

# Anti- and Pro-Oxidant Properties of Essential Oils

Subjects: **Microbiology**

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Natural products such as plant extracts and essential oils (EOs) have been viewed as potential candidates to combat antimicrobial resistance (AMR) due to their complex chemistry that carries inherent pro-oxidant and antioxidant properties. EOs and their constituents that hold pro-oxidant properties can induce oxidative stress by producing reactive oxygen species (ROS), leading to biological damage in target cells. In contrast, the antioxidant properties scavenge free radicals through offsetting ROS. Both pro-oxidant and antioxidant activities in EOs represent a promising strategy to tackle AMR.

pro-oxidant

antioxidant

antimicrobial resistance

essential oil

## 1. Introduction

Antimicrobial resistance (AMR) has caused significant detrimental effects on human health, contributing to increased mortality rates and infections due to the implementation of ineffectual antibiotic dosages [1]. According to the Centers for Disease Control and Prevention (CDC), human infections caused by antibiotic-resistant pathogens affect more than 2.8 million people, resulting in over 35,000 deaths annually in the United States [2]. The emergence of AMR is mainly due to the overprescription of antimicrobial agents by physicians and a lack of compliance by consumers [3]. Furthermore, easy accessibility and availability of antibiotics from over-the-counter pharmaceutical sales aggravate this issue. Over the past three decades, AMR rates have risen considerably while the pipeline of new antibiotics has declined significantly [4]. The administration of treatments during infections in the clinical setting is becoming more complicated; it remains unclear if researchers will be able to treat common infections in the future. As such, there is a pressing need to better understand underlying resistance mechanisms and uncover novel therapeutic strategies to fight against AMR [5].

Oxidative stress is defined as the disturbance of the balance between reactive oxygen species (ROS) production and antioxidants to neutralize their harmful effects [6]. This imbalance leads to molecular cell damage and complex biochemical mechanisms are required to regulate the entire process [4]. ROS can be produced intracellularly in biological pathways or induced extracellularly by exogenous procedures [7]. Generally, traditional antibiotics induce ROS to cause secondary damage to the pathogens or exerting bactericidal effects as the main mechanism. To increase survivability, pathogens may employ different strategies such as increased enzyme production and formation of oxidant scavengers to avoid cellular damage caused by ROS. In this regard, ROS is suggested to be used in clinical practice based on its antimicrobial activity against a wide spectrum of pathogens, including

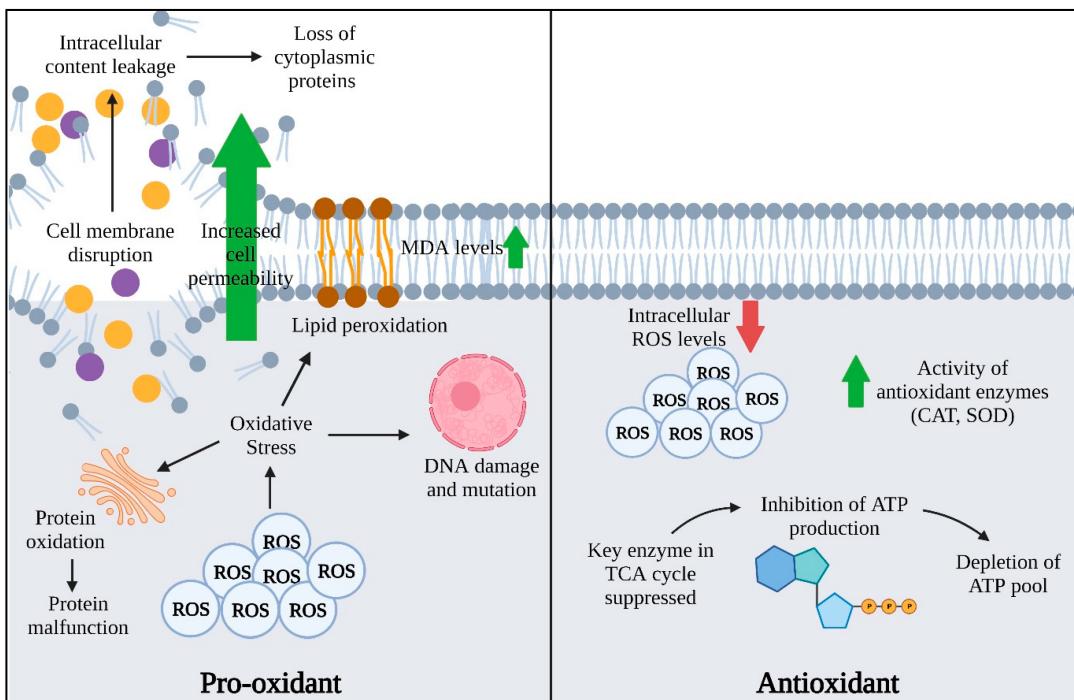
multidrug-resistant organisms. In clinical studies, surghoney reactive oxygen (SHRO) is the first ROS product that has shown great potency in controlling and eradicating bacterial bioburden and biofilm in chronic wounds, especially towards multidrug-resistant bacteria such as *Pseudomonas aeruginosa*, methicillin-resistant *Staphylococcus aureus* (MRSA), and vancomycin-resistant enterococci [8]. ROS can help to prevent and also to break down bacterial and fungal biofilms; these biofilms have remained a significant problem in many clinical settings by their increased resistance toward conventionally prescribed antimicrobials [9].

In addition to the promising outcomes with ROS therapy on infection models, researchers have diverted mining approaches towards medicinal plants, an option which seems to be more widely accepted by the public [6]. Natural products such as essential oil (EO) consisting of a plethora of chemical compounds are becoming a popular platform for researchers in drug discovery to improve antimicrobial efficacy and reduce the development of resistance [10].

## 2. Mechanisms of EOs in Combating AMR by Inducing ROS

Under aerobic conditions, antibiotic treatment generates ROS, which boosts the bactericidal effect and induces the production of ROS defense enzymes including catalase (CAT) and superoxide dismutase (SOD) [11]. There is much evidence of antibiotic-mediated ROS generation in *Escherichia coli*, *P. aeruginosa*, and *Acinetobacter baumannii* [12][13]. Polymyxin B is an example of bacterial antimicrobial peptides that were observed to induce rapid bacteria cell death through Fenton chemistry-mediated hydroxyl radical production in *A. baumannii* [14]. Similarly, superoxide and hydroxyl radical accumulation was detected in response to the treatment of *E. coli* with ampicillin and kanamycin [15]. This accumulation further amplified the oxidative damage and lethality of the antibiotics involved.

Eos are a composite mixture of volatile compounds which possess not only antioxidant activities but also act as pro-oxidants as they affect the cellular redox status while contributing to cellular damage (Figure 1) [16]. Mainly, EO can be grouped into three main groups—terpenes, terpenoids, and aromatic compounds [1]. Yang and colleagues (2020) demonstrated the synergism effect of lavender (*Lavandula angustifolia*) EO and meropenem in treating carbapenemase-producing *Klebsiella pneumoniae* (KPC-KP) cells [17]. Comparative proteomic analysis revealed the presence of oxidative stress, suggesting that lavender EO and meropenem generated ROS in KPC-KP cells. Further validation tests confirmed the high concentrations of ROS and malondialdehyde (MDA), indicating the presence of lipid peroxidation and explaining the observed membrane disruption in KPC-KP cells.



**Figure 1.** A schematic representation of pro-oxidant and antioxidant activities of EO against clinically important pathogens. Both pro-oxidant and antioxidant properties of EO act on different mechanisms of action that eventually lead to bacterial cell death.

More recently, Yang and colleagues (2021) revealed the accumulation of ROS and high levels of MDA in medium-treating KPC-KP cells with linalyl antranilate (LNA), a compound that originated from the lavender plant [6]. This implies that the generation of ROS is induced and lipid peroxidation is triggered. With treatment of terpene LNA, the abundance of membrane-related proteins in treated cells was reduced, suggesting a disrupted bacterial membrane that led to intracellular leakage and loss of cytoplasmic proteins. LNA has a different approach to killing bacterial cells and their antibacterial activity is lower in terms of effective concentration as compared with commercialized antibiotics. The ability of LNA in inducing ROS may facilitate the uptake and enhance the activity of antibiotics.

Yang and colleagues (2019) demonstrated the antibacterial activity of Cinnamon bark (*Cinnamomum verum*) EO by inducing oxidative stress in KPC-KP cells [10]. Accumulation of ROS disrupted the bacterial membrane and enabled influx of ROS into the cells, leading to intracellular content leakage. Overproduction of ROS subsequently contributed to bacterial cell death by malfunctioning the proteins involved in energy production such as the adenosine triphosphate (ATP synthase), the electron transport complex, and the NADH-quinone oxidoreductases. The proteomic profile revealed the loss of three essential proteins involved in bacterial cell wall synthesis upon treatment with cinnamon bark EO. Deleterious effects were also observed in genetic damage and impairment of DNA and membrane repair systems. Exposure of cinnamon bark EO released the DNA mismatch repair protein (MutS) and DNA ligase, indicating the presence of damage in genetic materials of KPC-KP cells.

Brun and colleagues (2019) revealed oxidative damage in *C. glabrata* by the exposure of tea tree EO, indicating that ROS production is associated with the disruption in mitochondria membrane and organelles [18]. Incubation with tea tree EO also reduced biofilm formation in both gram-positive and gram-negative bacteria, with observed significant biofilm inhibition in MRSA and *P. aeruginosa*. The antiviral activity of tea tree EO is similar to its antibacterial effects in which both infectivity and uptake of Herpes simplex virus type 1 (HSV-1) were reduced upon administration of Tea Tree EO.

Khan and colleagues (2017) performed several analyses on the ascitic fluid of a patient with urinary tract infection to determine the mechanism action of carvacrol (phenolic monoterpenoid) against extended-spectrum beta-lactamase *E. coli* [19]. In the presence of carvacrol (MIC: 450 µg/mL), high generation of ROS and bacterial membrane depolarization were observed. Carvacrol induced the highly oxidative stress environment in the bacterial cell, changed the membrane permeability, and increased the leakage of cellular contents (DNA and proteins) by disrupting the bacterial membrane integrity. Further scanning electron microscopy analysis revealed induction of structural disruption by the interaction between carvacrol and the lipid bilayer of *E. coli*.

Similar findings were found in the study conducted by Kim and colleagues (2019). Thymol and carvacrol extracted from Thyme White (*Thymus vulgaris*) and Summer Savory (*Satureja hortensis*) EOs showed strong fumigant antifungal activity against *Raffaelea quercus-mongolicae* and *Rhizoctonia solani* [20]. Exposure of *R. quercus-mongolicae* and *R. solani* to thymol and carvacrol induced the production of ROS in two phytopathogenic fungi, disrupted fungal cell membranes, and contributed to fungal cell death. Interestingly, thymol showed higher antifungal activity than carvacrol in *R. quercus-mongolicae* but this effect was not significantly observed in *R. solani*.

Lee and colleagues (2020) experimented with different EOs on the same fungi species. *Trans*-cinnamaldehyde, neral, and geranal extracted from cinnamon bark and lemongrass EOs were associated with the induction of ROS production in *R. quercus-mongolicae* and *R. solani* [21]. Further microscopy analysis revealed the importance of the stated constituents in fungal cell membrane disruption, leading to fungal cell death. **Table 1** summarizes the pro-oxidant activities of EOs.

**Table 1.** Recent insight of pro-oxidant activity of EOs against AMR.

Essential Oil/Essential Oil Constituents	Bacteria	Findings	References
Lavender	KPC-KP	<ul style="list-style-type: none"> <li>Comparative proteomic analysis revealed the presence of oxidative stress, suggesting that Lavender EO and meropenem generated ROS in KPC-KP cells.</li> </ul>	[17]

		<ul style="list-style-type: none"> <li>High concentrations of ROS and MDA were detected, indicating the presence of lipid peroxidation, and explaining the observed membrane disruption in KPC-KP cells.</li> </ul>	
LNA	KPC-KP	<ul style="list-style-type: none"> <li>Accumulation of ROS and high levels of MDA were detected, indicating the presence of lipid peroxidation.</li> <li>The abundance of membrane-related proteins in the treated cells was reduced, suggesting a disrupted bacterial membrane that led to intracellular leakage and loss of cytoplasmic proteins.</li> <li>the antibacterial activity of LNA is lower in terms of effective concentration as compared to commercialized antibiotics.</li> </ul>	[6]
Cinnamon bark	KPC-KP	<ul style="list-style-type: none"> <li>Oxidative stress was induced in KPC-KP cells.</li> <li>Accumulation of ROS disrupted the bacterial membrane and enabled the influx of ROS into the cells, leading to intracellular content leakage.</li> <li>Overproduction of ROS contributed to bacterial cell death by causing malfunction of the proteins involved in energy production such as the ATP synthase, the electron transport complex, and the NADH-quinone oxidoreductases.</li> <li>Proteomic profile revealed the loss of three essential proteins involved in bacterial cell wall synthesis.</li> <li>Deleterious effects were observed in genetic damage and impairment of DNA and membrane repair systems.</li> </ul>	[10]

		<ul style="list-style-type: none"> <li>Exposure of Cinnamon bark EO released the MutS and DNA ligase, indicating the presence of damage in the genetic materials of KPC-KP cells.</li> </ul>	
Tea Tree	<i>C. glabrata</i> , MRSA, HSV-1, <i>P. aeruginosa</i>	<ul style="list-style-type: none"> <li>Oxidative damage was observed in <i>C. glabrata</i>, indicating that ROS production is associated with disruption in mitochondria membrane and organelles.</li> <li>Incubation with Tea Tree EO inhibited biofilm formation in MRSA and <i>P. aeruginosa</i>.</li> <li>The infectivity and uptake of HSV-1 were reduced upon administration of Tea Tree EO.</li> </ul>	[18]
Carvacrol	Extended-spectrum beta-lactamase <i>E. coli</i>	<ul style="list-style-type: none"> <li>Carvacrol induced a high production of ROS and bacterial membrane depolarization with its MIC value at 450 µg/mL</li> <li>The highly oxidative stress environment in the bacterial cell changed membrane permeability and increased the leakage of cellular contents (DNA and proteins) by disrupting the bacterial membrane integrity.</li> <li>Further scanning electron microscopy analysis revealed the induction of structural disruption by the interaction between carvacrol and the lipid bilayer of <i>E. coli</i>.</li> </ul>	[19]
Thymol and Carvacrol	<i>R. quercus-mongolicae</i> , <i>R. solani</i>	<ul style="list-style-type: none"> <li>Thymol and carvacrol showed strong fumigant antifungal activity against <i>R. quercus-mongolicae</i> and <i>R. solani</i>.</li> <li>Generation of ROS is induced and this disrupted the fungal cell membrane, contributing to fungal cell death.</li> </ul>	[20]

		<ul style="list-style-type: none"> <li>Thymol showed higher antifungal activity than carvacrol in <i>R. quercus-mongolicae</i> but this effect does not significantly observe in <i>R. solani</i>.</li> </ul>	
<i>Trans</i> -cinnamaldehyde, Neral, and Geranal	<i>R. quercus-mongolicae</i> , <i>R. solani</i>	<ul style="list-style-type: none"> <li><i>Trans</i>-cinnamaldehyde, neral, and geranal induced ROS production in <i>R. quercus-mongolicae</i> and <i>R. solani</i>.</li> <li>Further microscopy analysis revealed the importance of the stated constituents in fungal cell membrane disruption, leading to fungal cell death.</li> </ul>	<a href="#">[21]</a>

### 3. Antioxidant Activities of EO in Mitigating AMR

EOs may act as pro-oxidants and antioxidants due to their complex chemistry [\[16\]](#). Cinnamon bark EO, which was discussed in the previous section, was found to comprise 13 compounds, of which 4 were non-antioxidant compounds that are believed to be responsible for the induction of oxidative stress by generating ROS [\[10\]](#). The presence of oxidative stress seems to conflict with the perception that the EO contains a high concentration of antioxidants.

Although earlier research has examined the antioxidant activity of EOs by free radical scavenging methods such as 2,2-diphenyl-1-picrylhydrazyl (DPPH) and 2,2'-azinobis-3-ethylbenzothiazoline-6-sulfonic acid (ABTS) assay [\[22\]](#)[\[23\]](#), there is limited literature focusing on the mechanism of antioxidant action of EOs.

The outer membrane component lipopolysaccharide (LPS) of bacteria such as *E. coli* play a crucial role in its antimicrobial susceptibility [\[24\]](#). Upon LPS binding, toll-like receptors on macrophages will activate the immune response's downstream signaling pathways, including NF $\kappa$ B and MAPK [\[25\]](#). In a review, de Lavor and colleagues (2018) highlighted the antioxidant activity of various EOs and their constituents in reducing ROS concentration, NF- $\kappa$ B expression, and proinflammatory cytokines synthesis after the interaction with bacteria LPS [\[26\]](#). Recent research by Pandur and colleagues (2022) revealed similar findings on the antioxidant capacity of thyme (*Thymus vulgaris* L.) EO and thymol [\[27\]](#). Thyme EO and thymol significantly reduced the level of ROS at LPS-treated human macrophage cell lines after 6 h and 24 h. Increased levels of CAT and SOD activities were also observed, indicating the antioxidant properties of thyme EO and thymol.

Tang and colleagues (2021) employed metabolomics and found that the ROS concentrations were significantly lower in MRSA, while there was an increase in the concentration of *Amomum villosum* EO compared with the control group [28]. *A. villosum* EO inhibited the growth of MRSA by reducing the intracellular ROS levels, suggesting a different inhibition mechanism of bacterial growth compared with antibiotics. The activity of key enzymes involved in the tricarboxylic acid cycle was suppressed, leading to the inhibition of ATP production in MRSA. Hence, the reduction of ROS levels contributed to energy metabolic dysfunction and bacterial cell killing.

## 4. Challenges and Limitations in Combating AMR Using Plant-Derived Antimicrobials

In recent decades, there has been an emerging increase in the usage of EOs and their main constituents. The market for natural products is expected to grow substantially due to rising consumer demand. However, side effects of EOs such as allergic reactions and skin inflammation are present even when diluted [29][30]. Fuentes and colleagues (2021) showed the adverse effects of EOs and their components at medium and high concentrations exposure [31]. In vivo studies demonstrated adverse effects with acute and long-term usage of carvacrol and thymol in animals including mice, rats, and rabbits. Exposure to carvacrol, thymol, and eugenol caused skin irritation, inflammation, ulcer formation, dermatitis, and slow healing in human subjects. Interaction between the lipophilic property of EOs and the hydrophobic parts of the cell contributed to the toxicity mechanism [32]. Thus, more toxicological research focusing on chronic exposure and combined exposure is necessary to elucidate possible risks to the biological system for the preservation of human health.

Moreover, studies with plant-derived antimicrobials are lacking detail regarding the involved mechanisms. The antimicrobial mechanism of EO, for example, centralized on bacterial cell membrane disruption, which resulted in increased cell permeability and cell death. Several approaches including genomic profiling, proteomic profiling, transcriptomic, and metabolomics should be included in future research to provide a glimpse into cellular physiology and their mode of action with data combined from other approaches [1]. In addition, genetic changes of target microorganisms against EOs can be observed through comparative analysis of gene expression between EO-treated and untreated cells [33]. A panel of genes that are involved in pathogenic process, stress response, basic metabolism, and transcription regulation were identified when Kovács and colleagues (2019) employed quantitative real-time polymerase chain reaction to compare the activities of peppermint (*Mentha piperita*) EO in *Campylobacter jejuni* [34]. A high expression of oxidative stress response protein was also found in the study.

The metabolomics method is another potential tool for studying the antimicrobial mechanism of EO extracts using both quantitative and qualitative approaches [28]. Tang and colleagues (2021) identified key metabolic pathways that influenced MRSA activity as discussed in the previous section. Transcriptomic and proteomic profiling is becoming a popular approach that helps to measure gene expression and protein abundance [35]. Techniques such as two-dimensional polyacrylamide gel electrophoresis (2D-PAGE), stable isotopic labeling method, and matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) aid in finding the effective interactions of EO constituents with their targets, which are mostly proteins in nature [36]. Transcriptomic approaches can then be used to further validate the proteomics data by examining the gene expression profile and

quantifying the protein abundance. With extensive knowledge of the mechanisms of plant-derived antimicrobials, development of novel antimicrobials for effective therapeutic usage that may help in reviving existing antibiotics is greatly anticipated in the near future.

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