Current Nanomaterials in Nanomedicine

Subjects: Nanoscience & Nanotechnology

Contributor: Chih-Hsin Lin

Anticancer nanomedicines have been studied over 30 years, but fewer than 10 formulations have been approved for clinical therapy today. Despite abundant options of anticancer drugs, it remains challenging to have agents specifically target cancer cells while reducing collateral toxicity to healthy tissue. Nanocompartments that can be selective toward points deeply within malignant tissues are a promising concept, but the heterogeneity of tumor tissue, inefficiency of cargo loading and releasing, and low uniformity of manufacture required from preclinical to commercialization are major obstacles. Technological advances have been made in this field, creating engineered nanomaterials with improved uniformity, flexibility of cargo loading, diversity of surface modification, and less inducible immune responses.

Keywords: nanoparticles; nanomedicines; nanomaterials; nanotechnology

1. Introduction

The National Institutes of Health (NIH) defines nanoparticles as structures ranging from 1 to 100 nm in at least one dimension, while current nanoparticles in therapeutic application are acceptable up to hundreds of nm. Considering the tissue junction between capillaries (150–200 µm), nanoscale structures exhibit unique properties to enhance reactive areas as well as across cell or tissue barriers [1]. For pharmacokinetic properties, the optimal size of nanoparticles is around 100 nm in a hydrodynamic diameter.

Currently, nanoparticles are applied to conventional drugs to improve their efficacy and reduce morbidity for advanced cancer therapies. Antitumor cargos are either capsuled or covalently linked to the nanocarrier. The advantage of covalent links is a precise number of therapeutical molecules for each nanoparticle, while the encapsulation of materials provides more flexibility. Many antitumor drugs are hydrophobic, posing challenges for physiological uptake (**Table 1**).

Table 1. Hydrophobic and hydrophilic anticancer drugs in clinical use.

Drug	Solubility (in Water; 25 °C)	Clinical Use
	Hydrophobic	
Docetaxel	insoluble (<0.3 µg/mL)	Breast, prostate, non-small cell lung cancer, carcinoma, and adenocarcinoma
Paclitaxel	insoluble (<0.3 µg/mL)	AIDS-related Kaposi sarcoma, breast, ovarian, and non-small cell lung cancer
Alitretinoin	0.6 μg/mL	Acute promyelocytic leukemia, and AIDS-related Kaposi sarcoma
Etoposide	0.03 mg/mL	Small cell lung and testicular cancer
Cisplatin	2.5 mg/ml	Testicular, ovarian, breast, glioblastoma, non-small cell lung cancer, malignant mesothelioma, and lymphoma
Methotrexate	2.6 mg/mL	ALL, breast, and lung, head and neck cancer, non-Hodgkin lymphoma, and osteosarcoma
Fludarabine	3.53 mg/mL	CLL
Doxorubicin	10 mg/mL	ALL, AML, neuroblastoma, soft tissue and bone sarcomas, breast, ovary, urinary bladder, thyroid, gastric, thyroid, gastric cancer, Hodgkin's disease
Irinotecan HCL	25 mg/mL	Colon, and rectal cancer
Cyclophosphamide	15.1 mg/mL	ALL, AML, CLL, CML, breast cancer, Hodgkin lymphoma, multiple myeloma, and neuroblastoma
Gemcitabine	51.3 mg/mL	Pancreatic, breast, ovarian, and non-small cell lung cancer

Drug Solubility (in Water; 25 °C) Clinical Use

Hydrophilic

Besides stabilizing anticancer agents, designed nanoparticles can also enhance the delivery efficacy by targeting cancer lesions. This concept led to variable nanoparticle designs fitting physicochemical properties via surface modification for a multitude of biomedical applications. The targeting ability of nanoparticles, either passive or active, is aimed for enhancement of drug concentration within the specific tissue of interest, such as tumors, while limiting toxicity to healthy organs. Passive targeting depends on pathophysiological characteristics of tumor vessels, enabling nanomaterials to accumulate in the microenvironment. In tumor tissue, fast angiogenesis with highly disorganized and loosened vessel structure leads to enlarged gap junctions between endothelial cells, resulting in enhanced permeability and retention (EPR) effect $^{[2]}$. The EPR effect allows diffusion of molecules less than 400 nm in diameter, which is suitable for nanoscale complex. The other phenomenon generally observed in tumor tissue is the Warburg effect, a local high metabolic and glycolysis rate result in an acidic environment $^{[3]}$. Designed pH-sensitive biocarrier could be stable at physiological pH = 7.4, but rapidly disassembled and released payload once it reaches an acidic microenvironment. Common design of pH-sensitive nanoparticle is based on polymers with pKa in the range of 6.5–7.2, such as poly(L-histidine) (PHis) and poly(β -amino esters).

Unlike passive targeting, active delivery incorporates other high-affinity molecules to recognize cells directly. Active targeting based on surface receptors on target cells has been widely explored since malignant cells upregulate certain tumor-preferred receptors. For example, transferrin receptor (TfR) and folate receptors (FRs) are physiologically expressed on various normal cells but overexpressed in many cancer types in response to their higher metabolic rate [4][5].

Conjugation is the process to join the recognition molecules with the therapeutical complex, including direct conjugation or indirect method via linker. One of the main challenges in conjugation design is homogeneity of the molecules. By using a hydrazone ligation, Dawson et al. synthesized viral nanoparticles and conjugated with VEGFR-1 ligand (F56f peptide) on benzaldehyde cowpea mosaic virus nanoparticle for tumor targeting and imaging ^[6]. Moreover, considering orientations of ligands or antibodies; thus, conjugation via linker chemistry is better than direct conjugation for targeting molecules to nanoparticle.

Overall, development of nanomedicine from past decades is a proof of concept to selectively increase the concentration of anticancer agents in tumor malignancy but minimize the side effect from healthy tissues (**Figure 1**).

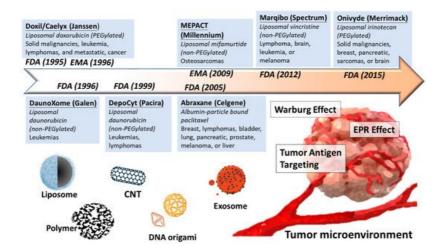


Figure 1. Development for nanomedicine reaching for tumor microenvironment. In past decades, plenty of nanocarriers are moving from preclinical bench work into clinical trial and finally approved for cancer therapy. The driving force of nanomedicine toward tumor microenvironment could be passive or active. Passive delivery relies on loose tumor vessels (EPR effect) and low pH (Warburg effect), while the active delivery can directly recognize tumor antigens by conjugating high-affinity molecules. Various novel and advanced materials of nanocarriers are designed for drug delivery, including liposome, polymer, CNT, DNA origami, and exosome.

2. Current Materials in Nanomedicine

2.1. Lipocomplex

Liposomal nanocomplex is the first delivery tool since the first discovery in the 1960s by A.D Bangham's group. Liposome formulation ranges from 50 to 200 nm with spherical vesicles composed of phospholipids, and steroids form bilayers in aqueous media can benefit as biocarriers [Z][8]. The properties of liposome were simply applied to increase the solubility of hydrophobic molecules and accelerate physiological metabolism in the beginning. For example, plenty of liposome formulations tried to fit numerous biochemical agents and provide less toxic than the free form. Liposomes were used to deliver lysophosphatidic acids and its analog which regulate normal or malignant blood cell differentiation and proliferation [9][10]. However, the liposomal formulations in this period faced a severe problem of short pharmacokinetic half-life, until the "stealth liposomes" was designed the 1990s. The second generation of liposome introduced the surface polyethylene glycol (PEG) coating, which highly improved stability and longer circulation time by alleviating the uptake of macrophages [11][12]. The PEGylation, constructed with a hydrophilic film on surface, can protect the liposome from clearance of reticuloendothelial system, making liposomal delivery clinical practical.

Several lipid complexes have been approved for clinical treatment after fifty years studying of lipocomplex (Table 2).

Table 2. Nanotherapeutics	approved for	oncological	therapy.
----------------------------------	--------------	-------------	----------

Name	ame Particle Base		Cancer Type	Approval
Liposome-based				
Doxil/Caelyx (Janssen)	PEGylated liposome	Doxorubicin	Ovarian, breast cancer, leukemia	FDA, 1995
DaunoXome (Galen)	Non-PEGylated liposome	Daunorubicin	HIV-related Kaposi sarcoma	FDA, 1996
DepoCyt (Pacira)	Non-PEGylated liposome	Cytarabine	AML, non-Hodgkin lymphoma	FDA,1999
Myocet (Teva UK)	Non-PEGylated liposome	Doxorubicin	Metastatic breast cancer	EMA, 2000
Marqibo (Spectrum)	Non-PEGylated liposome	Vincristine	Ph-ALL, Non-Hodgkin's lymphoma	FDA, 2012
Onivyde (Merrimack)	PEGylated liposome	irinotecan	Breast, pancreatic, sarcomas, or brain	FDA, 2015
Polymer-based				
Oncaspar (Sigma Tau)	PEGylation	L-asparaginase	ALL	FDA,1994
Abraxane (Celgene)	Albumin-bound polymer	Paclitaxel	Metastatic pancreatic cancer	FDA, 2005

2.2. Polymeric and Dendrimer Nanoparticles

Polymeric nanoparticles (PNPs) are structures with a diameter ranging from 10 to 100 nm, which was made from synthetic polymers (e.g., polycaprolactone and polyacrylate) or natural polymers (e.g., albumin, chitosan, and gelatin) [13] [14]. Clinical application of PNPs has reduced ionic surface to avoid the immunological response, while the immobilization of drug within PNPs can increase the drug stability as well. For example, docetaxel-loaded polymeric micelle (diameters < 30 nm) can reach poorly permeable pancreatic tumors in vivo [15]. The enhanced stability of the immobilized drug is attributed to the interaction with the polymer carriers to avoid the degradation. Once the complex reaches the target tissues, release mechanism would be triggered by tumor microenvironment. Several unique properties of tumor microenvironment have been used for cargo releasing, such as acidic, hyperthermia, and special enzymes secretion in the local environment. The pH-sensitive polymers are relative stable at a physiologic pH of 7.4 but can be rapidly destructured and can release active drugs in acidic tumor tissues. For example, poly(lactide-co-glycolide) (PLGA) polymer performed as 2–4-fold doxorubicin release in tumor-bearing tissue than circulation at pH 7.4 [16]. Moreover, thermosensitive polymeric, such as poly (N-isopropylacrylamide-co-allylamine) (PNIPAM-AAm-AA), could be a potential anticancer drug nanocarrier. Under the hyperthermia of tumor region, the hairy structure of PNIPAM-AAm-AA polymer would shrink, while the enclosed doxorubicin releases rapidly [17][18].

Dendrimer is a unique structure of polymer, which was first synthesized by Vogtle group in 1978, with branched 3D structure that provided a high degree of functional surface [19]. This multifunctional property provides the dendrimers more loading space for cargos and interaction with target cells. The cytotoxicity of dendrimer carrier depends on its surface area and the arms of dendrimer, while exchanging the amine groups into hydroxyl group may result in lower levels of cytotoxicity in vivo. The drug could be loaded into the internal structure of dendrimers or covalently linked to dendrimers molecule. Compared to the linear polymers with stochastic structures, dendrimers offer a well-defined size and structure,

performing a more precise polyvalence and molecular weight. The polyvalence defines the exact number of active groups on a single dendrimer. By controlling the number of covalent bonds within a single molecule, the quantity of drug loading could be adjusted. Noncovalent encapsulation is an alternative method only when payload is labile or poorly soluble. Poly(amido amide) (PAMAM), a very common dendrimer widely used in biomedical applications, is easily to have molecular conjugation through its branches of amine terminals [20]. Thioaptamer (TA)-modified PAMAM is developed to target CD44+ (TA receptor positive) breast cancer in vitro and in vivo by using ligand-receptor affinity [21]. Moreover, introducing a folic-acid conjugation has been reported to improve the delivery of PAMAM dendrimers loaded with 2-methoxyestradiol to target *KB* carcinoma cells overexpressing high-affinity folic acid receptors [22].

2.3. Carbon Nanomaterials

Carbon nanotubes (CNT), widely used as nanocarriers, are characterized by the unique structure with the rolling of a single (SWCNTs—single-walled carbon nanotubes) or multi (MWCNTs—multiwalled carbon nanotubes) sheet of graphite with an enormous surface area and an excellent electronic and thermal conductivity [23]. The compatibility of nanotube could improve biomedical reagent delivery with advanced chemical modification on its surface. SWCNT has a defined wall, whereas MWCNT mostly has structural defects which result in a less stable nanostructure [24]. SWCNTs is a one-dimensional nanomaterial composed of a single graphene layer of cylinder shape in a diameter of 1–2 nm and a length ranging from 50 nm to hundreds of µms. SWCNTs exhibit higher accumulation in tumor tissues physiologically, and their needle-like shape facilitates transmembrane penetration and internalization of therapeutic cargos. Moreover, a high surface area enhances ability to encapsulate and load cargo onto their surface or within their interior core via both covalent and noncovalent linkage. As drug carriers, there remain advantages and disadvantages of SWCNT relative to MWCNT. The stronger structure of SWCNT might be suitable for quality control of delivery, while the low stability of MWCNT makes it easier for further modification. Al Faraj et al. have recently demonstrated enhancement of delivery of doxorubicin by antibody-conjugated magnetic SWCNTs, which can also perform as a noninvasive imaging biomarker [25].

A. Pistone et al. have currently demonstrated hydroxyapatite-magnetite with MWCNT as a biocompatible magnetic drug delivery system in bone tissue engineering [27].

2.4. Nucleotide-Based Origami

DNA origami technique to build up uniform nanostructure was first named and introduced by PWK Rothemund in 2006. The method is to establish a scaffold which folds DNA into a desired shape using hundreds of short complement staple strands [28][29]. In the 2000s, DNA origami was widely investigated as candidates to serve as the next-generation drugdelivery vehicle [30]. Compared to other nanoscale methods for drug delivery, such as lipocomplex and inorganic nanoparticles, nucleotide-based origami performs several advantages: (i) uniformity of size, shape, and charge for each particle with self-assembled nanostructures; and (ii) precise control of the cargo loading on the scaffold by specific oligos or functional groups. The small DNA nanocarrier could serve as an effective delivery tool for anticancer drugs, RNA interference reagents, oligo-DNA, and antigen molecules, either in vitro or in vivo. Jiang et al. first showed a high level of doxorubicin loaded in DNA origami, and the complex exhibited prominent cytotoxicity in human breast cancer cells (MCF 7) and doxorubicin-resistant cancer cells [31].

2.5. Exosome-Derived Vehicle

Exosomes are cell-derived vesicles, ranging from 30 to 150 nm, that are present in many and perhaps all biological fluids for cellular communication. Exosomes were first described by Trams et al. and later substantiated by Johnstone et al., who observed intracellular interaction with small particles [32][33]. The main function of exosome was suggested as a route of cellular communication, which allows cells to exchange biomaterials, such as RNA, proteins, and lipid components. Since it is composed of partial cellular membrane, implying properties of high compatibility, low toxicity, and limited immunostimulation, exosomes are now regarded as a potential carrier of cargos to be delivered to the secondary cell. The lipid composition of exosome shares certain similarity to parental plasma membrane, but a different lipid raft composition with increase in sphingomyelin, phosphatidylserine, phosphatidylglycerol, lyso-phosphatidylethanolamine, and lyso-phosphatidylchoxline [34].

3. Conclusions

Nanocarriers are designed to improve the pharmacological and therapeutic properties from traditional free drugs. With growing knowledge of tumor heterogeneity and identified biomarkers, new nanomedicines are optimized with efficiency and selection to tumor lesions. From briefly prolonging circulation time to leading anticancer drugs toward lesions, the control of releasing would be the next step. Patients would benefit from the reduction of dosage index as concentrating therapeutic reagents pharmacologically to local tumor tissue and avoiding the universal side effect.

Increasing the need for a new strategy of disease treatment achieves the coordination of diagnosis and therapy by using advanced nanomaterials. The new direction of nanotechnology attempts to integrate therapeutics and diagnostics into a single nanomaterial, referred to as theranostics. The concept of theranostics provide the major applications in clinics which can improve targeted delivery, achieve gene delivery, and have the disease monitoring with the imaging platform by well-engineered nanoparticles. Currently, the pharmaceutical company Cristal Therapeutics is participating in a phase I clinical trial of CriPec® docetaxel combined with the imaging agent Zirconium-89 for PET imaging [35]. The platform evaluates the biodistribution and accumulation of the nanomedicine in solid tumors, leading to a better targeted therapy and follow-up prognosis. In the other clinical trial, Nanobiotix performs phase I/II trials for NBTXR3 comprising hafnium oxide nanoparticles as a radio-enhancer to kill tumor burden by locally additional radiation [36]. This also provides a new perspective to coordinate the imaging and radiology by advanced nanotechnology. Despite considerable development in this direction, nanomedicine of theranostics still faces challenges. The major challenge to successfully translate theranostic nanomedicine into routine clinics is the nano-bio interaction. The therapeutic nanoparticles generally have a larger window of treatment in patients which requires low tolerance of nano-bio interaction, while the diagnostic nanomaterials could be one-dose and real-time imaging every couple months. The cooperation of these different fields of nanomedicine requires further effort on developing innovative nanomaterials to achieve the goal.

Overall, most approved nanomedicines are those developed early and classic antineoplastic, meaning plenty of room for improvement. The next generation of nanomedicines will incorporate more diversity of new small-molecular compounds (pathway inhibitors, such as Rapamycin, a selective mTOR inhibitor) or gene therapeutic agents (siRNA, mRNA and gene editing). This flourishing field of nanoparticle delivery is expected to expand the versatility and potency of nanocarrier for cancer therapeutics. Given recent technical and material advancements in the past decades, smart and precise nanoparticles as drug carriers will revolutionize cancer therapy, not only significantly extending the patient's lifespan but improving their quality of life.

References

- 1. Capps, M.L.; Lawrence, I.E., Jr.; Burden, H.W. Cellular junctions in perifollicular contractile tissue of the rat ovary during the preovulatory period. Cell Tissue Res. 1981, 219, 133–141.
- 2. Maeda, H.; Matsumura, Y. Tumoritropic and lymphotropic principles of macromolecular drugs. Crit. Rev. Ther. Drug Car r. Syst. 1989, 6, 193–210.
- 3. Iyer, A.K.; Khaled, G.; Fang, J.; Maeda, H. Exploiting the enhanced permeability and retention effect for tumor targetin g. Drug Discov. Today 2006, 11, 812–818.
- 4. Cavanaugh, P.G.; Jia, L.; Zou, Y.; Nicolson, G.L. Transferrin receptor overexpression enhances transferrin responsiven ess and the metastatic growth of a rat mammary adenocarcinoma cell line. Breast Cancer Res. Treat. 1999, 56, 203–2
- 5. Lu, Y.; Sega, E.; Leamon, C.P.; Low, P.S. Folate receptor-targeted immunotherapy of cancer: Mechanism and therapeut ic potential. Adv. Drug Deliv. Rev. 2004, 56, 1161–1176.
- 6. Brunel, F.M.; Lewis, J.D.; Destito, G.; Steinmetz, N.F.; Manchester, M.; Stuhlmann, H.; Dawson, P.E. Hydrazone ligation strategy to assemble multifunctional viral nanoparticles for cell imaging and tumor targeting. Nano Lett. 2010, 10, 1093 –1097.
- 7. Bangham, A.D.; Horne, R.W.; Glauert, A.M.; Dingle, J.T.; Lucy, J.A. Action of saponin on biological cell membranes. Nat ure 1962, 196, 952–955.
- 8. Bangham, A.D.; Horne, R.W. Negative Staining of Phospholipids and Their Structural Modification by Surface-Active Agents as Observed in the Electron Microscope. J. Mol. Biol. 1964, 8, 660–668.
- 9. Benton, A.M.; Gerrard, J.M.; Michiel, T.; Kindom, S.E. Are lysophosphatidic acids or phosphatidic acids involved in stim ulus activation coupling in platelets? Blood 1982, 60, 642–649.
- Chiang, C.L.; Chen, S.S.A.; Lee, S.J.; Tsao, K.C.; Chu, P.L.; Wen, C.H.; Hwang, S.M.; Yao, C.L.; Lee, H. Lysophosphati dic Acid Induces Erythropoiesis through Activating Lysophosphatidic Acid Receptor 3. Stem Cells 2011, 29, 1763–177
 3.
- 11. Korgel, B.A.; van Zanten, J.H.; Monbouquette, H.G. Vesicle size distributions measured by flow field-flow fractionation c oupled with multiangle light scattering. Biophys. J. 1998, 74, 3264–3272.
- 12. Harris, J.M.; Martin, N.E.; Modi, M. Pegylation: A novel process for modifying pharmacokinetics. Clin. Pharmacokinet. 2 001, 40, 539–551.

- 13. Garay-Jimenez, J.C.; Turos, E. A convenient method to prepare emulsified polyacrylate nanoparticles from powders [co rrected] for drug delivery applications. Bioorganic Med. Chem. Lett. 2011, 21, 4589–4591.
- 14. Wong, B.S.; Teoh, S.H.; Kang, L. Polycaprolactone scaffold as targeted drug delivery system and cell attachment scaff old for postsurgical care of limb salvage. Drug Deliv. Transl. Res. 2012, 2, 272–283.
- 15. Li, Y.; Li, P.; Jin, M.; Jiang, C.; Gao, Z. Docetaxel-encapsulating small-sized polymeric micelles with higher permeability and its efficacy on the orthotopic transplantation model of pancreatic ductal adenocarcinoma. Int. J. Mol. Sci. 2014, 15, 23571–23588.
- 16. Hu, F.Q.; Zhang, Y.Y.; You, J.; Yuan, H.; Du, Y.Z. pH triggered doxorubicin delivery of PEGylated glycolipid conjugate mi celles for tumor targeting therapy. Mol. Pharm. 2012, 9, 2469–2478.
- 17. Shen, Z.; Wei, W.; Zhao, Y.J.; Ma, G.H.; Dobashi, T.; Maki, Y.; Su, Z.G.; Wan, J.P. Thermosensitive polymer-conjugated albumin nanospheres as thermal targeting anti-cancer drug carrier. Eur. J. Pharm. Sci. 2008, 35, 271–282.
- 18. Soleymani, M.; Edrissi, M.; Alizadeh, A.M. Thermosensitive polymer-coated La0.73Sr0.27MnO3 nanoparticles: Potentia I applications in cancer hyperthermia therapy and magnetically activated drug delivery systems. Polym. J. 2015, 47, 79 7–801.
- 19. Buhleier, E.; Wehner, W.; Vogtle, F. Cascade-Chain-Like and Nonskid-Chain-Like Syntheses of Molecular Cavity Topolo gies. Synthesis 1978, 2, 155–158.
- 20. Esfand, R.; Tomalia, D.A. Poly (amidoamine) (PAMAM) dendrimers: From biomimicry to drug delivery and biomedical a pplications. Drug Discov. Today 2001, 6, 427–436.
- 21. Fan, W.; Wang, X.; Ding, B.; Cai, H.; Wang, X.; Fan, Y.; Li, Y.; Liu, S.; Nie, S.; Lu, Q. Thioaptamer-conjugated CD44-tar geted delivery system for the treatment of breast cancer in vitro and in vivo. J. Drug Target. 2015, 1–13.
- 22. Wang, Y.; Guo, R.; Cao, X.; Shen, M.; Shi, X. Encapsulation of 2-methoxyestradiol within multifunctional poly (amidoam ine) dendrimers for targeted cancer therapy. Biomaterials 2011, 32, 3322–3329.
- 23. Madani, S.Y.; Mandel, A.; Seifalian, A.M. A concise review of carbon nanotube's toxicology. Nano Rev. 2013, 4, 21521.
- 24. He, H.; Pham-Huy, L.A.; Dramou, P.; Xiao, D.; Zuo, P.; Pham-Huy, C. Carbon nanotubes: Applications in pharmacy and medicine. BioMed Res. Int. 2013, 2013, 578290.
- 25. Al Faraj, A.; Cieslar, K.; Lacroix, G.; Gaillard, S.; Canot-Soulas, E.; Cremillieux, Y. In Vivo Imaging of Carbon Nanotube Biodistribution Using Magnetic Resonance Imaging. Nano Lett. 2009, 9, 1023–1027.
- 26. Al Faraj, A.; Shaik, A.P.; Shaik, A.S. Magnetic single-walled carbon nanotubes as efficient drug delivery nanocarriers in breast cancer murine model: Noninvasive monitoring using diffusion-weighted magnetic resonance imaging as sensitive imaging biomarker. Int. J. Nanomed. 2015, 10, 157–168.
- 27. Pistone, A.; Iannazzo, D.; Panseri, S.; Montesi, M.; Tampieri, A.; Galvagno, S. Hydroxyapatite-magnetite-MWCNT nano composite as a biocompatible multifunctional drug delivery system for bone tissue engineering. Nanotechnology 2014, 25, 425701.
- 28. O'Neill, P.; Rothemund, P.W.; Kumar, A.; Fygenson, D.K. Sturdier DNA nanotubes via ligation. Nano Lett. 2006, 6, 1379 –1383.
- 29. Rothemund, P.W. Folding DNA to create nanoscale shapes and patterns. Nature 2006, 440, 297-302.
- 30. Shih, W.M.; Quispe, J.D.; Joyce, G.F. A 1.7-kilobase single-stranded DNA that folds into a nanoscale octahedron. Natur e 2004, 427, 618–621.
- 31. Jiang, Q.; Song, C.; Nangreave, J.; Liu, X.; Lin, L.; Qiu, D.; Wang, Z.G.; Zou, G.; Liang, X.; Yan, H.; et al. DNA origami as a carrier for circumvention of drug resistance. J. Am. Chem. Soc. 2012, 134, 13396–13403.
- 32. Trams, E.G.; Lauter, C.J.; Salem, N.; Heine, U. Exfoliation of Membrane Ecto-Enzymes in the Form of Micro-Vesicles. Biochim. Biophys. Acta 1981, 645, 63–70.
- 33. Johnstone, R.M.; Adam, M.; Hammond, J.R.; Orr, L.; Turbide, C. Vesicle Formation during Reticulocyte Maturation—As sociation of Plasma-Membrane Activities with Released Vesicles (Exosomes). J. Biol. Chem. 1987, 262, 9412–9420.
- 34. Rappa, G.; Mercapide, J.; Anzanello, F.; Pope, R.M.; Lorico, A. Biochemical and biological characterization of exosome s containing prominin-1/CD133. Mol. Cancer 2013, 12, 1–7.
- 35. Zhang, Y.; Hong, H.; Cai, W. PET Tracers Based on Zirconium-89. Curr. Radiopharm. 2011, 4, 131-139.
- 36. Silva, C.O.; Pinho, J.O.; Lopes, J.M.; Almeida, A.J.; Gaspar, M.M.; Reis, C. Current trends in cancer nanotheranostics: Metallic, polymeric, and lipid-based systems. Pharmaceutics 2019, 11, 22.

Retrieved from https://encyclopedia.pub/entry/history/show/32371