Redefining Vaccine Effectiveness through Lessons from SARS-CoV-2

Subjects: Infectious Diseases

Contributor: Mohan E. Tulapurkar , Nouha H. Odeh , Abhay U. Andar , Renuka Roche , Joseph A. Roche

Anti-SARS-CoV-2 vaccines have played a pivotal role in reducing the risk of developing severe illness from COVID-19, thus helping end the COVID-19 global public health emergency. Intriguingly, as SARS-CoV-2 variants emerged, individuals who were fully vaccinated did get infected in high numbers, and viral loads in vaccinated individuals were as high as those in the unvaccinated. However, vaccines undoubtedly offered protection against severe illness even without conferring immunity in the classical sense. The lessons learned from anti-SARS-CoV-2 vaccines are a call to the medical community to revisit and redefine the concept of vaccine effectiveness. This endeavor is likely to increase increase vaccine confidence, and thus bolster global health education efforts and preventive care.

SARS-CoV-2 immunity

cytokines

infectious diseases

1. Introduction

On 5 May 2023, the World Health Organization (WHO) declared that coronavirus disease 2019 (COVID-19), which is linked to infection with the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) $^{[1]}$ is no longer a public health emergency of international concern (PHEIC) ^{[2][3][4]}. The WHO cautioned that the end of COVID-19's PHEIC status was not a license to stop being vigilant against the disease, but rather that COVID-19 should now be managed by local public health agencies along with other common infectious diseases [2][3][4]. The WHO justified its decision on the basis of data, which indicate that the proportion of infections leading to severe illness and deaths from COVID-19 have satisfactorily declined since the PHEIC was first declared in January of 2020 [2][3][4]. This is indeed a cause for celebration, given how overwhelmed healthcare delivery systems were during the early waves of COVID-19 [5][6][7][8][9][10][11][12][13]. The accelerated development, testing, and deployment of anti-SARS-CoV-2 vaccines (a.k.a., COVID-19 vaccines), which were deemed safe and effective by various local regulatory bodies, and ratified by the WHO and other global public health agencies, have played a pivotal role in reducing the likelihood of severe illness and death from COVID-19 [14][15]. Based on data from 185 countries, computational models suggest that during just the first year after the deployment of COVID-19 vaccines, there was about a 63% reduction in total deaths ^[16]. While celebrating the end of the COVID-19 PHEIC is warranted, it is important that the WHO and other health agencies critically assess the public health management of COVID-19 in order to more rapidly and effectively contain infectious disease outbreaks that might occur in the future 17.

2. SARS-CoV-2—Killer Virus or Just a Trigger for Kitchen Sink Inflammation?

SARS-CoV-2, which causes COVID-19, is a non-segmented, single-stranded, positive-sense RNA virus from the β genus of coronaviruses ^{[18][19]}. SARS-CoV-2 infection was first identified in Wuhan, Hubei Province, China in December of 2019, and was initially characterized as virally-induced pneumonia by clinicians before it was finally isolated in bronchoalveolar lavage fluid from patients ^{[19][20]}. The clinical presentation of COVID-19 ranges from asymptomatic infection to mild respiratory symptoms to severe viral pneumonia ^[21].

While SARS-CoV-2 initially infects and compromises the respiratory system, it also induces multiorgan dysfunction and damage ^{[22][23][24]}. One of the main factors implicated in COVID-19 multiorgan failure is a massive release of proinflammatory cytokines—formally known as cytokine release syndrome (CRS) but better known by the colloquial term "cytokine storms" ^{[25][26]}. The inflammatory reaction in COVID-19 is due to the overactivation of multiple cellular subtypes in the human body ^[27]. Inhaled viral particles bind to epithelial cells in the nasal mucosa or travel down the nasopharyngeal tract to reach the more distal areas of the airway. The effect of the virus in eliciting an inflammatory response through different cell types is detailed below, and this gives us an idea of how the virus wreaks havoc on homeostasis in the host. Local and systemic inflammation followed by systemic disruption of homeostasis leads to multiorgan symptoms, multiorgan damage, and the high case fatality rate associated with SARS-CoV-2 infection (**Figure 1**).

2.1. Epithelial System

The epithelial cells are the first line of defense against invading pathogens ^[28]. The viral particles encounter different kinds of epithelial cells as they travel from the nose and mouth, which are the most common points of entry, all the way down to the alveolar sacs—the sites of gaseous exchange (Figure 1) [29]. The nasal mucosa has different cell types, such as ciliated epithelial cells, mucous cells, and basal cells ^[30]. These cells express angiotensin converting enzyme-2 (ACE2) and the transmembrane protein serine protease-2 (TMPRSS2) on their plasma membranes [31][32][33]. ACE2, TMPRSS2, and other plasma membrane proteins aid in the docking of SARS-CoV-2 onto host cells and facilitate endocytosis of the virus ^[32]. Once the virus has entered host cells, the virus releases its genetic material into the cytoplasm and hijacks the host's cellular machinery to produce more viral particles [34][35] i.e., the cell is now infected with the virus. The release of viral particles from cells has been detected as early as one hour post-infection; however, 6-8 h post-infection is when significant viral load has been detected and infection of neighboring cells and loss of ciliated epithelium has been observed ^[36]. The loss of cilia on the epithelium results in reduced mucociliary clearance, which leads to increased local infection and progression of the disease [37][38]. This loss of clearance has been implicated in increased COVID-19 severity in individuals with pre-existing pulmonary inflammatory conditions like asthma ^[39], cystic fibrosis ^[40], and chronic obstructive pulmonary disease (COPD), where ciliary function is already impaired [41]. In nasal epithelial cells, there is a delayed or a muted interferon response that is observed after infection. The levels of type I (interferon alpha, beta) and type III (interferon lambda) interferons are lower in severe COVID-19 compared to mild cases [42][43][44].

As the virus moves down the airway, it encounters the bronchial epithelial cells, the mucus cells, and the club cells ^[30] (**Figure 1**). In the middle and lower airway, ciliated epithelial cells that become infected lose their cilia and have a denuded appearance, and the levels of proinflammatory signals in these cells correlate with the disease severity ^[45].

When the virus finally reaches the alveoli (the distal-most regions of the airway) where the epithelium, endothelium, and blood cells interface and enable gas exchange [46], it activates numerous signaling cascades in multiple cell types, which cause the most damage and destruction [47] (Figure 1). In alveoli, the virus primarily infects alveolar type 2 epithelial cells (a.k.a., AT2 cells or type 2 pneumocytes), potentially due to the abundance of ACE2 on their surface [48]. Infection of type 2 pneumocytes seems to be the initial step that triggers a domino of inflammatory signals ^[49]. It has been shown that alveolar type 1 cells can also be infected, but to a lesser extent ^[50]. The infection and damage of alveolar pneumocytes results in a cascade of physiological changes, beginning with an increased production of proinflammatory cytokines, such as interleukin (IL)-1 (IL-1), IL-6, IL-8, tumor necrosis factor-alpha (TNF-α), and elevated levels of C-reactive protein (CRP) and D-dimer [51][52][53][54][55]. These inflammatory cytokines result in recruitment of immune cells-mainly neutrophils and macrophages to loci of inflammation. The recruitment of immune cells further exacerbates the situation, as it results in the loss of barrier function of the underlying endothelial layer [56][57]. Alveolar macrophages, which are recruited to sites of damage have been shown to produce various chemokines, such as CCL2, CCL3, CCL7, CCL8, CCL13, CCL20, and cytokines, such as CXCL1, CXCL3, and CXCL10 [58][59]. Among these chemokines, CCL2 and CCL3 attract more monocytes and macrophages to the alveoli and induce CXCR1 gene expression in them; this promotes the production of tissue-damaging, proinflammatory reactive oxygen species ^[60]. The sum effect is largescale destruction of epithelial tissue, which promotes the recruitment of neutrophils that try to resolve the situation by forming neutrophil extracellular traps (NET) ^[61]. The heightened cellular signaling and subsequent tissue damage described above leads to alveolar flooding with the interstitial fluid and the blood, which leads to progressive hypoxia. Thus, the replication of the virus in the alveoli results in the progression towards acute respiratory distress syndrome (ARDS), triggering an imbalance between pro-coagulation and anticoagulation (i.e., pro-fibrinolysis) pathways, plus stimulating complement activation, damage to hyaline membranes, and the formation of clots in the small and large blood vessels [62][63].

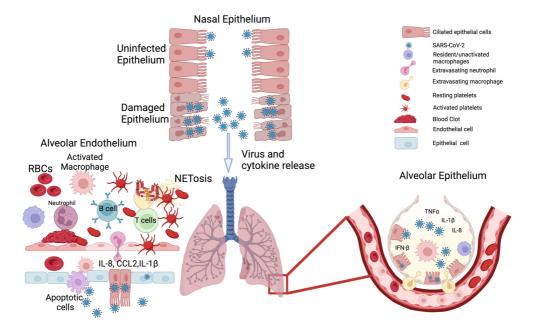


Figure 1. Pathophysiology of COVID-19 and "kitchen sink inflammation".

A. Nasal epithelium. The primary site of entry for SARS-CoV-2 is the nasal epithelium along with the oral mucosa. The virus binds to ciliated epithelial cells via ACE2 and is endocytosed, thus infecting the host cells. The virus uses host cell machinery to replicate and release more viral particles. The release of the viral particles results in the loss of epithelial barrier integrity, the release of inflammatory mediators, and the aggravation of symptoms. B. Alveolar epithelium. The Alveolar epithelium consists of ciliated type 2 alveolar epithelial cells (AT2) and type 1 non-ciliated (AT1) cells. Infection of the epithelial cells results in the production of a multitude of proinflammatory cytokines that promote extravasation/activation of the neutrophils and macrophages. This results in loss of barrier integrity, alveolar flooding, loss of functional surfactant, diffuse alveolar damage, and damage to the underlying endothelial cells. C. Endothelial cells. The loss of barrier integrity on the epithelial layer promotes further spread of damage to the endothelial layer. Secreted proinflammatory cytokines and chemokines recruit and activate a variety of immune cells (macrophages, neutrophils, and T and B cells). The exposed extracellular matrix also acts as a trigger for neutrophils, which undergo a process of cell death called NETosis; this leads to the aggregation and entanglement of platelets, thus promoting clot formation. Furthermore, T cells that are recruited to the site of injury can also aggravate endothelial injury.

2.2. Endothelial System

The endothelial cells line the innermost layers of the blood vessels (adjacent to the lumen) and tightly regulate the transport of nutrients, gases, metabolic wastes, bioactive molecules, and cells ^[64]. The endothelial cells lining the smaller capillaries express ACE2 and TMPRSS2, which as mentioned earlier are necessary for the host cell binding and internalization of SARS-CoV-2 ^[65]. The virus, on entering the endothelial cells, replicates and then disseminates through the blood stream to organs other than the lungs. The endothelial cells respond to the plethora of cytokines/chemokines that are produced by the epithelial cells upon infection. The predominant chemical signals that affect endothelial barrier function are IL-6 and TNF- α . Activation of the endothelial cells by IL-

6 or TNF-α results in the production of IL-8 (a major chemoattractant for neutrophils) and monocyte chemoattractant protein (MCP-1) and in the activation of the C5a complement ^[66]. Indeed, plasma IL-6 levels in patients with COVID-19 correlate with the disease severity ^[67]. The recruitment of overly activated immune cells, a hallmark of COVID-19, can result in endothelial cell death, which promotes vascular leak and initiation of microfoci for clot formation ^{[68][69]}. Activated endothelial cells can release large amounts of von Willebrand factor (VWF) and factor VIII, which play an active role in clot formation ^[70]. In addition to stimulating the formation of fibrin, endothelial cells also secrete plasminogen activator inhibitor-1 (PAI-1), which inhibits clot dissolution ^[71]. During the initial stage of infection, the loss of contact with the surrounding cells results in the formation of circulating endothelial cells (CECs), which can travel from one tissue to another. During the course of infection, there is a loss of endothelial barrier integrity, which results in fluid accumulation in the alveolar and pleural spaces. This is a result of the upregulation of interleukin 2 receptor (IL-2R) on the endothelial cells and the increased release of IL-2 from the activated T cells ^[72]. Thus, direct viral infection or indirect activation of endothelial cells causes a loss of barrier function, which then results in fluid accumulation, increased extravasation of immune cells, widespread microthrombosis, diffuse fibrin deposits, and secretion of PAI-1 that prevents clot dissolution—all working together to increase the likelihood of thromboembolisms ^{[73][74]} (Figure 1).

2.3. The Mismatch between Viral Load and Symptom Severity

We attempted to answer the question as to whether SARS-CoV-2 is a killer virus or just a trigger for "kitchen sink inflammation". The related expressions "to throw everything but the kitchen sink" or "to throw the kitchen sink" imply doing everything possible to address a problem, regardless of whether the solutions are likely to be good or bad [75] ^[76]. Due to the novelty of SARS-CoV-2 to the human immune system, when people were first infected, the immune response was indeed comparable to throwing an inflammatory kitchen sink at the viral invader, resulting in cellsignaling storms ^{[25][26][47][55][77][78][79][80][81][82][83]}. The fact that some individuals with high viral loads presented with minimal symptoms, and some individuals with severe symptoms had low viral loads suggest that, SARS-CoV-2 by itself is not a killer virus, but rather, it is a nonspecific and overly aggressive inflammatory response, which causes the most harm [84][85][86][87][88]. This is not to diminish the fact that SARS-CoV-2 infection has been implicated in the deaths of nearly seven million people, chronic illness in 10-20% of the survivors, and about 800 million cumulative cases in just over three years [89][90]. However, the available data point towards severe illness and deaths being linked to an inflammatory response, which overwhelms the host, rather than widespread virus-mediated killing of cells in the respiratory system and other vital systems of the body [88]. Interestingly, an overly aggressive immune response, rather than the pathogen itself, might underlie severe illness in various infectious diseases [91]. Therefore, public health agencies must invest resources in finding safe, effective, practical, and standardized therapies, which can work to combat a wide range of existing and novel pathogens. In hindsight, one of the simplest public health strategies to blunt the impact of SARS-CoV-2 would have been to assume that it would behave just like its predecessor SARS-CoV (implicated in the SARS outbreak of the early 2000s) and amplify the importance of commonsense preventative healthcare measures (e.g., hand hygiene, mask wearing, and avoiding unnecessary social activities) until safe and effective pharmacological measures were available [92][93][94].

The recognition of "self" from "non-self" is a fundamental aspect of immunity, and this function serves an important role in protecting the body from disease-causing elements ^[95]. Vaccination takes advantage of this by introducing a harmless version of the disease-causing pathogen, allowing the body to recognize and remember it. There are two phases of immunity that are activated upon infection or inoculation through vaccination: Innate immunity and adaptive immunity ^{[96][97][98]} (**Figure 2**).

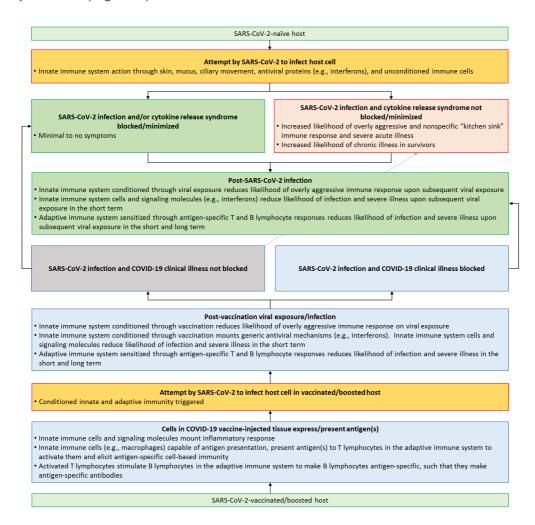


Figure 2. The role of vaccine-induced immune system conditioning in reducing COVID-19 illness severity.

References

- 1. Hu, B.; Guo, H.; Zhou, P.; Shi, Z.L. Characteristics of SARS-CoV-2 and COVID-19. Nat. Rev. Microbiol. 2021, 19, 141–154.
- World Health Organization (WHO). Statement on the Fifteenth Meeting of the Ihr (2005) Emergency Committee on the COVID-19 Pandemic; World Health Organization (WHO): Geneva, Switzerland, 2005; Available online: https://www.who.int/news/item/05-05-2023-statement-on-thefifteenth-meeting-of-the-international-health-regulations-(2005)-emergency-committee-regardingthe-coronavirus-disease-(COVID-19)-pandemic (accessed on 11 May 2023).

- 3. Wise, J. COVID-19: Who Declares End of Global Health Emergency. Bmj 2023, 381, 1041.
- 4. Lenharo, M. Who Declares End to COVID-19's Emergency Phase. Nature 2023.
- Ranney, M.L.; Valerie Griffeth, M.P.H.; Jha, A.K. Critical Supply Shortages—The Need for Ventilators and Personal Protective Equipment during the COVID-19 Pandemic. N. Engl. J. Med. 2020, 382, e41.
- Dey, A.K.; Bhan, N.; Rao, N.; Ghule, M.; Chatterji, S.; Raj, A. Factors Affecting Delayed and Non-Receipt of Healthcare During the COVID-19 Pandemic for Women in Rural Maharashtra, India: Evidence from a Cross-Sectional Study. EClinicalMedicine 2022, 53, 101741.
- Rogers, E.H.; Porter, J.N.; Ren, J.J.; Battista, J.J.; Grewal, N.P.; Valentin, N.I. Experiences from the Epicenter: An Observational Report on Converting a Post Anesthesia Care Unit to a COVID-19 Intensive Care Unit. Int. J. Surg. Glob. Health 2020, 3, e20.
- Davis, B.; Bankhead-Kendall, B.K.; Dumas, R.P. A Review of COVID-19's Impact on Modern Medical Systems from a Health Organization Management Perspective. Health Technol. 2022, 12, 815–824.
- Parotto, E.; Lamberti-Castronuovo, A.; Censi, V.; Valente, M.; Atzori, A.; Ragazzoni, L. Exploring Italian Healthcare Facilities Response to COVID-19 Pandemic: Lessons Learned from the Italian Response to COVID-19 Initiative. Front. Public Health 2022, 10, 1016649.
- 10. The Lancet Public, Health. COVID-19 in Spain: A Predictable Storm? Lancet Public Health 2020, 5, e568.
- 11. Sharma, A.; Gupta, P.; Jha, R. COVID-19: Impact on Health Supply Chain and Lessons to Be Learnt. J. Health Manag. 2020, 22, 248–261.
- 12. Usher, A.D. Medical Oxygen Crisis: A Belated COVID-19 Response. Lancet 2021, 397, 868–869.
- 13. Feinmann, J. How COVID-19 Revealed the Scandal of Medical Oxygen Supplies Worldwide. Bmj 2021, 373, n1166.
- Tenforde, M.W.; Self, W.H.; Adams, K.; Gaglani, M.; Ginde, A.A.; McNeal, T.; Ghamande, S.; Douin, D.J.; Talbot, H.K.; Casey, J.D.; et al. Association between Mrna Vaccination and COVID-19 Hospitalization and Disease Severity. JAMA 2021, 326, 2043–2054.
- Moghadas, S.M.; Vilches, T.N.; Zhang, K.; Wells, C.R.; Shoukat, A.; Singer, B.H.; Meyers, L.A.; Neuzil, K.M.; Langley, J.M.; Fitzpatrick, M.C.; et al. The Impact of Vaccination on COVID-19 Outbreaks in the United States. medRxiv 2021.
- Watson, O.J.; Barnsley, G.; Toor, J.; Hogan, A.B.; Winskill, P.; Ghani, A.C. Ghani. Global Impact of the First Year of COVID-19 Vaccination: A Mathematical Modelling Study. Lancet Infect. Dis. 2022, 22, 1293–1302.

- Torreele, E.; Kazatchkine, M.; Liu, J.; Dybul, M.; Cárdenas, M.; Singh, S.; Quigley, H.L.; McNab, C.; Sirleaf, E.J.; Mazzucato, M.; et al. Stopping Epidemics When and Where They Occur. Lancet 2023, 401, 324–328.
- Samudrala, P.K.; Kumar, P.; Choudhary, K.; Thakur, N.; Wadekar, G.S.; Dayaramani, R.; Agrawal, M.; Alexander, A. Virology, Pathogenesis, Diagnosis and in-Line Treatment of COVID-19. Eur. J. Pharmacol. 2020, 883, 173375.
- 19. Jin, Y.; Yang, H.; Ji, W.; Wu, W.; Chen, S.; Zhang, W.; Duan, G. Virology, Epidemiology, Pathogenesis, and Control of COVID-19. Viruses 2020, 12, 372.
- 20. Ciotti, M.; Ciccozzi, M.; Terrinoni, A.; Jiang, W.C.; Wang, C.B.; Bernardini, S. The COVID-19 Pandemic. Crit. Rev. Clin. Lab Sci. 2020, 57, 365–388.
- 21. Huang, H.; Cai, S.; Li, Y.; Li, Y.; Fan, Y.; Li, L.; Lei, C.; Tang, X.; Hu, F.; Li, F.; et al. Prognostic Factors for COVID-19 Pneumonia Progression to Severe Symptoms Based on Earlier Clinical Features: A Retrospective Analysis. Front. Med. 2020, 7, 557453.
- Morrow, A.J.; Sykes, R.; McIntosh, A.; Kamdar, A.; Bagot, C.; Bayes, H.K.; Blyth, K.G.; Briscoe, M.; Bulluck, H.; Carrick, D.; et al. A Multisystem, Cardio-Renal Investigation of Post-COVID-19 Illness. Nat. Med. 2022, 28, 1303–1313.
- Gupta, A.; Madhavan, M.V.; Sehgal, K.; Nair, N.; Mahajan, S.; Sehrawat, T.S.; Bikdeli, B.; Ahluwalia, N.; Ausiello, J.C.; Wan, E.Y.; et al. Extrapulmonary Manifestations of COVID-19. Nat. Med. 2020, 26, 1017–1032.
- 24. Mokhtari, T.; Hassani, F.; Ghaffari, N.; Ebrahimi, B.; Yarahmadi, A.; Hassanzadeh, G. COVID-19 and multiorgan failure: A narrative review on potential mechanisms. Histochem. J. 2020, 51, 613– 628.
- Que, Y.; Hu, C.; Wan, K.; Hu, P.; Wang, R.; Luo, J.; Li, T.; Ping, R.; Hu, Q.; Sun, Y.; et al. Cytokine Release Syndrome in COVID-19: A Major Mechanism of Morbidity and Mortality. Int. Rev. Immunol. 2022, 41, 217–230.
- Montazersaheb, S.; Khatibi, S.M.H.; Hejazi, M.S.; Tarhriz, V.; Farjami, A.; Sorbeni, F.G.; Farahzadi, R.; Ghasemnejad, T. Ghasemian Sorbeni, R. Farahzadi, and T. Ghasemnejad. COVID-19 Infection: An Overview on Cytokine Storm and Related Interventions. Virol. J. 2022, 19, 92.
- 27. Mattoo SU, S.; Kim, S.J.; Ahn, D.G.; Myoung, J. Escape and over-Activation of Innate Immune Responses by SARS-CoV-2: Two Faces of a Coin. Viruses 2022, 14, 530.
- 28. Marshall, J.S.; Warrington, R.; Watson, W.; Kim, H.L. An Introduction to Immunology and Immunopathology. Allergy Asthma Clin. Immunol. 2018, 14, 49.

- 29. Murgolo, N.; Therien, A.G.; Howell, B.; Klein, D.; Koeplinger, K.; Lieberman, L.A.; Adam, G.C.; Flynn, J.; McKenna, P.; Swaminathan, G.; et al. SARS-CoV-2 tropism, entry, replication, and propagation: Considerations for drug discovery and development. PLoS Pathog. 2021, 17, e1009225.
- 30. Gengler, I.; Wang, J.C.; Speth, M.M.; Sedaghat, A.R. Sinonasal pathophysiology of SARS-CoV-2 and COVID-19: A systematic review of the current evidence. Laryngoscope Investig. Otolaryngol. 2020, 5, 354–359.
- Wruck, W.; Adjaye, J. SARS-CoV-2 receptor ACE2 is co-expressed with genes related to transmembrane serine proteases, viral entry, immunity and cellular stress. Sci. Rep. 2020, 10, 21415.
- Sungnak, W.; Huang, N.; Becavin, C.; Berg, M.; Queen, R.; Litvinukova, M.; Talavera-Lopez, C.; Maatz, H.; Reichart, D.; Sampaziotis, F.; et al. SARS-CoV-2 entry factors are highly expressed in nasal epithelial cells together with innate immune genes. Nat. Med. 2020, 26, 681–687.
- 33. Collin, J.; Queen, R.; Zerti, D.; Dorgau, B.; Georgiou, M.; Djidrovski, I.; Hussain, R.; Coxhead, J.M.; Joseph, A.; Rooney, P.; et al. Co-expression of SARS-CoV-2 entry genes in the superficial adult human conjunctival, limbal and corneal epithelium suggests an additional route of entry via the ocular surface. Ocul. Surf. 2021, 19, 190–200.
- 34. V'kovski, P.; Kratzel, A.; Steiner, S.; Stalder, H.; Thiel, V. Coronavirus biology and replication: Implications for SARS-CoV-2. Nat. Rev. Microbiol. 2021, 19, 155–170.
- 35. Hillen, H.S.; Kokic, G.; Farnung, L.; Dienemann, C.; Tegunov, D.; Cramer, P. Structure of replicating SARS-CoV-2 polymerase. Nature 2020, 584, 154–156.
- 36. Gamage, A.M.; Tan, K.S.; Chan, W.O.Y.; Liu, J.; Tan, C.W.; Ong, Y.K.; Thong, M.; Andiappan, A.K.; Anderson, D.E.; Wang, Y.; et al. Infection of human Nasal Epithelial Cells with SARS-CoV-2 and a 382-nt deletion isolate lacking ORF8 reveals similar viral kinetics and host transcriptional profiles. PLoS Pathog. 2020, 16, e1009130.
- 37. Zhu, N.; Wang, W.; Liu, Z.; Liang, C.; Wang, W.; Ye, F.; Huang, B.; Zhao, L.; Wang, H.; Zhou, W.; et al. Morphogenesis and cytopathic effect of SARS-CoV-2 infection in human airway epithelial cells. Nat. Commun. 2020, 11, 3910.
- 38. Ozturk, E.O.; Aslan, M.; Bayindir, T. The effect of COVID-19 on nasal mucociliary clearance. Acta Otolaryngol. 2022, 142, 329–332.
- Thomas, B.; Rutman, A.; Hirst, R.A.; Haldar, P.; Wardlaw, A.J.; Bankart, J.; Brightling, C.E.; O'Callaghan, C. Ciliary dysfunction and ultrastructural abnormalities are features of severe asthma. J. Allergy Clin. Immunol. 2010, 126, 722–729.e2.
- 40. Leung, H.M.; Birket, S.E.; Hyun, C.; Ford, T.N.; Cui, D.; Solomon, G.M.; Shei, R.J.; Adewale, A.T.; Lenzie, A.R.; Fernandez-Petty, C.M.; et al. Intranasal micro-optical coherence tomography

imaging for cystic fibrosis studies. Sci. Transl. Med. 2019, 11, eaav3505.

- 41. Yaghi, A.; Zaman, A.; Cox, G.; Dolovich, M.B. Ciliary beating is depressed in nasal cilia from chronic obstructive pulmonary disease subjects. Respir. Med. 2012, 106, 1139–1147.
- Fiege, J.K.; Thiede, J.M.; Nanda, H.A.; Matchett, W.E.; Moore, P.J.; Montanari, N.R.; Thielen, B.K.; Daniel, J.; Stanley, E.; Hunter, R.C.; et al. Single cell resolution of SARS-CoV-2 tropism, antiviral responses, and susceptibility to therapies in primary human airway epithelium. PLoS Pathog. 2021, 17, e1009292.
- Galani, I.-E.; Rovina, N.; Lampropoulou, V.; Triantafyllia, V.; Manioudaki, M.; Pavlos, E.; Koukaki, E.; Fragkou, P.C.; Panou, V.; Rapti, V.; et al. Untuned antiviral immunity in COVID-19 revealed by temporal type I/III interferon patterns and flu comparison. Nat. Immunol. 2021, 22, 32–40.
- 44. Kim, Y.-M.; Shin, E.-C. Type I and III interferon responses in SARS-CoV-2 infection. Exp. Mol. Med. 2021, 53, 750–760.
- 45. Ziegler, C.G.; Miao, V.N.; Owings, A.H.; Navia, A.W.; Tang, Y.; Bromley, J.D.; Lotfy, P.; Sloan, M.; Laird, H.; Williams, H.B. Impaired local intrinsic immunity to SARS-CoV-2 infection in severe COVID-19. Cell 2021, 184, 4713–4733.e4722.
- 46. Lamers, M.M.; Haagmans, B.L. SARS-CoV-2 pathogenesis. Nat. Rev. Microbiol. 2022, 20, 270–284.
- 47. Bosmuller, H.; Matter, M.; Fend, F.; Tzankov, A. The pulmonary pathology of COVID-19. Virchows Arch. 2021, 478, 137–150.
- 48. Zou, X.; Chen, K.; Zou, J.; Han, P.; Hao, J.; Han, Z. Single-cell RNA-seq data analysis on the receptor ACE2 expression reveals the potential risk of different human organs vulnerable to 2019nCoV infection. Front. Med. 2020, 14, 185–192.
- Huang, J.; Hume, A.J.; Abo, K.M.; Werder, R.B.; Villacorta-Martin, C.; Alysandratos, K.D.; Beermann, M.L.; Simone-Roach, C.; Lindstrom-Vautrin, J.; Olejnik, J.; et al. SARS-CoV-2 Infection of Pluripotent Stem Cell-Derived Human Lung Alveolar Type 2 Cells Elicits a Rapid Epithelial-Intrinsic Inflammatory Response. Cell Stem Cell 2020, 27, 962–973.e967.
- 50. Rockx, B.; Kuiken, T.; Herfst, S.; Bestebroer, T.; Lamers, M.M.; Oude Munnink, B.B.; de Meulder, D.; van Amerongen, G.; van den Brand, J.; Okba, N.M.A.; et al. Comparative pathogenesis of COVID-19, MERS, and SARS in a nonhuman primate model. Science 2020, 368, 1012–1015.
- Queiroz, M.A.F.; Neves, P.; Lima, S.S.; Lopes, J.D.C.; Torres, M.; Vallinoto, I.; Bichara, C.D.A.; Dos Santos, E.F.; de Brito, M.; da Silva, A.L.S.; et al. Cytokine Profiles Associated with Acute COVID-19 and Long COVID-19 Syndrome. Front. Cell Infect. Microbiol. 2022, 12, 922422.
- 52. Wilson, J.G.; Simpson, L.J.; Ferreira, A.-M.; Rustagi, A.; Roque, J.; Asuni, A.; Ranganath, T.; Grant, P.M.; Subramanian, A.; Rosenberg-Hasson, Y.; et al. Cytokine profile in plasma of severe

COVID-19 does not differ from ARDS and sepsis. JCI Insight 2020, 5, 289.

- Huang, I.; Pranata, R.; Lim, M.A.; Oehadian, A.; Alisjahbana, B. C-reactive protein, procalcitonin, D-dimer, and ferritin in severe coronavirus disease-2019: A meta-analysis. Ther. Adv. Respir. Dis. 2020, 14, 1753466620937175.
- Bello, S.; Lasierra, A.B.; López-Vergara, L.; de Diego, C.; Torralba, L.; de Gopegui, P.R.; Lahoz, R.; Abadía, C.; Godino, J.; Cebollada, A.; et al. IL-6 and cfDNA monitoring throughout COVID-19 hospitalization are accurate markers of its outcomes. Respir. Res. 2023, 24, 125.
- 55. Frisoni, P.; Neri, M.; D'Errico, S.; Alfieri, L.; Bonuccelli, D.; Cingolani, M.; Di Paolo, M.; Gaudio, R.M.; Lestani, M.; Marti, M.; et al. Cytokine storm and histopathological findings in 60 cases of COVID-19-related death: From viral load research to immunohistochemical quantification of major players IL-1β, IL-6, IL-15 and TNF-α. Forensic. Sci. Med. Pathol. 2022, 18, 4–19.
- 56. Meyer, N.J.; Gattinoni, L.; Calfee, C.S. Acute respiratory distress syndrome. Lancet 2021, 398, 622–637.
- 57. Millar, F.R.; Summers, C.; Griffiths, M.J.; Toshner, M.R.; Proudfoot, A.G. The pulmonary endothelium in acute respiratory distress syndrome: Insights and therapeutic opportunities. Thorax 2016, 71, 462–473.
- 58. Grant, R.A.; Morales-Nebreda, L.; Markov, N.S.; Swaminathan, S.; Querrey, M.; Guzman, E.R.; Abbott, D.A.; Donnelly, H.K.; Donayre, A.; Goldberg, I.A.; et al. Circuits between infected macrophages and T cells in SARS-CoV-2 pneumonia. Nature 2021, 590, 635–641.
- 59. Chua, R.L.; Lukassen, S.; Trump, S.; Hennig, B.P.; Wendisch, D.; Pott, F.; Debnath, O.; Thurmann, L.; Kurth, F.; Volker, M.T.; et al. COVID-19 severity correlates with airway epitheliumimmune cell interactions identified by single-cell analysis. Nat. Biotechnol. 2020, 38, 970–979.
- 60. Rendeiro, A.F.; Ravichandran, H.; Bram, Y.; Chandar, V.; Kim, J.; Meydan, C.; Park, J.; Foox, J.; Hether, T.; Warren, S.; et al. The spatial landscape of lung pathology during COVID-19 progression. Nature 2021, 593, 564–569.
- 61. Galani, I.E.; Andreakos, E. Neutrophils in viral infections: Current concepts and caveats. J. Leukoc. Biol. 2015, 98, 557–564.
- 62. Biswas, S.; Thakur, V.; Kaur, P.; Khan, A.; Kulshrestha, S.; Kumar, P. Blood clots in COVID-19 patients: Simplifying the curious mystery. Med. Hypotheses 2021, 146, 110371.
- Tuculeanu, G.; Barbu, E.C.; Lazar, M.; Chitu-Tisu, C.E.; Moisa, E.; Negoita, S.I.; Ion, D.A. Coagulation Disorders in Sepsis and COVID-19—Two Sides of the Same Coin? A Review of Inflammation–Coagulation Crosstalk in Bacterial Sepsis and COVID-19. J. Clin. Med. 2023, 12, 601.

- 64. Bkaily, G.; Jacques, D. Morphological and Functional Remodeling of Vascular Endothelium in Cardiovascular Diseases. Int. J. Mol. Sci. 2023, 24, 1998.
- Hoffmann, M.; Kleine-Weber, H.; Schroeder, S.; Kruger, N.; Herrler, T.; Erichsen, S.; Schiergens, T.S.; Herrler, G.; Wu, N.H.; Nitsche, A.; et al. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. Cell 2020, 181, 271– 280.e278.
- 66. Xu, S.-W.; Ilyas, I.; Weng, J.-P. Endothelial dysfunction in COVID-19: An overview of evidence, biomarkers, mechanisms and potential therapies. Acta Pharmacol. Sin. 2023, 44, 695–709.
- Chen, X.; Zhao, B.; Qu, Y.; Chen, Y.; Xiong, J.; Feng, Y.; Men, D.; Huang, Q.; Liu, Y.; Yang, B.; et al. Detectable Serum Severe Acute Respiratory Syndrome Coronavirus 2 Viral Load (RNAemia) Is Closely Correlated with Drastically Elevated Interleukin 6 Level in Critically III Patients with Coronavirus Disease 2019. Clin. Infect. Dis. 2020, 71, 1937–1942.
- 68. Kaur, S.; Tripathi, D.M.; Yadav, A. The Enigma of Endothelium in COVID-19. Front. Physiol. 2020, 11, 989.
- 69. Mosleh, W.; Chen, K.; Pfau, S.E.; Vashist, A. Endotheliitis and Endothelial Dysfunction in Patients with COVID-19: Its Role in Thrombosis and Adverse Outcomes. J. Clin. Med. 2020, 9, 1862.
- 70. Mojzisch, A.; Brehm, M.A. The Manifold Cellular Functions of von Willebrand Factor. Cells 2021, 10, 92351.
- Huertas, A.; Montani, D.; Savale, L.; Pichon, J.; Tu, L.; Parent, F.; Guignabert, C.; Humbert, M. Endothelial cell dysfunction: A major player in SARS-CoV-2 infection (COVID-19)? Eur. Respir. J. 2020, 56, 2001634.
- 72. Eurelings, L.E.M.; Miedema, J.R.; Dalm, V.; van Daele, P.L.A.; van Hagen, P.M.; van Laar, J.A.M.; Dik, W.A. Sensitivity and specificity of serum soluble interleukin-2 receptor for diagnosing sarcoidosis in a population of patients suspected of sarcoidosis. PLoS ONE 2019, 14, e0223897.
- 73. Marini, J.J.; Gattinoni, L. Management of COVID-19 Respiratory Distress. JAMA 2020, 323, 2329–2330.
- 74. Suh, Y.J.; Hong, H.; Ohana, M.; Bompard, F.; Revel, M.P.; Valle, C.; Gervaise, A.; Poissy, J.; Susen, S.; Hekimian, G.; et al. Pulmonary Embolism and Deep Vein Thrombosis in COVID-19: A Systematic Review and Meta-Analysis. Radiology 2021, 298, E70–E80.
- 75. Simpson, R.; Mendis, D. A Corpus-Based Study of Idioms in Academic Speech. TESOL Q. 2003, 37, 419–441.
- Cambridge University Press & Assessment. Everything but the Kitchen Sink. Available online: https://dictionary.cambridge.org/us/dictionary/english/everything-but-the-kitchen-sink (accessed on 15 May 2023).

- 77. Roche, J.A.; Roche, R. A hypothesized role for dysregulated bradykinin signaling in COVID-19 respiratory complications. FASEB J. 2020, 34, 7265–7269.
- Wilczynski, S.A.; Wenceslau, C.F.; McCarthy, C.G.; Webb, R.C. A Cytokine/Bradykinin Storm Comparison: What Is the Relationship Between Hypertension and COVID-19? Am. J. Hypertens. 2021, 34, 304–306.
- 79. McCarthy, C.G.; Wilczynski, S.; Wenceslau, C.F.; Webb, R.C. A new storm on the horizon in COVID-19: Bradykinin-induced vascular complications. Vasc. Pharmacol. 2021, 137, 106826.
- Barvin, M.R.; Alvarez, C.; Miller, J.I.; Prates, E.T.; Walker, A.M.; Amos, B.K.; Mast, A.E.; Justice, A.; Aronow, B.; Jacobson, D. A mechanistic model and therapeutic interventions for COVID-19 involving a RAS-mediated bradykinin storm. eLife 2020, 9, e59177.
- 81. Hojyo, S.; Uchida, M.; Tanaka, K.; Hasebe, R.; Tanaka, Y.; Murakami, M.; Hirano, T. How COVID-19 induces cytokine storm with high mortality. Inflamm. Regen. 2020, 40, 37.
- 82. Hu, B.; Huang, S.; Yin, L. The cytokine storm and COVID-19. J. Med. Virol. 2021, 93, 250–256.
- Jiang, Y.; Rubin, L.; Peng, T.; Liu, L.; Xing, X.; Lazarovici, P.; Zheng, W. Cytokine storm in COVID-19: From viral infection to immune responses, diagnosis and therapy. Int. J. Biol. Sci. 2022, 18, 459–472.
- 84. Rabaan, A.A.; Tirupathi, R.; Sule, A.A.; Aldali, J.; Mutair, A.A.; Alhumaid, S.; Muzaheed; Gupta, N.; Koritala, T.; Adhikari, R.; et al. Viral Dynamics and Real-Time RT-PCR Ct Values Correlation with Disease Severity in COVID-19. Diagnostics 2021, 11, 1091.
- 85. Abdulrahman, A.; Mallah, S.I.; Alqahtani, M. COVID-19 viral load not associated with disease severity: Findings from a retrospective cohort study. BMC Infect. Dis. 2021, 21, 688.
- Hasanoglu, I.; Korukluoglu, G.; Asilturk, D.; Cosgun, Y.; Kalem, A.K.; Altas, A.B.; Kayaaslan, B.; Eser, F.; Kuzucu, E.A.; Guner, R. Higher viral loads in asymptomatic COVID-19 patients might be the invisible part of the iceberg. Infection 2021, 49, 117–126.
- 87. Kim, Y.; Cheon, S.; Jeong, H.; Park, U.; Ha, N.-Y.; Lee, J.; Sohn, K.M.; Kim, Y.-S.; Cho, N.-H. Differential Association of Viral Dynamics with Disease Severity Depending on Patients' Age Group in COVID-19. Front. Microbiol. 2021, 12, 712260.
- Dorward, D.A.; Russell, C.D.; Um, I.H.; Elshani, M.; Armstrong, S.D.; Penrice-Randal, R.; Millar, T.; Lerpiniere, C.E.B.; Tagliavini, G.; Hartley, C.S.; et al. Tissue-Specific Immunopathology in Fatal COVID-19. Am. J. Respir. Crit. Care Med. 2020, 203, 192–201.
- 89. World Health Organization (WHO). WHO Coronavirus (COVID-19) Dashboard. Available online: https://covid19.who.int/ (accessed on 15 May 2023).
- 90. World Health Organization (WHO). Coronavirus Disease (COVID-19): Post COVID-19 Condition. Available online: https://www.who.int/news-room/questions-and-answers/item/coronavirus-

disease-(COVID-19)-post-COVID-19-condition (accessed on 15 May 2023).

- Morris, G.; Bortolasci, C.C.; Puri, B.K.; Marx, W.; O'Neil, A.; Athan, E.; Walder, K.; Berk, M.; Olive, L.; Carvalho, A.F.; et al. The cytokine storms of COVID-19, H1N1 influenza, CRS and MAS compared. Can one sized treatment fit all? Cytokine 2021, 144, 155593.
- 92. Fung, T.S.; Liu, D.X. Similarities and Dissimilarities of COVID-19 and Other Coronavirus Diseases. Annu. Rev. Microbiol. 2021, 75, 19–47.
- 93. Hirano, T.; Murakami, M. COVID-19: A New Virus, but a Familiar Receptor and Cytokine Release Syndrome. Immunity 2020, 52, 731–733.
- Bo, Y.; Guo, C.; Lin, C.; Zeng, Y.; Li, H.B.; Zhang, Y.; Hossain, M.S.; Chan, J.W.M.; Yeung, D.W.; Kwok, K.O.; et al. Effectiveness of non-pharmaceutical interventions on COVID-19 transmission in 190 countries from 23 January to 13 April 2020. Int. J. Infect. Dis. 2021, 102, 247–253.
- 95. Schlee, M.; Hartmann, G. Discriminating self from non-self in nucleic acid sensing. Nat. Rev. Immunol. 2016, 16, 566–580.
- 96. Murphy, K.; Weaver, C. Janeway's Immunobiology; Garland Science: New York, NY, USA, 2016.
- 97. Kasuga, Y.; Zhu, B.; Jang, K.J.; Yoo, J.S. Innate immune sensing of coronavirus and viral evasion strategies. Exp. Mol. Med. 2021, 53, 723–736.
- 98. Sette, A.; Crotty, S. Adaptive immunity to SARS-CoV-2 and COVID-19. Cell 2021, 184, 861–880. Retrieved from https://encyclopedia.pub/entry/history/show/127444