Clinical Applications of MR Spectroscopy in Pediatrics

Subjects: Radiology, Nuclear Medicine & Medical Imaging Contributor: Stefan Blüml , Alexander Saunders , Benita Tamrazi

In vivo MR spectroscopy is a non-invasive methodology that provides information about the biochemistry of tissues. It is available as a "push-button" application on state-of-the-art clinical MR scanners. MR spectroscopy has been used to study various brain diseases including tumors, stroke, trauma, degenerative disorders, epilepsy/seizures, inborn errors, neuropsychiatric disorders, and others.

pediatrics brain disorders in vivo MR spectroscopy

1. Pediatric Brain Tumors

Pediatric brain tumors are the second most frequent malignancy of childhood (after leukemia) with approximately 2500 new diagnoses per year in the United States. They are the leading cause of death from cancer in pediatric oncology ^{[1][2]}. Furthermore, survivors often have severe neurological, neurocognitive, and psychosocial sequelae. In contrast to adult brain tumors, which are mostly astrocytomas, pediatric brain tumors originate from different cell types and are, thus, biologically more heterogeneous with different biochemical and metabolic features.

Initial MRS studies have focused on common infratentorial tumors (medulloblastomas, pilocytic astrocytomas, ependymomas) and reported that proton MRS can be used to help differentiate cerebellar tumors by looking at ratios of NAA, creatine, choline, and lactate ^[3]. Since then, several groups ^{[4][5][6][7][8][9][10][11][12]}, taking advantage of the availability of robust short echo-time (TE) MRS, have independently confirmed the value of in vivo MRS for improving the accuracy of initial diagnoses including for tumors outside the posterior fossa such as germ cell tumors, choroid plexus tumors, and high-grade gliomas (**Figure 1**, **Figure 2** and **Figure 3**). It should be noted that high-grade gliomas in adults ^[13], and significant metabolic heterogeneity can be observed across subjects, in individual patients in different areas of the lesion, and in serial studies.



Figure 1. MR spectra of common pediatric posterior fossa tumors. Metabolic profiles of posterior fossa pilocytic astrocytoma (**A**) are generally relatively predictable. Pilocytic astrocytomas show elevated lactate and lipids. There is signal consistent with N-acetylated sugars (N-acetyl (NA) at \approx 2 ppm and a broad signal from sugars at \approx 3.8 ppm), while creatine (Cr) and myo-inositol (ml) levels are low. Ependymomas (**B**) have less predictable profiles. Whereas lipids are often prominent, they are not elevated in every ependymoma. Similarly, levels of other metabolites, such as mI can vary considerably. Medulloblastomas (**C**) are embryonal tumors that can present with strikingly different metabolic profiles for individual patients. To what extent metabolic profiles correlate with the molecular subgroups is an area of active research ^[14]. Above, examples for group 3 (i), group 4 (ii), sonic hedgehog (iii), and WNT (iv) are shown. Taurine (Tau) and glycine (Glyc) are often (but not always) detectable in these tumors. Medulloblastomas are generally more cellular tumors with higher absolute metabolite levels. For example, average choline (Cho) levels are approximately medulloblastoma:ependymoma:pilocytic astrocytoma = 5:3:2 ^[10], which cannot be appreciated when spectra, that are scaled to their tallest peaks, are compared. All spectra were acquired on 3T scanners with SV-PRESS, TE = 35 ms, and TR = 2 s.



Figure 2. Examples for MR spectra of pediatric brain tumors outside the posterior fossa. Metabolic profiles of posterior fossa pilocytic astrocytoma (A) are generally relatively predictable. Pilocytic astrocytomas show elevated lactate and lipids. There is signal consistent with N-acetylated sugars (N-acetyl (NA) at ≈ 2 ppm and a broad signal from sugars at ≈3.8 ppm), while creatine (Cr) and myo-inositol (mI) levels are low. Ependymomas (B) have less predictable profiles. Whereas lipids are often prominent, they are not elevated in every ependymoma. Similarly, levels of other metabolites, such as mI can vary considerably. Medulloblastomas (C) are embryonal tumors that can present with strikingly different metabolic profiles for individual patients. To what extent metabolic profiles correlate with the molecular subgroups is an area of active research ^[14]. Above, examples for group 3 (i), group 4 (ii), sonic hedgehog (iii), and WNT (iv) are shown. Taurine (Tau) and glycine (Glyc) are often (but not always) detectable in these tumors. Medulloblastomas are generally more cellular tumors with higher absolute choline metabolite levels. For example, average (Cho) levels are approximately medulloblastoma:ependymoma:pilocytic astrocytoma = 5:3:2 $\frac{10}{10}$, which cannot be appreciated when spectra, that are scaled to their tallest peaks, are compared. All spectra were acquired on 3T scanners with SV-PRESS, TE = 35 ms. and TR = 2 s.



Figure 3. Pediatric high-grade gliomas. Metabolic profiles of pediatric high-grade gliomas present with considerable heterogeneity. For example, three patients with thalamic anaplastic astrocytoma show varying levels of myo-inositol (mI) and glycine (Glyc) at initial diagnoses. Choline (Cho) is moderate or even low in patients 1 and 2 but is prominent in patient 3, with higher Cho generally associated with more proliferative tumors ^[15]. Citrate (Cit) is readily detectable in patients 2 and 3 but absent in patient 1. Two spectra acquired from patient 4 (glioblastoma) at diagnosis exhibit remarkable metabolic heterogeneity with glutathione (GSH), Glyc, and lactate (Lac) all elevated in one region but unremarkable in a second spectrum. It is presently unclear to what extent metabolic features identify subtypes and whether this information can be exploited to optimize therapeutic approaches and patient management. Serial MRS in patient 5 (glioblastoma) demonstrate the transition of a solid lesion to a partially necrotic lesion with increased lipids and lactate (Lac). All spectra were acquired on 3T scanners using SV-PRESS, TE = 35 ms, and TR = 2 s.

Among pediatric brain tumors, diffuse intrinsic pontine gliomas (DIPGs) carry the worst prognosis. They are highly resistant to chemo- and radiation therapy and, due to their location in the brainstem, inoperable. Thus, with no effective therapy available, the average survival after diagnosis is less than one year ^[16]. In vivo MR spectroscopy studies showed that, at initial diagnoses, these tumors often present with metabolic profiles that are consistent with low-proliferative tumors. The metabolism of DIPG then evolves into a profile typical for high-grade gliomas consistent with the observation that, at autopsy, most DIPGs have progressed to glioblastoma (**Figure 4**) ^{[17][18]}. These changes may precede clinical deterioration and progression on MRI, and MR spectroscopy could thus provide non-invasive biomarkers that help with patient management ^[19].



Figure 4. Diffuse intrinsic pontine glioma (DIPG). DIPGs are readily diagnosed by conventional MR imaging. MR spectroscopy demonstrates a metabolic evolution from a more moderately abnormal profile at presentation (**A**) to a metabolic pattern that is consistent with high-grade aggressive behavior at progression (**C**). Transiently, albeit

still consistent with viable tumor, a pattern suggestive for a limited response to therapy may be observed, characterized by reduced choline (Cho) and increased myo-inositol (ml) (**B**). Metabolic changes consistent with progression may precede clinical deterioration and progression on MRI. All spectra were acquired on a 1.5T scanner using SV-PRESS with TE = 35 ms and TR = 1.5 s. Cit = citrate.

Citrate (Cit) is routinely detectable in DIPG. Citrate is also detectable in subgroups of tumors outside the brainstem ^[20]. Among grade II astrocytomas, high levels of Cit appeared to indicate a high risk for malignant progression ^[21]. However, Cit was not generally specific for poor outcome, as it was undetectable in a significant number of high-grade gliomas with poor outcomes. Harris et al. reported that myo-inositol in supratentorial pilocytic astrocytomas is higher than in posterior fossa pilocytic astrocytomas (cf. **Figure 1** and **Figure 2**). They also noted that, among optic or thalamic tumors, those that had low myo-inositol at presentation were at higher risk for progression ^[22]. It has also been suggested that elevated levels of glycine identify tumors with increased malignancy ^{[23][24][25]}.

Recently, molecular subtypes of common pediatric brain tumors associated with significant different clinical outcomes have been identified using whole-genome sequencing methods ^{[26][27]}. Future novel targeted therapies might be able to treat these tumors without the need and risks of surgical resection and biopsies. Nevertheless, there is a need for accurate and early in vivo diagnosis of molecular subtypes—a possible important clinical application for in vivo MRS. Indeed, first studies indicate that MRS might be able to assist with the non-invasive identification of the medulloblastoma subtypes wingless (WNT), sonic hedgehog (SHH), group 3, and group 4 ^[14]. In vivo MRS may also predict key molecular features of atypical teratoid/rhabdoid tumors (AT/RT) at initial diagnosis ^[28] and may help with assigning subtypes of ependymomas ^[29].

2. Perinatal Hypoxic–Ischemic Encephalopathy

Perinatal hypoxic–ischemic encephalopathy (HIE) is a significant cause of neonatal death and of long-term neurodevelopmental disabilities ^[30]. MR imaging of the newborn brain provides biomarkers of disease status and predictors for outcome ^{[31][32][33][34]}. MRS complements conventional MRI and diffusion MRI by providing direct measures of metabolites that reflect the severity of injury ^{[34][35][36][37][38][39][40][41]} (**Figure 5**). Meta-analyses that compared various imaging modalities showed high sensitivity (82%) and specificity (95%) for the lactate to NAA ratio (Lac/NAA) for predicting neurodevelopmental outcomes ^{[42][43]}. Lactate accumulates when oxidation of pyruvate in the TCA cycle is impaired or halted, whereas a reduction of NAA indicates neuronal and axonal injury.



Figure 5. Newborn hypoxic/ischemic injury. Typical MR spectra of the thalamus of acute mild newborn hypoxic/ischemic injury (HIE) with clinically unremarkable follow up (A,C) versus severe HIE followed by death or significant disability (B,D). Note that spectra (A) + (B) were acquired with a short echo time (TE = 35 ms), whereas spectra (C) + (D) were acquired with long TE = 288 ms. Metabolic markers of severe injury that have been consistently reported in the literature are elevated lactate (Lac) and lipids, reduced NAA, and elevated glutamine (Gln). The above spectra were scaled to the approximate absolute metabolite levels ((A) vs. (B) and (C) vs. (D)). Edema formation and/or cell death and depletion of intracellular metabolites in severe HIE may explain generally lower absolute concentrations. In long-TE spectra, signals from lipid, glutamate, and glutamine are suppressed resulting in a more unambiguous detection and quantitation of NAA and lactate, which may simplify the determination of the important Lac/NAA ratio. Spectra were acquired within 1 week of injury on a clinical 3T scanner with SV-PRESS, TR = 2 s, and TE as indicated above.

The metabolic profiles of HIE evolve as the injury evolves. Lactate is prominent and glutamine (GIn) is elevated at the very early stage (1–2 days) with NAA more moderately decreased. Edema formation may also have an overall impact on metabolite profiles and concentrations during the acute phase of HIE. With subsequent cell death in severe HIE, a more substantial reduction of NAA and increased lipids are observed (**Figure 6**). Kreis et al. suggested that MRS performed 3–4 days after injury had higher predictive value for outcome than when done at 1–2 days; however, their study was performed in older children ^[44]. Of note, propylene glycol (Pgc), when used as a vehicle for medications, can accumulate in tissue and can be misidentified as lactate as their signals are similar ^[45]. This is avoided by appreciating the different positions of their signals on the frequency axis at ≈1.14 ppm for Pgc vs. ≈1.33 ppm for Lac (**Figure 7**). Pgc is ultimately metabolized to lactate and is, therefore, a potential exogenous source of lactate.



Figure 6. Evolution of HIE in newborns. MR spectra of the thalamus (**A**) and parietal gray matter (**B**) in a newborn with severe HIE on days 2, 4, and 24 after injury. Lactate (Lac) is elevated whereas lipids are unremarkable on day 2 after injury in both brain regions. Four days after injury, small increases of lipids are noted with lactate remaining elevated. Lipids are prominent at day 24 in the more severely injured thalamus. At that time, lactate levels in both regions have decreased. The broad peak at 2.8 ppm originates from poly-unsaturated fatty acids (PUFAs). Note that *absolute* NAA is reduced on day 2 but then further declines with cell death particularly in the thalamus. Spectra were acquired on a clinical 3T scanner with SV-PRESS, TE = 35 ms, and TR = 2 s.



Figure 7. Lactate vs. propylene glycol. MRS of parietal white matter of a 1-month-old male with suspected HIE shows a prominent propylene glycol (Pgc) signal centered at ≈1.19 ppm but essentially unremarkable Lac at ≈1.33 ppm. Other metabolites are also unremarkable.

3. Inborn Errors of Metabolism

With MRS providing metabolic information, inborn errors of metabolism (IEMs) seem to be a tailormade application for MRS. However, IEMs are generally infrequently encountered indications for brain MRI studies when compared with, for example, brain tumors. In addition, albeit altogether IEMs constitute a significant portion of childhood disorders, individually, IEMs are rare diseases. Finally, since radiologists are often uncomfortable with MRS, it has been utilized infrequently, and it is not surprising that the number of MRS studies of IEM is small. Nevertheless, MR imaging studies, for IEM patients who undergo an MR examination, are often interpreted as unremarkable or report ambiguity. For these patients, with a suspected or a known neurometabolic disease, the addition of a brief MRS acquisition may be beneficial either by improving the accuracy of diagnoses or the phenotyping of a known disease [46]. Examples for MRS of inborn errors are shown in **Figures 8, 9**.



Figure 8. Mitochondrial disorders. MRS of parieto/occipital gray matter of mild form of MELAS (mitochondrial encephalomyopathy, lactic acidosis, and stroke-like episodes) with an unremarkable MRI is essentially normal with no evidence for elevated lactate (Lac) (**A**). In contrast, Lac is readily detectable in another MELAS patient with a borderline normal MRI. Note that NAA (relative to Cr) is reduced in this patient (**B**). MR spectra of two patients diagnosed with Leigh's syndrome (siblings) are shown on the right. In addition to Lac, alanine (Ala) is elevated. In both spectra, NAA is reduced, and glucose (Glc) seems to accumulate in the upper spectrum (**C**,**D**). Spectra were acquired with SV-PRESS, TE = 35 ms, TR = 1.5 s on a 1.5T scanner (**A**,**B**) and TE = 35 ms, TR = 2 s on a 3T scanner (**C**,**D**).



Figure 9. MR spectra of various inborn errors of metabolism. (**A**) Non-ketotic hyperglycinemia (NKH): A 15day-old male newborn examined to confirm suspicion of acute NKH. The elevated glycine (Glyc) signal is consistent with hyperglycinemia, an amino aciduria in which a defect of the enzyme that breaks down glycine results in the abnormal accumulation of glycine in tissue. Note that for a 15-day-old newborn, the NAA and Lac signals are within normal. Other metabolic features are also unremarkable when adjusted for age. For the above patient, the MR images were mostly unremarkable. (**B**) Sandhoff disease: A 14-month-old female presenting with global developmental delay and hypotonia with delayed myelination and diffuse white matter abnormalities. The signal at approximately ≈2.07 ppm has been assigned to N-acetylhexosamine (NHEX), specific for Sandhoff disease . In addition, elevated mI and reduced NAA is noted. (**C**) Canavan disease: Canavan disease is a leukodystrophy where a defect in aspartoacylase (ASPA), the enzyme that breaks down N-acetylaspartate (NAA), results in excessive accumulation of NAA. In above spectrum, myo-inositol (mI) is also elevated. The MR images of the 6-month-old male patient showed significant diffuse white and gray matter abnormalities. (**D**) Krabbe's leukodystrophy: Krabbe's leukodystrophy is a lipid storage disorder caused by a deficiency of galactocerebrosidase (GALC), the enzyme required for the breakdown of the sphingolipids, galactosylceremide and psychosine. MR images of a 3-year-old child with Krabbe's leukodystrophy demonstrate white matter dysmyelination and loss. MRS of white matter show a significant reduction of NAA and elevated ml. (E) Adrenoleukodystrophy (ALD): ALD is caused by mutations in the ABCD1 genes. In vivo MRS of affected the white matter in a 5-year-old male shows, relative to creatine (Cr), elevated lipids, depleted NAA, elevated choline (Cho), and elevated ml. Note that the spectrum carries some similarities with the spectra of gliosis and gliomas. (F) Metachromatic leukodystrophy (MLD): In MLD the accumulation of sulfatides causes the destruction of the myelin sheath. MR images show profoundly abnormal white matter. The MR spectrum shows elevated lipids and macromolecules (MM), elevated Lac, and reduced NAA. (G) Adenylosuccinate lyase deficiency (ASLD): ASLD causes the buildup of succinylaminoimidazole carboxamide riboside (SAICA riboside) and succinyladenosine (S-Ado), which are detectable at 7.5 and 8.3 ppm. In addition, in this patient, Lac is elevated, NAA is reduced, and Cho is elevated relative to Cr. The MR images of the 17-month-old female showed general volume loss and hypomyelination. (H) 3hydroxy-3-methylglutaric acid (HMG) CoA lyase deficiency: In HMG CoA lyase deficiency, cells are unable to process leucine and synthesize ketone bodies. The MR images of this 12-year-old female were mildly abnormal. The MR spectrum acquired in parietal white matter demonstrates accumulation of HMG and of 3-hydroxy isovaleric acid (OHIV). All spectra were acquired on clinical 1.5T (C.E.) or 3T scanners (A.B.D.F.G) with SV-PRESS sequence, TE = 35 ms, and TR = 1.5 s (1.5T) or TR = 2 s (3T).

4. Trauma

Traumatic brain injury (TBI) in children, including from child abuse (non-accidental trauma), is a leading cause of child death and neurologic complications in the United States ^{[47][48][49]}. In addition, possible adverse long-term effects from mild but repeated TBI (concussions) from sports or other activities are likely underreported and are an increasing concern in children ^{[50][51][52][53]}.

Computer tomography (CT) and MR imaging are the first choices for detecting bleeds and edema/swelling in acute and severe TBI. However, some aspects of acute and chronic injury or more mild but repetitive injury at a cellular level may be difficult to recognize by these methodologies. Metabolic patterns observed by in vivo MRS should be expected to be heterogeneous depending on the time after injury, the severity of the injury, the brain region examined, and the response of the brain to injury, which may vary during brain development.

Several groups independently concluded that MRS has value when performed early after an injury as it is helpful for evaluating the extent of injury and improves the accuracy of long-term prognosis ^{[54][55][56][57][58][59][60][61]}. It was reported that abnormal MRS in brain regions that were deemed to be normal by MRI, predicted outcome more accurately than the abnormalities of lesions ^{[62][63]}.

Severe, acute injury is recognized by elevated lactate and lipids and a reduction of the axonal/neuronal marker NAA and often mimics MRS patterns that are observed in hypoxic–ischemic injury, consistent with the interruption of blood perfusion and subsequent apoptosis and cell death (**Figure 10**). In more mild/moderate cases of traumatic

brain injury, NAA may be mildly or only transiently reduced, whereas Cho is often elevated, possibly reflecting axonal injury and subsequent repair processes [63][64][65][66][67].



Figure 10. MRS of suspected non-accidental trauma. A parietal white matter spectrum acquired from a 5-monthold male with subarachnoid hemorrhage but otherwise unremarkable MR imaging. The MR spectrum appears to be normal for age (**A**). Six-month-old with subdural hemorrhages and diffuse supratentorial volume loss. Choline (Cho) appears to be elevated suggestive for axonal injury (**B**). Two-month-old with acute subdural hemorrhage in the posterior fossa and diffusion abnormality consistent with acute infarct. Lactate is elevated and NAA is reduced. The elevated signal in the 2.2–2.5 ppm range is likely from glutamine (Gln) (**C**). All spectra were acquired on a 1.5T system with SV-PRESS, TE = 35 ms, TR= 1.5 s.

5. Infections, Inflammation

Acute abscesses can present with metabolic profiles that are strikingly unusual depending on the organism (bacteria, fungi) that is causing them (**Figure 11**A). The presence of cytosolic amino acids (e.g., leucine, isoleucine, and valine at 0.8 ppm) has been consistently reported both in aerobic and anaerobic pyogenic lesions with varying amounts of lactate and lipids. Acetate at 1.9 ppm and succinate at 2.4 ppm may also be observed as well as other signals that yet need to be assigned ^{[68][69][70][71]}. Abscesses that have been treated successfully may present with elevated Lac and lipids with all other metabolites being depleted (**Figure 11**B).



Figure 11. Acute and chronic infections. MR spectra of acute abscesses can be strikingly unusual. In the above example (**A**), common brain metabolites are absent, whereas prominent signals form succinate (Suc) and acetate (Act) as well the cytosolic amino acids leucine (Leu), isoleucine (ILeu), and valine (Val) are observed. lactate (Lac), alanine (Ala), and moderate amounts of lipids are also detectable. On the other hand, only lipids and lactate are observed in a shrinking abscess after 20 days of antibiotics treatment (**B**). A spectrum of acute cerebellitis shows elevated lipids and lactate as well as reduced N-acetylaspartate (NAA). Glutamine (Gln) is elevated, whereas myoinositol (ml) is low (**C**). In a spectrum acquired from a 2 ½-year-old child with a history of meningoencephalitis, lipids are unremarkable, lactate is close to normal, and both myo-inositol and glutamine are unremarkable. NAA is reduced, indicating some permanent neuronal/axonal injury (**D**). Spectra were acquired on 3T (**A**,**C**,**D**) and 1.5T (**B**) scanners with SV-PRESS, TE = 35 ms, and TR = 2 s (3T) or TR = 1.5 s (1.5T).

Otherwise, infectious or inflammatory conditions may present with a wide range of metabolic abnormalities depending on type, time of onset, extent, and treatments ^{[72][73][74][75][76][77][78][79][80][81]}. It has been suggested that MRS could be useful for differentiating infectious/demyelinating processes from tumors by the relative prominence of glutamine in short-TE spectra ^{[82][83]}. Of note, with long-TE MRS acquisitions (e.g., TE > 130 ms), the glutamine signal disappears and spectra need to be interpreted carefully ^[84]. Furthermore, when the ROI for a tumor study includes edema/inflammation adjacent to a tumor, the resulting spectrum will show a combination of metabolic features, complicating the interpretation.

6. Epilepsy

Epilepsy is a chronic disorder characterized by unprovoked seizures. Seizure cause is often unknown. Several groups have reported a decrease of the neuronal marker NAA, which could be attributed to loss of neurons but also to abnormal mitochondrial function as NAA is synthesized in the mitochondria ^{[85][86][87][88]}. NAA can recover to normal levels with disappearing seizure activity ^{[89][90]}. It has also been reported that lactate is elevated during seizure activity possibly due to increased energy demand that is met with partially anaerobic metabolism ^[91]. Tissue glutamate and GABA levels, being major excitatory and inhibitory neurotransmitters, have been investigated by several groups in patients with epilepsy. However, it is challenging to accurately quantify these metabolites in vivo since their MR signals are complex and overlapping ^{[92][93][94][95][96]}.

Clinically, focal abnormal metabolic features could be exploited for detecting seizure foci and for disease lateralization in temporal lobe epilepsy. Nevertheless, in practice, that would require examining the whole brain or a large section of the brain with multi-voxel spectroscopy, which is time-consuming and requires elaborate post-processing by experienced MR spectroscopists and is, therefore, currently not utilized.

Ketogenic diets have been effective in reducing seizure activities for a subgroup of patients. Albeit ketosis can readily be monitored with urine ketone levels, it should be noted that the accumulation of ketone bodies in the brain can be observed with in vivo MRS ^[97] (**Figure 12**).











Figure 12. Detection of ketone bodies in in vivo MRS (**A**) The MR spectrum of a 13-year-old boy with refractory epilepsy on ketogenic diet and unremarkable MR images shows a s shows a signal consistent with acetone (Acn) at 2.22 ppm and a doublet from β -hydroxybutyrate (β HB) centered at \approx 1.19 ppm. NAA is below normal for age. MR imaging in this patient was unremarkable. (**B**) The MR spectrum of a 11-year-old boy with abnormal MRI, a history of meningoencephalitis, and refractory epilepsy on ketogenic diet shows prominent Acn signal, elevated lactate (Lac) and glutamine (Gln), as well as reduced NAA. Spectra were acquired on a 3T scanner with PRESS, TE = 35 ms, and TR = 2s.

7. Neuropsychiatric Disorders

In vivo metabolic abnormalities have been detected in several neuropsychiatric disorders in children ^{[98][99][100]}. Unfortunately, some of the neurochemicals that are of interest for psychiatric disorders, including GABA and glutathione, cannot be quantified accurately in clinical settings. In addition, metabolites that can be assessed more reliably, such as NAA, Cr, Cho, and mI, show a large overlap in patients and controls. Thus, there is currently no role for MRS as a clinical tool for the diagnoses and monitoring of neuropsychiatric disorders in pediatrics, and MRS is restricted to academic research.

8. Metabolites

Chemicals detectable by MRS (**Table 1**) are small, mobile, and mostly *intracellular metabolites*, whereas large immobile macromolecules and phospholipids, myelin, proteins, RNA, and DNA are rendered "invisible" to MRS. Synthesis and breakdown of the small amino acids, carbohydrates, fatty acids, and lipids that contribute to cell metabolism is closely controlled by enzymes, and their concentrations are, thus, kept close to constant. Therefore, the MR spectra of normal in vivo brain biochemistry are remarkably robust and comparable across subjects and serially in individuals with no "Monday morning" vs. "Friday afternoon" metabolism.

Table 1. Metabolites detectable with clinical MR spectroscopy in the human brain.

Metabolite (Abbr.)	Functional Role and Remarks	Decreased ^a	Increased ^a
Acetate (Act)	Energy source, precursor of acetyl-CoA, common building block for biosynthesis	Disease correlate unknown	Infection/abscesses, brain death
Acetoacetate (AcAc)	Energy source, produced in the mitochondria of liver	Disease correlate unknown	Ketosis

	cells from acetoacetyl coenzyme A (CoA)		
Acetone (Acn)	Produced by decarboxylation of acetoacetate, singlet at 2.22 ppm more readily detectable than βHB (see below)	Disease correlate unknown	Ketosis
Alanine (Ala)	Amino acid, protein constituent, glucose–alanine cycle	Disease correlate unknown	Inborn errors; meningioma and subgroups of other tumors
Aspartate (Asp)	Excitatory neurotransmitter	Disease correlate unknown	Challenging to recognize due to complex signal and signal overlap with NAA and other chemicals
β-Hydroxybutyrate (βHB)	Produced by the decarboxylation of acetoacetate, doublet similar to lactate but at 1.19 ppm	Disease correlate unknown	Ketosis
Choline (Cho) = glycerophosphocholine + phosphocholine + free choline	Membrane/myelin synthesis/degradation, acetylcholine precursor, osmolyte	Liver disease; hypo- osmotic state; during cooling (hypometabolic?)	De novo synthesis of biomass, including tumors, brain growth, tissue repair; hyper-osmotic state
Citrate (Cit)	TCA cycle intermediate, produced when the	Disease correlate unknown	Newborns, subgroups of tumors, most common in

	glycolytic rate exceeds TCA activity, fatty acid synthesis		diffuse intrinsic brainstem gliomas
Creatine (Cr) = free creatine (fCR) + phosphocreatine (PCr)	Energy metabolism, energy storage PCr <-> fCr + ATP	Cells without creatine kinase, creatine deficiencies, some tumors	Subgroups of gliomas, gliosis?
γ-Aminobutyric acid (GABA)	Inhibitory neurotransmitter	Disease correlate unknown	Challenging to recognize due to complex signal and signal overlap with other chemicals
Glucose (Glc) (α and β isomers)	Principal fuel for cells	Hypoglycemia, detection challenging	Uncontrolled diabetes; hyperglycemia
Glutamate (Glu)	Excitatory neurotransmitter	Most tumors, hepatic encephalopathy, acute hypoxic/ischemic injury	Subgroup of seizures
Glutamine (Gln)	Part of the Glu–Gln neurotransmitter cycle; hyper ammonia detoxifier, fuel, osmolyte	Disease correlate unknown	Most tumors, edema (relative increase), demyelinating lesions, hepatic encephalopathy, acute hypoxic/ischemic injury
Glutathione (GSH)	Consists of glycine, cysteine, and glutamate. Present in reduced (predominant) and oxidized	Disease correlate unknown	Meningioma

	form. Marker of oxidative stress		
Glycine (Glyc)	Neurotransmitter inhibitory and excitatory, cellular migration and circuit formation, antioxidant	Disease correlate unknown	Medulloblastoma and other tumors; hyperglycinemia
Lactate (Lac)	Endpoint of anaerobic glycolysis, in normal brain present in cerebrospinal fluid at higher concentrations than in tissue	Disease correlate unknown	Inborn errors of energy metabolism, hypoxic/ischemic injury; tumors, cystic lesions, normal newborn
Lipids (Lip) with contributions from macromolecules (MM)	Indicators for cell membrane breakdown when elevated	Disease correlate unknown	Injury/cell death and tumor subgroups
Leucine (Leu), iso-leucine (ILeu), valine (Val)	Branched-chain amino acids (BCAA)	Disease correlate unknown	Elevated in inborn error of BCAA metabolism, acute abscesses
Myo-inositol (ml)	Glial marker, involved in phospholipid membrane metabolism, osmolyte	Liver disease, hepatic encephalopathy, osmotic imbalance	Normal newborns, astrocytes, subgroups of tumors (e.g., astrocytoma, ependymoma, choroid plexus papilloma), osmotic imbalance
N-acetylaspartate (NAA)	Marker for mature neurons and axons	Pathologies associated with neuronal/axonal damage/loss,	Canavan disease

		mitochondrial function?		
N-acetylaspartate glutamate (NAAG)	Neurotransmitter release modulator, small shoulder next to NAA, detectable in high-quality spectra	Disease correlate unknown	unknown	
Phenylalanine	Essential amino acid	Disease correlate unknown	Uncontrolled phenylketonuria (PKU, phenylalanine hydroxylase deficiency)	ko, C.; tem
Propylene glycol (Pgc)	Medication solvent (e.g., anticonvulsants), metabolizes to lactate, doublet similar to lactate but at 1.14 ppm	Disease correlate unknown	Frequently seen in newborns on medications, possibly because of underdeveloped blood– brain barrier	4), IVI– asms. Program Cancer
Scyllo-inositol (sI)	Symmetric sugar–alcohol isomer, osmolyte, inhibits amyloid-beta aggregation?	Disease correlate unknown in majority of population	Detectable under normal conditions in a subgroup of the population; glial tumors	۹ roradiol.
Succinate (Suc)	TCA cycle intermediate	Disease correlate unknown	Abscesses, infection	R.A. 388–
Taurine (Tau)	Osmolyte, modulator of neurotransmission	Decreasing with normal brain maturation	Newborns; medulloblastoma (group 3, group 4), germinoma, pineoblastoma, and possibly others	of Tarbell,

7. A Lazareff, J.; Olmstead, C.; Bockhorst, K.H.; Alger, J.R. Proton magnetic resonance spectroscopic imaging of pediatric low-grade astrocytomas. Child's Nerv. Syst. 1996, 12.

8. Peet, A.C.; Lateef, S.; MacPherson, L.; Natarajan, K.; Sgouros, S.; Grundy, R.G. Short echo time ^aThe accuracy for detecting some of the metabolites is low, even if they are present in the tissue, due to low 1 H magnetic resonance spectroscopy of childhood brain tumours. Child's Nerv. Syst. 2006, 23, concentrations and/or due to complex signals that overlap with signals from other chemicals. For these chemicals, 163–169. Sparings, NdR;th/iloopyd/.alhiamisade M.yiNlattly ajamaskible.atecf.ress/Macplaecsonaylbe Systembis, i6djvidual spectrandy, R.G.; Arvanitis, T.N.; Peet, A.C. Identification and characterisation of childhood cerebellar tumours by in vivo proton MRS. NMR Biomed. 2008, 21, 908–918.

- Panigrahy, A.; Krieger, M.; Gonzalez-Gomez, I.; Liu, X.; McComb, J.; Finlay, J.; Nelson, M.; Gilles, F.; Blüml, S. Quantitative Short Echo Time 1H-MR Spectroscopy of Untreated Pediatric Brain Tumors: Preoperative Diagnosis and Characterization. Am. J. Neuroradiol. 2006, 27, 560–572.
- Shiroishi, M.S.; Panigrahy, A.; Moore, K.R.; Nelson, M.D.; Gilles, F.H.; González-Gómez, I.; Blüml, S. Combined MRI and MRS improves pre-therapeutic diagnoses of pediatric brain tumors over MRI alone. Neuroradiology 2015, 57, 951–956.
- 12. Tamrazi, B.; Nelson, M.D.; Blüml, S. MRS of pilocytic astrocytoma: The peak at 2 ppm may not be NAA. Magn. Reson. Med. 2016, 78, 452–456.
- Jones, C.; Karajannis, M.A.; Jones, D.T.W.; Kieran, M.W.; Monje, M.; Baker, S.J.; Becher, O.J.; Cho, Y.-J.; Gupta, N.; Hawkins, C.; et al. Pediatric high-grade glioma: Biologically and clinically in need of new thinking. Neuro-Oncology 2016, 19, 153–161.
- Blüml, S.; Margol, A.S.; Sposto, R.; Kennedy, R.J.; Robison, N.J.; Vali, M.; Hung, L.T.; Muthugounder, S.; Finlay, J.L.; Erdreich-Epstein, A.; et al. Molecular subgroups of medulloblastoma identification using noninvasive magnetic resonance spectroscopy. Neuro Oncol. 2015, 18, 126–131.
- Shimizu, H.; Kumabe, T.; Shirane, R.; Yoshimoto, T. Correlation between Choline Level Measured by Proton MR Spectroscopy and Ki-67 Labeling Index in Gliomas. Am. J. Neuroradiol. 2000, 21, 659–665.
- 16. Clymer, J.; Kieran, M.W. The Integration of Biology Into the Treatment of Diffuse Intrinsic Pontine Glioma: A Review of the North American Clinical Trial Perspective. Front. Oncol. 2018, 8, 169.
- 17. Pan, E.; Prados, M. Pediatric CNS Tumors; Gupta, N., Haas-Kogen, D., Banerjee, A., Eds.; Springer: Berlin/Heidelberg, Germany; New York, NY, USA; Volume 3, pp. 49–61.
- 18. Yoshimura, J.; Onda, K.; Tanaka, R.; Takahashi, H. Clinicopathological Study of Diffuse Type Brainstem Gliomas: Analysis of 40 Autopsy Cases. Neurol. Med.-Chir. 2003, 43, 375–382.
- 19. Panigrahy, A.; Nelson, M.D.; Finlay, J.L.; Sposto, R.; Krieger, M.D.; Gilles, F.H.; Blüml, S. Metabolism of diffuse intrinsic brainstem gliomas in children. Neuro-Oncology 2008, 10, 32–44.
- 20. Seymour, Z.A.; Panigrahy, A.; Finlay, J.L.; Nelson, M.D., Jr.; Bluml, S. Citrate in pediatric CNS tumors? AJNR Am. J. Neuroradiol. 2008, 29, 1006–1011.
- Blüml, S.; Panigrahy, A.; Laskov, M.; Dhall, G.; Krieger, M.D.; Nelson, M.D.; Finlay, J.L.; Gilles, F.H. Elevated citrate in pediatric astrocytomas with malignant progression. Neuro-Oncology 2011, 13, 1107–1117.

- 22. Harris, L.M.; Davies, N.; MacPherson, L.; Lateef, S.; Natarajan, K.; Brundler, M.-A.; Sgouros, S.; English, M.W.; Arvanitis, T.; Grundy, R.G.; et al. Magnetic resonance spectroscopy in the assessment of pilocytic astrocytomas. Eur. J. Cancer 2008, 44, 2640–2647.
- 23. Davies, N.P.; Wilson, M.; Natarajan, K.; Sun, Y.; MacPherson, L.; Brundler, M.A.; Arvanitis, T.N.; Grundy, R.G.; Peet, A.C. Non-invasive detection of glycine as a biomarker of malignancy in childhood brain tumours using in-vivo 1H MRS at 1.5 tesla confirmed by ex-vivo high-resolution magic-angle spinning NMR. NMR Biomed. 2010, 23, 80–87.
- 24. Carapella, C.M.; Carpinelli, G.; Knijn, A.; Raus, L.; Caroli, F.; Podo, F. Potential Role of in vitro 1H Magnetic Resonance Spectroscopy in the Definition of Malignancy Grading of Human Neuroepithelial Brain Tumours. Acta Neurochir. Suppl. 1997, 68, 127–132.
- 25. Tzika, A.A.; Righi, V.; Andronesi, O.C.; Mintzopoulos, D.; Black, P.M. High-resolution magic angle spinning magnetic resonance spectroscopy detects glycine as a biomarker in brain tumors. Int. J. Oncol. 2009, 36, 301–306.
- Louis, D.N.; Perry, A.; Wesseling, P.; Brat, D.J.; Cree, I.A.; Figarella-Branger, D.; Hawkins, C.; Ng, H.K.; Pfister, S.M.; Reifenberger, G.; et al. The 2021 WHO Classification of Tumors of the Central Nervous System: A summary. Neuro-Oncology 2021, 23, 1231–1251.
- 27. A Northcott, P.; Dubuc, A.M.; Pfister, S.; Taylor, M.D. Molecular subgroups of medulloblastoma. Expert Rev. Neurother. 2012, 12, 871–884.
- Tamrazi, B.; Venneti, S.; Margol, A.; Hawes, D.; Cen, S.; Nelson, M.; Judkins, A.; Biegel, J.; Blüml, S. Pediatric Atypical Teratoid/Rhabdoid Tumors of the Brain: Identification of Metabolic Subgroups Using In Vivo 1H-MR Spectroscopy. Am. J. Neuroradiol. 2019, 40, 872–877.
- 29. Panwalkar, P.; Tamrazi, B.; Dang, D.; Chung, C.; Sweha, S.; Natarajan, S.K.; Pun, M.; Bayliss, J.; Ogrodzinski, M.P.; Pratt, D.; et al. Targeting integrated epigenetic and metabolic pathways in lethal childhood PFA ependymomas. Sci. Transl. Med. 2021, 13.
- 30. Lawn, J.; Shibuya, K.; Stein, C. No cry at birth: Global estimates of intrapartum stillbirths and intrapartum-related neonatal deaths. Bull. World Health Organ. 2005, 83, 409–417.
- Barkovich, A.J.; Hajnal, B.L.; Vigneron, D.; Sola, A.; Partridge, J.C.; Allen, F.; Ferriero, D.M. Prediction of neuromotor outcome in perinatal asphyxia: Evaluation of MR scoring systems. AJNR Am. J. Neuroradiol. 1998, 19, 143–149.
- Miller, S.; Ramaswamy, V.; Michelson, D.; Barkovich, A.J.; Holshouser, B.; Wycliffe, N.; Glidden, D.; Deming, D.; Partridge, J.C.; Wu, Y.W.; et al. Patterns of brain injury in term neonatal encephalopathy. J. Pediatr. 2005, 146, 453–460.
- 33. Groenendaal, F.; Veenhoven, R.H.; van der Grond, J.; Jansen, G.H.; Witkamp, T.D.; de Vries, L.S. Cerebral Lactate and N-Acetyl-Aspartate/Choline Ratios in Asphyxiated Full-Term Neonates

Demonstrated In Vivo Using Proton Magnetic Resonance Spectroscopy. Pediatr. Res. 1994, 35, 148–151.

- 34. Alderliesten, T.; De Vries, L.S.; Benders, M.J.N.L.; Koopman, C.; Groenendaal, F. MR Imaging and Outcome of Term Neonates with Perinatal Asphyxia: Value of Diffusion-weighted MR Imaging and H MR Spectroscopy. Radiology 2011, 261, 235–242.
- 35. Parmentier, C.E.J.; de Vries, L.S.; Groenendaal, F. Magnetic Resonance Imaging in (Near-)Term Infants with Hypoxic-Ischemic Encephalopathy. Diagnostics 2022, 12, 645.
- Miller, S.P.; Newton, N.; Ferriero, D.M.; Partridge, J.C.; Glidden, D.V.; Barnwell, A.; A Chuang, N.; Vigneron, D.B.; Barkovich, A.J. Predictors of 30-Month Outcome after Perinatal Depression: Role of Proton MRS and Socioeconomic Factors. Pediatr. Res. 2002, 52, 71–77.
- 37. Azzopardi, D.; Edwards, A.D. Magnetic resonance biomarkers of neuroprotective effects in infants with hypoxic ischemic encephalopathy. Semin. Fetal Neonatal Med. 2010, 15, 261–269.
- Cheong, J.; Cady, E.; Penrice, J.; Wyatt, J.; Cox, I.; Robertson, N. Proton MR Spectroscopy in Neonates with Perinatal Cerebral Hypoxic-Ischemic Injury: Metabolite Peak-Area Ratios, Relaxation Times, and Absolute Concentrations. Am. J. Neuroradiol. 2006, 27, 1546–1554.
- Barkovich, A.J.; Baranski, K.; Vigneron, D.; Partridge, J.C.; Hallam, D.K.; Hajnal, B.L.; Ferriero, D.M. Proton MR Spectroscopy for the Evaluation of Brain Injury in Asphyxiated, Term Neonates. Am. J. Neuroradiol. 1999, 20, 1399–1405.
- 40. Mitra, S.; Kendall, G.S.; Bainbridge, A.; Sokolska, M.; Dinan, M.; Uria-Avellanal, C.; Price, D.; McKinnon, K.; Gunny, R.; Huertas-Ceballos, A.; et al. Proton magnetic resonance spectroscopy lactate/N-acetylaspartate within 2 weeks of birth accurately predicts 2-year motor, cognitive and language outcomes in neonatal encephalopathy after therapeutic hypothermia. Arch. Dis. Child.-Fetal Neonatal Ed. 2018, 104, F424–F432.
- 41. Aida, N. 1H-MR Spectroscopy of the Early Developmental Brain, Neonatal Encephalopathies, and Neurometabolic Disorders. Magn. Reson. Med. Sci. 2022, 21, 9–28.
- Thayyil, S.; Chandrasekaran, M.; Taylor, A.; Bainbridge, A.; Cady, E.B.; Chong, W.K.K.; Murad, S.; Omar, R.Z.; Robertson, N.J. Cerebral Magnetic Resonance Biomarkers in Neonatal Encephalopathy: A Meta-analysis. Pediatrics 2010, 125, e382–e395.
- Shanmugalingam, S.; Thornton, J.S.; Iwata, O.; Bainbridge, A.; O'Brien, F.E.; Priest, A.N.; Ordidge, R.J.; Cady, E.B.; Wyatt, J.S.; Robertson, N.J. Comparative Prognostic Utilities of Early Quantitative Magnetic Resonance Imaging Spin-Spin Relaxometry and Proton Magnetic Resonance Spectroscopy in Neonatal Encephalopathy. Pediatrics 2006, 118, 1467–1477.
- Kreis, R.; Arcinue, E.; Ernst, T.; Shonk, T.K.; Flores, R.; Ross, B.D. Hypoxic encephalopathy after near-drowning studied by quantitative 1H-magnetic resonance spectroscopy. J. Clin. Investig. 1996, 97, 1142–1154.

- Cady, E.B.; Lorek, A.; Penrice, J.; Reynolds, E.O.; Iles, R.A.; Burns, S.P.; Coutts, G.A.; Cowan, F.M. Detection of propan-1,2-diol in neonatal brain by in vivo proton magnetic resonance spectroscopy. Magn. Reson. Med. 1994, 32, 764–767.
- 46. Whitehead, M.T.; Lai, L.M.; Blüml, S. Clinical 1H MRS in childhood neurometabolic diseases— Part 1: Technique and age-related normal spectra. Neuroradiology 2022, 64, 1101–1110.
- 47. Paul, A.R.; Adamo, M.A. Non-accidental trauma in pediatric patients: A review of epidemiology, pathophysiology, diagnosis and treatment. Transl. Pediatr. 2014, 3, 195–207.
- 48. Keenan, H.T.; Runyan, D.K.; Marshall, S.W.; Nocera, M.A.; Merten, D.F.; Sinal, S.H. A populationbased study of inflicted traumatic brain injury in young children. JAMA 2003, 290, 621–626.
- 49. Theodore, A.D.; Chang, J.J.; Runyan, D.K.; Hunter, W.M.; Bangdiwala, S.I.; Agans, R. Epidemiologic Features of the Physical and Sexual Maltreatment of Children in the Carolinas. Pediatrics 2005, 115, e331–e337.
- 50. Kay, T.; Harrington, D.E.R.A.; Anderson, T.; Berrol, S.; Cicerone, K. Definition of mild traumatic brain injury. J. Head Trauma Rehabil. 1993, 8, 86–87.
- Arbogast, K.B.; Curry, A.; Pfeiffer, M.R.; Zonfrillo, M.; Haarbauer-Krupa, J.; Breiding, M.J.; Coronado, V.G.; Master, C. Point of Health Care Entry for Youth With Concussion Within a Large Pediatric Care Network. JAMA Pediatr. 2016, 170, e160294.
- 52. Meehan, W.P., 3rd; Bachur, R.G. Sport-related concussion. Pediatrics 2009, 123, 114–123.
- 53. Field, M.; Collins, M.W.; Lovell, M.R.; Maroon, J. Does age play a role in recovery from sportsrelated concussion? A comparison of high school and collegiate athletes. J. Pediatr. 2003, 142, 546–553.
- 54. Makoroff, K.L.; Cecil, K.M.; Caré, M.; Ball, W.S. Elevated lactate as an early marker of brain injury in inflicted traumatic brain injury. Pediatr. Radiol. 2005, 35, 668–676.
- 55. Ashwal, S.; A Holshouser, B.; Shu, S.K.; Simmons, P.L.; Perkin, R.M.; Tomasi, L.G.; Knierim, D.S.; Sheridan, C.; Craig, K.; Andrews, G.H.; et al. Predictive value of proton magnetic resonance spectroscopy in pediatric closed head injury. Pediatr. Neurol. 2000, 23, 114–125.
- 56. Aaen, G.S.; Holshouser, B.A.; Sheridan, C.; Colbert, C.; McKenney, M.; Kido, D.; Ashwal, S. Magnetic Resonance Spectroscopy Predicts Outcomes for Children With Nonaccidental Trauma. Pediatrics 2010, 125, 295–303.
- 57. Haseler, L.J.; Arcinue, E.; Danielsen, E.R.; Bluml, S.; Ross, B.D. Evidence From Proton Magnetic Resonance Spectroscopy for a Metabolic Cascade of Neuronal Damage in Shaken Baby Syndrome. Pediatrics 1997, 99, 4–14.
- 58. A Holshouser, B.; Ashwal, S.; Luh, G.Y.; Shu, S.; Kahlon, S.; Auld, K.L.; Tomasi, L.G.; Perkin, R.M.; Hinshaw, D.B. Proton MR spectroscopy after acute central nervous system injury: Outcome

prediction in neonates, infants, and children. Radiology 1997, 202, 487-496.

- Ross, B.D.; Ernst, T.; Kreis, R.; Haseler, L.J.; Bayer, S.; Danielsen, E.; Bluml, S.; Shonk, T.; Mandigo, J.C.; Caton, W.; et al. 1H MRS in acute traumatic brain injury. J. Magn. Reson. Imaging 1998, 8, 829–840.
- 60. Holshouser, B.A.; Ashwal, S.; Shu, S.; Hinshaw, D.B., Jr. Proton MR spectroscopy in children with acute brain injury: Comparison of short and long echo time acquisitions. J. Magn. Reson. Imaging 2000, 11, 9–19.
- 61. Friedman, S.; Brooks, W.; Jung, R.; Chiulli, S.; Sloan, J.; Montoya, B.; Hart, B.; Yeo, R. Quantitative proton MRS predicts outcome after traumatic brain injury. Neurology 1999, 52, 1384.
- 62. Holshouser, B.A.; Tong, K.A.; Ashwal, S. Proton MR Spectroscopic Imaging Depicts Diffuse Axonal Injury in Children with Traumatic Brain Injury. Am. J. Neuroradiol. 2005, 26, 1276–1285.
- 63. Govindaraju, V.; Gauger, G.E.; Manley, G.T.; Ebel, A.; Meeker, M.; Maudsley, A.A. Volumetric Proton Spectroscopic Imaging of Mild Traumatic Brain Injury. Am. J. Neuroradiol. 2004, 25, 730– 737.
- Brooks, W.M.; Stidley, C.A.; Petropoulos, H.; Jung, R.E.; Weers, D.C.; Friedman, S.; Barlow, M.A.; Sibbitt, W.; Yeo, R.A. Metabolic and Cognitive Response to Human Traumatic Brain Injury: A Quantitative Proton Magnetic Resonance Study. J. Neurotrauma 2000, 17, 629–640.
- 65. Gasparovic, C.; Arfai, N.; Smid, N.; Feeney, D.M. Decrease and Recovery of N-Acetylaspartate/Creatine in Rat Brain Remote from Focal Injury. J. Neurotrauma 2001, 18, 241– 246.
- 66. Schuhmann, M.U.; Stiller, D.; Skardelly, M.; Thomas, S.; Samii, M.; Brinker, T. Long-Time in-Vivo Metabolic Monitoring Following Experimental Brain Contusion Using Proton Magnetic Resonance Spectroscopy. Acta Neurochir. Suppl. 2002, 81, 209–212.
- Cecil, K.M.; Hills, E.C.; Sandel, M.E.; Smith, D.H.; McIntosh, T.K.; Mannon, L.J.; Sinson, G.P.; Bagley, L.J.; Grossman, R.I.; Lenkinski, R.E. Proton magnetic resonance spectroscopy for detection of axonal injury in the splenium of the corpus callosum of brain-injured patients. J. Neurosurg. 1998, 88, 795–801.
- Pal, D.; Bhattacharyya, A.; Husain, M.; Prasad, K.; Pandey, C.; Gupta, R. In Vivo Proton MR Spectroscopy Evaluation of Pyogenic Brain Abscesses: A Report of 194 Cases. Am. J. Neuroradiol. 2009, 31, 360–366.
- Lai, P.-H.; Hsu, S.-S.; Ding, S.-W.; Ko, C.-W.; Fu, J.-H.; Weng, M.-J.; Yeh, L.-R.; Wu, M.-T.; Liang, H.-L.; Chen, C.-K.; et al. Proton magnetic resonance spectroscopy and diffusion-weighted imaging in intracranial cystic mass lesions. Surg. Neurol. 2007, 68, S25–S36.

- Luthra, G.; Parihar, A.; Nath, K.; Jaiswal, S.; Prasad, K.; Husain, N.; Husain, M.; Singh, S.; Behari, S.; Gupta, R. Comparative Evaluation of Fungal, Tubercular, and Pyogenic Brain Abscesses with Conventional and Diffusion MR Imaging and Proton MR Spectroscopy. Am. J. Neuroradiol. 2007, 28, 1332–1338.
- 71. Gupta, R.K.; Jain, K.K.; Mittal, S.K.; Kumar, S. Imaging features of central nervous system fungal infections. Neurol. India 2007, 55, 241–250.
- 72. Ferraz-Filho, J.R.; Santana-Netto, P.V.; Rocha-Filho, J.A.; Sgnolf, A.; Mauad, F.; Sanches, R.A. Application of magnetic resonance spectroscopy in the differentiation of high-grade brain neoplasm and inflammatory brain lesions. Arq. Neuro-Psiquiatr. 2009, 67, 250–253.
- 73. Keller, M.A.; Venkatraman, T.N.; Thomas, A.; Deveikis, A.; LoPresti, C.; Hayes, J.; Berman, N.; Walot, I.; Padilla, S.; Johnston-Jones, J.; et al. Altered neurometabolite development in HIVinfected children: Correlation with neuropsychological tests. Neurology 2004, 62, 1810–1817.
- 74. van der Voorn, J.P.; Pouwels, P.J.; Vermeulen, R.J.; Barkhof, F.; van der Knaap, M.S. Quantitative MR imaging and spectroscopy in congenital cytomegalovirus infection and periventricular leukomalacia suggests a comparable neuropathological substrate of the cerebral white matter lesions. Neuropediatrics 2009, 40, 168–173.
- 75. Takanashi, J.-I.; Sugita, K.; Ishii, M.; Aoyagi, M.; Niimi, H. Longitudinal MR imaging and proton MR spectroscopy in herpes simplex encephalitis. J. Neurol. Sci. 1997, 149, 99–102.
- 76. Cecil, K.M.; Jones, B.V.; Williams, S.; Hedlund, G.L. CT, MRI and MRS of Epstein-Barr virus infection: Case report. Neuroradiology 2000, 42, 619–622.
- 77. Cecil, K.M.; Lindquist, D.M. Infection and Encephalitis. In MR Spectroscopy of Pediatric Brain Disorders; Springer: New York, NY, USA, 2013; pp. 155–166.
- 78. Mader, I.; Wolff, M.; Nägele, T.; Niemann, G.; Grodd, W.; Küker, W. MRI and proton MR spectroscopy in acute disseminated encephalomyelitis. Child's Nerv. Syst. 2005, 21, 566–572.
- 79. Seo, H.-E.; Hwang, S.-K.; Choe, B.H.; Cho, M.-H.; Park, S.-P.; Kwon, S. Clinical Spectrum and Prognostic Factors of Acute Necrotizing Encephalopathy in Children. J. Korean Med. Sci. 2010, 25, 449–453.
- Gupta, R.K.; Roy, R.; Dev, R.; Husain, M.; Poptani, H.; Pandey, R.; Kishore, J.; Bhaduri, A.P. Finger printing of Mycobacterium tuberculosis in patients with intracranial tuberculomas by using in vivo, ex vivo, and in vitro magnetic resonance spectroscopy. Magn. Reson. Med. 1996, 36, 829–833.
- Gupta, R.K.; Husain, M.; Vatsal, D.K.; Kumar, R.; Chawla, S.; Husain, N. Comparative evaluation of magnetization transfer MR imaging and in-vivo proton MR spectroscopy in brain tuberculomas. Magn. Reson. Imaging 2002, 20, 375–381.

- Malhotra, H.; Jain, K.; Agarwal, A.; Singh, M.; Yadav, S.; Husain, M.; Krishnani, N.; Gupta, R. Characterization of tumefactive demyelinating lesions using MR imaging and in-vivo proton MR spectroscopy. Mult. Scler. J. 2008, 15, 193–203.
- Cianfoni, A.; Niku, S.; Imbesi, S. Metabolite Findings in Tumefactive Demyelinating Lesions Utilizing Short Echo Time Proton Magnetic Resonance Spectroscopy. Am. J. Neuroradiol. 2007, 28, 272–277.
- 84. Saindane, A.M.; Cha, S.; Law, M.; Xue, X.; Knopp, E.A.; Zagzag, D. Proton MR Spectroscopy of Tumefactive Demyelinating Lesions. Am. J. Neuroradiol. 2002, 23, 1378–1386.
- 85. Urenjak, J.; Williams, S.R.; Gadian, D.G.; Noble, M. Specific expression of N-acetylaspartate in neurons, oligodendrocyte-type-2 astrocyte progenitors, and immature oligodendrocytes in vitro. J. Neurochem. 1992, 59, 55–61.
- 86. Signoretti, S.; Marmarou, A.; Tavazzi, B.; Lazzarino, G.; Beaumont, A.; Vagnozzi, R. N-Acetylaspartate Reduction as a Measure of Injury Severity and Mitochondrial Dysfunction Following Diffuse Traumatic Brain Injury. J. Neurotrauma 2001, 18, 977–991.
- 87. Varho, T.; Komu, M.; Sonninen, P.; Lähdetie, J.; Holopainen, I.E. Quantitative 1H MRS and MRI Volumetry Indicate Neuronal Damage in the Hippocampus of Children with Focal Epilepsy and Infrequent Seizures. Epilepsia 2005, 46, 696–703.
- Miller, E.; Widjaja, E. Magnetic Resonance Spectroscopy in Epilepsy. In MR Spectroscopy of Pediatric Brain Disorders; Bluml, S., Panigrahy, A., Eds.; Springer: New York, NY, USA, 2013; pp. 175–192.
- Najm, I.M.; Wang, Y.; Hong, S.C.; Luders, H.O.; Ng, T.C.; Comair, Y.G. Temporal Changes in Proton MRS Metabolites After Kainic Acid-Induced Seizures in Rat Brain. Epilepsia 1997, 38, 87– 94.
- Baslow, M.H. Evidence supporting a role for N-acetyl-l-aspartate as a molecular water pump in myelinated neurons in the central nervous system: An analytical review. Neurochem. Int. 2002, 40, 295–300.
- 91. Najm, I.M.; Wang, Y.; Shedid, D.; Luders, H.O.; Ng, T.C.; Comair, Y.G. MRS metabolic markers of seizures and seizure-induced neuronal damage. Epilepsia 1998, 39, 244–250.
- 92. Woermann, F.G.; McLean, M.A.; Bartlett, P.A.; Parker, G.J.; Barker, G.J.; Duncan, J.S. Short echo time single-voxel 1H magnetic resonance spectroscopy in magnetic resonance imaging-negative temporal lobe epilepsy: Different biochemical profile compared with hippocampal sclerosis. Ann. Neurol. 1999, 45, 369–376.
- Simister, R.J.; McLean, M.A.; Barker, G.J.; Duncan, J.S. A Proton Magnetic Resonance Spectroscopy Study of Metabolites in the Occipital Lobes in Epilepsy. Epilepsia 2003, 44, 550– 558.

- 94. Sherwin, A.; Robitaille, Y.; Quesney, F.; Olivier, A.; Villemure, J.; Leblanc, R.; Feindel, W.; Andermann, E.; Gotman, J.; Ethier, R.; et al. Excitatory amino acids are elevated in human epileptic cerebral cortex. Neurology 1988, 38, 920.
- 95. Petroff, O.A.; Pleban, L.A.; Spencer, D.D. Symbiosis between in vivo and in vitro NMR spectroscopy: The creatine, N-acetylaspartate, glutamate, and GABA content of the epileptic human brain. Magn. Reson. Imaging 1995, 13, 1197–1211.
- 96. Pfund, Z.; Chugani, D.C.; Juhász, C.; Muzik, O.; Chugani, H.T.; Wilds, I.B.; Seraji-Bozorgzad, N.; Moore, G.J. Evidence for Coupling between Glucose Metabolism and Glutamate Cycling Using FDG PET and 1H Magnetic Resonance Spectroscopy in Patients with Epilepsy. J. Cereb. Blood Flow Metab. 2000, 20, 871–878.
- Seymour, K.J.; Bluml, S.; Sutherling, J.; Sutherling, W.; Ross, B.D. Identification of cerebral acetone by 1H-MRS in patients with epilepsy controlled by ketogenic diet. Magma 1999, 8, 33– 42.
- Horska, A.; Mahone, E.M. 1H Magnetic Resonance Spectroscopy of the Brain During Adolescence: Normal Brain Development and Neuropsychiatric Disorders. In MR Spectroscopy of Pediatric Brain Disorders; Bluml, S., Panigrahy, A., Eds.; Springer: New York, NY, USA, 2013; pp. 193–212.
- Levitt, J.G.; O'Neill, J.; Alger, J.R. Magnetic Resonance Spectroscopy Studies of Autistic Spectrum Disorders. In MR Spectroscopy of Pediatric Brain Disorders; Bluml, S., Panigrahy, A., Eds.; Springer: New York, NY, USA, 2013; pp. 213–227.
- 100. O'Neill, J.; Levitt, J.G.; Alger, J.R. Magnetic Resonance Spectroscopy Studies of Attention Deficit Hyperactivity Disorder. In MR Spectroscopy of Pediatric Brain Diseases; Bluml, S., Panigrahy, A., Eds.; Springer: New York, NY, USA, 2013; pp. 229–275.

Retrieved from https://encyclopedia.pub/entry/history/show/59648