

Veterinary Antiparasitic to Human Anticancer

Subjects: Oncology

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Cancer is an extensive disease and the most common cause of morbidity and mortality worldwide. It is characterized by a deregulation of the cell cycle, which primarily results in a progressive loss of control of cellular growth and differentiation. The repurposing of veterinary antiparasitic drugs for the treatment of cancer is gaining traction, as supported by existing literature. A prominent example is the proposal to implement the use of veterinary antiparasitics such as benzimidazole carbamates and halogenated salicylanilides as novel anticancer drugs. These agents have revealed pronounced anti-tumor activities and gained special attention for “double repositioning”, as they are repurposed for different species and diseases simultaneously, acting via different mechanisms depending on their target. As anticancer agents, these compounds employ several mechanisms, including the inhibition of oncogenic signal transduction pathways of mitochondrial respiration and the inhibition of cellular stress responses.

Keywords: drug repurposing ; antiparasitic ; benzimidazole carbamates ; halogenated salicylanilides ; cancer therapy

1. Introduction

Cancer is an extensive disease and the most common cause of morbidity and mortality worldwide. It is characterized by a deregulation of the cell cycle, which primarily results in a progressive loss of control of cellular growth and differentiation [1]. Although there are numerous ongoing studies on anticancer therapy, with many lead candidates at various phases of preclinical or clinical research, only 5% of potential anticancer therapies entering phase I clinical trials have been approved and have entered the market [2]. The standard cancer treatments include surgery, immunotherapy, radiation, and chemotherapy. Currently, chemotherapy is one of the most efficient and potent strategies used to treat malignant tumors. However, the development of multidrug resistance to chemotherapeutics has become a huge impediment to successful cancer treatment. Clearly, new therapeutic alternatives are required to improve cancer diagnosis and treatment. Prior to being marketed as a new drug, the lead compounds face many hurdles during preclinical and clinical studies to ensure their quality, safety, dosage, and efficacy. Clinical trials are costly and time-consuming, requiring ten to fifteen years of dedicated research. The entire development process of getting a single candidate compound onto the market is hindered by the exorbitant costs (approximately \$1–\$2.5 billion) associated with the necessary trials required for U.S. Food and Drug Administration (FDA) approval [3].

Drug repurposing has gained recognition in the last decade, enabling existing pharmaceutical products to be reconsidered for alternative applications. It has reduced the risk of a drug failing to reach the market, owing to the low burden of adverse effects, the attenuation of the economic load, and the expedition of the approval process [4]. It can also offer an improved risk versus reward trade-off as it shortens the timeline of the drug development process and is also economically feasible when compared to other drug development strategies [5]. Additionally, the preclinical results obtained from the use of repurposed drugs may expedite the process of the preclinical to clinical translation of cancer treatment [6].

2. BZ Carbamates

BZ antiparasitics are a group of heterocyclic aromatic organic compounds that are extensively used in both human and veterinary medicines to inhibit internal parasites. Some important BZ drugs include MZ, albendazole (ABZ), fenbendazole (FZ), flubendazole (FLU), triclabendazole, parbendazole, oxbendazole, and ricobendazole. In the last few years, some of these have been successfully investigated for various types of cancers worldwide.

2.1. Mechanism of Action of BZ Carbamates

The molecular mode of action of BZ carbamates involves inhibiting the polymerization of tubulin and facilitating the disruption of microtubules in parasite cells (**Figure 1**) [7]. An in vitro study using the extracts of helminthic and mammalian tubulin has implicated tubulin as the leading molecular target of BZ carbamates [8]. Tubulin is pivotal to cell motility,

proliferation, and division; the intercellular transport of organelles; the maintenance of cell shape; and the secretion process of cells in all living organisms [9]. By blocking microtubule elongation in worms, BZ carbamates perturb glucose uptake in cells. Eventually, the glycogen reserves are exhausted, and their energy management mechanisms are depleted, culminating in the death of the parasites [10].

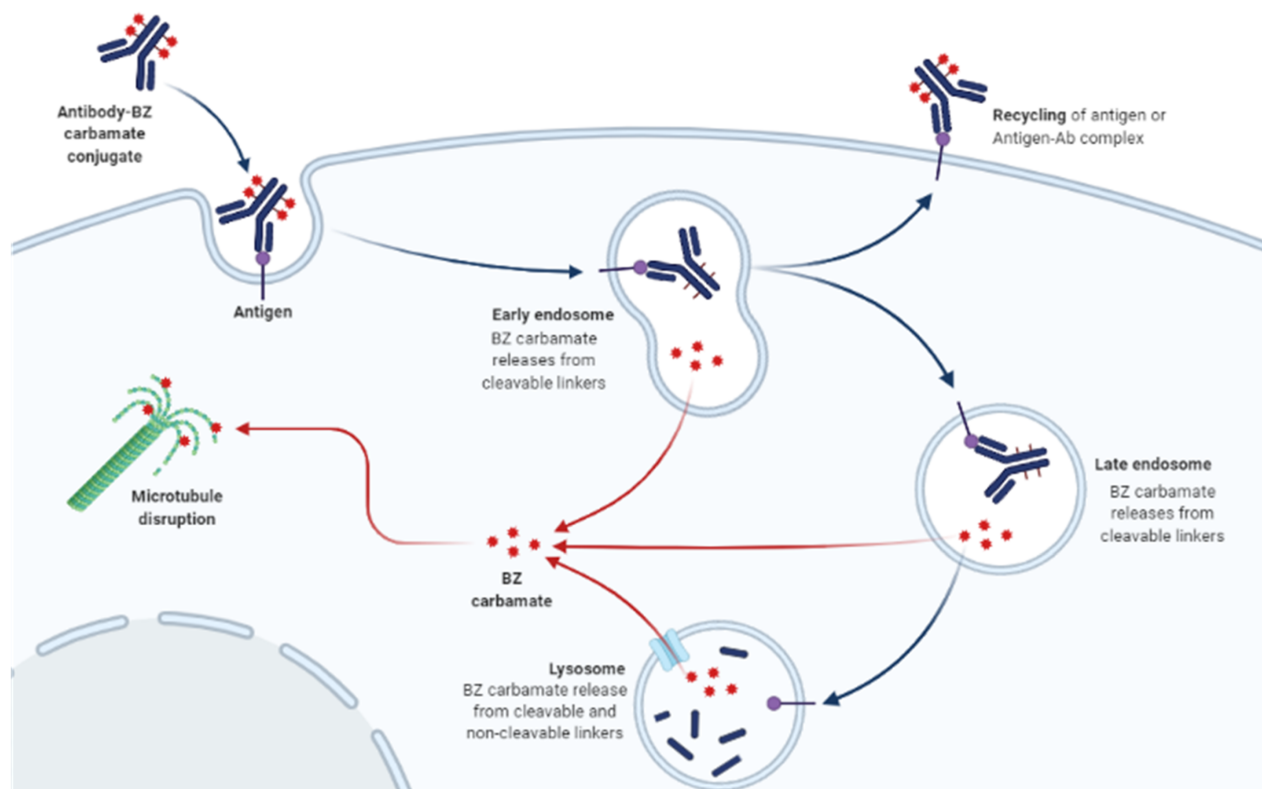


Figure 1. Mechanism of action of benzimidazole (BZ) carbamates targeting tubulin. Tubulin is the leading molecular target of BZ carbamates. They selectively bind to parasitic β -tubulin, promoting their immobilization and death. adapted from “Antibody-Drug Conjugate Drug Release”, by BioRender.com (2022). Retrieved from <https://app.biorender.com/biorender-templates>, accessed on 10 March 2022.

2.2. Anticancer Activity of BZ Carbamates

BZ carbamates are cancer cell-selective, causing minimal cytotoxicity in normal cells but increased cytotoxicity in different tumor cells. Several studies have reported that BZ carbamates inhibit the polymerization of mammalian tubulin in vitro. Whether the same effect would be observed in human cells, and if so, whether such targeted efforts could be effective against tumors, are some questions raised by these reports. Lacey et al. first addressed the activity of BZ carbamates against mouse leukemia cells L1210 in 1985 [11]. A more thorough inquiry into the antitumor effects of BZ carbamates was carried out; the most promising outcomes of this inquiry are summarized in **Table 1**. The general pharmacokinetic properties of BZ carbamates are as follows: slow absorption; wide distribution throughout the body; extensive hepatic metabolism; and excretion via urine and feces (**Figure 2a**). Their common side effects are fever, nausea, vomiting, abdominal discomfort, and hepatotoxicity. The low intestinal absorption rate of BZ carbamates may make it difficult for them to reach concentrations in the systemic circulation effective in treating cancers in humans. Increased bioavailability is necessary to enhance their antitumor effect, making them safe and well tolerable in human and veterinary use.

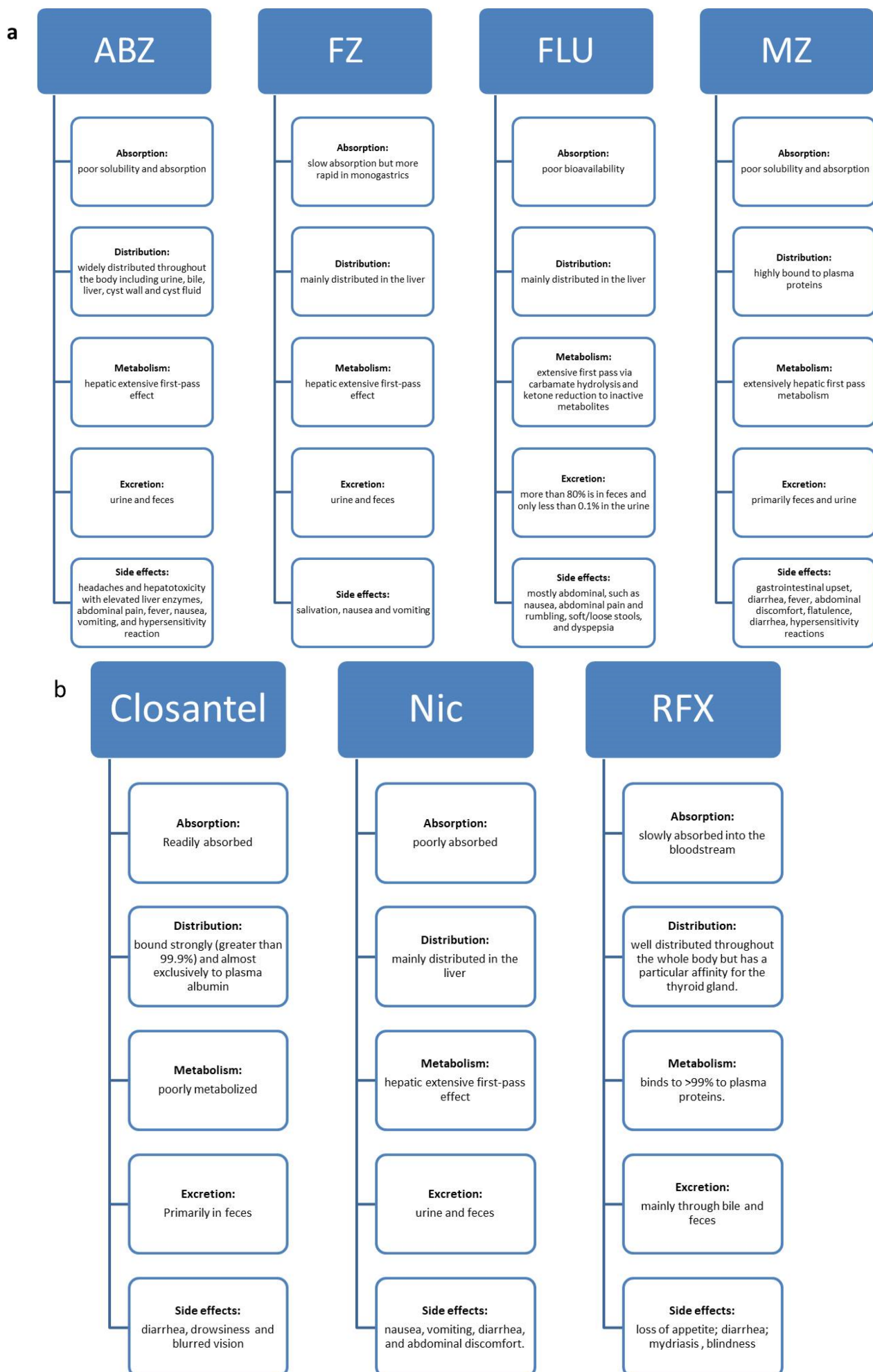


Figure 2. Pharmacokinetic properties and side effects of veterinary antiparasitic drugs. **(a)** The BZ carbamate drugs are poorly absorbed; have a wide distribution in the body; show extensive hepatic metabolism; and are excreted via feces and

urine. (b) The halogenated salicylanilides (HS) antiparasitic drugs show poor absorption, distribution throughout the body, are poorly metabolized and are excreted in bile, feces, or urine.

Table 1. Anticancer activity of BZ carbamates.

Cell Source	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
Human	Hep G2 and Hep3B	in vitro	<i>Mice</i>	ABZ	HCC	Cytotoxicity	[12]
Human	Hep G2 and Hep3B, PLC/PRF/5 and SKHEP-1	in vitro					
	SKHEP-1	in vivo	<i>Mice</i>	ABZ	HCC	Tubulin disruption	[13]
Rat	HTC, Novikoff	in vitro					
Mice	Hep1-6	in vitro					
Human	SW480, SW620, HCT8 and Caco2	in vitro	<i>Mice</i>	ABZ, RBZ, FLU	Intestinal cancer	Tubulin disruption	[14]
Human	HT-29	in vitro	<i>Mice</i>	ABZ	CRC	Apoptosis	[15]
Human	CEM/dEpoB300	in vitro	<i>Mice</i>	ABZ	Leukemia	Apoptosis	[16]
Human	1A9Pc TX22	in vitro	<i>Mice</i>	ABZ	OC	Apoptosis	[17]
Mouse	EMT6	in vitro in vivo	<i>Mice</i>	FZ	Mammary carcinoma	Cytotoxicity	[18]
Human	H460 and A549	in vitro in vivo	<i>nu/nu mice</i>	FZ	LC	microtubule disruption, p53 activation and down regulation of pivotal glycolytic enzymes	[19]
Human	P493-6	in vitro in vivo	<i>SCID mice</i>	FZ	Lymphoma	Tubulin disruption	[20]

Cell Source	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
Mice	EMT6	in vitro	<i>BALB/c Rw mice</i>	FZ	Mammary carcinoma	Tubulin disruption	[21]
		in vivo					
Human	OCI-AML-2	in vitro	<i>SCID mice</i>	FLU	Leukaemia and Myeloma	Tubulin disruption	[22]
		in vivo					
Human	MDA-MB-231, BT-549, SK-BR-3 and MCF-7	in vitro	<i>Mice</i>	FLU	BC	Tubulin disruption	[23]
		in vivo					
Human	TNBC cell lines MDA-MB-231 and MDA-MB-468	in vitro	<i>Mice</i>	FLU	BC	Apoptosis	[24]
		in vivo					
Human	BT474, SK-BR-3, MDA-MB-453, JIMT-1	in vitro	<i>BALB/c mice</i>	FLU	BC	Tubulin disruption	[25]
		in vivo				Apoptosis	
Human	HCT116, RKO and SW480	in vitro	<i>BALB/c mice</i>	FLU	CRC	Apoptosis	[26]
		in vivo					
Human	H295R and SW-13	in vitro	<i>Mice</i>	MZ	Adrenocortical carcinoma	Apoptosis	[27]
Human	H460, A549, H1299 and WI-38	in vitro	<i>Mice</i>	MZ	LC	Tubulin disruption,	[28]
		in vivo				Apoptosis	
Human	HCT 116 and RKO	in silico	-	MZ	CC	Tubulin disruption	[29]
Human	DLD-1, HCT-116, HT-29 and SW480	in vitro	<i>Mice</i>	MZ	CC	Tubulin disruption	[30]
Human	ACP-02, ACP-03 and AGP-01	in vitro	<i>Mice</i>	MZ	GC	Tubulin disruption	[31]
		in vivo					

Cell Source	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
Mouse	GL261	in vitro	<i>C57BL6 Mice</i>	MZ	Brain tumour	Tubulin disruption	[32]
		in vivo				Apoptosis	
Human	GBM U87-MG, D54, H80, H247, H392, H397, H502 and H566	in vitro	<i>C57BL/6 mice</i>	MZ	Brain cancer	Apoptosis	[33]
		in vivo					
Mouse	GL261						
Human	D425 MB	in vivo	<i>p53 mice</i>	MZ	Medullo-blastoma	Tubulin disruption	[34]
Human	293T and hTERT-RPE1	in vitro	<i>nu/nu athymic mice</i>	MZ	Medullo-blastoma	Hedgehog inhibitor	[35]
		in vivo					
Murine	CP2 and SP1	in vitro	<i>BALB/c mice</i>	MZ	PC	Tubulin disruption	[36]
		in vivo					
Human	KKU-M213	in vitro	<i>Nude mice</i>	MZ	Bile duct Cancer	Apoptosis	[37]
		in vivo					
Human	PANC-1	in vitro	<i>Mice</i>	MZ	Pancreatic cancer	-	[38]
Human	CAL27 and HCC15	in vitro	<i>Nude mice</i>	MZ	Head and neck cancer	Apoptosis	[39]
		in vivo					
Human	SK-Br-3	in vivo	<i>Mice</i>	MZ	BC	Tubulin disruption	[40]
Human	M-14 and SK-Mel-19	in vitro	<i>Mice</i>	MZ	Melanoma	Tubulin disruption	[41]
Human	MM622, MM540, D08, MM329, D17, and UACC1097	in vitro	<i>Mice</i>	MZ	Melanoma	Tubulin disruption	[42]
		in vivo					

Cell Source	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
Human	NRAS ^{Q61K}	in vitro	<i>Athymic mice</i>	MZ	Melanoma	Apoptosis	[43]
		in vivo					
		in silico					
Human	GL261	in vitro	<i>C57BL/6 mice</i>	MZ	Brain cancer	Tubulin disruption	[44]
		in vivo					
Human	Burkitt's lymphoma Ramos cells, HeLa cells, PANC-1 cells, and HepG2 cells	in vivo	<i>Zebra-fish</i>	Closantel	Lymphoma, cervical cancer, PC, and LC	Suppression of antiangiogenesis and Closantel	[45]
Human	Du146	in vitro	<i>Mice</i>	Nic	PC	Inhibition of STAT3 Pathway	[46]
Human	HEK293 cells	in vitro	<i>Mice</i>	Nic	PC and BC	Inhibition of Wnt/ β -catenin Pathway	[47]
Human	MCF7 and MDA-MB-231	in vitro	<i>NOD/SCID mice</i>	Nic	BC	Apoptosis and downregulation stem pathways	[48]
		in vivo					
Human	MDA-MB-231	in vitro	<i>BALB/c nude mice</i>	Nic and cisplatin	BC	Apoptosis and inhibition of Akt, ERK, and Src pathways	[49]
		in vivo					
Human	MDA-MB-468 and MCF-7	in vitro	<i>Mice</i>	Nic	BC	Inhibition of cell motility and STAT3 activity	[50]
Human	TNBC MDA-MB-231, MDA-MB-468 and Hs578T	in vitro	<i>Athymic nude mice</i>	Nic	BC	Inhibition of Wnt/ β -catenin Pathway	[51]
		in vivo					

Cell Source	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
Mouse	4T1	in vitro					
		in vivo	<i>BALB/c mice</i>	Nic	BC	Apoptosis and suppression of cell migration and invasion	[52]
Human	MDA-MB-231, MDA-MB-468 and MCF-7	in vitro					
Human	2LMP, SUM159, HCC1187, and HCC1143	in vitro	<i>NOD/SCID mice</i>	Nic	BC	Cytotoxicity	[53]
		in vivo					
Human	K562 and KBM5-T315l cells	in vitro	<i>NOD mice</i>	Nic	Chronic myelogenous leukemia	Inhibition of FOXM1/ β -catenin Pathway	[54]
		in vivo					
Human	HL-60, U937, OCI-AML3, Molm13, MV4-11, and U266 cells	in vitro	<i>BALB/c mice</i>	Nic	Acute myelogenous leukemia	Apoptosis and Inhibition of NF- κ B pathway	[55]
		in vivo					
	MCF7				Adeno-carcinoma		
		in vitro					
	HCC1954				Carcinoma		
Human	BT-474		<i>Mice</i>	Nic	Ductal Carcinoma	Inhibition of PI3K-dependent signalling	[56]
		in vivo					
	MDA-MB-361 and				Adeno-carcinoma		
	SKBR3 cell	in silico			Adeno-carcinoma		
Human	HCT116, SW620, and HT29	in vivo	<i>Mice</i>	Nic	CC	Inhibition of STAT3 phosphorylation	[57]
Human	HCT116, SW480, DLD1 and 293 cells	in vitro	<i>APC-MIN mice</i>	Nic	CC	Inhibition of Wnt/Snail-mediated EMT	[58]
		in vivo					

Cell Source	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
Human	HCT116, SW620, LS174T, SW480, and DLD-1	in vitro	<i>NOD/SCID mice</i>	Nic	CC	Inhibition of S100A4-induced metastasis formation	[59]
		in vivo					
		in situ					
Human	HT29, HCT116, CaCO2 and MCF-10A	in vitro	<i>NOD/SCID mice</i>	Nic	CC	Inhibition of Wnt/ β -catenin Pathway	[60]
		in vivo					
Human	HEK293T, U2OS, WIDR, DLD-1, CRC 240, COLO205, CRC57 and HCT116	in vitro	<i>Mice</i>	Nic	CC	Induction of autophagy and inhibition of Wnt/ β -catenin Pathway	[61]
Human	SW480 and SW620	in vitro	<i>Mice</i>	Nic	CC	Reduction of Wnt activity	[62]
Rodent	CC531	in vivo					
Murine	MC38	in vitro					
		in vivo					
Human	HCT116	in vitro	<i>APC^{min/+} mouse</i>	Nic-EN and oxyclozanide	CC	Mitochondrial uncoupling	[63]
Rodent	C2C12	in vitro	<i>SCID mice</i>	Nic	OC	Induction of metabolic shift to glycolysis	[64]
		in vivo					
Human	SKOV3 and CP70	in vitro					
		in vivo					
Human	OVCAR-3, SKOV-3 and A2780	in vitro	<i>NOD/</i>	Nic	OC	Inhibition of CP70sps and primary OTICs	[65]
		in vivo	<i>SCID mice</i>				
Human	SKOV3.ip1	in vitro	<i>Mice</i>	Nic	OC	Inhibition of Wnt/ β -catenin Pathway	[66]
		in vivo					

Cell Source	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
Human	SKOV3 and HO8910	in vitro in vivo	<i>Athymic Nude mice</i>	Nic	OC	Mitochondrial Respiration and aerobic glycolysis	[67]
Human	A2780ip2, A2780cp20, and SKOV3Trip2	in vitro in vivo	<i>SCID mice</i>	Nic	OC	Inhibition of Wnt/ β -catenin, mTOR and STAT3 pathways	[68]
Human	Tumorspheres	in vitro in vivo	<i>Mice</i>	Nic and its analogs in combination with carboplatin	OC	Cytotoxicity	[69]
Human	HepG2 and QGY7701	in vitro	<i>Mice</i>	Nic	HCC	Apoptosis and suppression of ATF3 expression	[70]
Human	NSCLC, NCI-H1299 and HCT116	in vitro	<i>Mice</i>	Nic	LC	Apoptosis through ROS-mediated p38 MAPK-c-Jun activation	[71]
Human	SK-Hep-1 and Huh7	in vitro	<i>Mice</i>	Nic	HCC	Inhibition of metastasis of HCC, and CD10	[72]
Human	HCC827, H1650, and H1975	in vitro in vivo	<i>Nu/Nu nude mice</i>	Nic	LC	Inhibition of STAT3 phosphorylation	[73]
Human	A549/DDP	in vitro	<i>Mice</i>	Nic combined with cisplatin (DDP)	Cisplatin-resistant LC	Apoptosis and reduction of c-myc protein	[74]
Human	HepG2, QGY-7703 and SMMC-7721	in vitro	<i>Mice</i>	Nic	HCC	Inhibition of cell growth and STAT3 pathway	[75]

Cell Source	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
Human	Lung adenocarcinoma (549, EKVX, H358, Hop62, H322M, H522, H838, and H23), large cell lung carcinoma (H460, Hop92), NCSLC (H1299, H810) and small cell LC (H82)	in vitro	<i>Mice</i>	Nic	LC	Reduction in proliferation and inhibition of S100A4 protein	[76]
Human	U-87 MG	in vitro	<i>Mice</i>	Nic	Glioblastoma	Cell toxicity and inhibition of Wnt/ β -catenin, PI3K/AKT, MAPK/ERK, and STAT3	[77]
Human	TS15-88, GSC11	in vitro in vivo	<i>Athymic nude mice</i>	Nic and/or temo-zolomide	Glioblastoma	Inhibition of the expression of epithelial-mesenchymal transition-related markers, Zeb1, N-cadherin, and β -catenin	[78]
Human	LN229, T98G, U87(MG), U138, and U373(MG)	in vitro in vivo	<i>Rag2^{-/-} Il2rg^{-/-} and SCID/Beige mice</i>	Nic	Glioblastoma	Cytotoxicity and diminished the pGBMs' malignant potential	[79]
Human	C4-2B, LNCaP and DU145	in vitro	<i>Mice</i>	Nic with enzalutamide	Enzalutamide resistance PC	Inhibition of migration, invasion and IL6-Stat3-AR pathway	[80]
Human	LNCaP, VcaP, CWR22Rv1, PC3 and HEK293	in vitro in vivo	<i>SCID mice</i>	Nic with enzalutamide	Castration-resistant PC	Inhibition of AR variant and enzalutamide-resistant tumor growth	[81]

Cell	Cell Lines	Procedure of Study	Species	Antiparasitics	Cancer Type	Target Pathway	Reference
References							
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32. PubMed, Google Scholar, and CTD databases were used to search for the data for the antitumor effects of BZ carbamates. ABZ—albendazole; BBZ—broad-spectrum albendazole; CO—colorectal cancer; GC—gastric cancer; HCC—hepatocellular carcinoma; ICD—immunogenic cell death; LC—lung cancer; MZ—mebendazole; Nic—niclosamide; Nic-EN—niclosamide, ethanolamine; OC—ovarian cancer; PC—prostate cancer; RBZ—ribendazole; RFX—rafoxanide. [99]
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Cancer	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Antiparasitics	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Albendazole	Title	Phase	Purpose	Status/Result	Identifier/Ref
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47. Zhu, X.-Y.; Xia, B.; Liu, H.-C.; Xu, Y.-Q.; Huang, C.-J.; Gao, J.-M.; Dong, Q.-Y.; Li, C.-Q. Cloasatel Suppresses Angiogenesis and Cancer Growth in Zebrafish Models. *ASSAY Drug Dev. Technol.* 2010, 14, 282–290.

ABZ	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Neutropenia	Title	Phase	Purpose	Status/Result	Identifier/Ref
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2400 mg/day	Title	Phase	Purpose	Status/Result	Identifier/Ref
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To determine the safety, tolerability, and the maximal tolerated dose.	Title	Phase	Purpose	Status/Result	Identifier/Ref
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52. Gyamfi, J.; Lee, R.; Kim, B.S.; Chao, T.K. Niclosamide reverses adipocyte induced epithelial-mesenchymal transition in breast cancer cells and suppresses the IL-6/STAT3 signalling axis. *Sci. Rep.* 2019, 9, 11336.

ABZ	Title	Phase	Purpose	Status/Result	Identifier/Ref
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53. Yin, L.; Gao, Y.; Zhang, X.; Wang, J.; Ding, D.; Zhang, Y.; Zhang, J.; Chen, H. Niclosamide as triple-negative breast cancer cells to ionizing radiation in association with the inhibition of Wnt/beta-catenin signaling. *Oncotarget* 2016, 7, 42126–42138.

tumors	Title	Phase	Purpose	Status/Result	Identifier/Ref
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54. Ye, T.; Xiong, Y.; Yan, Y.; Xia, Y.; Song, X.; Liu, L.; Li, D.; Wang, N.-Y.; Zhang, L.; Zhu, Y.; et al. The Anthelmintic Drug Niclosamide Induces Apoptosis, Impairs Metastasis and Reduces Immunosuppressive Cells in Breast Cancer Model. *PLoS ONE* 2014, 9, e85887.

of efficacy.	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Albendazole	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Adreno-cortical carcinoma	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Adrenocortical Carcinoma	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Drug Repositioning From Bench To Bedside: Tumour	Title	Phase	Purpose	Status/Result	Identifier/Ref
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MZ	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Mebendazole	Title	Phase	Purpose	Status/Result	Identifier/Ref
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Mebendazole In	Title	Phase	Purpose	Status/Result	Identifier/Ref
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62. Osada, T.; Chen, M.; Yang, X.; VanDeusen, J.B.; Fink, D.; Ghossein, B.M.; Clay, T.M.; Chen, W.; Morse, M.A.; et al. Anthelmintic Compound Niclosamide Downregulates Wnt Signaling and Elicits Antitumor Responses in Tumors with Activating APC Mutations. *Cancer Res.* 2011, 71, 4172–4182.

MZ	Title	Phase	Purpose	Status/Result	Identifier/Ref
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63. Wang, J.; Ren, X.-R.; Piao, H.; Zhao, S.; Osada, T.; Premont, R.T.; Mook, P.A.; Morse, M.A.; Lyster, H.K.; Chen, W. Niclosamide-induced Wnt signaling inhibition in colorectal cancer is mediated by autophagy. *Biochem. J.* 2019, 476, 535–546.

blastoma	Title	Phase	Purpose	Status/Result	Identifier/Ref
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64. Monin, M.B.; Krause, P.; Stelling, R.; Bocuk, D.; Niebert, S.; Klemm, F.; Pukrop, T.; Koenig, S. The anthelmintic niclosamide inhibits colorectal cancer cell lines via modulation of the canonical and noncanonical Wnt signaling

(Mebendazole)	Title	Phase	Purpose	Status/Result	Identifier/Ref
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65	Anticancer Effects of Niclosamide on Hepatic Metastasis of Colon Cancer. Cell Death Dis. 2018, 9, 215.	Anticancer Effects of Niclosamide on Hepatic Metastasis of Colon Cancer	Chen, M.; Swapna, E.V.T.; Tao, H.; Guo, H.; Colandrese, J.; Fadhil, N.; Monteiro, R.; Jin, S.	Phase I Study	Purpose	Status/Result	Identifier/Ref		
66	Lin, C.-K.; Bai, M.-Y.; Hu, T.-M.; Wang, Y.-C.; Chao, T.-K.; Weng, Y.-C.; Huang, R.-L.; Su, P.-H.; Lai, H.-C. Preclinical evaluation of a nanoformulated amebicide, niclosamide, in ovarian cancer. Oncotarget 2016, 7, 8993–9006.	Preclinical evaluation of a nanoformulated amebicide, niclosamide, in ovarian cancer	Lin, C.-K.; Bai, M.-Y.; Hu, T.-M.; Wang, Y.-C.; Chao, T.-K.; Weng, Y.-C.; Huang, R.-L.; Su, P.-H.; Lai, H.-C.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
67	Yo, Y.-T.; Lin, Y.-Y.; Wang, Y.-C.; Balch, C.; Huang, R.L.; Chan, M.; Sytwu, H.-K.; Chen, C.-K.; Chang, C.-C.; Nephew, K.P.; et al. Growth Inhibition of Ovarian Tumor-Initiating Cells by Niclosamide. Mol. Cancer Ther. 2012, 11, 1703–1712.	Growth Inhibition of Ovarian Tumor-Initiating Cells by Niclosamide	Yo, Y.-T.; Lin, Y.-Y.; Wang, Y.-C.; Balch, C.; Huang, R.L.; Chan, M.; Sytwu, H.-K.; Chen, C.-K.; Chang, C.-C.; Nephew, K.P.; et al.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
68	Arend, R.C.; Londono-Joshi, A.I.; Samant, R.S.; Li, Y.; Conner, M.; Hidalgo, B.; Alvarez, R.D.; Landen, C.N.; Straughn, J.M.; Buchsbaum, D.J. Inhibition of Wnt/beta-catenin pathway by niclosamide: A therapeutic target for ovarian cancer. Gynecol. Oncol. 2014, 134, 112–120.	Inhibition of Wnt/beta-catenin pathway by niclosamide: A therapeutic target for ovarian cancer	Arend, R.C.; Londono-Joshi, A.I.; Samant, R.S.; Li, Y.; Conner, M.; Hidalgo, B.; Alvarez, R.D.; Landen, C.N.; Straughn, J.M.; Buchsbaum, D.J.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
69	Shangguan, F.; Liu, Y.; Ma, L.; Qu, G.; Lv, Q.; An, J.; Yang, S.; Lu, B.; Cao, Q. Niclosamide inhibits ovarian carcinoma growth by interrupting cellular bioenergetics. J. Cancer 2020, 11, 3454–3466.	Niclosamide inhibits ovarian carcinoma growth by interrupting cellular bioenergetics	Shangguan, F.; Liu, Y.; Ma, L.; Qu, G.; Lv, Q.; An, J.; Yang, S.; Lu, B.; Cao, Q.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
70	Arend, R.C.; Londono-Joshi, A.I.; Samant, R.S.; Li, P.-K.; Landen, C.N.; Yang, E.S.; et al. Niclosamide and its analogs are potent inhibitors of Wnt/beta-catenin, mTOR and STAT3 signaling in ovarian cancer. Oncotarget 2016, 7, 86803–86815.	Niclosamide and its analogs are potent inhibitors of Wnt/beta-catenin, mTOR and STAT3 signaling in ovarian cancer	Arend, R.C.; Londono-Joshi, A.I.; Samant, R.S.; Li, P.-K.; Landen, C.N.; Yang, E.S.; et al.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
71	Walters Haygood, C.L.; Arend, R.C.; Samant, R.S.; Chettiar, S.; Regan, N.; Hassmann, C.J.; Li, P.-K.; Hidalgo, B.; Straughn, J.M.; Buchsbaum, D.J. Niclosamide Analogs for Treatment of Ovarian Cancer. Int. J. Gynecol. Cancer 2015, 25, 1377–1385.	Niclosamide Analogs for Treatment of Ovarian Cancer	Walters Haygood, C.L.; Arend, R.C.; Samant, R.S.; Chettiar, S.; Regan, N.; Hassmann, C.J.; Li, P.-K.; Hidalgo, B.; Straughn, J.M.; Buchsbaum, D.J.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
72	Weng, S.; Zhou, L.; Deng, Q.; Wang, J.; Yu, Y.; Zhu, J.; Yuan, Y. Niclosamide induced cell apoptosis via upregulation of ATF3 and activation of PERK in Hepatocellular carcinoma cells. BMC Gastroenterol. 2016, 16, 25.	Niclosamide induced cell apoptosis via upregulation of ATF3 and activation of PERK in Hepatocellular carcinoma cells	Weng, S.; Zhou, L.; Deng, Q.; Wang, J.; Yu, Y.; Zhu, J.; Yuan, Y.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
73	Lee, S.-L.; Son, A.-R.; Ahn, J.; Song, J.-Y. Niclosamide enhances ROS-mediated cell death through c-Jun activation. Biomed. Pharmacother. 2014, 89, 611–617.	Niclosamide enhances ROS-mediated cell death through c-Jun activation	Lee, S.-L.; Son, A.-R.; Ahn, J.; Song, J.-Y.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
74	Chien, M.-H.; Hsu, Y.-C.; Yang, S.-F.; Yang, Y.-C.; Lai, S.-Y.; Chen, W.-S.; Chen, M.-J.; Yeh, C.-B. Niclosamide and its analogs are potent inhibitors of Wnt/beta-catenin, mTOR and STAT3 signaling in ovarian cancer. Oncotarget 2016, 7, 86803–86815.	Niclosamide and its analogs are potent inhibitors of Wnt/beta-catenin, mTOR and STAT3 signaling in ovarian cancer	Chien, M.-H.; Hsu, Y.-C.; Yang, S.-F.; Yang, Y.-C.; Lai, S.-Y.; Chen, W.-S.; Chen, M.-J.; Yeh, C.-B.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
75	Li, R.; Hu, Z.; Sun, S.-Y.; Chen, Z.G.; Owonikoko, T.K.; Sica, G.L.; Ramalingam, S.S.; Curran, W.J.; Khuri, F.R.; Deng, X. Niclosamide Overcomes Acquired Resistance to Erlotinib or Suppression of STAT3 in Non-Small Cell Lung Cancer. Mol. Cancer Ther. 2013, 12, 2200–2212.	Niclosamide Overcomes Acquired Resistance to Erlotinib or Suppression of STAT3 in Non-Small Cell Lung Cancer	Li, R.; Hu, Z.; Sun, S.-Y.; Chen, Z.G.; Owonikoko, T.K.; Sica, G.L.; Ramalingam, S.S.; Curran, W.J.; Khuri, F.R.; Deng, X.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
76	Zuo, Y.; Yang, D.; Yu, Y.; Xiang, M.; Li, H.; Yang, J.; Li, J.; Jiang, D.; Zhou, H.; Xu, Z.; et al. Niclosamide enhances the cytotoxic effect of cisplatin in cisplatin-resistant human lung cancer cells via suppression of lung resistance-related protein and c-myc. Mol. Med. Rep. 2017, 17, 3497–3502.	Niclosamide enhances the cytotoxic effect of cisplatin in cisplatin-resistant human lung cancer cells via suppression of lung resistance-related protein and c-myc	Zuo, Y.; Yang, D.; Yu, Y.; Xiang, M.; Li, H.; Yang, J.; Li, J.; Jiang, D.; Zhou, H.; Xu, Z.; et al.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
77	Wang, C.; Zhou, X.; Xu, H.; Shi, X.; Zhao, J.; Yang, M.; Zhang, L.; Jin, X.; Hu, Y.; Li, X.; et al. Niclosamide Inhibits Cell Growth and Enhances Drug Sensitivity of Hepatocellular Carcinoma Cells via STAT3 Signaling Pathway. J. Cancer 2018, 9, 4150–4155.	Niclosamide Inhibits Cell Growth and Enhances Drug Sensitivity of Hepatocellular Carcinoma Cells via STAT3 Signaling Pathway	Wang, C.; Zhou, X.; Xu, H.; Shi, X.; Zhao, J.; Yang, M.; Zhang, L.; Jin, X.; Hu, Y.; Li, X.; et al.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
78	Stewart, R.L.; Carpenter, B.L.; West, D.S.; Knifley, T.; Liu, L.; Wang, C.; Weiss, H.L.; Gal, T.S.; Durbin, E.B.; Arnold, S.M.; et al. S100A4 drives non-small cell lung cancer invasion, associates with poor prognosis, and is effectively targeted by the FDA-approved anti-helminthic agent niclosamide. Oncotarget 2016, 7, 34630–34642.	S100A4 drives non-small cell lung cancer invasion, associates with poor prognosis, and is effectively targeted by the FDA-approved anti-helminthic agent niclosamide	Stewart, R.L.; Carpenter, B.L.; West, D.S.; Knifley, T.; Liu, L.; Wang, C.; Weiss, H.L.; Gal, T.S.; Durbin, E.B.; Arnold, S.M.; et al.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
79	Cheng, B.; Morales, L.D.; Zhang, Y.; Mito, S.; Tsun, A. Niclosamide induces p53 protein ubiquitination and inhibits multiple pro-survival signaling pathways in the human glioblastoma U-87 MG cell line. PLoS ONE 2017, 12, e0184324.	Niclosamide induces p53 protein ubiquitination and inhibits multiple pro-survival signaling pathways in the human glioblastoma U-87 MG cell line	Cheng, B.; Morales, L.D.; Zhang, Y.; Mito, S.; Tsun, A.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
80	Oh, H.-C.; Shim, J.-K.; Park, J.; Lee, J.-H.; Choi, R.J.; Kim, N.H.; Kim, H.S.; Moon, J.H.; Kim, E.H.; Chang, J.H.; et al. Combined effects of niclosamide and temozolomide against human glioblastoma tumorspheres. J. Cancer Res. Clin. Oncol. 2020, 146, 2817–2828.	Combined effects of niclosamide and temozolomide against human glioblastoma tumorspheres	Oh, H.-C.; Shim, J.-K.; Park, J.; Lee, J.-H.; Choi, R.J.; Kim, N.H.; Kim, H.S.; Moon, J.H.; Kim, E.H.; Chang, J.H.; et al.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
81	Wieland, A.; Trageser, D.; Gogol, S.; Reinartz, R.; Höfer, H.; Keller, M.; Leinhaas, A.; Schelle, R.; Normann, S.; Klaas, L.; et al. Anticancer Effects of Niclosamide in Human Glioblastoma. Clin. Cancer Res. 2013, 19, 4124–4136.	Anticancer Effects of Niclosamide in Human Glioblastoma	Wieland, A.; Trageser, D.; Gogol, S.; Reinartz, R.; Höfer, H.; Keller, M.; Leinhaas, A.; Schelle, R.; Normann, S.; Klaas, L.; et al.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
82	Liu, C.; Lou, W.; Armstrong, C.; Zhu, Y.; Evans, C.P.; Gao, A.C. Niclosamide suppresses cell migration and invasion in enzalutamide resistant prostate cancer cells via Stat3-AR axis inhibition. Prostate 2015, 75, 1341–1353.	Niclosamide suppresses cell migration and invasion in enzalutamide resistant prostate cancer cells via Stat3-AR axis inhibition	Liu, C.; Lou, W.; Armstrong, C.; Zhu, Y.; Evans, C.P.; Gao, A.C.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
83	Liu, C.; Lou, W.; Zhu, Y.; Nadiminty, N.; Schwartz, C.T.; Evans, C.P.; Gao, A.C. Niclosamide Inhibits Androgen Receptor Variants Expression and Overcomes Enzalutamide Resistance in Castration-Resistant Prostate Cancer. Clin. Cancer Res. 2014, 20, 3198–3210.	Niclosamide Inhibits Androgen Receptor Variants Expression and Overcomes Enzalutamide Resistance in Castration-Resistant Prostate Cancer	Liu, C.; Lou, W.; Zhu, Y.; Nadiminty, N.; Schwartz, C.T.; Evans, C.P.; Gao, A.C.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		
84	Chen, L.; Wang, L.; Shen, H.; Lin, H.; Li, D. Anthelmintic drug niclosamide sensitizes the responsiveness of cervical cancer cells to paclitaxel via oxidative stress-mediated mTOR inhibition. Biochem. Biophys. Res. Commun. 2017, 484,	Anthelmintic drug niclosamide sensitizes the responsiveness of cervical cancer cells to paclitaxel via oxidative stress-mediated mTOR inhibition	Chen, L.; Wang, L.; Shen, H.; Lin, H.; Li, D.	Phase I Study	To determine the safety and efficacy of	Recruiting	NCT01837862		

	Antiparasitic Type	Title	Phase	Purpose	Status/Result	Identifier/Ref
416–421.		Cancer				
85.	Antiparasitic	Wu, W.; Liu, H.; Yuan, J.; Yao, P. Targeting Wnt/beta-catenin by an anthelmintic drug niclosamide overcomes paclitaxel resistance in esophageal cancer. <i>Fundam. Clin. Pharmacol.</i> 2021, 35, 165–173.				
86.		Lee, M.; Chen, Y.; Hsu, Y.; Lin, B. Niclosamide inhibits the cell proliferation and enhances the responsiveness of esophageal cancer cells to chemotherapeutic agents. <i>Oncol. Rep.</i> 2019, 43, 549–561.				
87.		Liao, Z.; Nan, G.; Yan, Z.; Zeng, L.; Deng, Y.; Ye, J.; Zhang, Z.; Qian, M.; He, S.; Penduluri, S.; et al. The Anthelmintic Drug Niclosamide Inhibits the Proliferative Activity of Human Ovarian Cancer Cells by Targeting Multiple Signaling Pathways. <i>Curr. Cancer Drug Targets</i> 2015, 15, 726–738.			Completed (No result posted)	NCT02532114
88.		Satoh, K.; Zhang, L.; Zhang, Y.; Chelluri, R.; Boufraqueh, M.; Nilubol, N.; Patel, D.; Shen, M.; Kebebew, E. Identification of Niclosamide as a Novel Anticancer Agent for Adrenocortical Carcinoma. <i>Clin. Cancer Res.</i> 2016, 22, 3458–3466.				
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Salicylanilides are a very large group of compounds that show efficient activity against certain types of parasites. Their basic chemical structure consists of a salicylic acid ring and an anilide ring. Examples of HS drugs with potent anthelmintic activity are Nic, rafoxanide (RFX), and closanter.

3.1. Mechanism of Action of HS

The primary mechanism of action of HS was investigated in vitro using houseflies and rat liver mitochondria. The authors found an association with the uncoupling of oxidative phosphorylation that halts the production of ATP. This seems to happen through the suppression of the activity of two enzymes, succinate dehydrogenase and fumarate reductase, and thus impairs the motility of parasites and eventually causes death [98]. Several researchers have subsequently confirmed the proposed mechanism in vivo [45][99].

3.2. Anticancer Activity of HS

Several HS group drugs have been investigated for their effect on cancer in experimental and preclinical models. The pharmacokinetic properties and common side effects of HS drugs are shown in **Figure 2b**.

3.3. Anticancer Activity of HS in Clinical Models

Nic underwent clinical trials in patients with resectable colon cancer in 2017, but was terminated because of the low enrolment rate (NCT02687009). Two other clinical studies are currently underway to test the anticancer effects of Nic in patients with FAP (NCT04296851) and progression of metastases of colorectal cancer after therapy (NCT02519582). Although a phase I trial of Nic administered together with enzalutamide in patients with castration-resistant prostate cancer has concluded, anticipating the commencement of a phase 2 trial (NCT02532114), another phase I clinical trial is investigating the potent dose and side effects of Nic in combination with enzalutamide to treat castration-resistant prostate cancer patients (NCT03123978). A phase II clinical trial is also ongoing to evaluate the efficacy of abiraterone acetate, Nic, and prednisone in treating patients with hormone-resistant prostate cancer (NCT02807805).