

# Application in Wound Healing

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Wound healing requires careful, directed, and effective therapies to prevent infections and accelerate tissue regeneration. In light of these demands, active biomolecules with antibacterial properties and/or healing capacities have been functionalized onto nanostructured polymeric dressings and their synergistic effect examined. In this work, various antibiotics, nanoparticles, and natural extract-derived products that were used in association with electrospun nanocomposites containing cellulose, cellulose acetate and different types of nanocellulose (cellulose nanocrystals, cellulose nanofibrils, and bacterial cellulose) have been reviewed. The impact of these combinations in wound healing are here examined and explored.

Keywords: cellulose ; cellulose acetate ; nanocellulose ; biomolecules functionalization ; natural-extracts ; nanofibrous dressings ; antimicrobial surfaces ; tissue regeneration

## 1. Introduction

In wound care, infections are a major concern, since they delay the healing process, leading to tissue disfigurement or even patient death. *Staphylococcus aureus* and *Pseudomonas aeruginosa* are the most common bacteria that are isolated from chronic wounds, being *S. aureus* usually detected on top of the wound and *P. aeruginosa* in the deepest regions. They can express virulence factors and surface proteins that affect wound healing. The co-infection of *S. aureus* and *P. aeruginosa* is even more problematic, since the virulence is increased; both bacteria have intrinsic and acquired antibiotic resistance, making the clinical management of these infections a real challenge <sup>[1]</sup>. In fact, the World Health Organization considers *P. aeruginosa* as one of the organisms in urgent need for novel, highly effective antibacterial strategies that combat its prevalence. Multiple strains of *S. aureus*, including methicillin-resistant and vancomycin-resistant strains, have been identified as high priority microbes in the fight against antimicrobial resistance build up <sup>[2]</sup>. In addition to the above, other microorganisms, such as beta-hemolytic streptococci, and mixtures of Gram-negative species, such as *Escherichia coli* and *Klebsiella* strains, are also present in wounds. Bacterium native to human skin such as *Staphylococcus epidermidis* (Gram-positive), may also turn pathogenic when exposed to systemic circulation in the wound bed <sup>[3]</sup>. Therefore, immediate care of open wounds is pivotal in preventing infection <sup>[4]</sup>. To treat this problem, new alternatives of wound dressings have emerged with incorporated bioactive agents that are capable of fighting these infections and accelerating the healing process.

## 2. Application in Wound Healing: Synergistic Effect with Specialized Biomolecules

The performance of bioactive dressings processed via electrospinning is dependent on the polymer or polymer blends properties (i.e. hydrophilicity and hydrophobicity), drug solubility, drug-polymer synergy, and mat structure. Antimicrobial agent-loaded electrospun mats have shown superior performance to films produced by other techniques, in regard to water uptake (four to five times superior), water permeability, drug release rate, and antibacterial activity <sup>[5]</sup>.

Drugs, nanoparticles, and natural extracts (Table 1) are some of the antimicrobial agents that have been incorporated in nanofibrous dressings, in order to reduce the risk of infection <sup>[6]</sup>. These compounds have been used for their anti-inflammatory, pain-relieving, vasodilation, and antimicrobial features <sup>[7]</sup>.

**Table 1.** Examples of compounds incorporated in electrospun nanostructures containing cellulose or its derivatives.

Subtract	Drugs	Nanoparticles	Natural Extracts	Ref.
Cellulose	Tetracycline hydrochloride (TH) Ciprofloxacin (CIF) Donepezil hydrochloride (DNP)	Silver NPs (AgNPs) Zinc oxide NPs (ZnONPs)	Bromelain	[8][9][10][11][12][13][14]
CA	TH Ferulic acid (FA) Ibuprofen (IBU) Ketoprofen (KET) Amoxicillin Thymoquinone (TQ) Silver salt of sulfadiazine (SSD)	Silver Titanium dioxide Zinc oxide Copper	Cinnamon (CN); Lemongrass (LG); Peppermint (PM) Rosemary; Oregano Thymol Zein Asiaticoside (AC) Curcumin (Curc) Acid gallic Gingerol Garlic extract	[15][16][17][18][19][20][21][22][23][24][25][26] [27][28][29][30][31][32][33][34]
CNC	TH	ZnO AgNPs	Thymol	[35][36][37][38][39][40][41]
BC		Soy protein particles Graphene oxide (GO)	Tragacanth gum (TG)	[42][43][44]

Several researchers claim that producing cellulose-based electrospun mats is a big challenge due to its highly crystalline structure, long chain length, increased rigidity, and strong inter- and intramolecular hydrogen bonding [45]. Selecting a proper solvent, adding other complementary polymers, or converting cellulose into its derivatives can facilitate this task. The solvents or solvent systems most used for cellulose are the ionic liquids (ILs), aqueous alkali/solvents (NaOH/urea), and polar aprotic solvents in combination with electrolytes (DMAc/LiCl); however, these are not very volatile, not being completely removed during electrospinning and, thus, limiting the use of cellulose in large scale productions. A proper solvent system is also very important in attaining appropriate viscosity levels, required for a successful electrospinning process. In fact, this is such an important processing parameter that to guarantee proper polymer solubilization, heaters have been placed within the electrospinning apparatus generating a new system, the melt-electrospinning (minimize the viscosity of spinning dopes) [46]. The option of transforming cellulose into its derivatives, such as cellulose acetate (CA), cellulose acetate phthalate (CAP), *ethyl cellulose* (EC), carboxymethyl cellulose (CMC), hydroxypropylcellulose (HPC), among others, is by far the most recurrent alternative to reduce the complexity of processing cellulose via electrospinning. Besides, most of these derivatives require different pHs for solubilization, which is a great advantage for biomedical applications [47].

Modifications have been proposed to increase the effectiveness of immobilized drugs, natural compounds, peptides, or other biomolecules within a cellulose-based nanostructured surface. For example, Nada et al. activated CA by introducing azide functional groups on the residual -OH groups of the polymeric chains, enhancing the release kinetics of capsaicin and sodium diclofenac from the electrospun mat and, thus, promoting patient relief [48]. To confer biocidal properties to CA nanofibers, Jiang et al. modified their surface with 4,4'-diphenylmethane diisocyanate (MDI). This resulted in a 100% inactivation of *S. aureus* and a 95% of *E. coli* within 10 min of exposure, and complete death after a 30 min contact [49]. Nano complexes with cellulose nanocrystals (CNCs) were developed with cationic  $\beta$ -cyclodextrin (CD) containing curcumin by ionic association and used in the treatment of colon and prostate cancers [50]. Nanocellulose has also contributed to the development of new and more efficient strategies for these biomolecules' delivery. The three -OH groups that were present in each individual glucose unit originate a highly reactive structure, which allows interaction with other molecules or with enzymes and/or proteins, contributing to overcome the low solubility of most drugs in aqueous medium [51]. Besides, the -OH groups can also be tailored by physical adsorption, surface graft polymerization, and covalent bonding to further improve the performance of the biomolecules. As a consequence of the bonds established, strong polymer-filler interactions are generated, significantly increasing the mechanical properties of material [52].

Nonetheless, the in vivo behavior of nanocelluloses is still little explored. Studies have reported that its toxicity depends on the solution concentration and its surface charges. In recent literature, nanocelluloses have not shown any toxicity at concentrations lower than 1 mg/mL; however, there are studies that reveal a concentration-dependent apoptotic toxicity of cellulose nanofibrils (CNFs) at 2–5 mg/mL. Additionally, anionic nanocelluloses, e.g., carboxymethylated-CNF, have been reported to be more cytotoxic than cationic nanocelluloses, e.g., trimethylammonium-CNF [53]. Toxicity effects might arise from the diversity of chemical structures and properties between cellulose types and sources. Among nanocelluloses, bacterial cellulose (BC) is considered to be the most biocompatible and has already been applied in wound dressings [54]. Still, its electrospinnability is very challenging for the same structural reasons of cellulose [45].

The incorporation of BC into synthetic and natural polymers has been carried out to enhance their morphological features as well as physicochemical and biological performances. A wide variety of polymers, such as chitosan, polycaprolactone (PCL), polyethylene oxide (PEO), ethylene vinyl alcohol (EVOH), polyvinyl alcohol (PVA), polylactic acid (PLA), polyacrylonitrile (PAN), polyester, silk, and zein, have been blended with BC and processed by electrospinning. Functionalization with 3-aminopropyl triethoxysilane (APS) has been attempted to further enhance cell attachment and antibacterial properties of BC-containing electrospun membranes for wound healing. BC membranes grafted with two organosilanes and acetylated have also shown an improved moisture resistance and hydrophobicity [55]. Naeem et. al even synthesized in situ BC on CA-based electrospun mats in a process known by self-assembly to produce a new generation of wound dressings [56].

Even though CNF has already been applied as a reinforcing agent in many polymeric composites via electrospinning, no reports have been found regarding the incorporation of biomolecules along its fibers [57].

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## References

1. Serra, R.; Grande, R.; Butrico, L.; Rossi, A.; Caroleo, B.; Amato, B.; Gallelli, L.; Franciscis, S. De Chronic wound infections: The role of *Pseudomonas aeruginosa* and *Staphylococcus aureus*. *Expert Rev. Anti Infect. Ther.* 2015, 13, 605–613.
2. Dart, A.; Bhawe, M.; Kingshott, P. Antimicrobial Peptide-Based Electrospun Fibers for Wound Healing Applications. *Macromol. Biosci.* 2019, 1800488, 1–16.
3. Buch, P.J.; Chai, Y.; Goluch, D. Treating Polymicrobial Infections in Chronic Diabetic Wounds. *Clin. Microbiol. Rev.* 2019, 32, e00091-18.
4. Unnithan, A.R.; Barakat, N.A.M.; Pichiah, P.B.T.; Gnanasekaran, G.; Nirmala, R.; Cha, Y.; Jung, C.; El-newehy, M.; Yong, H. Wound-dressing materials with antibacterial activity from electrospun polyurethane—Dextran nanofiber mats containing ciprofloxacin HCl. *Carbohydr. Polym.* 2012, 90, 1786–1793.
5. Unnithan, A.R.; Gnanasekaran, G.; Sathishkumar, Y.; Lee, Y.S.; Kim, C.S. Electrospun antibacterial polyurethane-cellulose acetate-zein composite mats for wound dressing. *Carbohydr. Polym.* 2014, 102, 884–892.
6. Miguel, S.P.; Figueira, D.R.; Simões, D.; Ribeiro, M.P.; Coutinho, P.; Ferreira, P.; Correia, I.J. Electrospun polymeric nanofibres as wound dressings: A review. *Colloids Surf. B Biointerfaces* 2018, 169, 60–71.
7. Ambekar, R.S.; Kandasubramanian, B. Advancements in nano fi bers for wound dressing: A review. *Eur. Polym. J.* 2019, 117, 304–336.
8. Esmaeili, A.; Haseli, M. Electrospinning of thermoplastic carboxymethyl cellulose/poly (ethylene oxide) nano fibers for use in drug-release systems. *Mater. Sci. Eng. C* 2017, 77, 1117–1127.
9. Li, H.; Zhang, Z.; Godakanda, V.U.; Chiu, Y.; Angkawinitwong, U.; Patel, K.; Stapleton, P.G.; De Silva, R.M.; De Silva, K.M.N.; Zhu, L.; et al. The effect of collection substrate on electrospun ciprofloxacin-loaded poly (vinylpyrrolidone) and ethyl cellulose nanofibers as potential wound dressing materials. *Mater. Sci. Eng. C* 2019, 104, 109917.
10. Journal, A.I.; Gencturk, A.; Kahraman, E.; Güngör, S.; Özhan, G.; Özsoy, Y.; Sarac, A.S.; Kahraman, E.; Güngör, S.; Özhan, G.; et al. Polyurethane/hydroxypropyl cellulose electrospun nanofiber mats as potential transdermal drug delivery system: Characterization studies and in vitro assays. *Artif. Cells Nanomed. Biotechnol.* 2017, 45, 655–664.
11. Shi, D.; Wang, F.; Lan, T.; Zhang, Y.; Shao, Z.; Nanofiber, E.Á.; Ag, Á. Convenient fabrication of carboxymethyl cellulose electrospun nanofibers functionalized with silver nanoparticles. *Cellulose* 2016, 23, 1899–1909.
12. Jatoi, A.W.; Kim, I.S.; Ni, Q.Q. A comparative study on synthesis of AgNPs on cellulose nanofibers by thermal treatment and DMF for antibacterial activities. *Mater. Sci. Eng. C* 2019, 98, 1179–1195.
13. Darbasizadeh, B.; Fatahi, Y.; Feyzi-barnaji, B.; Arabi, M.; Motasadizadeh, H.; Farhadnejad, H.; Moraffah, F.; Rabiee, N. Crosslinked-polyvinyl alcohol-carboxymethyl cellulose/ZnO nanocomposite fibrous mats containing erythromycin (PVA-

14. De Melo Brites, M.; Cerón, A.A.; Costa, S.M.; Oliveira, R.C.; Ferraz, H.G.; Catalani, L.H.; Costa, S.A. Bromelain immobilization in cellulose triacetate nanofiber membranes from sugarcane bagasse by electrospinning technique. *Enzym. Microb. Technol.* 2020, 132, 109384.
15. Liao, N.; Rajan, A.; Kumar, M.; Prasad, A.; Tshool, S.; Park, C.; Sang, C. Electrospun bioactive poly (E-caprolactone)–cellulose acetate—Dextran antibacterial composite mats for wound dressing applications. *Colloids Surf. A Physicochem. Eng. Asp.* 2015, 469, 194–201.
16. Chen, Y.; Qiu, Y.; Chen, W.; Wei, Q. Electrospun thymol-loaded porous cellulose acetate fibers with potential biomedical applications. *Mater. Sci. Eng. C* 2020, 109, 110536.
17. Liakos, I.L.; Holban, A.M.; Carzino, R.; Lauciello, S.; Grumezescu, A.M. Electrospun Fiber Pads of Cellulose Acetate and Essential Oils with Antimicrobial Activity. *Nanomaterials* 2017, 7, 84.
18. Liu, X.; Yang, Y.; Yu, D.; Zhu, M.; Zhao, M.; Williams, G.R. Tunable zero-order drug delivery systems created by modified triaxial electrospinning. *Chem. Eng. J.* 2019, 356, 886–894.
19. Brako, F.; Luo, C.; Craig, D.Q.M.; Edirisinghe, M. An Inexpensive, Portable Device for Point-of-Need Generation of Silver-Nanoparticle Doped Cellulose Acetate Nanofibers for Advanced Wound Dressing. *Macromol. Mater. Eng.* 2018, 303, 1700586.
20. Liakos, I.; Rizzello, L.; Hajiali, H.; Brunetti, V.; Carzino, R.; Pompa, P.P. Fibrous wound dressings encapsulating essential oils as natural antimicrobial agents. *J. Mater. Chem. B* 2015, 3, 1583–1589.
21. Yang, Y.; Li, W.; Yu, D.; Wang, G.; Williams, G.R. Tunable drug release from nano fibers coated with blank cellulose acetate layers fabricated using tri-axial electrospinning. *Carbohydr. Polym.* 2019, 203, 228–237.
22. Wahab Jatoi, A.; Kim, I.S.; Ni, Q. Cellulose acetate nano fibers embedded with AgNPs anchored TiO<sub>2</sub> nanoparticles for long term excellent antibacterial applications. *Carbohydr. Polym.* 2019, 207, 640–649.
23. Yu, D.; Yu, J.; Chen, L.; Williams, G.R.; Wang, X. Modified coaxial electrospinning for the preparation of high-quality ketoprofen-loaded cellulose acetate nanofibers. *Carbohydr. Polym.* 2012, 90, 1016–1023.
24. Wahab, A.; Soo Kim, I.; Ogasawara, H.; Ni, Q. Characterizations and application of CA/ZnO/AgNP composite nano fibers for sustained antibacterial properties. *Mater. Sci. Eng. C* 2019, 105, 110077.
25. Castillo-Ortega, M.; Nájera-Luna, A.; Rodríguez-Félix, D.E.; Encinas, J.C.; Rodríguez-Félix, F.; Romero, J.; Herrera-Franco, P.J. Preparation, characterization and release of amoxicillin from cellulose acetate and poly (vinyl pyrrolidone) coaxial electrospun fibrous membranes Preparation, characterization and release of amoxicillin from cellulose acetate and poly (vinyl pyrrolidone). *Mater. Sci. Eng. C* 2011, 31, 1772–1778.
26. Quirós, J.; Gonzalo, S.; Jalvo, B.; Boltes, K.; Perdigón-melón, J.A.; Rosal, R. Electrospun cellulose acetate composites containing supported metal nanoparticles for antifungal membranes. *Sci. Total Environ.* 2016, 563, 912–920.
27. Gomaa, S.F.; Madkour, T.M.; Moghannem, S.; El-Sherbiny, I.M. New polylactic acid/cellulose acetate-based antimicrobial interactive single dose nanofibrous wound dressing mats. *Int. J. Biol. Macromol.* 2017, 105, 1148–1160.
28. Khan, M.Q.; Kharaghani, D.; Shahzad, A.; Saito, Y.; Yamamoto, T.; Ogasawara, H.; Kim, I.S. Fabrication of antibacterial electrospun cellulose acetate/silver-sulfadiazine nanofibers composites for wound dressings applications. *Polym. Test.* 2019, 74, 39–44.
29. Rajan, A.; Gnanasekaran, G.; Sathishkumar, Y.; Soo, Y.; Sang, C. Electrospun antibacterial—Cellulose acetate—Zein composite mats for wound dressing. *Carbohydr. Polym.* 2014, 102, 884–892.
30. Suwantong, O.; Ruktanonchai, U.; Supaphol, P. Electrospun cellulose acetate fiber mats containing asiaticoside or *Centella asiatica* crude extract and the release characteristics of asiaticoside. *Polymer* 2008, 49, 4239–4247.
31. Suwantong, O.; Opanasopit, P.; Ruktanonchai, U.; Supaphol, P. Electrospun cellulose acetate fiber mats containing curcumin and release characteristic of the herbal substance. *Polymer* 2007, 48, 7546–7557.
32. Phiriyawirut, M.; Phaeachamud, T. Gallic Acid-loaded Cellulose Acetate Electrospun Nanofibers: Thermal Properties, Mechanical Properties, and Drug Release Behavior. *Open J. Polym. Chem.* 2012, 2012, 21–29.
33. Chantarodsakun, T.; Vongsetskul, T. [6]-Gingerol-loaded cellulose acetate electrospun fibers as a topical carrier for controlled release. *Polym. Bull.* 2014, 71, 3163–3176.
34. Edikresnha, D.; Suciati, T.; Munir, M.M.; Khairurrijal, K. Polyvinylpyrrolidone/cellulose acetate electrospun composite nanofibres loaded by glycerine and garlic extract with in vitro antibacterial activity and release behaviour test. *RSC Adv.* 2019, 9, 26351–26363.

35. Cheng, M.; Qin, Z.; Hu, S.; Dong, S.; Ren, Z.; Yu, H. Achieving Long-Term Sustained Drug Delivery for Electrospun Biopolyester Nano fibrous Membranes by Introducing Cellulose Nanocrystals. *ACS Biomater. Sci. Eng.* 2017, 3, 1666–1676.
36. Hivechi, A.; Bahrami, S.H.; Siegel, R.A. Drug release and biodegradability of electrospun cellulose nanocrystal reinforced polycaprolactone. *Mater. Sci. Eng. C* 2019, 94, 929–937.
37. Yu, H.; Wang, C.; Abdalkarim, S. Cellulose nanocrystals/polyethylene glycol as bifunctional reinforcing/compatibilizing agents in poly (lactic acid) nanofibers for controlling long-term in vitro drug release. *Cellulose* 2017, 24, 4461–4467.
38. Abdalkarim, S.Y.H.; Yu, H.Y.; Wang, D.; Yao, J. Electrospun poly(3-hydroxybutyrate-co-3-hydroxy-valerate)/cellulose reinforced nanofibrous membranes with ZnO nanocrystals for antibacterial wound dressings. *Cellulose* 2017, 24, 2925–2938.
39. Alvarado, N.; Romero, J.; Torres, A.; Carol, L.; Guarda, A. Supercritical impregnation of thymol in poly (lactic acid) filled with electrospun poly (vinyl alcohol) cellulose nanocrystals nano fibers: Development an active food packaging material. *J. Food Eng. J.* 2018, 217, 1–10.
40. Lu, Z.; Gao, J.; He, Q.; Wu, J.; Liang, D.; Yang, H.; Chen, R. Enhanced antibacterial and wound healing activities of microporous chitosan-Ag/ZnO composite dressing. *Carbohydr. Polym.* 2017, 156, 460–469.
41. Tian, J.; Wong, K.K.Y.; Ho, C.; Lok, C.; Yu, W.; Che, C.; Chiu, J.; Tam, P.K.H. Topical Delivery of Silver Nanoparticles Promotes Wound Healing. *ChemMedChem* 2007, 2, 129–136.
42. Zhijiang, C.; Ping, X.; Shiqi, H.; Cong, Z. Soy protein nanoparticles modified bacterial cellulose electrospun nanofiber membrane scaffold by ultrasound-induced self-assembly technique: Characterization and cytocompatibility. *Cellulose* 2019, 26, 6133–6150.
43. Azarniya, A.; Eslahi, N.; Mahmoudi, N.; Simchi, A. Effect of graphene oxide nanosheets on the physico-mechanical properties of chitosan/bacterial cellulose nanofibrous composites. *Compos. Part A Appl. Sci. Manuf.* 2016, 85, 113–122.
44. Azarniya, A.; Tamjid, E.; Eslahi, N.; Simchi, A. Modification of bacterial cellulose/keratin nanofibrous mats by a tragacanth gum-conjugated hydrogel for wound healing. *Int. J. Biol. Macromol.* 2019, 134, 280–289.
45. Ardila, N.; Medina, N.; Arkoun, M.; Heuzey, M.C.; Ajji, A.; Panchal, C.J. Chitosan–bacterial nanocellulose nanofibrous structures for potential wound dressing applications. *Cellulose* 2016, 23, 3089–3104.
46. Xu, H.; Bronner, T.; Yamamoto, M.; Yamane, H. Regeneration of cellulose dissolved in ionic liquid using laser-heated melt-electrospinning. *Carbohydr. Polym.* 2018, 201, 182–188.
47. Rezaei, A.; Nasirpour, A.; Fathi, M. Application of Cellulosic Nanofibers in Food Science Using Electrospinning and Its Potential Risk. *Compr. Rev. Food Sci. Food Saf.* 2015, 14, 269–284.
48. Nada, A.A.; Hassan, F.; Abdellatif, H.; Soliman, A.A.F.; Shen, J.; Hudson, S.M. Fabrication and bioevaluation of a medicated electrospun mat based on azido-cellulose acetate via click chemistry. *Cellulose* 2019, 26, 9721–9736.
49. Li, R.; Jiang, Q.; Ren, X.; Xie, Z.; Huang, T. Electrospun non-leaching biocompatible antimicrobial cellulose acetate nanofibrous mats. *J. Ind. Eng. Chem.* 2015, 27, 315–321.
50. Ndong, G.M.A.; Granet, R.; Pierre, J.; Brégier, F.; Léger, D.Y.; Fidanzig-dugas, C.; Lequart, V.; Joly, N.; Liagre, B.; Chaleix, V.; et al. Development of curcumin—Cyclodextrin/cellulose nanocrystals complexes: New anticancer drug delivery systems. *Bioorg. Med. Chem. Lett.* 2015, 26, 10–14.
51. Löbmann, K.; Svagan, A.J. Cellulose nanofibers as excipient for the delivery of poorly soluble drugs. *Int. J. Pharm.* 2017, 533, 285–297.
52. Bacakova, L.; Pajorova, J.; Bacakova, M.; Skogberg, A.; Kallio, P.; Kolarova, K.; Svorcik, V. Versatile Application of Nanocellulose: From Industry to Skin Tissue Engineering and Wound Healing. *Nanomaterials* 2019, 9, 164.
53. Sheikhi, A.; Hayashi, J.; Eichenbaum, J.; Gutin, M.; Kuntjoro, N.; Khorsandi, D.; Khademhosseini, A. Recent advances in nanoengineering cellulose for cargo delivery. *J. Control. Release* 2019, 294, 53–76.
54. Lin, N.; Dufresne, A. Nanocellulose in biomedicine: Current status and future prospect. *Eur. Polym. J.* 2014, 59, 302–325.
55. Eslahi, N.; Mahmoodi, A.; Mahmoudi, N.; Zandi, N. Processing and Properties of Nanofibrous Bacterial Cellulose-Containing Polymer Composites: A Review of Recent Advances for Biomedical Applications Processing and Properties of Nanofibrous Bacterial Cellulose-Containing Polymer Composites: A Review of R. *Polym. Rev.* 2020, 60, 144–170.
56. Naeem, M.; Siddiqui, Q.; Leroy, A.; Khan, M.R.; Wei, Q. The production and characterization of microbial cellulose—Electrospun membrane hybrid nano-fabrics. *J. Ind. Text.* 2019.

57. Kalia, S.; Boufi, S.; Celli, A.; Kango, S. Nanofibrillated cellulose: Surface modification and potential applications. *Colloid Polym. Sci.* 2014, 292, 5–31.
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