

# SputnikV Vaccine

Subjects: Infectious Diseases

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SputnikV is a vaccine against SARS-CoV-2 developed by the Gamaleya National Research Centre for Epidemiology and Microbiology.

Keywords: SARS-CoV-2 ; SputnikV ; Spike protein ; humoral immune response ; cellular immunity

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## 1. Introduction

The outbreak of acute pneumonia in Wuhan, China, in December 2019, spread rapidly throughout the world, prompting WHO to declare a pandemic by 11 March 2020 <sup>[1]</sup>. Sequence analysis identified a novel coronavirus as the cause of infection <sup>[2]</sup>, which was later named severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) on 11 February 2020, by the International Committee on Taxonomy of Viruses (ICTV) <sup>[1]</sup>. Soon after, WHO officially named the disease coronavirus disease 2019 (COVID-19) <sup>[1]</sup>. The disease rapidly spread from China, leading to a global health emergency. The disease quickly spread, and there are more than 213 million cases documented worldwide, with 4,459,381 deaths confirmed as of 27 August 2021 <sup>[3]</sup>. COVID-19 has been present in Russia since March 2020, where initially, all confirmed cases were imported, but local transmission was quickly observed and documented <sup>[4]</sup>.

Initially, multiple approaches were suggested to control the disease using repurposed and novel drugs <sup>[5][6][7][8]</sup>. Some of the repurposed drugs had demonstrated efficacy in vitro <sup>[9]</sup>, yet they failed to translate into patient treatment due to high mortality <sup>[10]</sup>. Lopinavir was used for COVID-19 treatment, although the drug's efficacy was inconsistent <sup>[11][12]</sup>. As a result, limited options remained to treat COVID-19, leading to high morbidity and mortality. With the increasing number of cases registered daily and the inefficacy of administrative measures to prevent virus spread, prevention of infection using vaccines appeared to be one of the most effective approaches to dealing with the pandemic. Several different methodologies have been developed including mRNA-based <sup>[13][14][15]</sup> and adenovirus-based vectors from gorilla <sup>[16]</sup> and chimpanzee <sup>[17][18]</sup> adenoviruses.

The SputnikV vaccine developed by Gamaleya National Research Centre for Epidemiology and Microbiology (GRCEM) demonstrating 91.6% efficacy in a clinical trial <sup>[19]</sup>. This vaccine is recombinant adenovirus (rAd) 26 and rAd5 vector-based, expressing SARS-CoV-2 Spike (S) protein. The selection of this protein is based on its essential role in virus entry as it binds to the receptor and allows for membrane fusion <sup>[20][21]</sup>. Currently, the vaccine is routinely used to immunize Russian citizens <sup>[22]</sup>, where 22.9% of Russian citizens were fully vaccinated by August 2021 <sup>[23][24]</sup>. The vaccine is also authorized in over 60 countries worldwide as of 29 July 2021 <sup>[25]</sup>. Clinical trial studies have demonstrated that the vaccine is safe, and no adverse effects have been demonstrated <sup>[19]</sup>. There is currently limited knowledge on which immunogenic epitopes of SARS-CoV-2 S protein elicit an antibody response in immunized individuals and how these compare to antibodies produced in a natural infection.

## 2. Discussions on SputnikV Vaccine

Our data confirm the immunogenicity of the SputnikV vaccine, developed by GRCEM <sup>[19]</sup>, where humoral and T cell immune responses were detected in vaccinated individuals. Our data corroborate the previous report by Logunov et al., where anti-SARS-CoV-2 antibodies and activated T cells were detected in immunized individuals <sup>[26]</sup>. In our study, we further advanced the understanding of the SputnikV vaccine by demonstrating the S protein epitopes involved in the induction of an immune response. We have identified several immunogenic epitopes located in NTD, RBD, FP, HR1 and HR2. Sixteen peptides were found in S1, and fourteen peptides were in S2 domains. S1 and S2 contain several domains and epitopes, ranging from binding to the receptor, membrane fusion and entry <sup>[20]</sup>. Therefore, by targeting multiple regions, antibodies in immunized individuals can potentially interfere with the most important events in virus replication: entry to the target cell.

Interestingly, six peptides reacting with immunized serum were in the RBD of S1 responsible for binding to the ACE2 receptor [27]. It is the primary target for the most potent neutralizing antibodies [28][29]. Therefore, it is expected that antibodies induced by SputnikV could also be neutralizing, which was previously shown by Logunov et al. [26]. Additionally, RBD targeting vaccines were shown to be highly immunogenic [30][31] in part due to eliciting neutralizing antibodies against SARS-CoV. We also identified peptides in NTD, which were shown to contain epitopes producing neutralizing antibodies [32]. These antibodies appear to interfere with receptor binding or S protein conformation [33]. Our data suggest that neutralizing antibodies, previously detected in immunized individuals, could target RBD and NTD. Together, antibodies to RBD and NTD could provide broad coverage of epitopes in the RBD and FP of SARS-CoV-2, which will reduce the virus's potential to escape from a vaccine-induced immune response.

We also identified multiple peptides in the S2 domain located in FP, HR1 and HR2. Upon attachment to the receptor, FP becomes embedded into the target cell membrane and installs an anchor inside to initiate fusion with the viral envelope [34][35]. HR1 and HR2 also contribute to fusion by bringing viral and cellular membranes proximal to each other [36]. Therefore, antibodies targeting these regions could potentially interfere with membrane fusion. As HR1 and HR2 are highly conserved among SARS-CoV viruses [37][38], including SARS-CoV-2, antibodies to these regions of S protein could prevent virus escape from immune response induced by the vaccine.

Convalescent plasma was shown to effectively treat critically ill COVID-19 patients [39][40]. This treatment is a polyclonal mix of antibodies developed to many viral proteins, including the S protein. It appears that among all antibodies, those recognizing the S protein are the most important in preventing a severe form of COVID-19. This assumption is based on the observation of Dispinseri et al., demonstrating that compromised immune responses to the S protein were major traits of critical COVID-19 conditions [41]. We have identified multiple peptides recognized by sera from both immunized and convalescent COVID-19 individuals. These data suggest that the SputnikV vaccine induces a humoral immune response similar to that occurring naturally after infection. Therefore, our data could explain the protective efficacy of the SputnikV vaccine demonstrated by Logunov et al. [26]. We believe that selection of epitopes similar to those in naturally infected and recovered individuals is the mechanism of SputnikV protection against COVID-19.

Studies have demonstrated that a single aa replacement could help the virus escape elimination by neutralizing antibodies [42][43]. Concerns have been presented that some mutations increase the infectivity of SARS-CoV-2 [44][45] and that new variants of SARS-CoV-2 may mutate to escape vaccine-induced immunity [46]. Our analysis demonstrated that several peptides recognized in vaccinated individuals remained 100% conserved in SARS-CoV-2 strains worldwide. Not surprisingly, three peptides were located in NTD, a highly conserved region [47][48]. We also identified four peptides in the RBD, which were 90–100% conserved in all SARS-CoV-2 strains included in the analysis. RBD region mutations could affect the neutralizing capacity of antibodies in the later waves of pandemics [21]. Our finding of conserved peptides in NTD and FP provides evidence that SputnikV induced antibodies should retain antiviral efficacy even in the case of a mutation in the S protein. We also did a detailed analysis of S protein mutations in one of the fastest spreading SARS-CoV-2 viruses, B.1.617.2 (Delta variant), which is currently detected in 78 countries [3][49]. This variant has shown moderate resistance to vaccine-induced immunity [50]. Only four peptides recognized by the immunized serum contained the aa positions mutated in the Delta variant of SARS-CoV-2. This is 11.1% out of all reacting peptides and represents a small fraction. Therefore, we suggested that the humoral immune response induced by SputnikV will be effective against the Delta variant of SARS-CoV-2.

We have identified multiple regions on S protein-containing immunogenic epitopes. These regions could be grouped as aa 137–309, 341–471, 579–812 and 1084–1188. Interestingly, these regions are within those Grifoni A et al. predicted as associated with robust immune response [51]. A similar location of the peptides reacting to COVID-19 sera was demonstrated by Shrock et al. [52]. Many peptides that were recognized by convalescent COVID-19 sera were also recognized by vaccinated sera. These data provide strong evidence that the immune response induced by the vaccine is similar to that of a naturally developed response, after recovery from SARS-CoV-2 infection. Therefore, it could be suggested that the protective efficacies of convalescent and immunized sera are comparable.

Our results confirm that SputnikV induces a T cell immune response supporting findings by Logunov et al. [26]. We have found that the T cell immune response was active 42, 90 and 210 days after immunization, suggesting long term circulation of memory T lymphocytes. These lymphocytes are essential for protection against virus infection [53], providing long-lasting immunity. T cells were shown to be activated in convalescent COVID-19 patients [54]. The analysis also revealed that those activated lymphocytes contribute to the amelioration of clinical symptoms of the disease. Multiple studies have found activation of T cells in convalescent COVID-19, where the S protein appears to maintain this immune response [54][55][56]. Interestingly, several peptides (S10, S21, S23, S46 and S48) that activated T cells in SputnikV immunized individuals were similar to those described by Peng et al. in convalescent COVID-19 patients [54]. The

immunogenic epitopes appear in aa 166–180, 351–365, 381–395, 751–765 and 801–815 regions on the S protein. Additionally, we have identified two peptides (S21 and S23) that activated T cell responses in SputnikV vaccinated individuals as located in the RBD of the S protein, suggesting that this domain also contains T cell epitopes similar to that described by Ni et al. in convalescent COVID-19 patients [56]. These data indicate that the T cell response in immunized individuals resembles that of individuals that recovered from a SARS-CoV-2 infection. Our data also show that the SputnikV vaccine-induced T cell and humoral immune responses to the S protein of SARS-CoV-2 lasted out to seven months after vaccination. This suggests the long-term efficacy of this vaccine, which is a significant consideration with all vaccine candidates [57][58].

There is limited real-life data on the duration of the immune response elicited by the Sputnik V vaccine. Our data, therefore, collected on samples from 90 and 210 days after the first dose of SputnikV vaccine demonstrating long-term activation of both the humoral and T cell immune responses is of importance. These long-term immune responses could explain the efficacy of the SputnikV vaccine reported by Gonzalez et al. in an Argentinian cohort [59]. Authors have also shown that receiving even a single dose of vaccine reduced the duration of hospitalization and fatality, after exposure to SARS-CoV-2. In another study by Rossi et al., eliciting anti-SARS-CoV-2 neutralizing antibodies was demonstrated in 288 volunteers 21 days after the first dose vaccine [60]. Additionally, the efficacy of SputnikV was demonstrated in a cohort from Venezuela, where even a single dose was shown sufficient to induce neutralizing antibodies in previously SARS-CoV-2 positive individuals [61]. A 100% seroconversion was found in a Venezuelan cohort 6 weeks after the second dose of SputnikV vaccine, confirming induction of long-term immune response.

Cytokines regulate immune responses during natural infection and vaccination [62][63]. These cytokines also contribute to developing clinical symptoms of COVID-19 and post-immunization side effects [64][65]. We have found fifteen cytokines that remain upregulated in convalescent COVID-19 individuals one month after infection (IL-1 $\alpha$ , IL-3, IL-10, IL-12p70, CCL7, IFN- $\alpha$ 2, bFGF, LIF, TRAIL, IL-2, IL-4, IL-5, IL-12p40, IL-17 and MIF) indicating a strong activation of the immune response. These cytokines are associated with inflammation, activation of T and B cell immune response, and regeneration [66]. A set of cytokines were upregulated in SputnikV immunized individuals (IL-1 $\alpha$ , IL-3, IL-10, IL-12p70, CCL7, IFN- $\alpha$ 2, bFGF, LIF and TRAIL), where nine of them were similar to that in convalescent COVID-19. These data suggest similarity in mechanisms of an immune response activation in vaccinated and convalescent individuals.

It should be noted that convalescent COVID-19 sera had more cytokines upregulated 42 days after infection than immunized, indicating that infection generates a more robust immune response. This could be explained by the exposure of the immune system to multiple viral antigens during infection compared to only the S protein used in SputnikV. Additionally, differences in cytokine activation could be explained by the differing quantities of SARS-CoV-2 antigens encountered during infection and upon vaccination. Some findings support this suggestion where a higher viral load was closely related to severe COVID-19 [67][68]. Additionally, the simultaneous exposure to adenovirus antigens from the vector and SARS-CoV-2 antigens could affect cytokine activation in vaccinated individuals. The lesser number of cytokines activated in vaccinated compared to convalescent COVID-19 could also explain the lack of clinical symptoms of COVID-19, such as severe inflammation, high fever and tissue damage [64].

In summary, we have shown that vaccination with SputnikV induces a broad antibody response that recognizes a wide variety of epitopes on the S protein and excellent cellular response. Both of these responses are still measurable three months after vaccination, suggesting the long-term efficacy of this vaccine.

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## References

1. WHO. Director-General's Opening Remarks at the Media Briefing on COVID-19—11 March 2020. Available online: <https://www.who.int/director-general/speeches/detail/who-director-general-s-opening-remarks-at-the-media-briefing-on-covid-19---11-march-2020> (accessed on 12 January 2021).
2. Wu, F.; Zhao, S.; Yu, B.; Chen, Y.-M.; Wang, W.; Song, Z.-G.; Hu, Y.; Tao, Z.-W.; Tian, J.-H.; Pei, Y.-Y.; et al. A new coronavirus associated with human respiratory disease in China. *Nature* 2020, 579, 265–269.
3. World Health Organization. WHO Coronavirus (COVID-19) Dashboard. Available online: <https://covid19.who.int/> (accessed on 21 September 2021).
4. Komissarov, A.B.; Safina, K.R.; Garushyants, S.K.; Fadeev, A.V.; Sergeeva, M.V.; Ivanova, A.A.; Danilenko, D.M.; Lioznov, D.; Shneider, O.V.; Shvyrev, N.; et al. Genomic epidemiology of the early stages of the SARS-CoV-2 outbreak in Russia. *Nat. Commun.* 2021, 12, 649.

5. Sanders, J.M.; Monogue, M.L.; Jodlowski, T.Z.; Cutrell, J.B. Pharmacologic Treatments for Coronavirus Disease 2019 (COVID-19): A Review. *JAMA* 2020, 323, 1824–1836.
6. Beigel, J.H.; Tomashek, K.M.; Dodd, L.E.; Mehta, A.K.; Zingman, B.S.; Kalil, A.C.; Hohmann, E.; Chu, H.Y.; Luetkemeyer, A.; Kline, S.; et al. Remdesivir for the Treatment of Covid-19—final Report. *N. Engl. J. Med.* 2020, 383, 1813–1836.
7. Lloyd, E.; Gandhi, T.; Petty, L. Monoclonal Antibodies for COVID-19. *JAMA* 2021, 325, 1015.
8. WHO Solidarity Trial Consortium Repurposed Antiviral Drugs for Covid-19—Interim WHO Solidarity Trial Results. *N. Engl. J. Med.* 2021, 384, 497–511.
9. Yao, X.; Ye, F.; Zhang, M.; Cui, C.; Huang, B.; Niu, P.; Liu, D. In Vitro Antiviral Activity and Projection of Optimized Dosing Design of Hydroxychloroquine for the Treatment of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). *Clin. Infect Dis.* 2020, 71, 732–739.
10. Axfors, C.; Schmitt, A.M.; Janiaud, P.; van't Hooft, J.; Abd-Elsalam, S.; Abdo, E.F.; Abella, B.S.; Akram, J.; Amaravadi, R.K.; Angus, D.C.; et al. Mortality outcomes with hydroxychloroquine and chloroquine in COVID-19 from an international collaborative meta-analysis of randomized trials. *Nat. Commun.* 2021, 12, 2349.
11. Guan, W.-J.; Ni, Z.-Y.; Hu, Y.; Liang, W.-H.; Ou, C.-Q.; He, J.-X.; Liu, L.; Shan, H.; Lei, C.-L.; Hui, D.S.; et al. Clinical Characteristics of Coronavirus Disease 2019 in China. *N. Engl. J. Med.* 2020, 382, 1708–1720.
12. Young, B.E.; Ong, S.W.X.; Kalimuddin, S.; Low, J.G.; Tan, S.Y.; Loh, J. Epidemiologic Features and Clinical Course of Patients Infected With SARS-CoV-2 in Singapore. *JAMA* 2020, 323, 1488–1494.
13. Baden, L.R.; El Sahly, H.M.; Essink, B.; Kotloff, K.; Frey, S.; Novak, R.; Diemert, D.; Spector, S.A.; Rouphael, N.; Creech, C.B.; et al. Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. *N. Engl. J. Med.* 2021, 384, 403–416.
14. Polack, F.P.; Thomas, S.J.; Kitchin, N.; Absalon, J.; Gurtman, A.; Lockhart, S.; Gruber, W.C. Safety and efficacy of the BNT162b2 mRNA Covid-19 vaccine. *N. Engl. J. Med.* 2020, 383, 2603–2615.
15. Walsh, E.E.; Frenck, R.W.; Falsey, A.R.; Kitchin, N.; Absalon, J.; Gurtman, A.; Lockhart, S.; Neuzil, K.; Mulligan, M.J.; Bailey, R.; et al. Safety and Immunogenicity of Two RNA-Based Covid-19 Vaccine Candidates. *N. Engl. J. Med.* 2020, 383, 2439–2450.
16. Capone, S.; Raggioli, A.; Gentile, M.; Battella, S.; Lahm, A.; Sommella, A.; Contino, A.M.; Urbanowicz, R.A.; Scala, R.; Barra, F.; et al. Immunogenicity of a new gorilla adenovirus vaccine candidate for COVID-19. *Mol. Ther.* 2021, 29, 2412–2423.
17. Ewer, K.J.; Barrett, J.R.; Belij-Rammerstorfer, S.; Sharpe, H.; Makinson, R.; Morter, R.; Flaxman, A.; Wright, D.; Bellamy, D.; Bittaye, M.; et al. T cell and antibody responses induced by a single dose of ChAdOx1 nCoV-19 (AZD1222) vaccine in a phase 1/2 clinical trial. *Nat. Med.* 2021, 27, 270–278.
18. Ramasamy, M.N.; Minassian, A.M.; Ewer, K.J.; Flaxman, A.L.; Folegatti, P.M.; Owens, D.R. Safety and immunogenicity of ChAdOx1 nCoV-19 vaccine administered in a prime-boost regimen in young and old adults (COV002): A single-blind, randomised, controlled, phase 2/3 trial. *Lancet* 2020, 396, 1979–1993.
19. Logunov, D.Y.; Dolzhikova, L.V.; Shcheblyakov, D.V.; Tukhvatulin, A.I.; Zubkova, O.V.; Dzharullaeva, A.S.; Kovyrshina, K.V.; Nadezhda, L.L.; Grousova, D.M.; Erokhova, A.S. Safety and efficacy of a rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine: An interim analysis of a randomized controlled phase 3 trial in Russia. *Lancet* 2021, 397, 671–681.
20. Huang, Y.; Yang, C.; Xu, X.F.; Xu, W.; Liu, S.W. Structural and functional properties of SARS-CoV-2 spike protein: Potential antiviral drug development for COVID-19. *Acta Pharm. Sin.* 2020, 41, 1141–1149.
21. Li, C.; Tian, X.; Jia, X.; Wan, J.; Lu, L.; Jiang, S.; Lan, F.; Lu, Y.; Wu, Y.; Ying, T. The impact of receptor-binding domain natural mutations on antibody recognition of SARS-CoV-2. *Signal Transduct. Target. Ther.* 2021, 6, 132.
22. Baraniuk, C. Covid-19: What do we know about airborne transmission of SARS-CoV-2? *BMJ* 2021, 373, n1030.
23. Reuters. COVID-19 TRACKER. 2021. Available online: <https://graphics.reuters.com/world-coronavirus-tracker-and-maps/countries-and-territories/russia/> (accessed on 6 April 2021).
24. CRC. The Race to Vaccinate the World. 2021. Available online: <https://coronavirus.jhu.edu/vaccines/international> (accessed on 8 June 2021).
25. Statista. Number of Doses of the COVID-19 Vaccine Sputnik V Ordered from Russia or Agreed to Be produced Abroad as of 25 May 2021, by Country. 2021. Available online: <https://www.statista.com/statistics/1123927/sputnik-v-exports-from-russia-by-country/> (accessed on 5 March 2021).
26. Logunov, D.Y.; Dolzhikova, I.V.; Zubkova, O.V.; Tukhvatullin, A.I.; Shcheblyakov, D.V.; Dzharullaeva, A.S. Safety and immunogenicity of a rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine in two formulations:

Two open, non-randomized phase 1/2 studies from Russia. *Lancet* 2020, 396, 887–897.

27. Wong, S.K.; Li, W.; Moore, M.J.; Choe, H.; Farzan, M. A 193-amino acid fragment of the SARS coronavirus S protein efficiently binds angiotensin-converting enzyme 2. *J. Biol. Chem.* 2004, 279, 3197–3201.
28. Robbiani, D.F.; Gaebler, C.; Muecksch, F.; Lorenzi, J.C.; Wang, Z.; Cho, A.; Nussenzweig, M.C. Convergent antibody responses to SARS-CoV-2 in convalescent individuals. *Nature* 2020, 584, 437–442.
29. Shi, R.; Shan, C.; Duan, X.; Chen, Z.; Liu, P.; Song, J.; Yan, J. A human neutralizing antibody targets the receptor-binding site of SARS-CoV-2. *Nature* 2020, 584, 120–124.
30. Brink, E.N.V.D.; ter Meulen, J.; Cox, F.; Jongeneelen, M.A.C.; Thijsse, A.; Throsby, M.; Marissen, W.E.; Rood, P.M.L.; Bakker, A.B.H.; Gelderblom, H.R.; et al. Molecular and Biological Characterization of Human Monoclonal Antibodies Binding to the Spike and Nucleocapsid Proteins of Severe Acute Respiratory Syndrome Coronavirus. *J. Virol.* 2005, 79, 1635–1644.
31. Cao, Z.; Liu, L.; Du, L.; Zhang, C.; Jiang, S.; Li, T.; He, Y. Potent and persistent antibody responses against the receptor-binding domain of SARS-CoV spike protein in recovered patients. *Virol. J.* 2010, 7, 1–6.
32. Cerutti, G.; Guo, Y.; Zhou, T.; Gorman, J.; Lee, M.; Rapp, M.; Reddem, E.R.; Yu, J.; Bahna, F.; Bimela, J.; et al. Potent SARS-CoV-2 neutralizing antibodies directed against spike N-terminal domain target a single supersite. *Cell Host Microbe* 2021, 29, 819–833.
33. Zhou, H.; Chen, Y.; Zhang, S.; Niu, P.; Qin, K.; Jia, W.; Wang, X. Structural definition of a neutralization epitope on the N-terminal domain of MERS-CoV spike glycoprotein. *Nat. Commun.* 2019, 10, 3068.
34. Tang, T.; Bidon, M.; Jaimes, J.A.; Whittaker, G.R.; Daniel, S. Coronavirus membrane fusion mechanism offers a potential target for antiviral development. *Antivir. Res.* 2020, 178, 104792.
35. Ou, X.; Zheng, W.; Shan, Y.; Mu, Z.; Dominguez, S.R.; Holmes, K.V.; Qian, Z. Identification of the Fusion Peptide-Containing Region in Betacoronavirus Spike Glycoproteins. *J. Virol.* 2016, 90, 5586–5600.
36. Xia, S.; Zhu, Y.; Liu, M.; Lan, Q.; Xu, W.; Wu, Y.; Ying, T.; Liu, S.; Shi, Z.; Jiang, S.; et al. Fusion mechanism of 2019-nCoV and fusion inhibitors targeting HR1 domain in spike protein. *Cell. Mol. Immunol.* 2020, 17, 765–767.
37. Trigueiro-Louro, J.; Correia, V.; Figueiredo-Nunes, I.; Gíria, M.; Rebelo-de-Andrade, H. Unlocking COVID therapeutic targets: A structure-based rationale against SARS-CoV-2, SARS-CoV and MERS-CoV Spike. *Comput. Struct. Biotechnol. J.* 2020, 18, 2117–2131.
38. Xu, Y.; Zhu, J.; Liu, Y.; Lou, Z.; Yua, F.; Liu, Y.; Cole, D.K.; Ni, L.; Su, N.; Qin, L.; et al. Characterization of the Heptad Repeat Regions, HR1 and HR2, and Design of a Fusion Core Structure Model of the Spike Protein from Severe Acute Respiratory Syndrome (SARS) Coronavirus. *Biochemistry* 2004, 43, 14064–14071.
39. Montelongo-Jauregui, D.; Vila, T.; Sultan, A.S.; Jabra-Rizk, M.A. Convalescent serum therapy for COVID-19: A 19th century remedy for a 21st century disease. *PLOS Pathog.* 2020, 16, e1008735.
40. Kumar, N.; Kumar, S.; Patel, S. Convalescent Plasma Therapy In Critically Ill Patients With Late Stage Covid-19. *Chest* 2020, 158, A601.
41. Dispinseri, S.; Secchi, M.; Pirillo, M.F.; Tolazzi, M.; Borghi, M.; Brigatti, C.; De Angelis, M.L.; Baratella, M.; Bazzigaluppi, E.; Venturi, G.; et al. Neutralizing antibody responses to SARS-CoV-2 in symptomatic COVID-19 is persistent and critical for survival. *Nat. Commun.* 2021, 12, 2670.
42. He, Y.; Li, J.; Jiang, S. A single amino acid substitution (R441A) in the receptor-binding domain of SARS coronavirus spike protein disrupts the antigenic structure and binding activity. *Biochem. Biophys. Res. Commun.* 2006, 344, 106–113.
43. Mitsuki, Y.Y.; Ohnishi, K.; Takagi, H.; Oshima, M.; Yamamoto, T.; Mizukoshi, F. A single amino acid substitution in the S1 and S2 Spike protein domains determines the neutralization escape phenotype of SARS-CoV. *Microbes Infect.* 2008, 10, 908–915.
44. Korber, B.; Fischer, W.M.; Gnanakaran, S.; Yoon, H.; Theiler, J.; Abfalterer, W.; Foley, B.; Giorgi, E.E.; Bhattacharya, T.; Parker, M.D.; et al. Spike mutation pipeline reveals the emergence of a more transmissible form of SARS-CoV-2. *BioRxiv* 2020.
45. Galloway, S.E.; Paul, P.; MacCannell, D.R.; Johansson, M.A.; Brooks, J.T.; MacNeil, A.; Slayton, R.B.; Tong, S.; Silk, B.J.; Armstrong, G.L.; et al. Emergence of SARS-CoV-2 B.1.1.7 Lineage—United States, 29 December 2020–12 January 2021. *Morb. Mortal. Wkly. Rep.* 2021, 70, 95.
46. Rubin, R. COVID-19 Vaccines vs Variants—Determining How Much Immunity Is Enough. *JAMA* 2021, 325, 1241.
47. Agrawal, L.; Poullikkas, T.; Eisenhower, S.; Monsanto, C.; Bakku, R.K.; Chen, M.H.; Kalra, R.S. Bioinformatics-Based Analysis of SARS-CoV-2 Core Proteins for Potential Therapeutic Targets. *Antibodies* 2021, 10, 3.

48. Lei, K.; Zhang, X. Conservation analysis of SARS-CoV-2 spike suggests complicated viral adaptation history from bat to human. *Evol. Med. Public Health* 2020, 2020, 290–303.
49. GISAID. Tracking of Variants. 2021. Available online: <https://www.gisaid.org/hcov19-variants/> (accessed on 24 April 2021).
50. Callaway, E. Delta coronavirus variant: Scientists brace for impact. *Nature* 2021, 595, 17–18.
51. Grifoni, A.; Sidney, J.; Zhang, Y.; Scheuermann, R.H.; Peters, B.; Sette, A. A sequence homology and bioinformatic approach can predict candidate targets for immune responses to SARS-CoV-2. *Cell Host Microbe* 2020, 27, 671–680.e672.
52. Shrock, E.; Fujimura, E.; Kula, T.; Timms, R.T.; Lee, I.H.; Leng, Y.; Elledge, S.J. MGH COVID-19 Collection & Processing Team Viral epitope profiling of COVID-19 patients reveals cross-reactivity and correlates of severity. *Science* 2020, 370, 4250.
53. Schmidt, M.E.; Varga, S.M. The CD8 T Cell Response to Respiratory Virus Infections. *Front. Immunol.* 2018, 9, 678.
54. Peng, Y.; Mentzer, A.J.; Liu, G.; Yao, X.; Yin, Z.; Dong, D.; Dong, T. Broad and strong memory CD4+ and CD8+ T cells induced by SARS-CoV-2 in UK convalescent individuals following COVID-19. *Nat. Immunol.* 2020, 21, 1336–1345.
55. Grifoni, A.; Weiskopf, D.; Ramirez, S.I.; Mateus, J.; Dan, J.M.; Moderbacher, C.R.; Rawlings, S.A.; Sutherland, A.; Premkumar, L.; Jadi, R.S.; et al. Targets of T cell responses to SARS-CoV-2 coronavirus in humans with COVID-19 disease and unexposed individuals. *Cell* 2020, 181, 1489–1501.e1415.
56. Ni, L.; Ye, F.; Cheng, M.L.; Feng, Y.; Deng, Y.Q.; Zhao, H.; Dong, C. Detection of SARS-CoV-2-specific humoral and cellular immunity in COVID-19 convalescent individuals. *Immunity* 2020, 52, 971–977.e973.
57. Goel, R.R.; Painter, M.M.; Apostolidis, S.A.; Mathew, D.; Meng, W.; Rosenfeld, A.M.; Lundgreen, K.A.; Reynaldi, A.; Khoury, D.S.; Pattekar, A.; et al. mRNA Vaccination Induces Durable Immune Memory to SARS-CoV-2 with Continued Evolution to Variants of Concern. *Biorxiv* 2021.
58. Urbanowicz, R.A.; Tsoleridis, T.; Jackson, H.J.; Cusin, L.; Duncan, J.D.; Chappell, J.G.; Ollivere, B.J. Two doses of the SARS-CoV-2 BNT162b2 vaccine enhances antibody responses to variants in individuals with prior SARS-CoV-2 infection. *Sci. Transl. Med.* 2021, 13, 253.
59. González, S.; Olszevicki, S.; Salazar, M.; Calabria, A.; Regairaz, L.; Marín, L.; Campos, P.; Varela, T.; Martínez, V.V.G.; Ceriani, L.; et al. Effectiveness of the first component of Gam-COVID-Vac (Sputnik V) on reduction of SARS-CoV-2 confirmed infections, hospitalisations and mortality in patients aged 60-79: A retrospective cohort study in Argentina. *EClinicalMedicine* 2021, 40, 101126.
60. Rossi, A.H.; Ojeda, D.S.; Varese, A.; Sanchez, L.; Ledesma, M.M.G.L.; Mazzitelli, I.; Juliá, A.A.; Rouco, S.O.; Pallarés, H.M.; Navarro, G.S.C.; et al. Sputnik V Vaccine Elicits Seroconversion and Neutralizing Capacity to SARS CoV-2 after a Single Dose. *Cell Rep. Med.* 2021, 2, 100359.
61. Claro, F.; Silva, D.; Rodriguez, M.; Rangel, R.; de Waard, J.H. IgG Antibody response to the Sputnik V vaccine: Previous SARS-CoV-2 seropositive individuals might need just one vaccine dose. *Int. J. Infect. Dis.* 2021, 111, 261–266.
62. Mudd, P.A.; Crawford, J.C.; Turner, J.S.; Souquette, A.; Reynolds, D.; Bender, D.; Bosanquet, J.P.; Anand, N.J.; Striker, D.A.; Martin, R.S.; et al. Distinct inflammatory profiles distinguish COVID-19 from influenza with limited contributions from cytokine storm. *Sci. Adv.* 2020, 6, eabe3024.
63. Skibinski, D.A.G.; Jones, L.A.; Zhu, Y.O.; Xue, L.W.; Au, B.; Lee, B.; Naim, A.N.M.; Lee, A.; Kaliaperumal, N.; Low, J.G.H.; et al. Induction of Human T-cell and Cytokine Responses Following Vaccination with a Novel Influenza Vaccine. *Sci. Rep.* 2018, 8, 18007.
64. Del Valle, D.M.; Kim-Schulze, S.; Huang, H.H.; Beckmann, N.D.; Nirenberg, S.; Wang, B.; Lavin, Y.; Swartz, T.H.; Madduri, D.; Stock, A.; et al. An inflammatory cytokine signature predicts COVID-19 severity and survival. *Nat. Med.* 2020, 26, 1636–1643.
65. Waltuch, T.; Gill, P.; Zinns, L.E.; Whitney, R.; Tokarski, J.; Tsung, J.W.; Sanders, J.E. Features of COVID-19 post-infectious cytokine release syndrome in children presenting to the emergency department. *Am. J. Emerg. Med.* 2020, 38, 2246.e3–2246.e6.
66. Holdsworth, S.R.; Gan, P.Y. Cytokines: Names and Numbers You Should Care About. *Clin. J. Am. Soc. Nephrol.* 2015, 10, 2243–2254.
67. Liu, Y.; Soh, W.T.; Kishikawa, J.I.; Hirose, M.; Nakayama, E.E.; Li, S.; Arase, H. An infectivity-enhancing site on the SARS-CoV-2 spike protein targeted by antibodies. *Cell* 2021, 184, 3452–3466.e18.

68. Zheng, S.; Fan, J.; Yu, F.; Feng, B.; Lou, B.; Zou, Q.; Liang, T. Viral load dynamics and disease severity in patients infected with SARS-CoV-2 in Zhejiang province, China, January-March 2020: Retrospective cohort study. *BMJ* 2020, 369, m1443.
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