# **Natural Herbals and Spices in Wound Healing**

Subjects: Integrative & Complementary Medicine

Contributor: Stefania Vitale, Sara Colanero, Martina Placidi, Giovanna Di Emidio, Carla Tatone, Fernanda Amicarelli, Anna Maria

D'Alessandro

Herbal medicines have been employed in folk medicine to accelerate wound healing since ancient times. Many plants and various preparations thereof have been used traditionally in relation to wound treatment, especially due to their immense potential to affect wound healing. Plant-based extracts and/or isolates support tissue regeneration through a variety of mechanisms, which often work together to improve the whole healing process. Currently, the efficacy of many of these herbs is well documented together with their mechanisms of action. Therefore, natural products as well as their pure compounds are an emerging source of alternative medicinal compounds for the management of various diseases, among which is wound healing.

Keywords: wound healing; herbs; medicinal plants; secondary metabolites; natural wound dressing

### 1. Achillea millefolium

Achillea millefolium L. (yarrow) is an important species of Asteraceae family with common utilization in the traditional medicine of several cultures from Europe to Asia for the treatment of various disorders including wounds, cuts, abrasions, and diabetic ulcers [1][2][3][4]. Essential oils, the most active part of the yarrow flower, are used in wound therapy as a hemostatic [5][6][7]. The most representative metabolites in volatile oil extracted from Achillea flowers are monoterpenes (90% of the essential oils). However, there are reports on the presence of higher levels of sesquiterpenes compared with monoterpenes [8][9][10]. A wide range of other chemical compounds has also been reported in the methanolic extract of A. millefolium aerial parts. Among them, flavonoids such as chlorogenic acid, rutin, luteolin-glucoside, and apigeninglucoside account for over 90% of the total extract components [11]. In different in vitro and in vivo experiments, active compounds derived from A. millefolium showed antioxidant and anti-inflammatory, mostly attributed to the presence of flavonoids [4][12][13][14][15][16][17]. Despite yarrow being extensively used for wound healing and skin inflammatory disorders in traditional medicine [17], modern studies of yarrow extract effects on wound healing are lacking.

In a recent paper, Dorjsembe et al.  $^{[18]}$  investigated the effect of the alcoholic extract of *Achillea asiatica flowers* on cutaneous wound healing, in in vitro and in vivo models, showing that 3% YE treatment significantly increased epithelialization and accelerated wound healing in a rat model. This effect was associated with an increase in  $\beta$ -catenin, and Akt expressions  $^{[18]}$ . Furthermore, in in vitro experiments, YE treatments (25–50 mg/mL) stimulated collagen expression by activating transforming growth factor- $\beta$  (TGF- $\beta$ ) in Hs68 fibroblasts and reduced Nitric Oxide (NO) and prostaglandin E2 released, in RAW 264.7 macrophages, reflecting anti-inflammatory activity. Based on these results, a beneficial positive effect of yarrow flower extract on wound healing can be hypothesized  $^{[18]}$ .

#### 2. Aloe vera

Aloe vera L. or Aloe barbadensis Miller (family Xanthorrhoeaceae) is a perennial green herb with bright yellow tubular flowers. The mucilaginous gel has been extensively used in pharmacological and cosmetic applications [19]. It has more than 75 different compounds, including vitamins A, C, E, and B12, enzymes (amylase, catalase, and peroxidase), minerals (zinc, copper, selenium, and calcium), sugars (glucomannans, acetylated mannans, polymannans), polyphenols (anthraquinones), sterols (lupeol and campesterol), and hormones (auxins and gibberellins) [20][21][22]. Traditionally, the therapeutic uses of *A. vera* have ranged across a broad list of conditions, as do its associated pharmacological activities. This medicinal plant has been employed to treat different skin problems such as rejuvenation, wound healing, and other dermatologic conditions, i.e., burns and inflammatory processes. Indeed, despite its widespread use as a folk remedy, scientific studies on its physiological function in wound repair have only recently been conducted [21]. Numerous *A. vera* gel-based cosmetics and medicinal products that are marketed are made from the mucilaginous tissue of *A. vera* leaves. However, Aloe gel has been linked to a variety of beneficial effects and therapeutic indications for skin inflammation [21].

Individual components of *A. vera* gel may promote wound healing in animal models, and specific glycoproteins are responsible for the beneficial effects when the gel is applied to acute wounds in various animal models; however, controlled clinical trials in humans demonstrated no benefit when *A. vera* was incorporated into topical therapy, and one study reported delayed wound healing [23][24][25]. Recently, topical use of *A. vera* in rat cutaneous wound models has shown that it significantly increases the rate of wound contraction, epithelialization, and maturation. Moreover, it reduces inflammation, decreases scar tissue size, and increases alignment and organization of regenerated scar tissue, increasing the concentration of collagen and glycosaminoglycans compared to the control lesions [26].

To clarify the mechanism by which *A. vera* modulates inflammation, Hormozi and colleagues  $^{[27]}$  exposed mouse embryonic fibroblast cells to different concentrations (50–150 µg/mL) of *A. vera* gel for 12 and 24 h  $^{[27]}$  and found that Transforming growth factor -beta1 (TGF1) and Fibroblast Growth Factor basic Protein (bFGF) genes were found upregulated during the first 12 h and down-regulated at the end of incubation. This suggests that *A. vera* exerts beneficial effects by stimulating collagen deposition, fibroblast proliferation, and angiogenesis and inhibiting the overproduction and accumulation of matrix proteins that cause hypertrophic scarring  $^{[27]}$ .

In in vitro models of human skin fibroblasts and keratinocytes [28][29][30][31], *A. vera* accelerated wound healing by strongly stimulating fibroblast and keratinocyte proliferation and moderately stimulating cell migration. Surprisingly, *A. vera* also protected keratinocytes against preservative-induced death [32]. These previously unknown protective actions may account for some of the beneficial benefits of *A. vera* in wound healing.

According to a recent review *A. vera* compounds such as aloesin can modulate the inflammatory response  $\frac{[31]}{1}$ . Aloesin promotes leukocyte extravasation as well as cytokine and growth factor release during the early stages of wound healing. As a result, aloesin appears to influence both fibroblast and leukocyte migration  $\frac{[27]}{1}$ . Aloesin is thought to influence leukocyte migration by phosphorylating Cdc42 and Rac1, two signaling proteins that coordinate and regulate actin dynamics and cell polarization  $\frac{[28]}{1}$ . In the presence of aloesin, TNF-alpha, Interleukin 1 beta (IL-1beta), Interleukin 6 (IL-6), and TGF-1 are pro-inflammatory markers that mediate leukocyte signaling, migration, and phagocytosis  $\frac{[29]}{1}$ . Recently, a study by De Oliveira et al.  $\frac{[30]}{1}$  established that the healing property of *A. vera* extract can be observed at a dose of 50 mg/kg, but at the same dose, mutagenic and cytotoxic effects are observed in peripheral blood  $\frac{[30]}{1}$ .

Aside from flavonoids such as aloin and emodin, polysaccharides such as glucomannan, acetylated polymannan, acemannan, and mannose-6-phosphate appear to promote wound healing  $\frac{[22]}{2}$ . Some studies showed that acemannan appears to stimulate macrophages and boost bactericidal activity  $\frac{[24][25]}{2}$ , the expression of keratinocyte growth factor-1 (KGF-1), cell proliferation, as well as vascular endothelial growth factor (VEGF), an important molecule in the formation of new blood vessels, and type I collagen synthesis  $\frac{[33][34]}{2}$ . Studies in skin fibroblasts revealed that acemannan not only increased proliferation but also shifted the cell cycle from the G1 to S phase, increasing the expression of proliferation markers such as cyclin D1 in a dose-dependent manner  $\frac{[35][36]}{2}$ .

Some studies have found no significant differences between *A. vera* treatment and control groups in terms of wound healing, with little evidence of decreasing microorganism concentrations or improving scar maturation in burn wounds, no significance in fibroblast migration dynamics, and no difference between sample groups in terms of wound repair [31]. This is likely due to a problem of standardization of extract concentrations. Some of these studies may have used aloe extracts with insufficient concentrations of active compounds with known anti-inflammatory and proliferative effects, or they may have been combined with other unsuitable alternative therapies.

#### 3. Bletilla striata

Bletilla striata is a member of the *Orchidaceae* family and has been used in Traditional Chinese Medicine for over 1500 years to promote wound healing and treat alimentary canal mucosal damage, chapped skin, ulcers, bleeding, bruises, and burns [37]. Phytochemical research on *B. striata* yielded the identification of 192 monomeric compounds. These compounds, extracted from *B. striata* tubers, are primarily triterpenoids: Phenanthrenes, biphenanthrenes, dihydrophenanthrenes, anthocyanins, quinones, steroids, glucosides, bibenzyls, and phenolic acids [38]. According to pharmacology studies, the plant has many biological activities, including antioxidative, anti-inflammatory, and immune regulatory effects [38]. *Bletilla striata* also contains several polysaccharides, which have been identified as the major active components in its dried tubers, and are responsible for antimicrobial, anti-aging, antioxidative, and antiviral activity [39]. Natural polysaccharides have been used in a variety of biomaterials in recent years due to their high biocompatibility, low toxicity, and pharmaceutical biomedical activity. *Bletilla striata* polysaccharides (BSP) can be used as natural biomaterials for drug delivery and wound dressing in addition to promoting wound healing [40]. The most common polysaccharide in BSP is glucomannan, which is made up of D-glucose and D-mannose and has a high molecular weight [41]. BSP plays a

critical role in the three main phases of wound healing: Inflammation, granulation tissue proliferation, and repair  $^{[42]}$ . During the inflammation phase, BSP promotes the expression of inflammatory mediators such as tumor necrosis factor (TNF)-, interleukin (IL)-1, and interferon (IFN)-, increases NO, and promotes neutrophil, monocyte, and macrophage chemotaxis  $^{[43]}$ . By controlling the expression of TNF-, BSP can reduce the inflammatory reaction in the wound and prevent damage to remaining cells during the phases of granulation tissue proliferation and repair  $^{[44]}$ . These actions promote epithelial cell growth, fibroblast proliferation, and wound healing by contracting the wound  $^{[37][45]}$ . Yue and colleagues  $^{[46]}$  measured the response induced by BSP pretreatment in terms of ROS levels and proinflammatory cytokines after Ang II stimulation with and without BSP and investigated the signaling pathways potentially involved in the anti-oxidative stress and anti-inflammatory functions mediated by BSP in a human mesangial cell model in vitro (HMCs) study. BSP was found to inhibit the generation of Ang II-induced reactive oxygen species (ROS) as well as the activation of pro-inflammatory cytokines such as interleukin 6 (IL-6) and tumor necrosis factor (TNF), in a dose-dependent manner (5–80  $\mu$ g/mL). Furthermore, BSP effectively inhibited NADPH oxidase 4 upregulation (NOX4). Indeed, NOX4 knockdown significantly reduced Ang II-induced TLR2 overexpression and blocked TLR2 expression, impairing BSP's anti-inflammatory effects. Furthermore, BSP has anti-oxidative and anti-inflammatory properties that can help stop ROS production and the production of proinflammatory cytokines  $^{[46]}$ .

BSP derived from *B. striata* has been used in wound dressings for many years, with outstanding biocompatibility, biodegradability, and gelling properties [44]. BSP hydrogel demonstrated excellent performance in the absorption of wound fluids and exudates and was able to provide water to the wound, thus creating a suitable fluid environment for the wound bed [44]. Surprisingly, recent research has revealed that BSP has hemostatic activity, likely mediating blood clotting and fibrinolysis [47][48][49]. BSP has been made into a new type of hemostatic agent that can be used as a drug delivery vehicle and wound dressing. [24].

Ding et al.  $^{[50]}$  created a chemically cross-linked composite bilayer as a spongy wound dressing and tested it on dermal wounds in mice. The upper layer was made of chitosan-Ag nanoparticles that were cross-linked with genipin, extracted from gardenia fruits  $^{[51]}$ , and exerted antibacterial activity. The lower layer was a hybrid cross-linked chitosan with genipin and partially oxidized BSP that revealed potent cell proliferation activities. Based on its ability to accelerate the healing rate of cutaneous wounds, this novel composite bilayer has a great deal of potential in wound-dressing applications  $^{[52]}$ . Y. Song and colleagues  $^{[52]}$  discovered that the fibrous residues of polysaccharide extraction from *B. striata* used in traditional medicine have a high phenolic content, strong DPPH (2,2-diphenyl-1-picrylhydrazyl) scavenging activity, ferric-reducing antioxidant capacity, and tyrosinase inhibition activity  $^{[52]}$ . After 15 days of treatment, the wound shrinkage percent in mice treated with mix ointment containing 1% residue was statistically higher than in animals treated with saline solution, Bletilla phenolic ointment, or partially oxidized BSP ointment. Based on these findings, the authors concluded that wound-healing functions of *B. striata* are due not only to the composition of the polysaccharides, but also to the activity of the polyphenols  $^{[52]}$ .

In a more recent study, Cheng et al. [53] combined BSP and the ethanol extract of *B. striata* (EEB) to create new composite sponges through simple mixing and freeze-drying processes. When EEB (25–50% in H<sub>2</sub>O) was added to the BSP sponge, it influenced blood coagulation factors, shortened hemostasis time, and improved antiseptic properties for wound healing. The degradation, biocompatibility, hemostatic, bacteriostatic, and wound-healing-promotion properties were also tested in vivo and in vitro [53]. This plant could be used to make plant-based components for wound dressing and drug delivery systems on a large scale.

### 4. Calendula officinalis

Calendula officinalis, belonging to the Asteraceae family, is a common garden plant used medicinally in Europe, China, the United States, and India. It has numerous common names in use, including Marigold and Pot Marigold. Traditionally, it has been used externally to treat small wounds, burns, and other skin problems [54][55]. C. officinalis can be used in the form of an infusion, tincture, liquid extract, cream, or ointment for numerous uses, including the treatment of herpes, wounds, scars, and skin and hair products [54][55][56][57].

Over the past decade, chemical and pharmacological studies have found that *C. officinalis* possesses many secondary metabolites with various pharmacological properties that contribute to its medicinal use [56][57][58][59][60][61][62]. The most active components are triterpenoids, both in their free and esterified forms, flavonoids, coumarines, quinones, volatile oil, carotenoids, polyunsaturated fatty acids, such as calendic acid, and amino acids [31][60].

Most of the research on the role of *C. officinalis* in healing acute wounds came from in vitro (fibroblasts and keratinocytes cells) and in vivo (rodent animal models) studies [59][62][63][64][65][66][67]. Clinical studies have focused on chronic wounds

[60], and only one clinical trial looked at acute wounds [61]. *Calendula* flower alcohol extracts (CFE) have shown anti-inflammatory and antiedematous properties in vitro. Furthermore, they increased human fibroblast and keratinocyte proliferation and migration, promoted angiogenesis in the chorioallantois membrane model, and decreased collagenase activity [66]. CFE influenced the inflammatory phase in keratinocytes by activating the transcription factor NF-kB and increasing the amount of chemokine IL-8 at both transcriptional and translational levels, whereas it had a minor effect on keratinocyte migration. CFE induces fibroblast proliferation [59][67] and upregulates the expression of connective tissue growth factor (CTGF) and alpha-smooth muscle actin (alpha-SMA) during the early stages of wound healing [67]. In a previous in vivo study, CFE reduced the presence of fibrin and hyperemia while increasing collagen deposition in a rat cutaneous wound model. These findings indicate that CFE has anti-bacterial properties, stimulates fibroplasia and angiogenesis, and positively influences the inflammatory and proliferative phases of the healing process of cutaneous wounds [67]. Terpenoids seem to be primarily responsible for all these effects.

Recently, ethanolic CFE was loaded onto chitosan nanofibers as a wound-healing dressing [68], and potent antibacterial properties were observed, with more than 90% reduction of Gram-positive and Gram-negative bacteria. In vitro studies with fibroblast cells demonstrated this mixed film increased cell proliferation, growth, and adhesion and inhibited cell collagenase activity with a subsequent increase in collagen production. In vivo studies of rat wounds revealed that this dressing has an excellent wound-healing capacity (87.5% wound closure after 14 days) by enhancing collagen synthesis, re-epithelialization, and tissue remodeling [68]. In all the studies examined, CFE was associated with a statistically significant improvement in wound healing according to the outcomes measured [59][65][66][67][68].

## 5. Casearia sylvestris

De Campos et al. [77], using a burn wound model in rats, carried out daily wound treatments with a biofilm impregnated with *C. sylvestris* extract and observed significant extension of the healing area, neovascularization, fibroblast proliferation, and epithelialization, concluding that *C. sylvestris* may have a potential therapeutic benefit in second-degree scald burn injuries. This effect has been ascribed to rutin, a *C. sylvestris* bioactive compound with powerful antioxidant properties [71]. When *Casearia sylvestris* leaves extract was tested on wound healing in beef cattle, cattle, a positive effect on the macroscopic aspect of cutaneous lesions in cattle was observed during the first two treatment days [78].

In a recent paper, the authors developed membranes for wound healing dressing by combining a natural latex from *Hevea brasiliensis* (*Euphorbiaceae*) providing angiogenic action, with leaf ethanolic extract of *Casearia sylvestris Sw.* (diterpene concentrated fraction and casearin J) characterized by anti-inflammatory and wound-healing activities [79]. Permeation and retention assays demonstrated the dermal penetration of phenolic compounds from the membrane with ethanolic extract and penetration of casearin-like clerodane diterpenes from all membranes, indicating that these topical systems have great potential for therapeutical application of *C. sylvestris* components [79].

#### 6. Crocus sativus

Crocus sativus L., from the family of *Iridaceae*, is commonly known as saffron, which is the name of a spice derived from the *C. sativus* stigmas dried. Saffron is used for medicinal purposes in Chinese, Ayurvedic, Persian, and Unani traditional medicines [80]. The therapeutic properties of saffron used for healing purposes could be found in Materia Medica, written by a Greek physician (*Pedanius Dioscorides*) in the first century A.D. [81]. Modern pharmacological studies have demonstrated that saffron extract or its bioactive constituents, such as apocarotenoids, mono-terpenoids, flavonoids, phenolic acids, and phytosterols, have a wide range of therapeutic effects [82]. Major carotenoids contained in saffron extracts (SE) include crocin, crocetin, picrocrocin, and safranal [83]. Crocin, the digentiobiosyl ester of crocetin, is one of the few water-soluble carotenoids found in nature [83]. These compounds have been shown to have a wide spectrum of biological activities, including several antioxidant and anti-inflammatory properties [82][83][84][85]. Recent studies have shown that the effects of crocin, crocetin, and safranal against oxidative stress include the reduction in lipid peroxidation

(malondialdehyde levels) and nitric oxide levels, and an increase in the levels of glutathione, antioxidant enzymes (superoxide dismutase, catalase, and glutathione peroxidase), and thiol content [86][87][88][89].

Despite the large number of papers indicating that saffron and its carotenoids have potent antioxidant and anti-inflammatory properties, only a few studies have reported these effects in wound healing  $^{[90][91]}$ . In an in vitro study, anti-inflammatory properties and ROS scavenging activity of crocin were investigated in human epidermal keratinocyte and human dermal fibroblasts,  $^{[92]}$ . Crocin inhibited squalene peroxidation and UVA-induced cell membrane arachidonic acid peroxidation in a dose-dependent manner (31-113-195  $\mu$ M) and, from 300  $\mu$ M to 1 mM, prevented the release of inflammatory mediators, modulating the expression of NF-kB-related genes and glycosylation-related genes  $^{[92]}$ . Given the impact of UV exposure on skin quality  $^{[93]}$ , this study suggests that crocin can be regarded as a molecule with potential beneficial effects against skin aging.

More recently, an in vivo study was conducted to compare the effect of saffron extract to silver sulfadiazine (SSD), the most widely used topical treatment for burn injuries, in a burn wound model in rats. The animals treated with 20% saffron pomade showed more significant wound closure than the SSD-treated and untreated groups (83.04  $\pm$  1.36% versus 57.57  $\pm$  2.8% versus 35.53  $\pm$  3.5%, respectively, on day 7, p < 0.001), including re-epithelialization and wound contraction [94]. Based on the scratch test, in vitro experiments confirmed the positive role of saffron in fibroblast migration and proliferation during the remodeling phase, as well as regenerative and anti-scarring properties of saffron [94].

In the production of saffron spices, copious amounts of floral bio residues are generated and wasted. Interestingly, polyphenols and anthocyanins found in the sepals and stamens of C. sativus have been used as a source of antioxidant and anti-inflammatory molecules with beneficial effects against skin pathologies  $\frac{[95][96]}{}$ . Khorasani and colleagues  $\frac{[91]}{}$  previously evaluated the efficacy of saffron pollen extract in cream in the treatment of thermally induced burn wounds in rats. When compared to those treated with silver sulfadiazine, the results showed that saffron cream accelerated wound healing and reduced healing time. Furthermore, wounds treated with saffron cream had fewer inflammatory cells than the other groups, and histological studies of wound sections confirmed the formation of new epiderma, and fewer inflammatory cells infiltrated the dermis. These effects were most likely mediated by antioxidant and anti-inflammatory mechanisms, though the exact mechanism has yet to be determined  $\frac{[91]}{}$ .

Zeka et al.  $^{[90]}$  developed a new hydrogel enriched with antioxidants derived from saffron flowers. When kaempferol or 10  $\mu$ M crocin-enriched hydrogels were used in fibroblast cell cultures isolated from newborn mice (as a model for skin), it was found that cells grew faster when compared to the control  $^{[90]}$ . Although more research is needed to understand the mechanisms of action and pathways involved in wound healing, the antioxidant potential of compounds extracted from saffron petals and other floral bio residues can be effectively used in wound management.

## 7. Curcuma longa

*Curcuma longa* is an herbaceous, perennial, rhizomatous plant belonging to the *Zingiberacee* family (one of the many species of the *Curcuma* genus). The spice is obtained from the rhizome of *Curcuma longa* (commonly called "turmeric") and contains polyphenolic antioxidant compounds  $^{[97]}$ . In Ayurvedic and folk medicine, turmeric has been used for a long time to treat various inflammatory diseases. The anti-inflammatory effect of turmeric is due to its ability to decrease the production of histamine and prolong the action of the natural anti-inflammatory adrenal hormone, cortisol  $^{[98]}$ .

Turmeric's effects on health are generally centered upon its principal component, called "curcumin," an orange-yellow-colored, lipophilic polyphenol compound <sup>[99]</sup> (**Figure 3**). Curcumin is one of the three curcuminoids present in turmeric, making up 2 to 5% of the spice <sup>[99]</sup> and approximately 77% of the singular extract <sup>[99]</sup>. The structure of curcumin (1,7-bis (4-hydroxy-3methoxyphenyl)-1,6-hepadiene-3,5-dione) was first described by Milobedska et al. in 1910 <sup>[100]</sup>. Curcumin is a highly pleiotropic molecule <sup>[101]</sup>. In recent times, it has been extensively studied as an anti-cancer <sup>[102]</sup>, anti-aging <sup>[103]</sup>, antiviral <sup>[104]</sup>, antibacterial <sup>[105]</sup>, and wound-healing agent <sup>[106]</sup>. The wound-healing potential of curcumin is attributed to its anti-inflammatory <sup>[107]</sup>, antioxidant, and radical scavenging <sup>[108]</sup> activities. Curcumin acts in all wound-healing stages. In the inflammation stage, curcumin was shown to inhibit the activity of NF-kB and the production of TNF-alpha and IL-1 <sup>[109]</sup> <sup>[110]</sup>.

Radical-scavenging activities of curcumin have been reported in studies on wound healing, although this compound can act as a pro-oxidant when used at high doses [111][112][113][114]. In in vitro studies on keratinocytes and fibroblasts [115][116], curcumin provided optimal protection against hydrogen peroxide. In the proliferative stage, curcumin enhances fibroblast migration, granulation tissue formation, collagen deposition, and re-epithelialization. Curcumin improves wound contraction during the remodeling stage by increasing TGFbeta-production and, as a result, fibroblast proliferation. The

suppression of acute and chronic inflammation also occurs by blocking the formation of cyclooxygenases, lipoxygenases, and inducible nitric oxide synthase enzymes [115].

Curcumin was used in a recent study to optimize the survival and tolerance capacity of bone marrow mesenchymal stem cells, which were used to improve tissue repair in chronic wounds and burns, in terms of self-renewal and paracrine factor production [116].

Furthermore, curcumin was explored for potent antiherpetic actions against herpes simplex virus type 1 and type 2, and the associated inflammation connected to these diseases  $^{[104]}$ . Indeed, clinically, HSV-1 and HSV-2 infections cause lesions on the mucocutaneous junctions of the face and genitalia. Vesicular lesions can sometimes ulcerate, leaving recalcitrant wounds that are difficult to treat  $^{[104]}$ . Until now, the foundation of treatment has been the eradication of viral infection. Little attention has been paid to the outcome of the viral infection and the wounds that resulted, specifically whether this represents an epidermal or dermal injury. Curcumin represents an opportunity for dressing and healing these lesions.

Unfortunately, the use of curcumin is limited by its low bioavailability, rapid metabolism, poor solubility, and light sensitivity [101]. A large number of recent papers [117][118][119][120][121][122][123][124][125][126][127] reported the incorporation of curcumin into nanocomposite materials such as nanofibers, nanoparticles, hydrogels, or novel combinations of these. Recently, numerous researchers have developed biocompatible and biodegradable composite nano-fibrous materials and hydrogel systems containing curcumin for wound healing applications, for example as an anti-scar or in chronic wounds [115][118]. These scaffold systems include polyvinyl pyrrolidone (PVP) [118], cerium nitrate hexahydrate (Ce(NO3)3·6H2O) [119], nanofibers of carboxymethyl guargum (CMGG) [120], electrospun amine-functionalized SBA-15 (Santa Barbara Amorphous) incorporated with PVA (Poly Vinyl Alcohol) [121], nanofibers of cellulose acetate (CA) integrated with graphene oxide, TiO2 [122], electrospun fibers comprising SBA-15 (Santa Barbara Amorphous), amine-functionalized SBA-15 polycaprolactone (PCL) [123], nanofibers of electrospun poly(ε-caprolactone) (PCL), Chitosan (CS) [124], thermo-sensitive sodium alginate hydrogel cross-linked with poly(N-isopropylacrylamide), and curcumin (Alg-g-pNIPAM) copolymers [125]. All these nanofibrous scaffolds exhibited high porosity and biocompatible characteristics that enhance the solubility of this hydrophobic drug, aiding its release in a controlled manner [125]. A novel curcumin-loaded sandwich-like nanofibrous membrane was prepared using sequential electrospinning. The nanofibrous membrane in the sub-layer consisting of gelatin, chitosan, and PCL provided efficient hemostatic activity and was effective in absorbing exudate and keeping the wound moist. The curcumin-loaded nanofibrous membrane in the mid-layer released curcumin, therefore reducing wound oxidative stress and inflammation. Finally, the top layer, consisting of Ag nanoparticles, silk fibroin, and PCL, released Ag nanoparticles, which acted against external bacteria [125]. In a further study, when the eggshell membrane, a highly proteinaceous thin layer present between the egg white and shell, was added to the bottom layer made up of PVA with curcumin nanoparticles, antioxidant and anti-inflammatory action was implemented with positive effects on the migration of dermal fibroblasts [126]. This patch contributed to wound healing by providing exudate absorption, releasing therapeutical components, and supporting the deposition of extracellular matrix [126][127].

## 8. Glycyrrhiza glabra L.

Glycyrrhiza glabra is a herbaceous perennial legume of the Fabaceae bean family, from whose root is extracted sweet and aromatic flavoring known as licorice. The licorice plant is widely used as an herbal remedy  $\frac{[128]}{}$  and in skin-care products  $\frac{[129]}{}$ . Numerous articles have reported pharmaceutical therapeutic properties, such as antifungal, antibacterial, antiviral, and antioxidant properties, but these are only some of the possible therapeutic properties  $\frac{[130]}{}$ . Licorice root extract has been used for years as an effective medication, especially in gastric ulcers. The main biologically active components of licorice include terpenoids such as triterpene saponins, chalcones, and glycyrrhizin, flavonoids, and isoflavonoids, which are responsible for the observed activities  $\frac{[128]}{}$ .

Some studies have demonstrated antiulcer effects of G. glabra in the healing of gastric [131], oral [132], and colitis mucosal ulcers [133], as well as corneal neovascularization [134], but only a few studies have reported its wound-healing activity on full-thickness dermal wounds [135][136][137][138][139]. Previous research has shown that antioxidant and anti-inflammatory compounds of G. glabra, such as triterpenes and flavonoids, reduce free radicals and pus in the wound area, aiding in wound healing [136][137].

Zangeneh et al. [138] discovered that 3% *G. glabra* aqueous extract ointment significantly accelerates wound-healing activity in rats by reducing the wound area, total cells, macrophage, lymphocyte, and neutrophil levels while increasing wound contracture, fibrocyte, hexuronic acid, and hydroxyproline levels when compared to the basal ointment and control groups.

According to the findings of Siriwattanasatorn et al.  $^{[139]}$ , ethanolic extract of *G. glabra* promoted proliferation and accelerated wound closure by inhibiting superoxide anion and nitric oxide production with IC<sub>50</sub> values of 28.62  $\pm$  1.91 and 46.35  $\pm$  0.43  $\mu$ g/mL, respectively. The bioactive compound, glycyrrhizin, exhibited antioxidant activities with an IC<sub>50</sub> value of 40.85  $\pm$  2.30  $\mu$ g/mL but had no activity against nitric oxide production inhibition  $^{[139]}$ .

More recently, Hanafy et al. [140] studied the effect of *G. glabra* extract on full-thickness wound healing in a Guinea pig model. In this study, it was found that the topical application of a cream containing 5–10% *w/w G. glabra* extract accelerated wound healing with statistically significant differences in the re-epithelialization of treated wounds, which were likely due to the presence of flavonoids [140].

### 9. Malva sylvestris

The use of *Malva sylvestris*, a species of the mallow genus *Malva* in the family, has been documented since long ago. *M. sylvestris* is recommended for acne and skincare, as an antiseptic and emollient [141][142][143], and as an antimicrobial and anti-inflammatory agent for burn and cut wound healing [144][145][146]. The healing capabilities of this plant relate to the mucilage and flavonoids found in the leaves and flowers [147]. Indeed, *M. sylvestris* flowers extract contains anthocyanin, malvidin, flavones, flavonols, malvin, malvaline, niacin, and folic acid, which are responsible for their pharmacological and biological activities [146][147][148].

Afshar et al. [149] investigated the wound-healing ability of M. sylvestris using a wound mouse model. The researchers reported that mice treated with silver sulfadiazine were less capable of wound healing than mice treated with 1% M. sylvestris extract, where increased collagen synthesis was observed [149]. Another in vitro study investigated the effect of M. sylvestris or silver sulfadiazine on wound healing using a second-degree burn model [150]. The results clearly demonstrated that both 5% and 10% M. sylvestris cream was superior to sulfadiazine in terms of reducing the time required for complete wound healing [150]. Nevertheless, the molecular mechanism underlying these actions requires further investigation.

Recent research has examined the effect of *M. sylvestris* extract inserted into a novel polyurethane (PU)-based nanofibers used as dressing for diabetic wounds [151]. When animal wounds treated with *M. sylvestris* nanofibers were compared to control groups, the wound area was shown to be significantly decreased. A polymer blend containing 15% *w/w* herbal extract of *M. sylvestris* was used to monitor the herbal compound's progressive release over an 85-h period. In comparison to the control group, treatments with extract-loaded wound dressings were significantly more effective at reducing acute and chronic inflammation, increasing collagen deposition and neovascularization, and demonstrated acceptable antibacterial activity against *Staphylococcus aureus* and *Escherichia coli* [151].

## 10. Plantago

*Plantago L.* (*Plantaginaceae*) is a worldwide genus including approximately 260 species of annual and perennial herbs and shrubs [152]. Phytochemical studies have shown that the *Plantago* genus contains a great number of compounds such as acteosides, iridoids, glucosides, phenylethanoid glycosides, flavonoids, tannins, triterpenes, saponins, sterols, and phenyl carboxylic acid derivatives [153]. The genus *Plantago* is widely used in folk medicine as an anti-inflammatory agent for various diseases, including the treatment of wounds [154]. Fresh plantago leaves were shown to have antibacterial effects and to be beneficial for wound healing. Crushed leaves are used to treat chronic wounds, abscesses, and acne [155]. Not many studies have investigated the effects of *Plantago* species on wound healing.

An in vitro investigation of human oral epithelial cell lines H400 indicated the anti-inflammatory properties of P. major are due to the synergistic actions of polyphenols and water-soluble substances (i.e., polysaccharides) [155]. Genc et al. investigated the wound healing ability of P. subulata in vitro using fibroblasts (L929 cell line), whereas the anti-inflammatory activity was evaluated using macrophages (RAW 264.7 cell line) [156]. Treatments with different concentrations (200, 400, 800, and 1000  $\mu$ g/mL) of the aqueous fraction of P. subulata methanolic extract (PHE) exerted protection against oxidative stress in fibroblasts, although not in correlation with PHE concentrations. Preincubation of macrophages with 50 and 200  $\mu$ g/mL PHE reduced the production of NO, PGE2, and TNF- $\alpha$  induced by lipopolysaccharide [156]. Acteoside was thought to be the chemical responsible for these effects [156].

Recently, a clinical investigation assessed the efficacy of a gel comprising *A. vera* and *P. major* in treating diabetic foot ulcers [157]. At the conclusion of the randomized open-label controlled trial, patients treated with a topical gel containing 10% plantago hydroalcoholic extract applied to the wound once daily for two weeks showed a significant reduction in wound size when compared to the control group [157]. The application of *P. major* topical gel accelerates the healing of

diabetic foot ulcers and pressure ulcers by reducing the wound's erythema. Additionally, the proportion of patients who completed the wound-healing process was higher in those who received *P. major* dressing [157]. However, this study was limited by its relatively small sample size, absence of placebo and blinding (due to technical issues), short duration of intervention, and use of novel dressings in the control groups (because of different characteristics of ulcers necessitating this). These results showed that Plantago and its compounds may be good candidates for future drug studies.

### 11. Salvia officinalis

Salvia officinalisL. (also called **sage**, garden sage, or common sage) is a perennial evergreen subshrub with woody stems, grayish leaves, and blue to purplish flowers  $\frac{[158][159]}{[159]}$ . Salvia is an important genus consisting of approximately 900 species in the family Lamiaceae (formerly Labiatae). Salvia species are native to the Mediterranean region and are cultivated worldwide for use in folk medicine and culinary purposes. The name Salvia is derived from "Salvere", a Latin word meaning "to feel well and healthy"  $\frac{[160]}{[160]}$ . The genus Salvia has a wide range of medicinal uses, and it is traditionally used to treat more than sixty health illnesses  $\frac{[160][161]}{[160]}$ .

According to the analysis of essential oils taken from aerial sections of the plant, these species contain approximately 120 phytocomponents [161][162][163][164][165], including fatty acids, carbohydrates, alkaloids, glycoside saponins, terpenes, phenolic acids, flavonoids, and polyacetylenes [161][164]. The biological actions of oil and aqueous extracts include anti-inflammatory, antibacterial, and antioxidant activity [165][166][167]. The terpenoids, e.g., 1,8-cineole, oxygenated sesquiterpene viridiflorol, camphor, nonacosane, and pentacosane, were identified as the major constituents of the essential oil [168]. Numerous studies have demonstrated that the composition of sage essential oil undergoes considerable changes in relation to seasonal variation, genetic diversity, defense in plant sections, and developmental stage differences [167][169]

Numerous investigations have established the influence of *S. officinalis* on wound healing  $\frac{[169][170][171][172][173]}{[172][173]}$ . Farahpour et al.  $\frac{[171]}{[171]}$  demonstrated that 2% and 4% (*w/w*) topical ointments of sage oil on mouse full-thickness cutaneous wounds dramatically decreased pro-inflammatory cytokines and the overall bacteria count when compared to a control group  $\frac{[171]}{[171]}$ . The data in this study indicated that this treatment suppressed the pro-inflammatory response by decreasing mRNA expression levels of IL-6, IL-1, and TNF-alpha, and promoted fibroblast proliferation by increasing cyclin-D1 expression  $\frac{[171]}{[171]}$ . Furthermore, *S. officinalis* has been shown to stimulate VEGF transcription and secretion and FGF-2 expression at the wound edge and in basal keratinocytes  $\frac{[171]}{[171]}$ . In in vivo experiments using a wound model in rats, topical application of 1, 3, and 5% *S. officinalis* hydroethanolic leaf extracts significantly increased, particularly at higher doses, the percentage of wound contraction, the re-epithelialization period, the breaking strength ratio, and upregulated hydroxyproline content compared to the control group  $\frac{[171]}{[171]}$ . Additionally, *S. officinalis* significantly increased new vessel formation and fibroblast distribution  $\frac{[172]}{[171]}$ .

Another study has looked at the effects of ethanol leaf extracts of two different *Salvia* species, *Salvia kronenburgii* (SK) and *Salvia euphratica* (SE), at two different concentrations (0.5% and 1% (w/w) on incision and excision wound models in diabetic rats <sup>[173]</sup>. In addition, these extracts were tested for their ability to fight off bacteria, viruses, and fungi. They worked best against *Staphylococcus aureus* and *Bacillus subtilis*, as well as *Escherichia coli*, *Aeromonas hydrophila*, and *Mycobacterium tuberculosis*. <sup>[173]</sup>. Moreover, both concentrations of SK and SE ointments were found to be as effective as the reference drug, showing significant wound closure ratios with a range of 96.9–99.9% in excision and incision wound models after the 14th day <sup>[173]</sup>. These results suggest that Salvia species, which are common in nature, may be promising antimicrobial and wound-healing agents for the treatment of infectious diseases as an alternative to synthetic drugs with high costs and side effects.

## 12. Rosmarinus officinalis (Syn. Salvia rosmarinus)

Rosmarinus officinalis L., commonly known as rosemary, is a member of the *Lamiaceae* family. Recent evolutionary research has reported that the genus *Rosmarinus* has merged with the genus *Salvia*. On this basis, *Rosmarinus officinalis* was changed into *Salvia Rosmarinus* [174]. Rosemary is a fragrant, needle-like-leaved plant that is widely cultivated worldwide and used in folk medicine. Due to the presence of carnosol/carnosic and ursolic acids, rosemary has therapeutic properties and is used in the pharmaceutical and cosmetics industries, primarily for its antioxidant and anti-inflammatory properties [175]. Other uses, including treatments for wound healing, skin cancer, and mycoses, have also been investigated [176][177][178]. Potential applications in the treatment of non-pathological skin conditions, such as ultraviolet damage and aging, have been shown [177][178][179][180].

Rosemary contains a large number of secondary metabolites that have been identified using ultra- and high-performance liquid chromatography and gas chromatography to reveal significant amounts of phenolic compounds (diterpenoids and flavonoids) and volatile chemicals [181][182]. The flavonoids discovered in rosemary (eriocitrin, luteolin 3'-O-D-glucuronide, hesperidin, diosmin, isoscutellarein 7-O-glucoside, hispidulin 7-O-glucoside, and genkwanin) were found in various parts of the plant over time [182].

In an in vivo study, the group treated with rosemary essential oils demonstrated significant improvements in healing, angiogenesis, and granulation tissue formation when compared to the control group [179]. Another study reported that diabetic and non-diabetic rats treated topically with 10% *R. officinalis* oil experienced rapid wound healing [183]. Using a rat excision wound model, a recent in vivo study examined the wound healing potential of three chitosan-based topical formulations containing either tea tree essential oil, rosemary essential oil, or a combination of the two oils [184]. When compared to groups treated with individual essential oils or the control group, topical application of a chitosan-based product containing a blend of both oils increased wound shrinkage significantly. Additionally, histopathological examination revealed complete re-epithelialization and activation of hair follicles when the two essential oils were combined. Monoterpene content of essential oils contributed significantly to their antioxidant and wound-healing properties [184]. In conclusion, rosemary extract has a great deal of therapeutic potential and could help wounds heal at different stages.

#### References

- Cavalcanti, A.M.; Baggio, C.H.; Freitas, C.S.; Rieck, L.; de Sousa, R.S.; Da Silva-Santos, J.E.; Mesia-Vela, S.;
  Marques, M.C.A. Safety and Antiulcer Efficacy Studies of Achillea millefolium L. after Chronic Treatment in Wistar Rats.
  J. Ethnopharmacol. 2006, 107, 277–284.
- 2. Benedek, B.; Kopp, B. Achillea millefolium L. s.l. Revisited: Recent Findings Confirm the Traditional Use. Wien. Med. Wochenschr. 2007, 157, 312–314.
- 3. Chávez-Silva, F.; Cerón-Romero, L.; Arias-Durán, L.; Navarrete-Vázquez, G.; Almanza-Pérez, J.; Román-Ramos, R.; Ramírez-Ávila, G.; Perea-Arango, I.; Villalobos-Molina, R.; Estrada-Soto, S. Antidiabetic Effect of Achillea millefollium through Multitarget Interactions: α-Glucosidases Inhibition, Insulin Sensitization and Insulin Secretagogue Activities. J. Ethnopharmacol. 2018, 212, 1–7.
- 4. Ali, S.I.; Gopalakrishnan, B.; Venkatesalu, V. Pharmacognosy, Phytochemistry and Pharmacological Properties of Achillea millefolium L.: A Review. Phytother. Res. 2017, 31, 1140–1161.
- 5. Başer, K.H.; Demirci, B.; Demirci, F.; Koçak, S.; Akıncı, Ç.; Malyer, H.; Güleryüz, G. Composition and Antimicrobial Activity of the Essential Oil of Achillea multifida. Planta Med. 2002, 68, 941–943.
- 6. Benedek, B.; Rothwangl-Wiltschnigg, K.; Rozema, E.; Gjoncaj, N.; Reznicek, G.; Jurenitsch, J.; Kopp, B.; Glasl, S. Yarrow (Achillea millefolium L. s.l.): Pharmaceutical Quality of Commercial Samples. Pharm.-Int. J. Pharm. Sci. 2008, 63, 23–26.
- 7. Falk, A.J.; Smolenski, S.J.; Bauer, L.; Bell, C.L. Isolation and Identification of Three New Flavones from Achillea millefolium L. J. Pharm. Sci. 1975, 64, 1838–1842.
- 8. Nemeth, E. Essential Oil Composition of Species in the Genus Achillea. J. Essent. Oil Res. 2005, 17, 501–512.
- 9. Nemeth, E.; Bernath, J. Biological Activities of Yarrow Species (Achillea Spp.). Curr. Pharm. Des. 2008, 14, 3151–3167.
- 10. Bakkali, F.; Averbeck, S.; Averbeck, D.; Idaomar, M. Biological Effects of Essential Oils—A Review. Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc. 2008, 46, 446–475.
- 11. Vitalini, S.; Beretta, G.; Iriti, M.; Orsenigo, S.; Basilico, N.; Dall'Acqua, S.; Iorizzi, M.; Fico, G. Phenolic Compounds from Achillea millefolium L. and Their Bioactivity. Acta Biochim. Pol. 2011, 58, 203–209.
- 12. De Souza, P.; Gasparotto, A.; Crestani, S.; Stefanello, M.É.A.; Marques, M.C.A.; da Silva-Santos, J.E.; Kassuya, C.A.L. Hypotensive Mechanism of the Extracts and Artemetin Isolated from Achillea millefolium L. (Asteraceae) in Rats. Phytomedicine 2011, 18, 819–825.
- 13. Csupor-Löffler, B.; Hajdú, Z.; Zupkó, I.; Réthy, B.; Falkay, G.; Forgo, P.; Hohmann, J. Antiproliferative Effect of Flavonoids and Sesquiterpenoids from Achillea millefolium s.l. on Cultured Human Tumour Cell Lines. Phytother. Res. 2009, 23, 672–676.
- 14. Lemmens-Gruber, R.; Marchart, E.; Rawnduzi, P.; Engel, N.; Benedek, B.; Kopp, B. Investigation of the Spasmolytic Activity of the Flavonoid Fraction of Achillea millefolium s.l. on Isolated Guinea-Pig Ilea. Arzneimittelforschung 2006, 56, 582–588.

- 15. Khan, A.; Gilani, A.H. Blood Pressure Lowering, Cardiovascular Inhibitory and Bronchodilatory Actions of Achillea millefolium. Phytother. Res. 2011, 25, 577–583.
- 16. Arias-Durán, L.; Estrada-Soto, S.; Hernández-Morales, M.; Chávez-Silva, F.; Navarrete-Vázquez, G.; León-Rivera, I.; Perea-Arango, I.; Villalobos-Molina, R.; Ibarra-Barajas, M. Tracheal Relaxation through Calcium Channel Blockade of Achillea millefolium Hexanic Extract and Its Main Bioactive Compounds. J. Ethnopharmacol. 2020, 253, 112643.
- 17. Tadić, V.; Arsić, I.; Zvezdanović, J.; Zugić, A.; Cvetković, D.; Pavkov, S. The Estimation of the Traditionally Used Yarrow (Achillea millefolium L. Asteraceae) Oil Extracts with Anti-Inflamatory Potential in Topical Application. J. Ethnopharmacol. 2017, 199, 138–148.
- 18. Dorjsembe, B.; Lee, H.J.; Kim, M.; Dulamjav, B.; Jigjid, T.; Nho, C.W. Achillea Asiatica Extract and Its Active Compounds Induce Cutaneous Wound Healing. J. Ethnopharmacol. 2017, 206, 306–314.
- 19. Long, V. Aloe vera in Dermatology—The Plant of Immortality. JAMA Dermatol. 2016, 152, 1364.
- 20. Zeng, W.M.; Parus, A.; Barnes, C.W.; Hiro, M.E.; Robson, M.C.; Payne, W.G. Aloe vera—Mechanisms of Action, Uses, and Potential Uses in Plastic Surgery and Wound Healing. Surg. Sci. 2020, 11, 312.
- 21. Sánchez-Machado, D.I.; López-Cervantes, J.; Sendón, R.; Sanches-Silva, A. Aloe vera: Ancient Knowledge with New Frontiers. Trends Food Sci. Technol. 2017, 61, 94–102.
- 22. Salehi, B.; Albayrak, S.; Antolak, H.; Kręgiel, D.; Pawlikowska, E.; Sharifi-Rad, M.; Uprety, Y.; Tsouh Fokou, P.V.; Yousef, Z.; Amiruddin Zakaria, Z.; et al. Aloe Genus Plants: From Farm to Food Applications and Phytopharmacotherapy. Int. J. Mol. Sci. 2018, 19, 2843.
- 23. Gallagher, J.; Gray, M. Is Aloe vera Effective for Healing Chronic Wounds? J. Wound Ostomy Cont. Nurs. Off. Publ. Wound Ostomy Cont. Nurses Soc. 2003, 30, 68–71.
- 24. Roberts, D.B.; Travis, E.L. Acemannan-Containing Wound Dressing Gel Reduces Radiation-Induced Skin Reactions in C3H Mice. Int. J. Radiat. Oncol. Biol. Phys. 1995, 32, 1047–1052.
- 25. Zhang, L.; Tizard, I.R. Activation of a Mouse Macrophage Cell Line by Acemannan: The Major Carbohydrate Fraction from Aloe vera Gel. Immunopharmacology 1996, 35, 119–128.
- 26. Oryan, A.; Mohammadalipour, A.; Moshiri, A.; Tabandeh, M.R. Topical Application of Aloe vera Accelerated Wound Healing, Modeling, and Remodeling: An Experimental Study. Ann. Plast. Surg. 2016, 77, 37–46.
- 27. Hormozi, M.; Assaei, R.; Boroujeni, M.B. The Effect of Aloe vera on the Expression of Wound Healing Factors (TGFβ1 and BFGF) in Mouse Embryonic Fibroblast Cell: In Vitro Study. Biomed. Pharmacother. Biomed. Pharmacother. 2017, 88, 610–616.
- 28. Wahedi, H.M.; Jeong, M.; Chae, J.K.; Do, S.G.; Yoon, H.; Kim, S.Y. Aloesin from Aloe vera Accelerates Skin Wound Healing by Modulating MAPK/Rho and Smad Signaling Pathways in Vitro and in vivo. Phytomed. Int. J. Phytother. Phytopharm. 2017, 28, 19–26.
- 29. Yamao, M.; Naoki, H.; Kunida, K.; Aoki, K.; Matsuda, M.; Ishii, S. Distinct Predictive Performance of Rac1 and Cdc42 in Cell Migration. Sci. Rep. 2015, 5, 17527.
- 30. De Oliveira, A.C.L.; Tabrez, S.; Shakil, S.; Khan, M.I.; Asghar, M.N.; Matias, B.D.; da Silva Batista, J.M.A.; Rosal, M.M.; de Lima, M.M.D.F.; Gomes, S.R.F. Mutagenic, Antioxidant and Wound Healing Properties of Aloe vera. J. Ethnopharmacol. 2018, 227, 191–197.
- 31. Oliveira, R.N.; Mancini, M.C.; de Oliveira, F.C.S.; Passos, T.M.; Quilty, B.; Thiré, R.M.d.S.M.; McGuinness, G.B. FTIR Analysis and Quantification of Phenols and Flavonoids of Five Commercially Available Plants Extracts Used in Wound Healing. Matér. Rio Jan. 2016, 21, 767–779.
- 32. Teplicki, E.; Ma, Q.; Castillo, D.E.; Zarei, M.; Hustad, A.P.; Chen, J.; Li, J. The Effects of Aloe vera on Wound Healing in Cell Proliferation, Migration, and Viability. Wounds Compend. Clin. Res. Pract. 2018, 30, 263–268.
- 33. Rossiter, H.; Barresi, C.; Pammer, J.; Rendl, M.; Haigh, J.; Wagner, E.F.; Tschachler, E. Loss of Vascular Endothelial Growth Factor a Activity in Murine Epidermal Keratinocytes Delays Wound Healing and Inhibits Tumor Formation. Cancer Res. 2004, 64, 3508–3516.
- 34. Chantarawaratit, P.; Sangvanich, P.; Banlunara, W.; Soontornvipart, K.; Thunyakitpisal, P. Acemannan Sponges Stimulate Alveolar Bone, Cementum and Periodontal Ligament Regeneration in a Canine Class II Furcation Defect Model. J. Periodontal Res. 2014, 49, 164–178.
- 35. Xing, W.; Guo, W.; Zou, C.-H.; Fu, T.-T.; Li, X.-Y.; Zhu, M.; Qi, J.-H.; Song, J.; Dong, C.-H.; Li, Z.; et al. Acemannan Accelerates Cell Proliferation and Skin Wound Healing through AKT/MTOR Signaling Pathway. J. Dermatol. Sci. 2015, 79, 101–109.

- 36. Lin, L.-X.; Wang, P.; Wang, Y.-T.; Huang, Y.; Jiang, L.; Wang, X.-M. Aloe vera and Vitis Vinifera Improve Wound Healing in an in vivo Rat Burn Wound Model. Mol. Med. Rep. 2016, 13, 1070–1076.
- 37. Wang, X.; Jia, W.; Zhao, A.; Wang, X. Anti-Influenza Agents from Plants and Traditional Chinese Medicine. Phytother. Res. PTR 2006, 20, 335–341.
- 38. Xu, D.; Pan, Y.; Chen, J. Chemical Constituents, Pharmacologic Properties, and Clinical Applications of Bletilla striata. Front. Pharmacol. 2019, 10, 1168.
- 39. Jiang, F.; Li, W.; Huang, Y.; Chen, Y.; Jin, B.; Chen, N.; Ding, Z.; Ding, X. Antioxidant, Antityrosinase and Antitumor Activity Comparison: The Potential Utilization of Fibrous Root Part of Bletilla striata (Thunb.) Reichb.f. PLoS ONE 2013, 8, e58004.
- 40. He, X.; Wang, X.; Fang, J.; Zhao, Z.; Huang, L.; Guo, H.; Zheng, X. Bletilla striata: Medicinal Uses, Phytochemistry and Pharmacological Activities. J. Ethnopharmacol. 2017, 195, 20–38.
- 41. Liu, F.Q.; Wang, Y.P.; Han, D.; Bi, Y.J.; Li, H.B.; Liu, G. Extraction of Bletilla striata Polysaccharide and Its Relative Molecular Mass Determination and Structure Study. Chin. Tradit. Pat. Med. 2013, 35, 2291–2293.
- 42. Süntar, I.; Küpeli Akkol, E.; Keles, H.; Yesilada, E.; Sarker, S.D.; Baykal, T. Comparative Evaluation of Traditional Prescriptions from Cichorium intybus L. for Wound Healing: Stepwise Isolation of an Active Component by in vivo Bioassay and Its Mode of Activity. J. Ethnopharmacol. 2012, 143, 299–309.
- 43. Diao, H.; Li, X.; Chen, J.; Luo, Y.; Chen, X.; Dong, L.; Wang, C.; Zhang, C.; Zhang, J. Bletilla striata Polysaccharide Stimulates Inducible Nitric Oxide Synthase and Proinflammatory Cytokine Expression in Macrophages. J. Biosci. Bioeng. 2008, 105, 85–89.
- 44. Luo, Y.; Diao, H.; Xia, S.; Dong, L.; Chen, J.; Zhang, J. A Physiologically Active Polysaccharide Hydrogel Promotes Wound Healing. J. Biomed. Mater. Res. A 2010, 94A, 193–204.
- 45. Yu, Y.; Tan, M.; Chen, H.; Wu, Z.; Xu, L.; Li, J.; Cao, J.; Yang, Y.; Xiao, X.; Lian, X.; et al. Non-Thermal Plasma Suppresses Bacterial Colonization on Skin Wound and Promotes Wound Healing in Mice. J. Huazhong Univ. Sci. Technol. Med. Sci. 2011, 31, 390–394.
- 46. Yue, L.; Wang, W.; Wang, Y.; Du, T.; Shen, W.; Tang, H.; Wang, Y.; Yin, H. Bletilla striata Polysaccharide Inhibits Angiotensin II-Induced ROS and Inflammation via NOX4 and TLR2 Pathways. Int. J. Biol. Macromol. 2016, 89, 376–388.
- 47. Wang, W.; Meng, H. Cytotoxic, Anti-Inflammatory and Hemostatic Spirostane-Steroidal Saponins from the Ethanol Extract of the Roots of Bletilla striata. Fitoterapia 2015, 101, 12–18.
- 48. Hung, H.-Y.; Wu, T.-S. Recent Progress on the Traditional Chinese Medicines That Regulate the Blood. J. Food Drug Anal. 2016, 24, 221–238.
- 49. Zhao, F.; Yang, X.; Xu, D.; Dong, L.; Li, J.; Wang, Y.; Liao, S. Hemostatic effect and mechanism of a non-polysaccharide fraction of Bletilla striata. Chin. Pharmacol. Bull. 2016, 1121–1126.
- 50. Ding, L.; Shan, X.; Zhao, X.; Zha, H.; Chen, X.; Wang, J.; Cai, C.; Wang, X.; Li, G.; Hao, J.; et al. Spongy Bilayer Dressing Composed of Chitosan-Ag Nanoparticles and Chitosan-Bletilla striata Polysaccharide for Wound Healing Applications. Carbohydr. Polym. 2017, 157, 1538–1547.
- 51. Tsai, C.C.; Huang, R.N.; Sung, H.W.; Liang, H.C. In Vitro Evaluation of the Genotoxicity of a Naturally Occurring Crosslinking Agent (Genipin) for Biologic Tissue Fixation. J. Biomed. Mater. Res. 2000, 52, 58–65.
- 52. Song, Y.; Zeng, R.; Hu, L.; Maffucci, K.G.; Ren, X.; Qu, Y. In vivo Wound Healing and in Vitro Antioxidant Activities of Bletilla striata Phenolic Extracts. Biomed. Pharmacother. Biomed. Pharmacother. 2017, 93, 451–461.
- 53. Cheng, W.; Zhou, F.; Zhu, B.; Ding, X.; Lu, J.; Qian, C.; Ye, X.; Ding, Z. Characterization and Evaluation of Bletilla striata Polysaccharide/Ethanol Extract Composite Multifunctional Sponges. Mater. Des. 2021, 206, 109806.
- 54. Basch, E.; Bent, S.; Foppa, I.; Haskmi, S.; Kroll, D.; Mele, M.; Szapary, P.; Ulbricht, C.; Vora, M.; Yong, S. Marigold (Calendula officinalis L.). J. Herb. Pharmacother. 2006, 6, 135–159.
- 55. John, R.; Jan, N. Calendula officinalis—An Important Medicinal Plant with Potential Biological Properties. Proc. Indian Natl. Sci. Acad. 2017, 93.
- 56. Leach, M.J. Calendula officinalis and Wound Healing: A Systematic Review. Wounds Compend. Clin. Res. Pract. 2008, 20, 236–243.
- 57. Muley, B.P.; Khadabadi, S.S.; Banarase, N.B. Phytochemical Constituents and Pharmacological Activities of Calendula officinalis Linn (Asteraceae): A Review. Trop. J. Pharm. Res. 2009, 8.
- 58. Arora, D.; Rani, A.; Sharma, A. A Review on Phytochemistry and Ethnopharmacological Aspects of Genus Calendula. Pharmacogn. Rev. 2013, 7, 179–187.

- 59. Shafeie, N.; Naini, A.T.; Jahromi, H.K. Comparison of Different Concentrations of Calendula officinalis Gel on Cutaneous Wound Healing. Biomed. Pharmacol. J. 2015, 8, 979–992.
- 60. Givol, O.; Kornhaber, R.; Visentin, D.; Cleary, M.; Haik, J.; Harats, M. A Systematic Review of Calendula officinalis Extract for Wound Healing. Wound Repair Regen. 2019, 27, 548–561.
- 61. Eghdampour, F.; Jahdie, F.; Kheyrkhah, M.; Taghizadeh, M.; Naghizadeh, S.; Hagani, H. The Impact of Aloe Vera and Calendula on Perineal Healing after Episiotomy in Primiparous Women: A Randomized Clinical Trial. J. Caring Sci. 2013, 2, 279–286.
- 62. Fronza, M.; Heinzmann, B.; Hamburger, M.; Laufer, S.; Merfort, I. Determination of the Wound Healing Effect of Calendula Extracts Using the Scratch Assay with 3T3 Fibroblasts. J. Ethnopharmacol. 2009, 126, 463–467.
- 63. Fonseca, Y.M.; Catini, C.D.; Vicentini, F.T.M.C.; Nomizo, A.; Gerlach, R.F.; Fonseca, M.J.V. Protective Effect of Calendula officinalis Extract against UVB-Induced Oxidative Stress in Skin: Evaluation of Reduced Glutathione Levels and Matrix Metalloproteinase Secretion. J. Ethnopharmacol. 2010, 127, 596–601.
- 64. Dinda, M.; Dasgupta, U.; Singh, N.; Bhattacharyya, D.; Karmakar, P. Pl3K-Mediated Proliferation of Fibroblasts by Calendula officinalis Tincture: Implication in Wound Healing. Phytother. Res. PTR 2015, 29, 607–616.
- 65. Dinda, M.; Mazumdar, S.; Das, S.; Ganguly, D.; Dasgupta, U.B.; Dutta, A.; Jana, K.; Karmakar, P. The Water Fraction of Calendula Officinalis Hydroethanol Extract Stimulates In Vitro and In vivo Proliferation of Dermal Fibroblasts in Wound Healing. Phytother. Res. PTR 2016, 30, 1696–1707.
- 66. Nicolaus, C.; Junghanns, S.; Hartmann, A.; Murillo, R.; Ganzera, M.; Merfort, I. In Vitro Studies to Evaluate the Wound Healing Properties of Calendula officinalis Extracts. J. Ethnopharmacol. 2017, 196, 94–103.
- 67. Parente, L.M.L.; de Lino Júnior, R.S.; Tresvenzol, L.M.F.; Vinaud, M.C.; de Paula, J.R.; Paulo, N.M. Wound Healing and Anti-Inflammatory Effect in Animal Models of Calendula officinalis L. Growing in Brazil. Evid. Based Complement. Alternat. Med. 2012, 2012, e375671.
- 68. Kharat, Z.; Amiri Goushki, M.; Sarvian, N.; Asad, S.; Dehghan, M.M.; Kabiri, M. Chitosan/PEO Nanofibers Containing Calendula Officinalis Extract: Preparation, Characterization, in Vitro and in vivo Evaluation for Wound Healing Applications. Int. J. Pharm. 2021, 609, 121132.
- 69. Ferreira, P.M.P.; Costa-Lotufo, L.V.; Moraes, M.O.; Barros, F.W.A.; Martins, A.M.A.; Cavalheiro, A.J.; Bolzani, V.S.; Santos, A.G.; Pessoa, C. Folk Uses and Pharmacological Properties of Casearia sylvestris: A Medicinal Review. An. Acad. Bras. Ciênc. 2011, 83, 1373–1384.
- 70. Bueno, P.C.P.; Passareli, F.; Anhesine, N.B.; Torres, R.B.; Cavalheiro, A.J. Flavonoids from Casearia sylvestris Swartz Variety Lingua (Salicaceae). Biochem. Syst. Ecol. 2016, 68, 23–26.
- 71. Albano, M.N.; da Silveira, M.R.; Danielski, L.G.; Florentino, D.; Petronilho, F.; Piovezan, A.P. Anti-Inflammatory and Antioxidant Properties of Hydroalcoholic Crude Extract from Casearia sylvestris Sw. (Salicaceae). J. Ethnopharmacol. 2013, 147, 612–617.
- 72. Pierri, E.G.; Castro, R.C.; Vizioli, E.O.; Ferreira, C.M.R.; Cavalheiro, A.J.; Tininis, A.G.; Chin, C.M.; Santos, A.G. Anti-Inflammatory Action of Ethanolic Extract and Clerodane diterpenes from Casearia sylvestris. Rev. Bras. Farmacogn. 2017, 27, 495–501.
- 73. dos Santos, A.G.; Ferreira, P.M.P.; Vieira Júnior, G.M.; Perez, C.C.; Gomes Tininis, A.; Silva, G.H.; da Bolzani, V.S.; Costa-Lotufo, L.V.; do Pessoa, C.O.; Cavalheiro, A.J. Casearin X, Its Degradation Product and Other Clerodane diterpenes from Leaves of Casearia sylvestris: Evaluation of Cytotoxicity against Normal and Tumor Human Cells. Chem. Biodivers. 2010, 7, 205–215.
- 74. Basile, A.C.; Sertié, J.A.A.; Panizza, S.; Oshiro, T.T.; Azzolini, C.A. Pharmacological Assay of Casearia sylvestris. I: Preventive Anti-Ulcer Activity and Toxicity of the Leaf Crude Extract. J. Ethnopharmacol. 1990, 30, 185–197.
- 75. Maistro, E.L.; Carvalho, J.C.T.; Mantovani, M.S. Evaluation of the Genotoxic Potential of the Casearia sylvestris Extract on HTC and V79 Cells by the Comet Assay. Toxicol. In Vitro 2004, 18, 337–342.
- 76. Heymanns, A.C.; Albano, M.N.; da Silveira, M.R.; Muller, S.D.; Petronilho, F.C.; Gainski, L.D.; Cargnin-Ferreira, E.; Piovezan, A.P. Macroscopic, Biochemical and Hystological Evaluation of Topical Anti-Inflammatory Activity of Casearia sylvestris (Flacourtiaceae) in Mice. J. Ethnopharmacol. 2021, 264, 113139.
- 77. de Campos, E.P.; Trombini, L.N.; Rodrigues, R.; Portella, D.L.; Werner, A.C.; Ferraz, M.C.; de Oliveira, R.V.M.; Cogo, J.C.; Oshima-Franco, Y.; Aranha, N.; et al. Healing Activity of Casearia sylvestris Sw. in Second-Degree Scald Burns in Rodents. BMC Res. Notes 2015, 8, 269.
- 78. Lipinski, L.C.; de Wouk, A.F.P.; da Silva, N.L.; Perotto, D.; Ollhoff, R.D. Effects of 3 Topical Plant Extracts on Wound Healing in Beef Cattle. Afr. J. Tradit. Complement. Altern. Med. AJTCAM 2012, 9, 542–547.

- 79. Carvalho, F.A.; Uchina, H.S.; Borges, F.A.; Oyafuso, M.H.; Herculano, R.D.; Gremião, M.P.D.; Santos, A.G. Natural Membranes of Hevea Brasiliensis Latex as Delivery System for Casearia sylvestris Leaf Components. Rev. Bras. Farmacogn. 2018, 28, 102–110.
- 80. Kumar, R.; Singh, V.; Devi, K.; Sharma, M.; Singh, M.K.; Ahuja, P.S. State of Art of Saffron (Crocus sativus L.) Agronomy: A Comprehensive Review. Food Rev. Int. 2008, 25, 44–85.
- 81. Christodoulou, E.; Kadoglou, N.P.E.; Kostomitsopoulos, N.; Valsami, G. Saffron: A Natural Product with Potential Pharmaceutical Applications. J. Pharm. Pharmacol. 2015, 67, 1634–1649.
- 82. Abu-Izneid, T.; Rauf, A.; Khalil, A.A.; Olatunde, A.; Khalid, A.; Alhumaydhi, F.A.; Aljohani, A.S.M.; Sahab Uddin, M.; Heydari, M.; Khayrullin, M.; et al. Nutritional and Health Beneficial Properties of Saffron (Crocus sativus L): A Comprehensive Review. Crit. Rev. Food Sci. Nutr. 2020, 62, 2683–2706.
- 83. Alonso, G.L.; Zalacain, A.; Carmona, M. Saffron. In Handbook of Herbs and Spices; Elsevier: Amsterdam, The Netherlands, 2012; pp. 469–498.
- 84. Festuccia, C.; Mancini, A.; Gravina, G.L.; Scarsella, L.; Llorens, S.; Alonso, G.L.; Tatone, C.; Di Cesare, E.; Jannini, E.A.; Lenzi, A. Antitumor Effects of Saffron-Derived Carotenoids in Prostate Cancer Cell Models. BioMed Res. Int. 2014, 2014.
- 85. Colapietro, A.; Mancini, A.; D'Alessandro, A.M.; Festuccia, C. Crocetin and Crocin from Saffron in Cancer Chemotherapy and Chemoprevention. Anti-Cancer Agents Med. Chem. Former. Curr. Med. Chem.-Anti-Cancer Agents 2019, 19, 38–47.
- 86. Del-Angel, D.S.; Martínez, N.L.H.; Cruz, M.E.G.; Urrutia, E.C.; Riverón-Negrete, L.; Abdullaev, F. Saffron extract ameliorates oxidative damage and mitochondrial dysfunction in the rat brain. Acta Hortic. 2007, 739, 359–366.
- 87. Cerdá-Bernad, D.; Valero-Cases, E.; Pastor, J.-J.; Frutos, M.J. Saffron Bioactives Crocin, Crocetin and Safranal: Effect on Oxidative Stress and Mechanisms of Action. Crit. Rev. Food Sci. Nutr. 2020, 62, 3232–3249.
- 88. Nanda, S.; Madan, K. The Role of Safranal and Saffron Stigma Extracts in Oxidative Stress, Diseases and Photoaging: A Systematic Review. Heliyon 2021, 7, e06117.
- 89. Li, S.; Liu, X.; Lei, J.; Yang, J.; Tian, P.; Gao, Y. Crocin Protects Podocytes Against Oxidative Stress and Inflammation Induced by High Glucose Through Inhibition of NF-KB. Cell. Physiol. Biochem. Int. J. Exp. Cell. Physiol. Biochem. Pharmacol. 2017, 42, 1481–1492.
- 90. Zeka, K.; Ruparelia, K.C.; Sansone, C.; Macchiarelli, G.; Continenza, M.A.; Arroo, R.R.J. New Hydrogels Enriched with Antioxidants from Saffron Crocus Can Find Applications in Wound Treatment and/or Beautification. Skin Pharmacol. Physiol. 2018, 31, 95–98.
- 91. Khorasani, G.; Hosseinimehr, S.J.; Zamani, P.; Ghasemi, M.; Ahmadi, A. The Effect of Saffron (Crocus sativus) Extract for Healing of Second-Degree Burn Wounds in Rats. Keio J. Med. 2008, 57, 190–195.
- 92. Fagot, D.; Pham, D.M.; Laboureau, J.; Planel, E.; Guerin, L.; Nègre, C.; Donovan, M.; Bernard, B.A. Crocin, a Natural Molecule with Potentially Beneficial Effects against Skin Ageing. Int. J. Cosmet. Sci. 2018, 40, 388–400.
- 93. Krutmann, J.; Bouloc, A.; Sore, G.; Bernard, B.A.; Passeron, T. The Skin Aging Exposome. J. Dermatol. Sci. 2017, 85, 152–161.
- 94. Alemzadeh, E.; Oryan, A. Effectiveness of a Crocus sativus Extract on Burn Wounds in Rats. Planta Med. 2018, 84, 1191–1200.
- 95. Gigliobianco, M.R.; Cortese, M.; Peregrina, D.V.; Villa, C.; Lupidi, G.; Pruccoli, L.; Angeloni, C.; Tarozzi, A.; Censi, R.; Di Martino, P. Development of New Extracts of Crocus sativus L. By-Product from Two Different Italian Regions as New Potential Active Ingredient in Cosmetic Formulations. Cosmetics 2021, 8, 51.
- 96. Chichiriccò, G.; Ferrante, C.; Menghini, L.; Recinella, L.; Leone, S.; Chiavaroli, A.; Brunetti, L.; Di Simone, S.; Ronci, M.; Piccone, P.; et al. Crocus sativus By-Products as Sources of Bioactive Extracts: Pharmacological and Toxicological Focus on Anthers. Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc. 2019, 126, 7–14.
- 97. Hassan, A. Curcuma Longa, Turmeric: A Monograph. Aust. J. Med. Herbal. 2006, 18, 66-76.
- 98. Memarzia, A.; Khazdair, M.R.; Behrouz, S.; Gholamnezhad, Z.; Jafarnezhad, M.; Saadat, S.; Boskabady, M.H. Experimental and Clinical Reports on Anti-Inflammatory, Antioxidant, and Immunomodulatory Effects of Curcuma Longa and Curcumin, an Updated and Comprehensive Review. BioFactors Oxf. Engl. 2021, 47, 311–350.
- 99. Anamika, B. Extraction of Curcumin. J. Env. Sci Toxicol Food Technol 2012, 1, 1–16.
- 100. Miłobędzka, J.; Kostanecki, S.; Lampe, V. Zur Kenntnis des Curcumins. Berichte Dtsch. Chem. Ges. 1910, 43, 2163–2170.

- 101. Jantarat, C. Bioavailability enhancement techniques of herbal medicine: A case example of curcumin. Int. J. Pharm. Pharm. Sci. 2013, 5, 493–500.
- 102. Agrawal, D.K.; Mishra, P.K. Curcumin and Its Analogues: Potential Anticancer Agents. Med. Res. Rev. 2010, 30, 818–860.
- 103. Lima, C.F.; Pereira-Wilson, C.; Rattan, S.I.S. Curcumin Induces Heme Oxygenase-1 in Normal Human Skin Fibroblasts through Redox Signaling: Relevance for Anti-Aging Intervention. Mol. Nutr. Food Res. 2011, 55, 430–442.
- 104. Šudomová, M.; Hassan, S.T. Nutraceutical curcumin with promising protection against herpesvirus infections and their associated inflammation: Mechanisms and pathways. Microorganisms 2021, 9, 292.
- 105. Mun, S.-H.; Joung, D.-K.; Kim, Y.-S.; Kang, O.-H.; Kim, S.-B.; Seo, Y.-S.; Kim, Y.-C.; Lee, D.-S.; Shin, D.-W.; Kweon, K.-T.; et al. Synergistic Antibacterial Effect of Curcumin against Methicillin-Resistant Staphylococcus Aureus. Phytomedicine Int. J. Phytother. Phytopharm. 2013, 20, 714–718.
- 106. Akbik, D.; Ghadiri, M.; Chrzanowski, W.; Rohanizadeh, R. Curcumin as a Wound Healing Agent. Life Sci. 2014, 116, 1–7.
- 107. Liang, G.; Yang, S.; Zhou, H.; Shao, L.; Huang, K.; Xiao, J.; Huang, Z.; Li, X. Synthesis, Crystal Structure and Anti-Inflammatory Properties of Curcumin Analogues. Eur. J. Med. Chem. 2009, 44, 915–919.
- 108. Ak, T.; Gülçin, I. Antioxidant and Radical Scavenging Properties of Curcumin. Chem. Biol. Interact. 2008, 174, 27–37.
- 109. Thaloor, D.; Miller, K.J.; Gephart, J.; Mitchell, P.O.; Pavlath, G.K. Systemic Administration of the NF-KB Inhibitor Curcumin Stimulates Muscle Regeneration after Traumatic Injury. Am. J. Physiol.-Cell Physiol. 1999, 277, C320–C329.
- 110. Thangapazham, R.L.; Sharad, S.; Maheshwari, R.K. Skin Regenerative Potentials of Curcumin. BioFactors Oxf. Engl. 2013, 39, 141–149.
- 111. Apel, K.; Hirt, H. Reactive Oxygen Species: Metabolism, Oxidative Stress, and Signal Transduction. Annu. Rev. Plant Biol. 2004, 55, 373–399.
- 112. Fujisawa, S.; Atsumi, T.; Ishihara, M.; Kadoma, Y. Cytotoxicity, ROS-Generation Activity and Radical-Scavenging Activity of Curcumin and Related Compounds. Anticancer Res. 2004, 24, 563–569.
- 113. Phan, T.T.; See, P.; Lee, S.T.; Chan, S.Y. Protective Effects of Curcumin against Oxidative Damage on Skin Cells in Vitro: Its Implication for Wound Healing. J. Trauma 2001, 51, 927–931.
- 114. Barzegar, A.; Moosavi-Movahedi, A.A. Intracellular ROS Protection Efficiency and Free Radical-Scavenging Activity of Curcumin. PLoS ONE 2011, 6, e26012.
- 115. Tejada, S.; Manayi, A.; Daglia, M.; Nabavi, F.S.; Sureda, A.; Hajheydari, Z.; Gortzi, O.; Pazoki-Toroudi, H.; Nabavi, M.S. Wound Healing Effects of Curcumin: A Short Review. Curr. Pharm. Biotechnol. 2016, 17, 1002–1007.
- 116. Wang, X.; Shen, K.; Wang, J.; Liu, K.; Wu, G.; Li, Y.; Luo, L.; Zheng, Z.; Hu, D. Hypoxic Preconditioning Combined with Curcumin Promotes Cell Survival and Mitochondrial Quality of Bone Marrow Mesenchymal Stem Cells, and Accelerates Cutaneous Wound Healing via PGC-1α/SIRT3/HIF-1α Signaling. Free Radic. Biol. Med. 2020, 159, 164–176.
- 117. Mohanty, C.; Das, M.; Sahoo, S.K. Emerging Role of Nanocarriers to Increase the Solubility and Bioavailability of Curcumin. Expert Opin. Drug Deliv. 2012, 9, 1347–1364.
- 118. Pandey, V.K.; Ajmal, G.; Upadhyay, S.N.; Mishra, P.K. Nano-Fibrous Scaffold with Curcumin for Anti-Scar Wound Healing. Int. J. Pharm. 2020, 589, 119858.
- 119. Orsu, P.; Haider, H.Y.; Koyyada, A. Bioengineering for Curcumin Loaded Carboxymethyl Guargum/Reduced Graphene Oxide Nanocomposites for Chronic Wound Healing Applications. Int. J. Pharm. 2021, 606, 120928.
- 120. Rathinavel, S.; Korrapati, P.S.; Kalaiselvi, P.; Dharmalingam, S. Mesoporous Silica Incorporated PCL/Curcumin Nanofiber for Wound Healing Application. Eur. J. Pharm. Sci. Off. J. Eur. Fed. Pharm. Sci. 2021, 167, 106021.
- 121. Prakash, J.; Venkataprasanna, K.S.; Bharath, G.; Banat, F.; Niranjan, R.; Venkatasubbu, G.D. In-Vitro Evaluation of Electrospun Cellulose Acetate Nanofiber Containing Graphene Oxide/TiO2/Curcumin for Wound Healing Application. Colloids Surf. Physicochem. Eng. Asp. 2021, 627, 127166.
- 122. Fahimirad, S.; Abtahi, H.; Satei, P.; Ghaznavi-Rad, E.; Moslehi, M.; Ganji, A. Wound Healing Performance of PCL/Chitosan Based Electrospun Nanofiber Electrosprayed with Curcumin Loaded Chitosan Nanoparticles. Carbohydr. Polym. 2021, 259, 117640.
- 123. Zakerikhoob, M.; Abbasi, S.; Yousefi, G.; Mokhtari, M.; Noorbakhsh, M.S. Curcumin-Incorporated Crosslinked Sodium Alginate-g-Poly (N-Isopropyl Acrylamide) Thermo-Responsive Hydrogel as an in-Situ Forming Injectable Dressing for Wound Healing: In Vitro Characterization and in vivo Evaluation. Carbohydr. Polym. 2021, 271, 118434.

- 124. Rathinavel, S.; Indrakumar, J.; Korrapati, P.S.; Dharmalingam, S. Synthesis and Fabrication of Amine Functionalized SBA-15 Incorporated PVA/Curcumin Nanofiber for Skin Wound Healing Application. Colloids Surf. Physicochem. Eng. Asp. 2022, 637, 128185.
- 125. Chen, K.; Pan, H.; Ji, D.; Li, Y.; Duan, H.; Pan, W. Curcumin-Loaded Sandwich-like Nanofibrous Membrane Prepared by Electrospinning Technology as Wound Dressing for Accelerate Wound Healing. Mater. Sci. Eng. C Mater. Biol. Appl. 2021, 127, 112245.
- 126. Pillai, M.M.; Dandia, H.; Checker, R.; Rokade, S.; Sharma, D.; Tayalia, P. Novel Combination of Bioactive Agents in Bilayered Dermal Patches Provides Superior Wound Healing. Nanomed. Nanotechnol. Biol. Med. 2022, 40, 102495.
- 127. Gorain, B.; Pandey, M.; Leng, N.H.; Yan, C.W.; Nie, K.W.; Kaur, S.J.; Marshall, V.; Sisinthy, S.P.; Panneerselvam, J.; Molugulu, N.; et al. Advanced Drug Delivery Systems Containing Herbal Components for Wound Healing. Int. J. Pharm. 2022, 617, 121617.
- 128. Komes, D.; Belščak-Cvitanović, A.; Jurić, S.; Bušić, A.; Vojvodić, A.; Durgo, K. Consumer Acceptability of Liquorice Root (Glycyrrhiza glabra L.) as an Alternative Sweetener and Correlation with Its Bioactive Content and Biological Activity. Int. J. Food Sci. Nutr. 2016, 67, 53–66.
- 129. Jeon, J.S.; Kim, H.T.; Kim, M.G.; Oh, M.S.; Hong, S.R.; Yoon, M.H.; Shin, H.C.; Shim, J.H.; Afifi, N.A.; Hacımüftüoğlu, A.; et al. Simultaneous Detection of Glabridin, (-)-α-Bisabolol, and Ascorbyl Tetraisopalmitate in Whitening Cosmetic Creams Using HPLC-PAD. Chromatographia 2016, 79, 851–860.
- 130. Pastorino, G.; Cornara, L.; Soares, S.; Rodrigues, F.; Oliveira, M.B.P.P. Liquorice (Glycyrrhiza glabra): A Phytochemical and Pharmacological Review. Phytother. Res. 2018, 32, 2323–2339.
- 131. Memariani, Z.; Hajimahmoodi, M.; Minaee, B.; Khodagholi, F.; Yans, A.; Rahimi, R.; Amin, G.; Moghaddam, G.; Toliyat, T.; Sharifzadeh, M. Protective Effect of a Polyherbal Traditional Formula Consisting of Rosa damascena Mill., Glycyrrhiza glabra L. and Nardostachys jatamansi DC., Against Ethanol-Induced Gastric Ulcer. Iran. J. Pharm. Res. IJPR 2017, 16, 694–707.
- 132. Najeeb, V.D.; Al-Refai, A.S. Antibacterial Effect and Healing Potential of Topically Applied Licorice Root Extract on Experimentally Induced Oral Wounds in Rabbits. Saudi J. Oral Sci. 2015, 2, 10.
- 133. Chen, X.; Fang, D.; Li, L.; Chen, L.; Li, Q.; Gong, F.; Fang, M. Glycyrrhizin Ameliorates Experimental Colitis through Attenuating Interleukin-17-Producing T Cell Responses via Regulating Antigen-Presenting Cells. Immunol. Res. 2017, 65, 666–680.
- 134. Shah, S.L.; Wahid, F.; Khan, N.; Farooq, U.; Shah, A.J.; Tareen, S.; Ahmad, F.; Khan, T. Inhibitory Effects of Glycyrrhiza glabra and Its Major Constituent Glycyrrhizin on Inflammation-Associated Corneal Neovascularization. Evid. Based Complement. Alternat. Med. 2018, 2018.
- 135. Oloumi, M.M.; Derakhshanfar, A.; Nikpour, A. Healing Potential of Liquorice Root Extract on Dermal Wounds in Rats. J. Vet. Res. 2007, 62, 147–154.
- 136. Li, B.; Wang, J.H.-C. Fibroblasts and Myofibroblasts in Wound Healing: Force Generation and Measurement. J. Tissue Viability 2011, 20, 108–120.
- 137. Geethalakshmi, R.; Sakravarthi, C.; Kritika, T.; Arul Kirubakaran, M.; Sarada, D.V.L. Evaluation of Antioxidant and Wound Healing Potentials of Sphaeranthus Amaranthoides Burm.f. BioMed Res. Int. 2013, 2013, 607109.
- 138. Zangeneh, A.; Pooyanmehr, M.; Zangeneh, M.M.; Moradi, R.; Rasad, R.; Kazemi, N. Therapeutic Effects of Glycyrrhiza glabra Aqueous Extract Ointment on Cutaneous Wound Healing in Sprague Dawley Male Rats. Comp. Clin. Pathol. 2019, 28, 1507–1514.
- 139. Siriwattanasatorn, M.; Itharat, A.; Thongdeeying, P.; Ooraikul, B. In Vitro Wound Healing Activities of Three Most Commonly Used Thai Medicinal Plants and Their Three Markers. Evid. Based Complement. Alternat. Med. 2020, 2020, e6795383.
- 140. Hanafi, N.; Amiri, F.T.; Shahani, S.; Enayatifard, R.; Ghasemi, M.; Karimpour, A.A. Licorice Cream Promotes Full-Thickness Wound Healing in Guinea Pigs. Marmara Pharm. J. 2022, 22, 411–421.
- 141. Barros, L.; Carvalho, A.M.; Ferreira, I.C.F.R. Leaves, Flowers, Immature Fruits and Leafy Flowered Stems of Malva sylvestris: A Comparative Study of the Nutraceutical Potential and Composition. Food Chem. Toxicol. 2010, 48, 1466–1472.
- 142. Quave, C.L.; Plano, L.R.W.; Pantuso, T.; Bennett, B.C. Effects of Extracts from Italian Medicinal Plants on Planktonic Growth, Biofilm Formation and Adherence of Methicillin-Resistant Staphylococcus Aureus. J. Ethnopharmacol. 2008, 118, 418–428.
- 143. DellaGreca, M.; Cutillo, F.; D'Abrosca, B.; Fiorentino, A.; Pacifico, S.; Zarrelli, A. Antioxidant and Radical Scavenging Properties of Malva sylvestris. Nat. Prod. Commun. 2009, 4, 893–896.

- 144. Leporatti, M.L.; Ghedira, K. Comparative Analysis of Medicinal Plants Used in Traditional Medicine in Italy and Tunisia. J. Ethnobiol. Ethnomedicine 2009, 5, 31.
- 145. Pirbalouti, A.G.; Azizi, S.; Koohpayeh, A.; Hamedi, B. Wound Healing Activity of Malva sylvestris and Punica Granatum in Alloxan-Induced Diabetic Rats. Acta Pol. Pharm. 2010, 67, 511–516.
- 146. Pirbalouti, A.G.; Koohpyeh, A. Wound Healing Activity of Extracts of Malva sylvestris and Stachys lavandulifolia. Int. J. Biol. 2010, 3, 174.
- 147. Gasparetto, J.C.; Martins, C.A.F.; Hayashi, S.S.; Otuky, M.F.; Pontarolo, R. Ethnobotanical and Scientific Aspects of Malva sylvestris L.: A Millennial Herbal Medicine. J. Pharm. Pharmacol. 2012, 64, 172–189.
- 148. Mousavi, S.M.; Hashemi, S.A.; Behbudi, G.; Mazraedoost, S.; Omidifar, N.; Gholami, A.; Chiang, W.-H.; Babapoor, A.; Pynadathu Rumjit, N. A Review on Health Benefits of Malva sylvestris L. Nutritional Compounds for Metabolites, Antioxidants, and Anti-Inflammatory, Anticancer, and Antimicrobial Applications. Evid. Based Complement. Alternat. Med. 2021, 2021, e5548404.
- 149. Afshar, M.; Ravarian, B.; Zardast, M.; Moallem, S.A.; Fard, M.H.; Valavi, M. Evaluation of Cutaneous Wound Healing Activity of Malva sylvestris Aqueous Extract in BALB/c Mice. Iran. J. Basic Med. Sci. 2015, 18, 616.
- 150. Nasiri, E.; Hosseinimehr, S.J.; Azadbakht, M.; Akbari, J.; Enayati-Fard, R.; Azizi, S. Effect of Malva sylvestris Cream on Burn Injury and Wounds in Rats. Avicenna J. Phytomed. 2015, 5, 341–354.
- 151. Almasian, A.; Najafi, F.; Eftekhari, M.; Ardekani, M.R.S.; Sharifzadeh, M.; Khanavi, M. Polyurethane/Carboxymethylcellulose Nanofibers Containing Malva sylvestris Extract for Healing Diabetic Wounds: Preparation, Characterization, in Vitro and in vivo Studies. Mater. Sci. Eng. C Mater. Biol. Appl. 2020, 114, 111039.
- 152. Janković, T.; Zdunić, G.; Beara, I.; Balog, K.; Pljevljakušić, D.; Stešević, D.; Šavikin, K. Comparative Study of Some Polyphenols in Plantago Species. Biochem. Syst. Ecol. 2012, 42, 69–74.
- 153. Beara, I.N.; Lesjak, M.M.; Jovin, E.D.; Balog, K.J.; Anackov, G.T.; Orcić, D.Z.; Mimica-Dukić, N.M. Plantain (Plantago L.) Species as Novel Sources of Flavonoid Antioxidants. J. Agric. Food Chem. 2009, 57, 9268–9273.
- 154. Amakura, Y.; Yoshimura, A.; Yoshimura, M.; Yoshida, T. Isolation and Characterization of Phenolic Antioxidants from Plantago Herb. Molecules 2012, 17, 5459–5466.
- 155. Zubair, M.; Widén, C.; Renvert, S.; Rumpunen, K. Water and Ethanol Extracts of Plantago Major Leaves Show Anti-Inflammatory Activity on Oral Epithelial Cells. J. Tradit. Complement. Med. 2019, 9, 169–171.
- 156. Genc, Y.; Harput, U.S.; Saracoglu, I. Active Compounds Isolated from Plantago subulata L. via Wound Healing and Antiinflammatory Activity Guided Studies. J. Ethnopharmacol. 2019, 241, 112030.
- 157. Ghanadian, M.; Soltani, R.; Homayouni, A.; Khorvash, F.; Jouabadi, S.M.; Abdollahzadeh, M. The Effect of Plantago Major Hydroalcoholic Extract on the Healing of Diabetic Foot and Pressure Ulcers: A Randomized Open-Label Controlled Clinical Trial. Int. J. Low Extrem. Wounds 2022.
- 158. Wang, M.; Li, J.; Rangarajan, M.; Shao, Y.; LaVoie, E.J.; Huang, T.-C.; Ho, C.-T. Antioxidative Phenolic Compounds from Sage (Salvia officinalis). J. Agric. Food Chem. 1998, 46, 4869–4873.
- 159. Venskutonis, P.R. Effect of Drying on the Volatile Constituents of Thyme (Thymus Vulgaris L.) and Sage (Salvia officinalis L.). Food Chem. 1997, 59, 219–227.
- 160. Kamatou, G.P.P.; Makunga, N.P.; Ramogola, W.P.N.; Viljoen, A.M. South African Salvia Species: A Review of Biological Activities and Phytochemistry. J. Ethnopharmacol. 2008, 119, 664–672.
- 161. Topçu, G. Bioactive Triterpenoids from Salvia Species. J. Nat. Prod. 2006, 69, 482–487.
- 162. Bozin, B.; Mimica-Dukic, N.; Samojlik, I.; Jovin, E. Antimicrobial and Antioxidant Properties of Rosemary and Sage (Rosmarinus officinalis L. and Salvia officinalis L., Lamiaceae) Essential Oils. J. Agric. Food Chem. 2007, 55, 7879–7885.
- 163. Länger, R.; Mechtler, C.; Jurenitsch, J. Composition of the Essential Oils of Commercial Samples of Salvia officinalis L. and S. Fruticosa Miller: A Comparison of Oils Obtained by Extraction and Steam Distillation. Phytochem. Anal. 1996, 7, 289–293.
- 164. Hayouni, E.A.; Chraief, I.; Abedrabba, M.; Bouix, M.; Leveau, J.-Y.; Mohammed, H.; Hamdi, M. Tunisian Salvia officinalis L. and Schinus molle L. Essential Oils: Their Chemical Compositions and Their Preservative Effects against Salmonella Inoculated in Minced Beef Meat. Int. J. Food Microbiol. 2008, 125, 242–251.
- 165. Badiee, P.; Nasirzadeh, A.R.; Motaffaf, M. Comparison of Salvia officinalis L. Essential Oil and Antifungal Agents against Candida Species. J. Pharm. Technol. Drug Res. 2012, 1.
- 166. Ghorbani, A.; Esmaeilizadeh, M. Pharmacological Properties of Salvia officinalis and Its Components. J. Tradit. Complement. Med. 2017, 7, 433–440.

- 167. Perry, N.B.; Anderson, R.E.; Brennan, N.J.; Douglas, M.H.; Heaney, A.J.; McGimpsey, J.A.; Smallfield, B.M. Essential Oils from Dalmatian Sage (Salvia officinalis L.): Variations among Individuals, Plant Parts, Seasons, and Sites. J. Agric. Food Chem. 1999, 47, 2048–2054.
- 168. Raal, A.; Orav, A.; Arak, E. Composition of the Essential Oil of Salvia officinalis L. from Various European Countries. Nat. Prod. Res. 2007, 21, 406–411.
- 169. de Macedo, L.M.; dos Santos, É.M.; Militão, L.; Tundisi, L.L.; Ataide, J.A.; Souto, E.B.; Mazzola, P.G. Rosemary (Rosmarinus officinalis L., Syn Salvia rosmarinus Spenn.) and Its Topical Ap-plications: A Review. Plants 2020, 9, 651.
- 170. Scrima, M.; Melito, C.; Merola, F.; Iorio, A.; Vito, N.; Giori, A.M.; Ferravante, A. Evaluation of Wound Healing Activity of Salvia Haenkei Hydroalcoholic Aerial Part Extract on in Vitro and in vivo Experimental Models. Clin. Cosmet. Investig. Dermatol. 2020, 13, 627–637.
- 171. Farahpour, M.R.; Pirkhezr, E.; Ashrafian, A.; Sonboli, A. Accelerated Healing by Topical Administration of Salvia officinalis Essential Oil on Pseudomonas Aeruginosa and Staphylococcus Aureus Infected Wound Model. Biomed. Pharmacother. Biomed. Pharmacother. 2020, 128, 110120.
- 172. Karimzadeh, S.; Farahpour, M.R. Topical Application of Salvia officinalis Hydroethanolic Leaf Extract Improves Wound Healing Process. Indian J. Exp. Biol. 2017, 55, 98–106.
- 173. Güzel, S.; Özay, Y.; Kumaş, M.; Uzun, C.; Özkorkmaz, E.G.; Yıldırım, Z.; Ülger, M.; Güler, G.; Çelik, A.; ÇamLıca, Y.; et al. Wound Healing Properties, Antimicrobial and Antioxidant Activities of Salvia Kronenburgii Rech. f. and Salvia Euphratica Montbret, Aucher & Rech. f. Var. Euphratica on Excision and Incision Wound Models in Diabetic Rats. Biomed. Pharmacother. Biomed. Pharmacother. 2019, 111, 1260–1276.
- 174. Taxonomy. Available online: https://www.uniprot.org/taxonomy/39367 (accessed on 29 March 2022).
- 175. Al-Sereiti, M.R.; Abu-Amer, K.M.; Sen, P. Pharmacology of Rosemary (Rosmarinus Officinalis Linn.) and Its Therapeutic Potentials. Indian J. Exp. Biol. 1999, 37, 124–130.
- 176. Ribeiro-Santos, R.; Carvalho-Costa, D.; Cavaleiro, C.; Costa, H.S.; Albuquerque, T.G.; Castilho, M.C.; Ramos, F.; Melo, N.R.; Sanches-Silva, A. A Novel Insight on an Ancient Aromatic Plant: The Rosemary (Rosmarinus Officinalis L.).

  Trends Food Sci. Technol. 2015, 45, 355–368.
- 177. Begum, A.; Sandhya, S.; Shaffath Ali, S.; Vinod, K.R.; Reddy, S.; Banji, D. An In-Depth Review on the Medicinal Flora Rosmarinus Officinalis (Lamiaceae). Acta Sci. Pol. Technol. Aliment. 2013, 12, 61–73.
- 178. Rašković, A.; Milanović, I.; Pavlović, N.; Ćebović, T.; Vukmirović, S.; Mikov, M. Antioxidant Activity of Rosemary (Rosmarinus Officinalis L.) Essential Oil and Its Hepatoprotective Potential. BMC Complement. Altern. Med. 2014, 14, 225.
- 179. Abu-Al-Basal, M.A. Healing Potential of Rosmarinus officinalis L. on Full-Thickness Excision Cutaneous Wounds in Alloxan-Induced-Diabetic BALB/c Mice. J. Ethnopharmacol. 2010, 131, 443–450.
- 180. Yu, M.-H.; Choi, J.-H.; Chae, I.-G.; Im, H.-G.; Yang, S.-A.; More, K.; Lee, I.-S.; Lee, J. Suppression of LPS-Induced Inflammatory Activities by Rosmarinus officinalis L. Food Chem. 2013, 136, 1047–1054.
- 181. Mena, P.; Cirlini, M.; Tassotti, M.; Herrlinger, K.A.; Dall'Asta, C.; Del Rio, D. Phytochemical Profiling of Flavonoids, Phenolic Acids, Terpenoids, and Volatile Fraction of a Rosemary (Rosmarinus Officinalis L.) Extract. Molecules 2016, 21, 1576.
- 182. del Baño, M.J.; Lorente, J.; Castillo, J.; Benavente-García, O.; Marín, M.P.; Del Río, J.A.; Ortuño, A.; Ibarra, I. Flavonoid Distribution during the Development of Leaves, Flowers, Stems, and Roots of Rosmarinus Officinalis. Postulation of a Biosynthetic Pathway. J. Agric. Food Chem. 2004, 52, 4987–4992.
- 183. Umasankar, K.; Nambikkairaj, B.; Backyavathy, D.M. Effect of Topical Treatment of Rosmarinus Officinalis Essential Oil on Wound Healing in Streptozotocin Induced Diabetic Rats. Nat. Environ. Pollut. Technol. 2012, 11, 5.
- 184. Labib, R.M.; Ayoub, I.M.; Michel, H.E.; Mehanny, M.; Kamil, V.; Hany, M.; Magdy, M.; Moataz, A.; Maged, B.; Mohamed, A. Appraisal on the Wound Healing Potential of Melaleuca Alternifolia and Rosmarinus officinalis L. Essential Oil-Loaded Chitosan Topical Preparations. PLoS ONE 2019, 14, e0219561.