Chlorella vulgaris Biomass

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Chlorella vulgaris biomass means the biomass made of Chlorella vulgaris, which is a kind of microalgae. Under appropriate conditions, microalgae convert solar energy into chemical energy stored as starch or lipids, which are precursors for bioethanol and biodiesel production. Given the higher photosynthetic efficiency, higher biomass production per unit area and faster growth rate compared to energy crops, microalgae are good alternative as feedstock for biofuel production. An additional advantage of microalgae is the lack of competition for nutrients with food crops. Furthermore, biomass production can be located on marginal lands. The negative environmental impact associated with the cultivation of microalgae for energy purposes is described as potentially negligible.

Keywords: sodium biocarbonite; carbon dioxide; biofixation; microalgal biomass; lipids content

1. Introduction

The main source of energy in the world, also used for fuel production, is still crude oil $^{[1]}$. Limited fossil fuel resources and adverse environmental impact due to greenhouse gas emissions increased interest in advanced fuel production technologies $^{[2]}$. The primary feedstocks used for their production are obtained from energy crops or lignocellulosic wastes. Less conventional sources include the biomass of macroalgae and microalgae $^{[3][4]}$.

Microalgae are unicellular or multicellular simple organisms that are metabolically diverse, but most of them are photoautotrophs ^[5]. A valuable property of theirs is that their fast biomass growth, which per hectare is several times higher compared to terrestrial plants ^[6], but just like plants, microalgae require nutrients, light and carbon dioxide to grow ^[7].

The main problems of algal biofuel production are related to cultivation costs and biomass dehydration processes $\frac{[3][9]}{[9]}$. To increase the cost-effectiveness of biomass production, nutrients contained in municipal wastewater $\frac{[10][11]}{[12]}$ or in aquaculture wastewater $\frac{[12]}{[12]}$, are used in cultivation. Microalgae have a high ability to remove nitrogen and phosphorus compounds, thus biomass production can be more sustainable and can be used in the bioremediation of the aquatic environment $\frac{[13]}{[13]}$.

Commercial cultivation of microalgae requires the supply of significant amounts of inorganic carbon for photosynthesis $\frac{[14]}{2}$. This is mainly provided by carbon dioxide from the air and eventually from industrial emissions $\frac{[15]}{2}$. This may be a method for its biological sequestration [16], especially considering that some microalgae are capable of assimilating up to 1.83 Mg of CO₂ during the production of 1 Mg of their biomass $\frac{[17]}{}$. Higher CO₂ concentration promotes lipid accumulation in the cells [18]. Atmospheric carbon dioxide concentration depends on anthropogenic activities [19] and industrial development stage [20] and may increase above 400 PPM [21]. Concentrations of CO₂ may be an important factor to limit growth and development of some microalgal species [22], however, minimum and maximum concentrations CO₂ dissolved in the culture medium for microalgae cultivation vary from one species to another [23]. Some microalgae can grow under atmospheric air [24], others in extremely high CO₂ concentrations ranging from 40 to 100 vol% [25]. Compressed CO₂ can also be used in cultivation [26], but this increases the costs, which are related to the capture, compression, transport or storage of this gas [27]. The cost of using compressed carbon dioxide in microalgae culture can account for up to half of the total cost of biomass production [28]. An alternative option may be using of bicarbonate salts [29]. These compounds have much higher solubility in water than CO2 and higher efficiency in biomass production compared to compressed carbon dioxide [22]. A review of the recent progress in bicarbonate-based microalgae cultivation suggested potential to significantly reduce production cost. The use of sodium bicarbonate reduces the cost of carbon supply, increases the accumulation of valuable components, and is energetically efficient [30]. New technologies make it possible to produce sodium bicarbonate from carbon dioxide, including from industrial CO2 emissions, which are responsible for the negative effects of climate change [31].

2. Utilisation of CO₂ from Sodium Bicarbonate to Produce *Chlorella vulgaris* Biomass in Tubular Photobioreactors for Biofuel Purposes

2.1. Biomass Production with CO₂ from Sodium Bicarbonate

The effect of the NaHCO₃ dose on the growth dynamics of *C. vulgaris* is shown in **Figure 1**A.

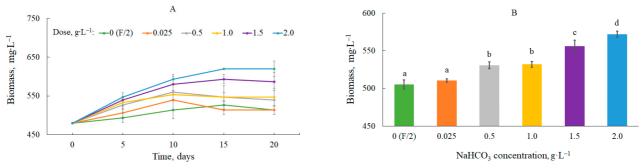


Figure 1. Dynamics of changes in biomass content (**A**) and average biomass content in the culture (**B**). Mean over each column not marked with the same letter is significantly different at $p \le 0.05$.

The biomass productivity changed as a function of the bicarbonate dose and time (**Table 1**). The highest values were observed at the beginning of the study at a dose of 2.0 g·L⁻¹ (13.3 \pm 2.3 mg·L⁻¹·d⁻¹). The productivity decreased with time. The changes could be due to the time-limited availability of nutrients as observed in batch cultures [32] or to the level of carbon dioxide utilisation.

Table 1. Biomass	productivity	' in relation i	to bicarbonate	aose.

Level of Sodium Bicarbonate (g·L ⁻¹)	Biomass Produc	Biomass Productivity (mg·L ⁻¹ ·d ⁻¹)			
	Day 5	Day 10	Day 15	Day 20	
0 (F/2)	2.7 ± 2.3	3.3 ± 2.3	3.1 ± 0.8	1.7 ± 0.6	
0.025	5.3 ± 2.3	6.0 ± 0.0	2.2 ± 0.8	1.7 ± 0.6	
0.5	9.3 ± 2.3	8.0 ± 2.0	4.4 ± 0.8	3.0 ± 0.0	
1.0	10.7 ± 2.3	7.3 ± 1.2	4.4 ± 2.0	3.3 ± 1.2	
1.5	12.0 ± 0.0	10.0 ± 2.0	7.6 ± 0.8	5.3 ± 1.2	
2.0	13.3 ± 2.3	11.3 ± 1.2	9.3 ± 0.0	7.0 ± 1.0	

3.2. Effect of Sodium Bicarbonate on Lipid Accumulation in Microalgal Biomass

The presence of bicarbonate in the culture medium, in excess of carbon storage in algal cells, can promote lipid accumulation $^{[33]}$. According to **Figure 2**, adding low concentrations of bicarbonate from 0.5 g·L⁻¹ to 1.5 g·L⁻¹ had no distinct effect on the lipid production of *C. vulgaris*. From own research indicate that lipid synthesis in cells requires a higher dose of inorganic carbon in the medium. In the present study, a significant increase in lipids in the presence of NaHCO₃, compared to the control 0 (F/2) object, was observed in the culture at a highest dose of 2.0 g·L⁻¹. The lipid content was 26 ± 4% and this was 8% higher than the values obtained without bicarbonate. Li et al. $^{[34]}$ observed an increase in lipid content in *C.vulgaris* cells with increasing NaHCO₃ dose, but a decrease in the amount of biomass. At a dose of 160 mM the authors obtained approx. 450 mg·g⁻¹ of lipids. A linear dose-dependent increase in lipid content of algal biomass was reported by Bywaters and Fritsen $^{[35]}$. A too-high concentration of NaHCO₃ may adversely affect lipid accumulation in microalgae cells. Significantly reduced lipid accumulation capacity in *C. pyrenoidosa* biomass, after introduction of 200 mM NaHCO₃, was observed by Sampathkumar and Gothandam $^{[36]}$. This is also confirmed by Pimolrat et al. $^{[37]}$, who analyzed the effect of NaHCO₃ at doses ranging from 0.05 to 5 g·L⁻¹ on the stimulation of triacylglycerol production in *Chaetoceros gracilis* cells.

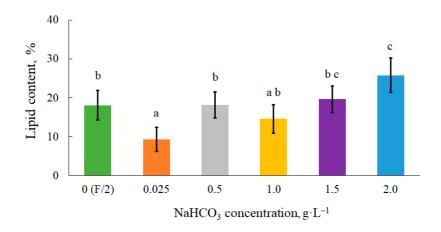


Figure 2. Lipid content in microalgal biomass. Mean over each column not marked with the same letter is significantly different at $p \le 0.05$.

2.3. Carbon Content and CO₂ Fixation Rate in Microalgal Biomass

The carbon content in microalgal biomass ranged from $0.832 \pm 0.127~g \cdot dw^{-1}$ in control 0 (F/2) object to $1.322~0.062~g \cdot dw^{-1}$ at a dose $2.0~g \cdot L^{-1}$ NaHCO $_3$. With increasing carbon in biomass, there was an increase in CO $_2$ fixation, which in the study ranged from $0.139 \pm 0.047~g \cdot L^{-1} \cdot d^{-1}$ to $0.925 \pm 0.073~g \cdot L^{-1} \cdot d^{-1}$. A high carbon content and rate of fixation indicate a high potential for CO $_2$ sequestration in *C. vulgaris* biomass $^{[38]}$. Similar results were reported by Prabakaran and Ravindran $^{[39]}$, who cultured three different algae strains (*Chlorella sp.*, *Ulothrix sp.* and *Chlorococcum sp.*) and obtained the highest carbon content and CO $_2$ fixation rate for *Chlorella sp.* at $0.486~g \cdot dw^{-1}$ and $0.68~g \cdot mL^{-1}d^{-1}$, respectively. Some authors indicate a linear relationship between NaHCO $_3$ dose and carbon accumulation in algal biomass $^{[33][34]}$. Mokashi et al. $^{[40]}$ applied bicarbonate in *C. vulgaris* cultures at dose from $0.25~to~1~g \cdot L^{-1}$ and determined the highest carbon content and CO $_2$ fixation rate of $0.497~g \cdot dw^{-1}$ and $0.69~g \cdot mL^{-1}d^{-1}$, respectively, at the highest dose. The level of carbon dioxide fixation varies depending on the microalgae strain and the carbon source (**Table 2**). The efficiency of the process carried out at the technical scale was higher compared to the results obtained by other authors, regardless of whether sodium bicarbonate or carbon dioxide was used in the microalgae cultivation.

Table 2. CO₂ fixation of microalgae biomass.

Strain	Carbon Source and Dose	Experimental Scale	CO ₂ Fixation, g·L ⁻¹ ·d ⁻¹	References
Chlorella vulgaris	NaHCO _{3,} 2.0 g·L ⁻¹	100 L	0.93	This study
Chlorella vulgaris	NaHCO _{3,} 1.0 g·L ⁻¹	100 mL	0.69	[<u>40</u>]
Chlorella sp.	NaHCO _{3,} 7.5 g·L ⁻¹	500 mL	0.21	[<u>41</u>]
Scenedesmus obliquus	CO ₂ , 10%	1 L	0.26	[<u>42</u>]
Scenedesmus almeriensis	CO ₂ , 3%	28.5 L	0.24	[<u>43</u>]

The optical density used to determine biomass growth does not always correlate with actual biomass content $\frac{[44]}{1}$; however, in the presented study, there was a significant and positive correlation between these parameters (r = 0.863) and moreover between the amount of biomass and the carbon content of microalgae cells (r = 0.785), as well as CO_2 fixation rate (r = 0.806).

3. Conclusions

We confirmed that the addition of NaHCO $_3$ to culture medium provides effective carbon source and facilitates cell growth and *C. vulgaris* biomass production. The highest biomass content (572 \pm 4 mg·L $^{-1}$) and productivity (7.0 \pm 1.0 mg·L $^{-1}$ ·d $^{-1}$) were obtained with bicarbonate at a dose of 2.0 g·L $^{-1}$. Under these conditions, the average optical density in culture was also the highest (OD $_{680}$ 0.181 \pm 0.00). An increase in NaHCO $_3$ dose increased lipid accumulation, carbon content in microalgae cells and carbon dioxide fixation rate. The highest values were observed at the highest dose of NaHCO $_3$. The average lipid content of the biomass was 26 \pm 4%. The carbon content of the biomass increased to 1.322 \pm 0.062 g·dw⁻¹, while the rate of CO $_2$ fixation increased to 0.925 \pm 0.073 g·L $^{-1}$ ·d $^{-1}$. There was a positive correlation between the biomass amount and the optical density and between the biomass, the carbon content and the CO $_2$ fixation rate.

The experiments were carried out in photobioreactors used in the industrial production of microalgae biomass and therefore the results obtained showed the real values that are possible to achieve at this scale. The lipid content in the biomass increased with the increasing dose of sodium bicarbonate. Future research should focus on determining the maximum dose of NaHCO₃ for optimal microalgal growth. It is important for the economic sustainability of microalgae cultivation for fuel purposes. The commercial production of microalgae biomass is carried out as a semi-continuous or continuous culture, so the correlation between the NaHCO₃ dose and the overaccumulation of Na⁺ ions and the possibility of limiting microalgal growth should be verified.

References

- 1. Bhatia, S.K.; Bhatia, R.K.; Jeon, J.M.; Pugazhendhi, A.; Awasthi, M.K.; Kumar, D.; Kumar, G.; Yoon, J.J.; Yang, Y.H. An overview on advancements in biobased transesterification methods for biodiesel production: Oil resources, extraction, biocatalysts, and process intensification technologies. Fuel 2021, 285, 119117.
- 2. Stephens, E.; Ross, I.L.; Mussgnug, J.H.; Wagner, L.D.; Borowitzka, M.A.; Posten, C.; Kruse, O.; Hankamer, B. Future prospects of microalgal biofuel production systems. Trends Plant Sci. 2010, 15, 554–564.
- 3. Chisti, Y. Biodiesel from microalgae. Biotechnol. Adv. 2007, 25, 294-306.
- 4. Arumugam, M.; Agarwal, A.; Arya, M.C.; Ahmed, Z. Influence of nitrogen sources on biomass productivity of microalgae Scenedesmus bijugatus. Biores. Technol. 2013, 131, 246–249.
- 5. Ratledge, C.; Cohen, Z. Microbial and algal oils: Do they have a future for biodiesel or as commodity oils? Lipid Technol. 2008, 20, 155–160.
- 6. Beer, L.L.; Boyd, E.S.; Peters, J.W.; Posewitz, M.C. Engineering algae for biohydrogen and biofuel production. Curr. Opin. Biotechnol. 2009, 20, 264–271.
- 7. Yuan, C.; Wang, S.; Cao, B.; Hu, Y.; Abomohra, A.E.-F.; Wang, Q.; Qian, L.; Liu, L.; Liu, X.; He, Z.; et al. Optimization of hydrothermal co-liquefaction of seaweeds with lignocellulosic biomass: Merging 2nd and 3rd generation feedstocks for enhanced bio-oil production. Energy 2019, 173, 413–422.
- 8. Rawat, I.; Kumar, R.; Mutanda, T.; Bux, F. Biodiesel from microalgae: A critical evaluation from laboratory to large scale production. Appl. Energy. 2012, 103, 444–467.
- 9. Sahoo, N.K.; Gupta, S.K.; Rawat, I.; Ansari, F.A.; Singh, P.; Naik, S.N.; Bux, F. Sustainable dewatering and drying of self-flocculating microalgae and study of cake properties. J. Clean. Prod. 2017, 159, 248–256.
- 10. Mohd Udaiyappan, A.F.; Abu Hasan, H.; Takriff, M.S.; Sheikh Abdullah, S.R. A review of the potentials, challenges and current status of microalgae biomass applications in industrial wastewater treatment. J. Water Process Eng. 2017, 20, 8–21.
- Shahid, A.; Malik, S.; Zhu, H.; Xu, J.; Nawaz, M.Z.; Nawaz, S.; Asraful Alam, M.; Mehmood, M.A. Cultivating microalgae inwastewater for biomass production, pollutant removal, and atmospheric carbon mitigation: A review. Sci. Total Environ. 2020, 704, 135303.
- 12. Hawrot-Paw, M.; Koniuszy, A.; Gałczyńska, M. Sustainable Production of Monoraphidium Microalgae Biomass as a Source of Bioenergy. Energies 2020, 13, 5975.
- 13. Xiaoning, L.; Guangyao, C.; Yi, T.; Jun, W. Application of effluent from WWTP in cultivation of four microalgae for nutrients removal and lipid production under the supply of CO2. Renew. Energy 2020, 149, 708–715.
- 14. Umetani, I.; Janka, E.; Sposób, M.; Hulatt, C.J.; Kleiven, S.; Bakke, R. Bicarbonate for microalgae cultivation: A case study in a chlorophyte, Tetradesmus wisconsinensis isolated from a Norwegian lake. J. Appl. Phycol. 2021, 33, 1341–1352.
- 15. Zhang, X. Microalgae removal of CO2 from flue gas; IEA Clean Coal Centre: London, UK, 2015.
- 16. Klinthong, W.; Yang, Y.-H.; Huang, C.-H.; Tan, C.-S. A review: Microalgae and their applications in CO2 capture and renewable energy. Aerosol Air Qual. Res. 2015, 15, 712–742.
- 17. Ho, S.H.; Chen, C.Y.; Lee, D.J.; Chang, J.S. Perspectives on microalgal CO2-emission mitigation systems—A review. Biotechnol Adv. 2011, 29, 189–198.
- 18. Rodolfi, L.; Zittelli, G.C.; Bassi, N.; Padovani, G.; Biondi, N.; Bonini, G.; Tredici, M.R. Microalgae for oil: Strain selection, induction of lipid synthesis and outdoor mass cultivation in a low-cost photobioreactor. Biotech. Bioeng. 2009, 102, 100–112.

- 19. Hussain, F.; Zahir, S.Z.; Zhou, W.; Iqbal, M. Microalgae screening under CO2 stress: Growth and micro-nutrients removal efficiency. J. Photochem. Photobiol. B Biol. 2017, 170, 91–98.
- 20. Ou, J.P.; Liu, X.P.; Li, X.; Chen, Y.M. Quantifying the relationship between urban forms and carbon emissions using panel data analysis. Landsc. Ecol. 2013, 28, 1889–1907.
- 21. Bezyk, Y.; Sówka, I.; Górka, M.; Blachowski, J. GIS-Based Approach to Spatio-Temporal Interpolation of Atmospheric CO2 Concentrations in Limited Monitoring Dataset. Atmosphere 2021, 12, 384.
- 22. Markou, G.; Vandamme, D.; Muylaert, K. Microalgal and cyanobacterial cultivation: The supply of nutrients. Water Res. 2014, 65, 186–202.
- 23. Salih, F. Microalgae Tolerance to High Concentrations of Carbon Dioxide: A Review. J. Environ. Prot. 2011, 2, 648-654.
- 24. Pourjamshidian, R.; Abolghasemi, H.; Esmaili, M.; Amrei, H.D.; Parsa, M.; Rezaei, S. Carbon dioxide biofixation by Chlorella sp in a bubble column reactor at different flow rates and CO2 concentrations. Brazilian J. Chem. Eng. 2019, 36, 639–645.
- 25. Solovchenko, A.; Khozin-Goldberg, I. High-CO2 tolerance in microalgae: Possible mechanisms and implications for biotechnology and bioremediation. Biotechnol. Lett. 2013, 35, 1745–1752.
- 26. Fulke, A.B.; Chambhare, K.; Giripunje, M.D.; Sangolkar, L.; Krishnamurthi, K.; Juwarkar, A.A.; Chakrabarti, T. Potential of wastewater grown algae for biodiesel production and CO2 sequestration. Afr. J. Biotechnol. 2013, 12, 2939–2948.
- 27. Chi, Z.; O'Fallon, J.V.; Chen, S. Bicarbonate produced from carbon capture for algae culture. Trends Biotechnol. 2011, 29, 537–541.
- 28. De Farias Silva, C.E.; Gris, B.; Sforza, E.; La Rocca, N.; Bertucco, A. Effects of sodium bicarbonate on biomass and carbohydrate production in Synechococcus pcc 7002. Chem. Eng. Trans. 2016, 49, 241–246.
- 29. Chisti, Y. Constraints to commercialization of algal fuels. J. Biotechnol. 2013, 167, 201–214.
- 30. Zhu, C.; Chen, S.; Ji, Y.; Schwaneberg, U.; Chi, Z. Progress toward a bicarbonate-based microalgae production system. Trends Biotechnol. 2021.
- 31. Cai, Y.; Wang, W.; Li, L.; Wang, Z.; Wang, S.; Ding, H.; Zhang, Z.; Sun, L.; Wang, W. Effective Capture of Carbon Dioxide Using Hydrated Sodium Carbonate Powders. Materials 2018, 11, 183.
- 32. Patyna, A.; Biłos, Ł.; Płaczek, M.; Witczak, S. Productivity of microalgae Chlorella vulgaris in laboratory condition. Ecol. Eng. 2017, 18, 99–105.
- 33. White, D.A.; Pagarette, A.; Rooks, P.; Ali, S.T. The effect of sodium bicarbonate supplementation on growth and biochemical composition of marine microalgae cultures. J. Appl. Phycol. 2013, 25, 153–165.
- 34. Li, J.; Li, C.; Lan, C.G.; Liao, D. Effects of sodium bicarbonate on cell growth, lipid accumulation, and morphology of Chlorella vulgaris. Microb. Cell Factories 2018, 17, 111.
- 35. Bywaters, K.F.; Fritsen, C.H. Biomass and neutral lipid production in geothermal microalgal consortia. Front. Bioeng. Biotechnol. 2015, 2, 1–11.
- 36. Sampathkumar, S.J.; Gothandam, K.M. Sodium bicarbonate augmentation enhances lutein biosynthesis in green microalgae Chlorella pyrenoidosa. Biocatal. Agric. Biotechnol. 2019, 22, 101406.
- 37. Pimolrat, P.; Direkbusarakom, S.; Chinajariyawong, C.; Powtongsook, S. The effect of sodium bicarbonate concentrations on growth and biochemical composition of Chaetoceros gracilis Schutt. Kasetsart Univ. Fish. Res. Bull. 2010, 34, 40–47.
- 38. Ryu, H.J.; Oh, K.K.; Kim, Y.S. Optimization of the influential factors for the improvement of CO2 utilization efficiency and CO2 mass transfer rate. J. Ind. Eng. Chem. 2009, 15, 471–475.
- 39. Prabakaran, P.; Ravindran, A.D. Lipid extraction and CO2 mitigation by microalgae. J. Biochem. Technol. 2012, 4, 469–472.
- 40. Mokashi, K.; Shetty, V.; George, S.; Sibi, G. Sodium Bicarbonate as Inorganic Carbon Source for Higher Biomass and Lipid Production Integrated Carbon Capture in Chlorella vulgaris. Achiev. Life Sci. 2016, 10, 111–117.
- 41. Nayak, M.; Suh, W.I.; Lee, B.; Chang, K.C. Enhanced carbon utilization efficiency and FAME production of Chlorella sp. HS2 through combined supplementation of bicarbonate and carbon dioxide. Energy Convers. Manag. 2018, 156, 45–52.
- 42. Tang, D.; Han, W.; Li, P.; Miao, X.; Zhong, J. CO2 biofixation and fatty acid composition of Scenedesmus obliquus and Chlorella pyrenoidosa in response to different CO2 levels. Bioresour. Technol. 2011, 102, 3071–3076.
- 43. Molino, A.; Mehariya, S.; Karatza, D.; Chianese, S.; Iovine, A.; Casella, P.; Marino, T.; Musmarra, D. Bench-Scale Cultivation of Microalgae Scenedesmus almeriensis for CO2 Capture and Lutein Production. Energies 2019, 12, 2806.

44. Aussant, J.; Guihéneuf, F.; Stengel, D.B. Impact of temperature on fatty acid composition and nutritional value in eight species of microalgae. Appl. Microbiol. Biotechnol. 2018, 102, 5279–5297.

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