

Genetic Loci Underlying Awn Morphology in Barley

Subjects: **Genetics & Heredity**

Contributor: Biguang Huang , Weiren Wu , Zonglie Hong

Awns are a transformed leaf blade that extends from the lemmas of the florets, and can contribute significantly to the grain filling via active photosynthesis. Barley awns are highly diverse in morphology, ranging from long to short or awnless types, and from straight to hooded or crooked shapes. The morphological diversity, importance to grain yield and ease of study of barley awns have intrigued plant geneticists for several decades. A large set of genetic loci associated with the development of awns have been identified genetically and mapped to chromosomes.

awn

barley

gene mapping

genetic epistasis

1. Awnness or Awnlessness

To be, or not to be, that is the question. With this soliloquy, William Shakespeare once pondered, through Hamlet's words, upon a state of being or not being, a forever lasting philosophic question about life and death. A similar balance about awnness or awnlessness should have puzzled our human ancestors some 8000 years ago when barley (*Hordeum vulgare* L.) was domesticated. Wild grass species, including wild barley (*Hordeum spontaneum*), are mostly awned, whereas their cereal crop relatives vary. Domestication has led to the disappearance of awns from most popular cultivars of rice and wheat, while, on the contrary, most modern barley varieties still preserve awns. The awn on the barley spike has no nutritional value and can cause lacerations in the mouth of animals. One of the most important breeding goals for forage barley is the elimination of the awn without reducing the grain yield. A better understanding of the genetic basis for awnness is key to advancing breeding programs aimed at barley cultivars with special awn types. Although environmental conditions affect awn development to a certain degree, the question of 'to awn or not to awn' is largely answered by the genetic composition of the barley genome. The inheritance of the awn trait in barley has been a topic of genetic study for several decades, primarily due to the importance of the awn to grain yield and its hindrance in forage quality and palatability ^[1]. The specific objectives of this review are to provide an update of research work conducted by various groups in the field of awn development and inheritance over the last several decades, with a focus on the recent progress in genetic loci underlying awnness that have been identified, mapped or cloned in barley. The complexity of gene–gene interactions and pleiotropic effects of awn loci are analyzed and the perspectives of future research work in this field are presented in this review.

In addition, the awn is not an essential organ for the survival and reproduction of barley, allowing awn shape to vary dramatically through natural selections and chemical mutagenesis. The awn protrudes from the canopy in fields and morphological differences are easy to score, leading to large collections of abundant germplasm

resources of awn mutants. In addition, barley represents an excellent model for genetic research of crops: it is diploid with a low chromosome number ($2n = 14$), can be both inbred and outbred easily, and adapts to diverse climate conditions [2][3]. The completion of the barley genome sequencing project [4][5][6] has facilitated identification and cloning of agronomically important genes, such as the row-type locus VRS3 [7][8].

2. Morphological Diversity of Barley Awns

The diversity and development of awns in barley have long been a topic of genetic research and now are subjects of intense molecular biology and genomic studies [9][10]. Based on the presence of awns, barley germplasms can be divided into awnless and awned types, with the latter subdivided further into hooded, leafy, straight and crooked phenotypes (**Figure 2**). The straight awns (single or triple) can be long, short or awnlet, while the hooded ones can be classified as either normal hooded, elevated hooded, or subjacent hooded. Furthermore, barley awns do not behave consistently in spikelets between the central and lateral rows of the same spike. For example, when the central row develops long awned spikelets, the lateral row can be short awned or even awnless. When both the central and lateral rows develop long awns, the awns in the lateral rows are generally a little shorter than those of the central rows. Hooded awns have their own unique structures, which can sometimes develop inverse, fertile flowers capable of producing seeds [9][11]. The striking inverse flower of hooded awns is thought to be the result of the transformation of the awn into reiterative inflorescence axes [12]. Subjacent hooded awns are similar to the hooded ones, but differ by bearing distal awns without an epiphyllous flower, reflecting differing degrees of phenotypic penetrance [13]. In anatomy, the awn is a transformed leaf blade, as illustrated in the leafy lemma mutant (**Figure 1K**). Barley germplasms from the Tibet plateau have been shown to exhibit highly diverse awn characteristics, ranging from long awns on central rows but long, short, or no awns at all on lateral rows. Similarly, other cultivars can have short awns on the central row with short or awnless on the lateral row, hooded awns on the central and hooded, short or awnless on the lateral, to completely awnless on both the central and lateral rows [14][15].



Figure 1. Morphological diversity of awns in barley cultivars and mutants. (A), 6-rowed spike with long awns. (B), awnless 2-rowed spike. (C), 6-rowed spike with stigma-like short, crooked awns. (D), Hooded spike. (E), Hooded spike with long awns. (F,G), 2-rowed and 6-rowed spikes with long awns. (H), 6-rowed spike having long awns on the central spikelet and awnless lateral spikelets. (I), Front- and side-views of a triple awn. (J), 6-rowed spikelets with elevated hoods. (K), spikelet with leafy awn. (L), awnless spikelet. (M), 6-rowed spikelets with stigma-like short, crooked awns. (N), hooded floret with an extra floret. (O), subjacent hooded floret. st, stigma-like short, crooked awn; h, hooded awn; ef, extra floret; eh, elevated hooded; leafy, leafy awn; le, lemma; ep, extra palea, el, extra lemma.

3. Mapped Genetic Loci for Awns in Barley

A three-letter code for genes and the new Triticeae system for chromosome designation have been adopted in **Table 1** and **Table 2** to describe barley awnness loci [16]. Previous designations of barley chromosomes 1 through 7 correspond to 7H, 2H, 3H, 4H, 1H, 6H and 5H, respectively [17]. The 12 awnness loci identified so far (**Table 1** and **Table 2**) underlie a spectrum of awn phenotypes, including awnlessness (Lks1, Lsa1), short awns (lks2,

lks5/lel2, lks6, and ari-a), reduced lateral spikelet appendage (lr) , leafy lemma (lel1), hooded lemma (Kap1), subjacent hooded lemma (sbk1), short, crooked awn (sca1), and triple awns (trp1). Their chromosome locations and their relative distances to known morphological marker genes are shown in **Figure 3** .

Table 1. Cloned awnness genes in barley.

Gene Names	Kap1 (hooded lemma 1)	vrs1 (six-rowed spike 1)	lks2 (short awn 2)
Other Names	K, HvKnox3	lr, HvHox1	lk2, lk4, ari-d, ubs4
Awn Phenotype	Hooded lemma	Reduced lateral spikelet appendage	Short awn
Chromosome	4HS	2HL	7HL
Cloning Method	Homologous cloning	Positional cloning	Positional cloning
Mutation Types	Insertion in intron 4	Missense, nonsense, and splicing site changes	Missense mutation
Protein Length ^a	364 aa	222 aa	344 aa
Protein Type	KNOX family transcription factor	HD-ZIP family transcription factor	SRS family transcription factor
Orthologs	Rice OSH15 and maize KNOTTED-1	OsHox12 and OsHox14 in rice	Os06g0712600 in rice
References	[18] [19]	[20] [21]	[22] [23]

^a aa, amino acid residues.

Table 2. Mapped genetic loci for awnness in barley.

Locus	Other Names	Awn Phenotypes	Chr ^a	Ref. ^b
sbk1	sk, cal-a	Subjacent hood	2HS	[13] [24]
lel1	lel	Leafy lemma	2HL	[3] [25]
trp1	tr	Triple awned lemma	2HL	[19] [26]
sca1	sca	Short, crooked awn	3HS	[27]
lks5	lk5, ari-c, lel2	Short awn	4HS	[23] [26]
ari-a	lk7	Short awn	3HS	[28] [29]

Locus	Other Names	Awn Phenotypes	Chr ^a	Ref. ^b
lks6	Lks.q	Short awn	ND ^c	[3]
Lks1	Lk	Awnless	2HL	[30]
Lsa1	Lsa	Awnless on lateral rows	2H	[31]

^a Location of the short (S) and long (L) arms of chromosomes. ^b Reference (ref.) sources. ^c ND: chromosome location not determined.

Lks1 (awnless) was once considered to be the same as vrs1, since they are closely linked on 2HL. This linkage has also complicated the fine mapping and gene cloning of Lks1. The awn inhibitor gene B1, which is dominant and inhibits awn development, has recently been cloned in wheat by several independent groups [32][33][34][35][36]. The wheat B1 gene encodes a putative C2H2 zinc-finger protein with an EAR domain that is characteristic of transcriptional repressors. The cloning of wheat B1 may facilitate cloning of barley Lks1, because the synteny of their locations on chromosomes between barley and wheat may provide useful hints for gene cloning.

The length of awns is controlled by the interactions of several awn genes, including Lr1 (for reduced length of lateral awns), lks5 (or lk5, for short awn 5) and lks2 (or lk and lk4, for short awns) [37][38]. The genetic effect of Lr1 is more prominent than that of Lks5, while Lks2 and Lks5 act additively to regulate awn length [31][37][38]. Thus, the interactions among these three awn genes play critical roles in the determination of awn length in barley.

Of the 12 awnness loci, both the awnless gene Lks1 and the hooded awn gene Kap1 or sbk1 may become important targets for feed barley improvements. Because the awn is an undesirable trait in feed barley, it has been proposed that the awnless gene Lks1 could be introduced to feed barley cultivars. However, this strategy is currently met with the challenge that the available Lks1 mutant is associated with reduced grain yield. The Kap1 gene appears to be a more suitable target gene for feed barley, because its mutant does not exhibit reductions in grain yield. In the USA, new barley cultivars containing Lks1 or Kap1 have been released for forage uses. If the reduction of grain yield cannot be resolved in awnless barley cultivars, one of the alternative approaches is to target short awn or hooded awn genes for improvement in forage breeding programs.

4. Pleiotropism of Awn Genes

Genetic pleiotropism describes the effect of a single gene on multiple, unrelated traits. The mutation of a pleiotropic gene may thus lead to changes in several seemingly unrelated attributes of an organism. For mapping analysis, it is critical to distinguish the pleiotropic effect of one gene on multiple traits from a close linkage of two independent loci. For a long time, it has been believed that the awnless gene Lks1 on chromosome 2H may be allelic to the row-type locus vrs1, and that the awnless phenotype and the six-row spike trait were just the result of pleiotropic effects of the same locus. This assumption was made on the basis of the lack of recombination between the two loci when a line with a dominant short awn gene Lks1 and the two-rowed gene Vrs1 is crossed with another line having long awns and six-rowed spikes (lks1/lks1 vrs1/vrs1) [30][39][40]. However, further genetic analysis has suggested that the

two loci are independent but closely linked and lie near a paracentric chromosome inversion that may inhibit recombination between them [\[41\]](#).

The pleiotropic effects of short awn genes are especially common. A QTL, qAL/SL/GD3.1, has been shown to control all three traits of grain density, spike length and awn length [\[42\]](#). The dwarf gene uzu1 has pleiotropic effects on the length of leaves, culms, rachis internodes, awns, glumes and kernels [\[43\]](#). A dwarfing gene closely linked to uzu1 on chromosome 3H, is pleiotropic for short spikes, short awns and high grain density [\[44\]](#). Allelism tests show that lks2, which reduces the awn length to about one half of the normal length (Lks2), is allelic to the unbranched style 4 (ubs4), which also displays sparse stigma hairs [\[22\]](#). The short awn gene lks5 on chromosome 4H has pleiotropic effects on the spikelets and produces extra florets [\[45\]](#).

Understanding the pleiotropism of awn genes will facilitate elucidation of how they function in awn development. Most awned varieties in barley and wheat have higher yields than awnless ones because of the grain-filling function of awns in photosynthesis [\[46\]](#)[\[47\]](#). Despite the overall higher potential of awned cultivars, there are awnless or short awned varieties that have good performance in yield in some ecological areas [\[48\]](#). This unexpected high yield of half-awned or awnless varieties has been attributed to the high kernel number per spike, not to grain size [\[49\]](#). Awnless or short awn traits normally decrease the size of grains but increase the number of spikelets/florets per spike [\[49\]](#). However, the awned trait usually reduces the grain number and increases the grain size and harvestable yield [\[48\]](#)[\[50\]](#). The wheat awnless gene B1 also decreases kernel weight, kernel length and test weight, but increases the number of spikelets per spike [\[33\]](#). Overexpression of the barley HvFT3 accelerates the initiation of spikelet primordia [\[32\]](#), and HvFT3 is believed to act upstream of the row type genes vrs1, vrs4 and int-c [\[51\]](#). Therefore, awn genes not only regulate awn development, but also play important roles in other physiological and developmental processes such as grain filling and specification of spikelet initiation and row types.

5. Concluding Remarks

The recent completion of the barley genome sequencing project, together with the application of novel sequencing and bioinformatic technologies, will greatly facilitate fine mapping and gene cloning of awn loci in barley. More and more awn genes will soon be cloned and characterized at the molecular biology and biochemistry levels. A better understanding of the molecular nature of awn genes would pave a solid foundation for further investigations on gene-gene interactions and molecular mechanisms of awn and grain development. These studies would eventually benefit barley breeders in developing new cultivars with improved yield and adaptability to various environmental conditions.

References

1. Ntakirutimana, F.; Xie, W. Morphological and genetic mechanisms underlying awn development in monocotyledonous grasses. *Genes* 2019, 10, 573.

2. Saisho, D.; Takeda, K. Barley: Emergence as a new research material of crop science. *Plant Cell Physiol.* 2011, 52, 724–727.
3. Druka, A.; Franckowiak, J.; Lundqvist, U.; Bonar, N.; Alexander, J.; Houston, K.; Radovic, S.; Shahinnia, F.; Vendramin, V.; Morgante, M.; et al. Genetic dissection of barley morphology and development. *Plant Physiol.* 2011, 155, 617–627.
4. The International Barley Genome Sequencing Consortium. A physical, genetic and functional sequence assembly of the barley genome. *Nature* 2012, 491, 711–717.
5. Mascher, M.; Gundlach, H.; Himmelbach, A.; Beier, S.; Twardziok, S.O.; Wicker, T.; Radchuk, V.; Dockter, C.; Hedley, P.E.; Russell, J.; et al. Chromosome conformation capture ordered sequence of the barley genome. *Nature* 2017, 544, 427–433.
6. Monat, C.; Padmarasu, S.; Lux, T.; Wicker, T.; Gundlach, H.; Himmelbach, A.; Ens, J.; Li, C.; Muehlbauer, G.J.; Schulman, A.H.; et al. TRITEX: Chromosome-scale sequence assembly of Triticeae genomes with open-source tools. *Genome Biol.* 2019, 20, 284.
7. van Esse, G.W.; Walla, A.; Finke, A.; Koornneef, M.; Pecinka, A.; von Korff, M. Six-rowed spike3 (VRS3) is a histone demethylase that controls lateral spikelet development in barley. *Plant Physiol.* 2017, 174, 2397–2408.
8. Bull, H.; Casao, M.C.; Zwirek, M.; Flavell, A.J.; Thomas, W.T.B.; Guo, W.; Zhang, R.; Rapazote-Flores, P.; Kyriakidis, S.; Russell, J.; et al. Barley SIX-ROWED SPIKE3 encodes a putative Jumonji C-type H3K9me2/me3 demethylase that represses lateral spikelet fertility. *Nat. Commun.* 2017, 8, 936.
9. Terzi, V.; Tumino, G.; Pagani, D.; Rizza, F.; Ghizzoni, R.; Morcia, C.; Stanca, A.M. Barley developmental mutants: The high road to understand the cereal spike morphology. *Diversity* 2017, 9, 21.
10. Schulte, D.; Close, T.J.; Graner, A.; Langridge, P.; Matsumoto, T.; Muehlbauer, G.; Sato, K.; Schulman, A.H.; Waugh, R.; Wise, R.P.; et al. The International Barley Sequencing Consortium—at the threshold of efficient access to the barley genome. *Plant Physiol.* 2009, 149, 142–147.
11. Stanca, A.M.; Tumino, G.; Pagani, D.; Rizza, F.; Alberici, R.; Lundqvist, U.; Morcia, C.; Tondelli, A.; Terzi, V. The “Italian” barley genetics mutant collection: Conservation, development of new mutants and use. In *Advance in Barley Sciences*; Zhang, G., Li, C., Liu, X., Eds.; Springer: Dordrecht, The Netherlands, 2013; pp. 30–35.
12. Stebbins, G.L.; Yagil, E. The morphogenetic effects of the hooded gene in barley. I. The course of development in hooded and awned genotypes. *Genetics* 1966, 54, 727–741.
13. Pozzi, C.; Faccioli, P.; Terzi, V.; Stanca, A.M.; Cerioli, S.; Castiglioni, P.; Fink, R.; Capone, R.; Müller, K.J.; Bossinger, G.; et al. Genetics of mutations affecting the development of a barley floral bract. *Genetics* 2000, 154, 1335–1346.

14. Xu, T. Classification and identification of varieties of Chinese cultivated barley. *Sci. Agric. Sin.* 1982, 6, 39–47.
15. Sun, L. Varieties of cultivated barley and their distribution in China. *Sci. Agric. Sin.* 1988, 21, 25–31.
16. Franckowiak, J.D.; Kleinhofs, A.; Lundqvist, U. Descriptions of barley genetic stocks for 2016. *Barley Genet. Newsl.* 2016, 46, 1–47.
17. Burnham, C.R.; Hagberg, A. Cytogenetic notes on chromosomal interchanges in barley. *Hereditas* 1956, 42, 467–482.
18. Müller, K.J.; Romano, N.; Gerstner, O.; Garcia-Maroto, F.; Pozzi, C.; Salamini, F.; Rohde, W. The barley hooded mutation caused by a duplication in a homeobox gene intron. *Nature* 1995, 374, 727–730.
19. Pozzi, C.; di Pietro, D.; Halas, G.; Roig, C.; Salamini, F. Integration of a barley (*H. vulgare*) molecular linkage map with the position of genetic loci hosting 29 developmental mutants. *Heredity* 2003, 90, 390–396.
20. Komatsuda, T.; Pourkheirandish, M.; He, C.F.; Azhaguvel, P.; Kanamori, H.; Perovic, D.; Stein, N.; Graner, A.; Wicker, T.; Tagiri, A.; et al. Six-rowed barley originated from a mutation in a homeodomain leucine zipper I-class homeobox gene. *Proc. Natl. Acad. Sci. USA* 2007, 104, 1424–1429.
21. Leonard, W.H. Inheritance of reduced lateral spikelet appendages in the Nudihaxtoni variety of barley. *J. Am. Soc. Agron.* 1942, 34, 211–221.
22. Yuo, T.; Yamashita, Y.; Kanamori, H.; Matsumoto, T.; Lundqvist, U.; Sato, K.; Ichii, M.; Jobling, S.A.; Taketa, S. A SHORT INTERNODES (SHI) family transcription factor gene regulates awn elongation and pistil morphology in barley. *J. Exp. Bot.* 2012, 63, 5223–5232.
23. Taketa, S.; Yuo, T.; Sakurai, Y.; Miyake, S.; Ichii, M. Molecular mapping of the short awn 2 (*lks2*) and dense spike 1 (*dsp1*) genes on barley chromosome 7H. *Breed. Sci.* 2011, 61, 80–85.
24. Gustafsson, Å.; Hagberg, A.; Lundqvist, U.; Persson, G. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 1969, 62, 409–414.
25. Graner, A.; Jahoor, A.; Schondelmaier, J.; Siedler, H.; Pillen, K.; Fischbeck, G.; Wenzel, G.; Herrmann, R.G. Construction of an RFLP map of barley. *Theor. Appl. Genet.* 1991, 83, 250–256.
26. Rossini, L.; Vecchietti, A.; Nicoloso, L.; Stein, N.; Franzago, S.; Salamini, F.; Pozzi, C. Candidate genes for barley mutants involved in plant architecture: An in silico approach. *Theor. Appl. Genet.* 2006, 112, 1073–1085.
27. Konishi, T.; Hayashi, J.; Myriya, I.; Takahashi, R. Inheritance and linkage studies in barley. VII. Location of six new mutant genes on chromosome 3. *Ber. Ohara Inst. Landwirtsch. Biol.*

- Okayama Univ. 1984, 18, 251–264.
28. Kucera, J.; Lundqvist, U.; Gustafsson, A. Induction of brevistaristatum mutants in barley. *Hereditas* 1975, 80, 263–278.
 29. Persson, G. An attempt to find suitable genetic markers for the dense ear loci in barley II. *Hereditas* 1969, 63, 1–28.
 30. Robertson, D.W.; Immer, F.R.; WLebe, G.A.; Stevens, H.L. The location of two genes for mature plant characters in barley in linkage group I. *J. Am. Soc. Agron.* 1944, 36, 66–72.
 31. Huang, B.; Huang, D.; Hong, Z.; Owie, S.O.; Wu, W. Genetic analysis reveals four interacting loci underlying awn trait diversity in barley (*Hordeum vulgare*). *Sci. Rep.* 2020, 10, 12535.
 32. Huang, D.; Zheng, Q.; Melchikart, T.; Bekkaoui, Y.; Konkin, D.J.F.; Kagale, S.; Martucci, M.; You, F.M.; Clarke, M.; Adamski, N.M.; et al. Dominant inhibition of awn development by a putative zinc-finger transcriptional repressor expressed at the B1 locus in wheat. *New Phytol.* 2020, 225, 340–355.
 33. DeWitt, N.; Guedira, M.; Lauer, E.; Sarinelli, M.; Tyagi, P.; Fu, D.; Hao, Q.; Murphy, J.P.; Marshall, D.; Akhunova, A.; et al. Sequence-based mapping identifies a candidate transcription repressor underlying awn suppression at the B1 locus in wheat. *New Phytol.* 2020, 225, 326–339.
 34. Würschum, T.; Jähne, F.; Phillips, A.L.; Langer, S.M.; Longin, C.F.H.; Tucker, M.R.; Leiser, W.L. Misexpression of a transcriptional repressor candidate provides a molecular mechanism for the suppression of awns by Tipped 1 in wheat. *J. Exp. Bot.* 2020, 71, 3428–3436.
 35. Wang, D.; Yu, K.; Jin, D.; Sun, L.; Chu, J.; Wu, W.; Xin, P.; Gregova, E.; Li, X.; Sun, J.; et al. Natural variations in the promoter of Awn Length Inhibitor 1 (ALI-1) are associated with awn elongation and grain length in common wheat. *Plant J.* 2020, 101, 1075–1090.
 36. Niu, J.; Zheng, S.; Shi, X.; Si, Y.; Tian, S.; He, Y.; Ling, H. Fine mapping and characterization of the awn inhibitor B1 locus in common wheat (*Triticum aestivum* L.). *Crop J.* 2020, 8, 613–622.
 37. Myler, J.L. Awn inheritance in barley. *J. Agric. Res.* 1942, 65, 405–412.
 38. Litzenberger, S.C.; Green, J.M. Inheritance of awns in barley. *Agron. J.* 1951, 43, 117–123.
 39. Franckowiak, J.D. Revised linkage maps for morphological markers in barley, *Hordeum vulgare*. *Barley Genet. Newsl.* 1997, 26, 9–21.
 40. Sameri, M.; Takeda, K.; Komatsuda, T. Quantitative trait loci controlling agronomic traits in recombinant inbred lines from a cross of oriental- and occidental-type barley cultivars. *Breed. Sci.* 2006, 56, 243–252.
 41. Tsuchiya, T.; Singh, R.J. Another case of paracentric inversion in a genetic stock, Engleawnless. *Barley Genet. Newsl.* 1972, 2, 110–111.

42. Chen, G.D.; Li, H.B.; Zheng, Z.; Wei, Y.M.; Zheng, Y.L.; McIntyre, C.L.; Zhou, M.X.; Liu, C.J. Characterization of a QTL affecting spike morphology on the long arm of chromosome 3H in barley (*Hordeum vulgare* L.) based on near isogenic lines and a NIL-derived population. *Theor. Appl. Genet.* 2012, 125, 1385–1392.
43. Gruszka, D.; Szarejko, I.; Maluszynski, M. New allele of HvBRI1 gene encoding brassinosteroid receptor in barley. *J. Appl. Genet.* 2011, 52, 257–268.
44. Wang, J.M.; Yang, J.M.; McNeil, D.L.; Zho, M.X. Identification and molecular mapping of a dwarfing gene in barley (*Hordeum vulgare* L.) and its correlation with other agronomic traits. *Euphytica* 2010, 175, 331–342.
45. Ramage, R.T. Alleles at the short-awned loci lk2 on chromosome 1 and lk5 on chromosome 4. *Barley Genet. Newsl.* 1986, 16, 22–23.
46. Kjack, J.L.; Witters, R.E. Physiological activity of awns in isolines of Atlas barley 1. *Crop Sci.* 1974, 14, 243–248.
47. Maydup, M.L.; Antonietta, M.; Graciano, C.; Guiamet, J.J.; Tambussi, E.A. The contribution of the awns of bread wheat (*Triticum aestivum* L.) to grain filling: Responses to water deficit and the effects of awns on ear temperature and hydraulic conductance. *Field Crop. Res.* 2014, 167, 102–111.
48. Rebetzke, G.J.; Bonnett, D.G.; Reynolds, M.P. Awns reduce grain number to increase grain size and harvestable yield in irrigated and rainfed spring wheat. *J. Exp. Bot.* 2016, 67, 2573–2586.
49. Qualset, C.Q.; Schaller, C.W.; Williams, J.C. Performance of isogenic lines of barley as influenced by awn length, linkage blocks, and environment. *Crop Sci.* 1965, 5, 489–494.
50. Haas, M.; Schreiber, M.; Mascher, M. Domestication and crop evolution of wheat and barley: Genes, genomics, and future directions. *J. Integr. Plant Biol.* 2019, 61, 204–225.
51. Mulki, M.A.; Bi, X.; von Korff, M. FLOWERING LOCUS T3 controls spikelet initiation but not floral development. *Plant Physiol.* 2018, 178, 1170–1186.

Retrieved from <https://encyclopedia.pub/entry/history/show/37172>