 47, 523–529.
- 21. Ito, K.; Maruyama, H.; Hirose, H.; Kido, K.; Koyama, K.; Kataoka, K.; Saruta, T. Exogenous insulin dose-dependently suppresses glucopenia-induced glucagon secretion from perfused rat pancreas. Metabolism 1995, 44, 358–362.
- 22. Sindelar, D.K.; Chu, C.A.; Rohlie, M.; Neal, D.W.; Swift, L.L.; Cherrington, A.D. The role of fatty acids in mediating the effects of peripheral insulin on hepatic glucose production in the conscious dog. Diabetes 1997, 46, 187–196.
- 23. Obici, S.; Zhang, B.B.; Karkanias, G.; Rossetti, L. Hypothalamic insulin signaling is required for inhibition of glucose production. Nat. Med. 2002, 8, 1376–1382.
- 24. Baron, A.D.; Brechtel, G.; Wallace, P.; Edelman, S.V. Rates and tissue sites of non-insulin- and insulin-mediated glucose uptake in humans. Am. J. Physiol. 1988, 255 Pt 1, E769–E774.
- 25. DeFronzo, R.A. Pathogenesis of type 2 diabetes mellitus. Med. Clin. N. Am. 2004, 88, 787-835.
- 26. DeFronzo, R.A. Lilly lecture 1987. The triumvirate: Beta-cell, muscle, liver. A collusion responsible for NIDDM. Diabetes 1988, 37, 667–687.
- 27. Defronzo, R.A. Banting Lecture. From the triumvirate to the ominous octet: A new paradigm for the treatment of type 2 diabetes mellitus. Diabetes 2009, 58, 773–795.
- 28. Abdulla, H.; Smith, K.; Atherton, P.J.; Idris, I. Role of insulin in the regulation of human skeletal muscle protein synthesis and breakdown: A systematic review and meta-analysis. Diabetologia 2016, 59, 44–55.
- Romijn, J.A.; Coyle, E.F.; Sidossis, L.S.; Gastaldelli, A.; Horowitz, J.F.; Endert, E.; Wolfe, R.R. Regulation of endogenous fat and carbohydrate metabolism in relation to exercise intensity and duration. Am. J. Physiol. 1993, 265 Pt 1, E380–E391.
- Karakelides, H.; Asmann, Y.W.; Bigelow, M.L.; Short, K.R.; Dhatariya, K.; Coenen-Schimke, J.; Kahl, J.; Mukhopadhyay, D.; Nair, K.S. Effect of insulin deprivation on muscle mitochondrial ATP production and gene transcript levels in type 1 diabetic subjects. Diabetes 2007, 56, 2683–2689.
- Birsoy, K.; Festuccia, W.T.; Laplante, M. A comparative perspective on lipid storage in animals. J. Cell Sci. 2013, 126 Pt 7, 1541–1552.
- 32. Cignarelli, A.; Genchi, V.A.; Perrini, S.; Natalicchio, A.; Laviola, L.; Giorgino, F. Insulin and Insulin Receptors in Adipose Tissue Development. Int. J. Mol. Sci. 2019, 20, 759.
- 33. Favero, G.; Paganelli, C.; Buffoli, B.; Rodella, L.F.; Rezzani, R. Endothelium and its alterations in cardiovascular diseases: Life style intervention. BioMed Res. Int. 2014, 2014, 801896.
- 34. Barac, A.; Campia, U.; Panza, J.A. Methods for evaluating endothelial function in humans. Hypertension 2007, 49, 748–760.

- 35. Potenza, M.A.; Addabbo, F.; Montagnani, M. Vascular actions of insulin with implications for endothelial dysfunction. Am. J. Physiol. Endocrinol. Metab. 2009, 297, E568–E577.
- 36. Hom, F.G.; Goodner, C.J.; Berrie, M.A. A [3H]2-deoxyglucose method for comparing rates of glucose metabolism and insulin responses among rat tissues in vivo. Validation of the model and the absence of an insulin effect on brain. Diabetes 1984, 33, 141–152.
- 37. Hasselbalch, S.G.; Knudsen, G.M.; Videbaek, C.; Pinborg, L.H.; Schmidt, J.F.; Holm, S.; Paulson, O.B. No effect of insulin on glucose blood-brain barrier transport and cerebral metabolism in humans. Diabetes 1999, 48, 1915–1921.
- 38. Havrankova, J.; Schmechel, D.; Roth, J.; Brownstein, M. Identification of insulin in rat brain. Proc. Natl. Acad. Sci. USA 1978, 75, 5737–5741.
- 39. Ramnanan, C.J.; Edgerton, D.S.; Cherrington, A.D. Evidence against a physiologic role for acute changes in CNS insulin action in the rapid regulation of hepatic glucose production. Cell Metab. 2012, 15, 656–664.
- 40. Ramnanan, C.J.; Kraft, G.; Smith, M.S.; Farmer, B.; Neal, D.; Williams, P.E.; Lautz, M.; Farmer, T.; Donahue, E.P.; Cherrington, A.D.; et al. Interaction between the central and peripheral effects of insulin in controlling hepatic glucose metabolism in the conscious dog. Diabetes 2013, 62, 74–84.
- 41. Benedict, C.; Hallschmid, M.; Schultes, B.; Born, J.; Kern, W. Intranasal insulin to improve memory function in humans. Neuroendocrinology 2007, 86, 136–142.
- 42. Craft, S.; Baker, L.D.; Montine, T.J.; Minoshima, S.; Watson, G.S.; Claxton, A.; Arbuckle, M.; Callaghan, M.; Tsai, E.; Plymate, S.R.; et al. Intranasal insulin therapy for Alzheimer disease and amnestic mild cognitive impairment: A pilot clinical trial. Arch. Neurol. 2012, 69, 29–38.
- 43. Taouis, M.; Torres-Aleman, I. Editorial: Insulin and The Brain. Front. Endocrinol. 2019, 10, 299.
- 44. Reger, M.A.; Watson, G.S.; Frey, W.H., 2nd; Baker, L.D.; Cholerton, B.; Keeling, M.L.; Belongia, D.A.; Fishel, M.A.; Plymate, S.R.; Schellenberg, G.D.; et al. Effects of intranasal insulin on cognition in memory-impaired older adults: Modulation by APOE genotype. Neurobiol. Aging 2006, 27, 451–458.
- Reger, M.A.; Watson, G.S.; Green, P.S.; Wilkinson, C.W.; Baker, L.D.; Cholerton, B.; Fishel, M.A.; Plymate, S.R.; Breitner, J.C.; DeGroodt, W.; et al. Intranasal insulin improves cognition and modulates beta-amyloid in early AD. Neurology 2008, 70, 440–448.
- 46. Hallschmid, M.; Benedict, C.; Born, J.; Fehm, H.L.; Kern, W. Manipulating central nervous mechanisms of food intake and body weight regulation by intranasal administration of neuropeptides in man. Physiol. Behav. 2004, 83, 55–64.
- 47. Farber, S.J.; Berger, E.Y.; Earle, D.P. Effect of diabetes and insulin of the maximum capacity of the renal tubules to reabsorb glucose. J. Clin. Investig. 1951, 30, 125–129.
- 48. Singh, S.; Sharma, R.; Kumari, M.; Tiwari, S. Insulin receptors in the kidneys in health and disease. World J. Nephrol. 2019, 8, 11–22.
- 49. Akhtar, M.; Taha, N.M.; Nauman, A.; Mujeeb, I.B.; Al-Nabet, A. Diabetic Kidney Disease: Past and Present. Adv. Anat. Pathol. 2020, 27, 87–97.
- 50. Malekzadeh, B.O.; Erlandsson, M.C.; Tengvall, P.; Palmquist, A.; Ransjo, M.; Bokarewa, M.I.; Westerlund, A. Effects of implant-delivered insulin on bone formation in osteoporotic rats. J. Biomed. Mater. Res. A 2018, 106, 2472–2480.
- Thrailkill, K.M.; Lumpkin, C.K., Jr.; Bunn, R.C.; Kemp, S.F.; Fowlkes, J.L. Is insulin an anabolic agent in bone? Dissecting the diabetic bone for clues. Am. J. Physiol. Endocrinol. Metab. 2005, 289, E735–E745.
- 52. Pierard, G.E.; Seite, S.; Hermanns-Le, T.; Delvenne, P.; Scheen, A.; Pierard-Franchimont, C. The skin landscape in diabetes mellitus. Focus on dermocosmetic management. Clin. Cosmet. Investig. Dermatol. 2013, 6, 127–135.
- 53. Napolitano, M.; Megna, M.; Monfrecola, G. Insulin resistance and skin diseases. Sci. World J. 2015, 2015, 479354.
- Davidovici, B.B.; Sattar, N.; Prinz, J.; Puig, L.; Emery, P.; Barker, J.N.; van de Kerkhof, P.; Stahle, M.; Nestle, F.O.; Girolomoni, G.; et al. Psoriasis and systemic inflammatory diseases: Potential mechanistic links between skin disease and co-morbid conditions. J. Investig. Dermatol. 2010, 130, 1785–1796.
- 55. Toussirot, E.; Aubin, F.; Dumoulin, G. Relationships between Adipose Tissue and Psoriasis, with or without Arthritis. Front. Immunol. 2014, 5, 368.