

Neurotoxicity in Acute Lymphoblastic Leukemia Treatment

Subjects: [Hematology](#) | [Cell Biology](#)

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Immunotherapy is a milestone in the treatment of poor-prognosis pediatric acute lymphoblastic leukemia (ALL) and is expected to improve treatment outcomes and reduce doses of conventional chemotherapy without compromising the effectiveness of the therapy. However, both chemotherapy and immunotherapy cause side effects, including neurological ones.

acute lymphoblastic leukemia

neurotoxicity

immunotherapy

chemotherapy

1. Introduction

Five-year overall survival of ALL has increased over the past decades and now exceeds over 96% ^[1]. Chemotherapy is a crucial part of treating ALL and involves many cytotoxic drugs, which inhibit cancer cells from growing rapidly, but they also damage healthy cells, resulting in a wide range of adverse effects. However, with the current high rate of survival, it would be difficult to improve results with only conventional chemotherapy which has reached its maximum of tolerance and could no longer be pushed to improved results. The target should be to search for the use of intensive multimodal treatment regimens, including high-dose chemotherapy and next-generation drugs ^[2]. Precision medicine with immunotherapy and other molecularly targeted treatments offers unique opportunities to customize treatment intensity ^[3]. Their advantages also include reducing the need for hematopoietic stem cell transplantation (HSCT), decreasing the burden of toxicities, and fighting persistent residual disease. Recently approved agents for ALL include blinatumomab, inotuzumab ozogamicin (InO), and CAR T-cell therapy, which are expected to improve treatment outcomes and reduce doses of conventional chemotherapy without compromising the effectiveness of the therapy. Nevertheless, the benefits of aggressive chemotherapy versus target therapy for different patient groups remain unclear and all the strategies cause adverse events, such as neurotoxicity, hepatotoxicity, gastrointestinal complication, and secondary malignancies, making neither of these therapies an ideal treatment ^[4].

2. Neurotoxicity of Conventional Therapy

The optimal treatment doses are determined based on tolerability, response assessment, and drug pharmacodynamics and pharmacogenomics. The clinical characteristics of the patient and the biological features of the leukemia are the main factors that determine the choice of specific chemotherapeutics. Treatment protocol consists of phases such as induction, intensification, consolidation, and maintenance ^[5]. Currently, first-line

treatment protocols include a variety of medication combinations which involve the use of VCR, L-ASP, corticosteroids, antimetabolites (cytarabine and MTX), and anthracyclines [3]. However, the dose intensity of conventional chemotherapy has been pushed to its limits, and because children absorb and metabolize drugs differently than adults, toxicity is a key issue in pediatric chemotherapy. To determine further treatment development, attention must be given to some of the unique neurotoxicities associated with MTX, VCR, L-ASP, and their molecular background (Table 1).

Table 1. Treatments used in ALL and associated neurotoxicity.

Phase of Treatment	Drugs	Toxicity-Related Gene	Mechanism of Neurotoxicity	Neurotoxicity	References
Induction	Vincristine	ABCC11 ¹ , ABCC2 ² , ABCC4 ³ , ABCC5 ⁴ , ABCB1 ⁵ , ABCC10 ⁶ , CEP72 ⁷ , SLC5A7 ⁸ , TUBB1 ⁹ , TUBB2A ¹⁰ , TUBB2B ¹¹ , TUBB3 ¹² , TUBB4A ¹³ , MAP4 ¹⁴ , CYP3A4 ¹⁵ , CYP2C9 ¹⁶ , CYP3A5 ¹⁷ , CEP72 ¹⁸	Interferes with the assembly of microtubule structures leading to cell apoptosis. It affects the peripheral nerves but can also contribute to dysfunction of the cranial nerves and autonomic nervous system.	Peripheral neuropathy, sensory neuropathy, sensory/factile impairment, numbness, and tingling in the hands and feet, paresthesia, decreased balance, tendon weakening, visual and hearing problems.	[10]
	L-asparaginase	ZBTB1 ¹⁹ , GRIA1 ²⁰ , HLA-DRB1 ²¹	L-asparaginase produces three neurotoxic agents: ammonia, L-aspartic acid, and glutamic acid. These two amino acids can induce cell death in CNS neurons by excessive stimulation through NMDA (N-methyl-D-aspartate) receptor, leading to a major intracellular calcium influx and apoptosis.	Myelosuppression, encephalopathy, hepatic toxicity.	[20,24]
Consolidation	Methotrexate (intravenous infusion and intrathecally)	DHFR19bp ²² , MTHFR 677C > T ²³ , MTHFR 677TT ²⁴ , SLC19A1 ²⁵ , TYMS ²⁶ , ADORA2A ²⁷	Methotrexate is an antimetabolite that inhibits Dihydrofolate Reductase and thus tetrahydrofolate formation. This affects the synthesis	Transverse myelopathy-symptoms include back pain with subsequent weakness, sensory loss and bladder or	[11,12,13]

3. Neurotoxicity of Immunotherapy

Several of the newly discovered molecular alterations have led to the development of approaches that focus on the dysregulation of cellular pathways [19]. Despite tremendous advances in the treatment of ALL, no drug has shown

Phase of Treatment	Drugs	Toxicity-Related Gene	Mechanism of Neurotoxicity	Neurotoxicity	References
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adolescent patients with R/R B-ALL. Patients in the InO arm received 1.8 mg/m² intravenously each cycle, for a maximum of six cycles. Patients in the chemotherapy arm received cytarabine with mitoxantrone, FLAG (fludarabine, cytarabine, and granulocyte colony-stimulating factor [GCSF]), or high-dose cytarabine, as

1 ABCB1, ATP Binding Cassette Subfamily C Member 1; 2 ABCG2, ATP Binding Cassette (ABC) Family 9 Member 2; 3 ABCG4, ATP Binding Cassette Subfamily G Member 4; 4 ABCG5, ATP Binding Cassette Subfamily G Member 5; 5 ABCG10, ATP Binding Cassette Subfamily G Member 10; 6 CYP2C8, Cytochrome P450 Family 2 Subfamily C Member 8; 7 CYP2C9, Cytochrome P450 Family 2 Subfamily C Member 9; 8 SLC5A7, Solute Carrier Family 5 Member 7; 9 TUBB1, Tubulin Beta Class V; 10 TUBB2A, Tubulin Beta Class IIa; 11 TUBB2B, Tubulin Beta Class IIa; 12 TUBB3, Tubulin Beta Class III; 13 TUBB4A, Tubulin Beta Class IVa; 14 MAP4, Microtubule-Associated Protein 4; 15 CYP3A4, Cytochrome P450 Family 3 Subfamily A Member 4; 16 CYP2C19, Cytochrome P450 Family 2 Subfamily C Member 19; 17 CYP3A5, Cytochrome P450 Family 3 Subfamily A Member 5; 18 CYP2E1, Cytochrome P450 Family 2 Subfamily B Member 1; 19 CYP2D6, Cytochrome P450 Family 2 Subfamily D Member 6; 20 CYP2C18, Cytochrome P450 Family 2 Subfamily C Member 18; 21 CYP2C17, Cytochrome P450 Family 2 Subfamily C Member 17; 22 CYP2C12, Cytochrome P450 Family 2 Subfamily C Member 12; 23 CYP2C13, Cytochrome P450 Family 2 Subfamily C Member 13; 24 MTHFR 677C > T, Methylenetetrahydrofolate Reductase polymorphism; 25 SCL29A1, Solute Carrier Family 29, member 1; 26 TYMS, Thymidylate Synthetase; 27 ADORA2A, Adenosine A2a Receptor; 28 DCK, Deoxycytidine Kinase; 29 NT5C2, 5'-Nucleotidase, Cytosolic II; 30 CDA, Cytidine Deaminase; 31 RRM1, Ribonucleotide Reductase Catalytic Subunit 1; 32 GIT1, G Protein-Coupled Receptor Kinase Interacting ArfGAP 1; 33 NT5C3, 5'-Nucleotidase, Cytosolic IIIA; 34 ENT1, Equilibrative nucleoside transporter 1; 35 CNS, central nervous system.

3.3. CAR T-Cell Therapy

The last resort for acute or refractory ALL in patients up to 25 years of age are two CAR T products M1; 32 GIT1, G Protein-Coupled Receptor Kinase Interacting ArfGAP 1; 33 NT5C3, 5'-Nucleotidase, Cytosolic IIIA; 34 ENT1, Equilibrative nucleoside transporter 1; 35 CNS, central nervous system. States and Europe [31]. There are three generations of CARs (Figure 1). A CAR T-cell's basic structure usually consists of a tumor-targeting domain derived from a monoclonal antibody linked to a CD3 zeta chain that serves as an intracellular signaling domain. A co-stimulatory endodomain, either 4-1BB or CD28, is also present in second-generation CARs [32]. A third-generation CAR T-cell's purpose is to increase T-cell proliferation and persistence by combining signaling domains such as 4-1BB, OX40 (CD134), inducible T-cell costimulator (ICOS), and CD27, to boost the cytotoxic effect [33].

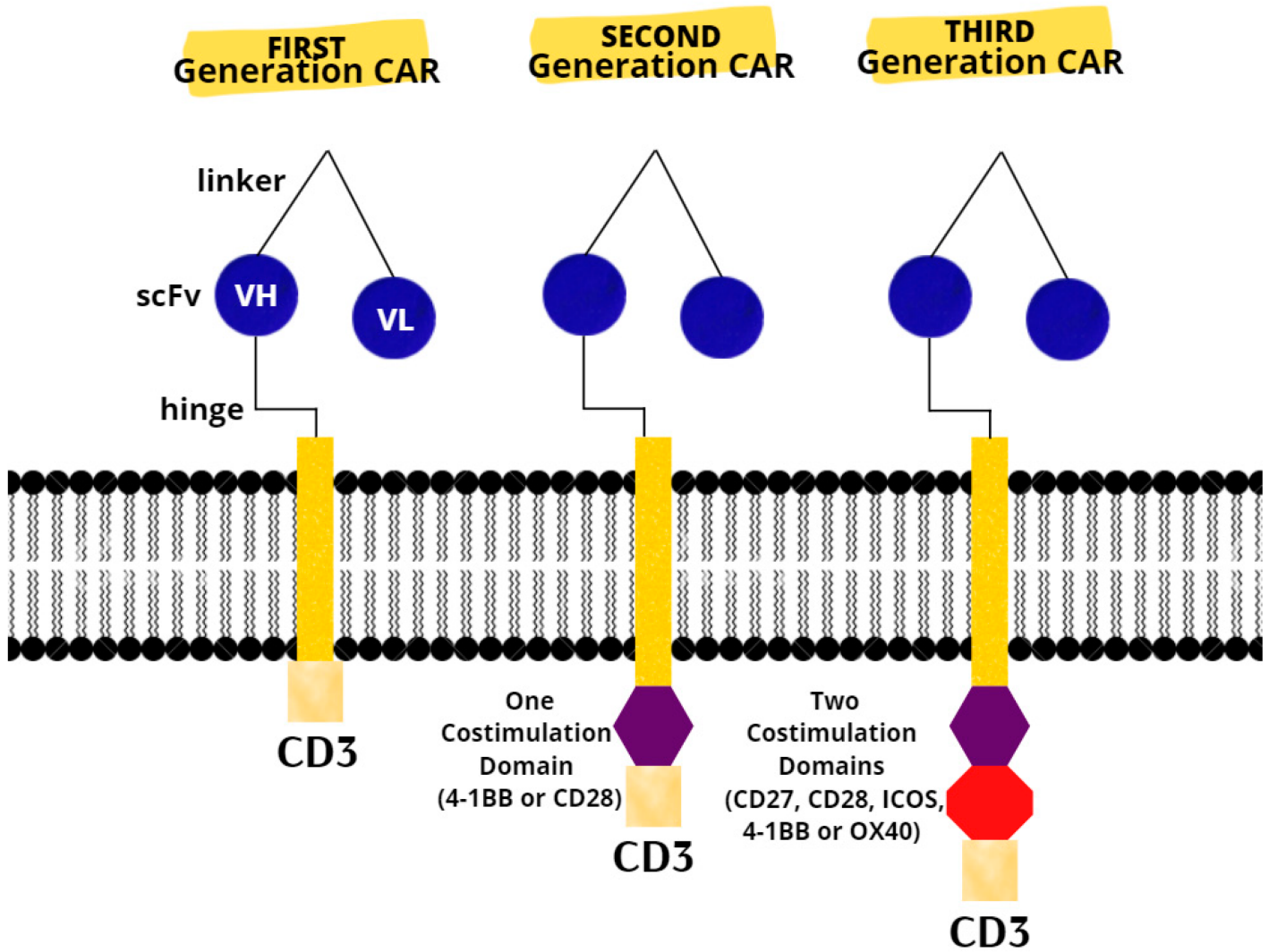


Figure 1. Chimeric antigen receptors. Next-generation CARs have additional modifications to their intracellular stimulatory domains. CD3, cluster of differentiation 3; ICOS, Inducible T-cell costimulator; scFv, single-chain fragment variable; VH, heavy chain variable gene segment; VL, variable region.

Although the exact mechanism of neurotoxicity is unknown, evidence shows endothelial activation and increased blood–brain barrier permeability, which results in a high cytokine concentration in the cerebrospinal fluid. These cause more endothelial cell and pericyte activation, which, if severe, might result in cerebral edema or other CAR T neurotoxicity manifestations (**Figure 2**). In contrast to the previously assumed mediators derived from T lymphocytes, after CAR T infusion the researchers observed a significant increase in the level of cytokines, including granulocyte-macrophage colony growth factor (GM-CSF), IL-10, IL-6, and IL-1 and IL-6 generated from host macrophages that have been shown to mediate neurotoxicity. Depending on the kinetics of T-cell proliferation, CRS with CAR T might develop shortly after infusion or be a delayed response that occurs days or weeks later ^[34].

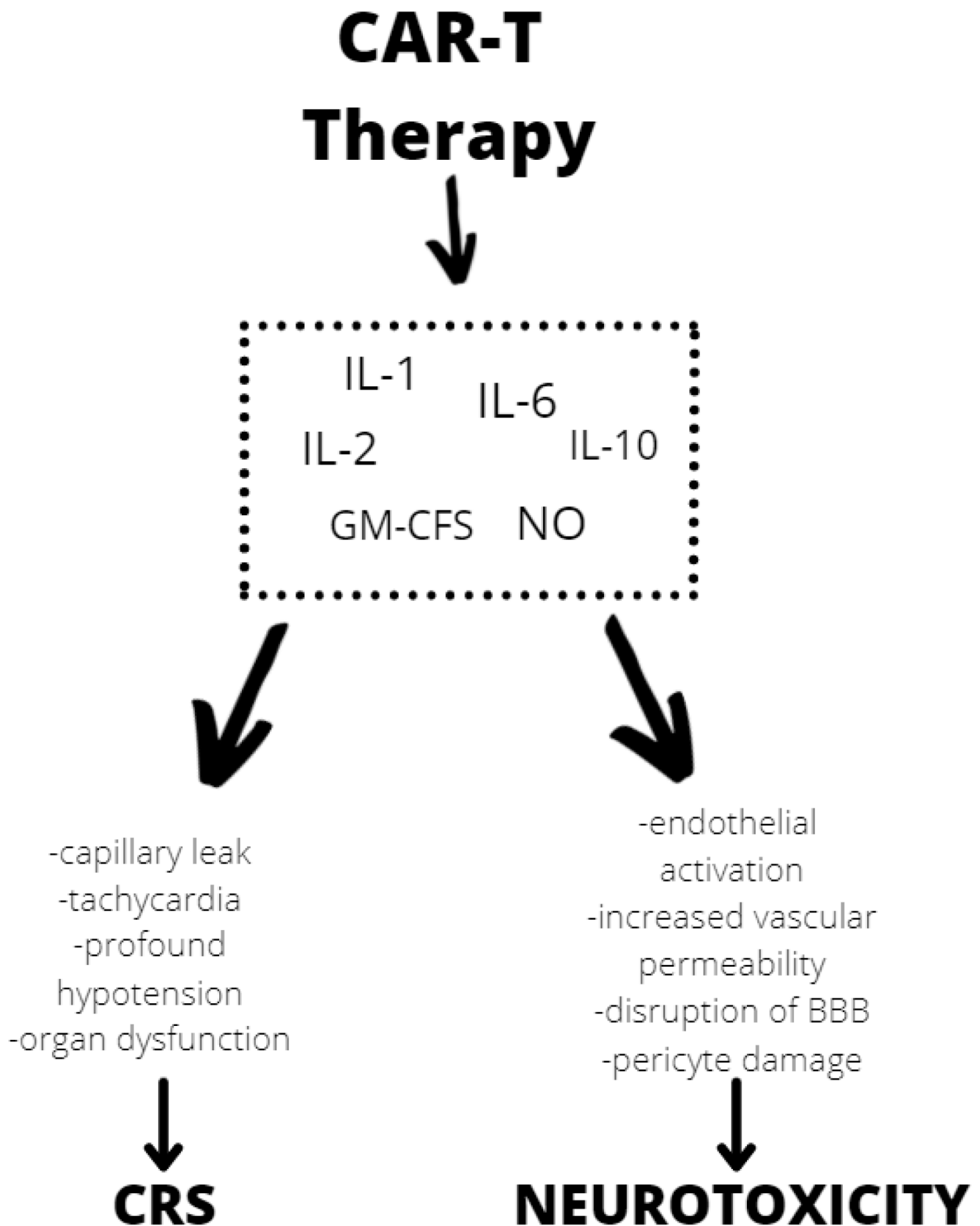


Figure 2. Mechanisms of neurotoxicity and cytokine release syndrome (CRS) caused by CAR T therapy. BBB—blood–brain barrier, GM-CFS—granulocyte-macrophage colony stimulating factor, NO—nitric oxide.

The evaluation and classification of these toxicities vary greatly between clinical trials and institutions, making it difficult to compare the safety of different medicines and to create effective care methods for these toxicities. The first patients treated with CD19 CAR T-cells experienced similar toxicities to those observed with the immunomodulatory drug theralizumab (TGN1412), including aggressive behavior, rigidity, fever, poor concentration, and psychosis [35]. Supraphysiologic cytokine increase was found to be responsible for the great majority of symptoms in the first pediatric ALL patient treated with CAR T-cell therapy, implying that these toxicities were caused by CRS. Encephalopathy, agitation, aphasia, tremor, lethargy, delirium, difficulty concentrating, seizures, and in rare cases even cerebral edema are all symptoms of ICANS [36]. In addition, headache is a very common symptom that may or may not indicate neurotoxicity (**Figure 3**). Over the years, many scales have been developed, such as CTCAE (Common Terminology Criteria for Adverse Events) version 4.03, CTCAE version 5.0, Lee criteria, and Peen criteria et al., which redefined the scoring criteria for CRS. Many CAR T-cell groups have adopted the Lee criterion, in part because it was the first to link a specific grade to a suggested therapy protocol. As previously indicated, neurotoxicity is a common side effect of CAR T-cell and other T-cell-engaging therapies. Neurotoxicity associated with immune effector cells, unlike classic CRS symptoms, do not often respond to tocilizumab treatment. This fact is not surprising, given that when tocilizumab is administered I.V., large amounts of the drug do not accumulate in the CSF. Symptoms of ICANS can be more diverse than those of CRS. Many patients with neurotoxicity have a stereotypic evolution of a specific set of symptoms. The earliest manifestations of ICANS are mild difficulty with expressive speech (especially in naming objects), dysgraphia, impaired attention, tremor, apraxia, and mild lethargy [37]. Early detection of marker changes such as peak C-reactive protein (CRP), ferritin on day 3 but not peak ferritin, and fever can have a huge impact on the prognosis of patients who develop neurotoxicity. In people at risk of neurotoxicity, the number of cytokines such as IL-6, IL-10, granulocyte-macrophage colony stimulating factor, IL-15, IL-2, and the TNF receptor should also be determined. However, it is important to realize that none of these markers are unique to chemotherapy or immunotherapy or CRS induced neurotoxicity. Only the combination of several ingredients gives a picture of whether a given patient is at risk of neurotoxicity. High-dose methylprednisolone is typically used in the most severe cases of CAR T-cell-related neurotoxicity. Most neurotoxic patients also have CRS, which can be treated with tocilizumab, an IL-6 receptor antibody, and/or corticosteroids to suppress T-cell activation, but the effect of these medications on neurotoxicity is uncertain [38].

CRS grading based on ICANS

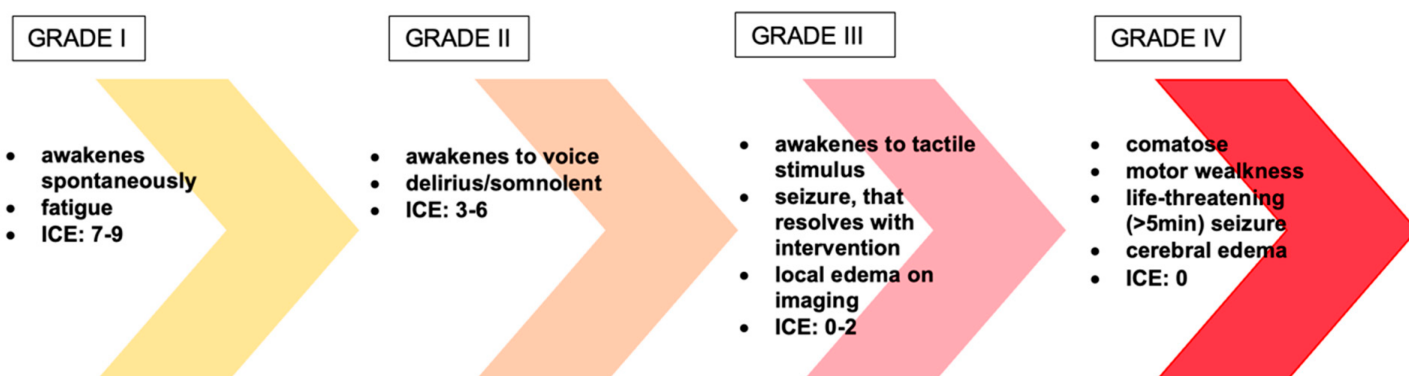


Figure 3. The management of ICANS is based on a grading system. CRS—cytokine release syndrome, ICANS—immune effector cell-associated neurotoxicity syndrome, ICE—Immune Effector Cell-Associated Encephalopathy score.

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