Multidrug Resistant *Pseudomonas aeruginosa* in Lebanon

Subjects: Infectious Diseases

Contributor: Issmat Kassem, Marwan Osman, Fouad Dabboussi

Pseudomonas aeruginosa is a common cause of healthcare-associated infections and chronic airway diseases in non-clinical settings. *P. aeruginosa* is intrinsically resistant to a variety of antimicrobials and has the ability to acquire resistance to others, causing increasingly recalcitrant infections and elevating public health concerns. It is showed that the bacterium was predominant in lesions of patients on mechanical ventilation and in burn patients and those with diabetic foot infections and hematological malignancies. It is also found that carbapenem resistance in *P. aeruginosa* isolates in Lebanon involved both enzymatic and non-enzymatic mechanisms but depended predominantly on VIM-2 production (40.7%). Additionally, MDR *P. aeruginosa* was detected in animals, where a study reported the emergence of carbapenemase-producing *P. aeruginosa* in livestock in Lebanon.

Keywords: Pseudomonas aeruginosa; MDR; carbapenemase; Lebanon; Antibiotic Resistance; Infectious Diseases

1. Introduction

Pseudomonas aeruginosa is a known bacterial pathogen that can cause severe infections in humans. *P. aeruginosa* is also notable for its intrinsic and acquired resistance to a broad spectrum of clinically-relevant antibiotics. The prevalence of community- and healthcare-associated multidrug-resistant (MDR) *P. aeruginosa* infections has been increasing over the last few decades, becoming a global public health threat and leading to difficult-to-treat infections [1]. Subsequently, the World Health Organization (WHO) has classified *P. aeruginosa* among the prominent drug-resistant bacteria that require urgent research and the development of effective therapeutic interventions [2].

Despite the importance of *P. aeruginosa*, the prevalence and spread of MDR strains of this opportunistic pathogen have not been fully investigated in Lebanon. This is especially important, because AMR has been steadily increasing and potentially causing significant morbidity and mortality in Lebanon [3][4][5][6]. Furthermore, limited studies on AMR in Lebanon have revealed insufficient knowledge and inadequate practices related to antibiotics and AMR in general, including weak stewardship in clinical settings, over-the-counter access to antimicrobials in community pharmacies and a lack of awareness to the dangers of inappropriate use in the Lebanese community at large [2][8]. Additionally, the healthcare system in Lebanon has been affected by many emerging and significant challenges, including a collapsing national economy, the COVID-19 pandemic, and the burden of providing services for ~1.5 million Syrian refugees [9]. The devaluation of the Lebanese currency and the significant decrease in the quality of life have resulted in a serious shortage in healthcare professionals, including infectious diseases specialists and clinical microbiologists due to immigration. Consequently, the United Nations Economic and Social Commission for Western Asia (ESCWA) reported that more than 82% of the Lebanese population has limited access to healthcare services [10].

The economic crisis has also affected drug availability and in vitro diagnostics for various medical applications, including screening for antimicrobial resistance, while AMR surveillance and research programs (in clinical settings and the environment) became difficult to conduct [9][11]. For example, the American University of Beirut Medical Center (AUBMC), one of Lebanon's largest tertiary care hospitals, has been running perilously short of medical supplies, affecting more than 500 critical items. The hospital now rations antibiotics. As a result, many patients are delaying treatment, leading to a higher risk of invasive infections and prolonged hospital stays [12]. This is not surprising when considering that Lebanon relies heavily on imports to meet its needs of medical and research supplies, and the severe devaluation of the Lebanese currency has limited purchases from foreign suppliers [9]. Predictably, the repercussions of a failing economy extend beyond the medical sector and impact agricultural practices and the environment, resulting in widespread pollution of critical resources in the country [11]. For example, prior to the crisis, Lebanon was already witnessing an emergence of resistance to last resort antibiotics in food industries, poultry farms, aquaculture, and surface water (irrigation, rivers, and sea), partially due to a debilitated infrastructure and suboptimal agricultural practices [5][6][13][14][15][16][17][18][19][20].

Therefore, AMR has been amplified due to many interacting reasons that include a weakened ability to tackle domestic and agricultural waste and maintain good animal farming practices. Taken together, the aforementioned observations suggest that the emergence of AMR and complicated infectious diseases in Lebanon will increase further.

In response to elevated concerns about AMR and infectious diseases in Lebanon, researchers aimed to gather existing evidence on antibiotic-resistant *P. aeruginosa* in order to address the knowledge gaps regarding its epidemiology in Lebanon, a developing East Mediterranean country that is facing unprecedented economic and healthcare challenges.

2. Mechanisms of Antimicrobial Resistance in Pseudomonas aeruginosa

P. aeruginosa is resistant to numerous antimicrobials, which is facilitated by different mechanisms that include (1) restricting outer membrane permeability, (2) the expression of many efflux systems (e.g., constitutive MexAB-OprM, inducible MexXY-OprM), (3) the production of naturally-occurring antimicrobial-inactivating enzymes such as the hydrolytic β-lactamase enzymes (bla_{AmpC} and bla_{OXA-50}) and the aminoglycoside modifying enzyme (AME) APH(3')-IIb, and (4) mutations and enzymes that modify the targets of the antimicrobials (**Figure 1**) [21][22]. P. aeruginosa is also capable of developing antimicrobial resistance via horizontal gene transfer and the acquisition of resistance genes. It is important to note that both intrinsic and acquired resistance mechanisms play an important role in the evolution of MDR P. aeruginosa. For example, carbapenem resistance mechanisms in P. aeruginosa include the overexpression of AmpC enzyme, the acquisition of extended-spectrum β-lactamase (ESBL) and/or carbapenemase encoding genes through horizontal gene transfer, reduction in membrane permeability (e.g., mutations in the outer membrane porin, OprD), overexpression of mexAB-oprM efflux pump, and/or modification of penicillin binding proteins (PBPs) P (23)[24][25].

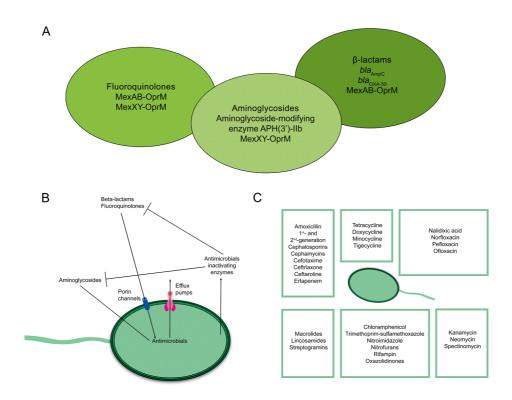


Figure 1. Antimicrobial resistance in Pseudomonas aeruginosa.

Similarly, aminoglycosides resistance in *P. aeruginosa* has been associated with an array of resistance mechanisms. For example, *P. aeruginosa* can inactivate aminoglycosides using AMEs which include acetyltransferases (AAC), nucleotidyltransferases (ANT), and phosphotransferases (APH). Numerous AMEs that were reported in the literature confer resistance to different aminoglycosides. Specifically, (i) AAC(3)-X, AAC(6')-Ib, ANT(4')-I, ANT(4')-II, APH(3')-IIIa, and APH(3')-VIb display resistance against amikacin, (ii) AAC(3), AAC(6')-I, AAC(6')-II, and ANT(2") confer resistance to gentamicin, and (iii) AAC(3)-II, ANT(2"), ANT(4')-I, and ANT(4')-II inactivate tobramycin. Resistance to aminoglycosides also includes overexpression of efflux pumps (particularly the MexXY-OprM complex), modification of 16S ribosomal RNA by methylases (e.g., *rmtA* and *rmtB*; preventing aminoglycosides from effectively binding to ribosomes), and decreased permeability [26].

Notably, *P. aeruginosa* can potentially develop resistance to fluoroquinolones, colistin, and fosfomycin. For example, although fluoroquinolones are frequently used to control infections with this bacterium, mutations in quinolone-resistance associated genes (i.e., *gyrA*, *gyrB*, *parC* and *parE*) along with the overexpression of resistance–nodulation–division efflux

pumps (i.e., MexAB-OprM, MexCD-OprJ, MexEF-OprN, and MexXY-OprM) [27] are prevalent determinants that contribute to fluoroquinolone resistance in *P. aeruginosa*.

Colistin (polymyxin E) has been used as a last resort for the treatment of MDR and extensively drug resistant (XDR) P. aeruginosa infections [28]. However, in recent years, resistance to colistin has emerged around the globe, complicating the clinical management of certain MDR infections [29]. The drug's consumption in Lebanese hospitals has increased 5×10^{12} between 2010 and 2017; highlighting the importance of this drug in treating a variety of recalcitrant infections as well as the increasing number of complicated and MDR infections in Lebanon [19]. Resistance to colistin is commonly facilitated by mutations in genes associated with the modification of the lipid A of LPS and/or through the acquisition of the mobile colistin resistant (mcr) genes [30]. The emergence, spread, and notable transmissibility of mcr have raised public health concerns over the loss of the efficacy of colistin in treating MDR P. aeruginosa and other bacterial pathogens. Although different mcr genes have been widely reported in Enterobacterales, only $ext{mcr} 1$ $ext{mcr} 2$ and $ext{mcr} 3$ have been identified in sporadic P. $ext{aeruginosa}$ isolates so far.

Due to the limited use of and the relatively low level of reported fosfomycin resistance in P. aeruginosa, this drug has been revisited to control antimicrobial-resistant P. aeruginosa strains [35]. Similarly to the antimicrobials discussed above, resistance to fosfomycin in P. aeruginosa can develop and is mainly associated with the gene encoding the inactivating enzyme, FosA [36], or the inactivation of the fosfomycin transport protein (GIpT) [37].

Taken together, it is perhaps clear that the antimicrobial options for treating and controlling *P. aeruginosa* infections are becoming increasingly limited, mainly due to the ability of this bacterium to develop resistance. This is predicted to have a serious impact on the emergence and spread of *P. aeruginosa* infections, especially in resource-limited countries like Lebanon.

3. Epidemiology of Pseudomonas aeruginosa Resistance in Lebanon

It is important to assess the epidemiology and the molecular mechanisms of resistance of MDR *P. aeruginosa* in Lebanon in order to guide empirical treatment choices. However, information on AMR in general and MDR *P. aeruginosa* in particular is lacking. For this purpose, researchers screened the literature to provide accessible and science-based evidence on the scope of this problem in Lebanon. Consequently, PubMed, Science Direct, Scopus, and the Google Scholar databases were mined for epidemiological studies on *P. aeruginosa* in hospital and/or extra-hospital settings that were published up to December 2021. Researchers used a combination of the following terms: "*Pseudomonas*", "*aeruginosa*", "Lebanon", "Susceptibility", "Resistance", "Antimicrobial", "Antibiotic", "AMR", "Epidemiology", "Imipenem", "Meropenem", and "Carbapenem". Indexed original articles in English and French of any epidemiological design and sampling strategy and of any enrollment timing (retrospective, prospective, or cross-sectional) were included. Other types of reports, such as case reports, case series, and narrative and systematic reviews were excluded (**Figure 2**). Studies were eligible for inclusion if they reported original information regarding the epidemiology of *P. aeruginosa* and its resistance to antibiotics in Lebanon. After importation of the search results, two authors (A.A. Dabbousi and F. Dabboussi) independently screened the citations for their relevance using the title and abstract and all qualified citations were retained for full-text assessment to confirm eligibility. Backward reference screening was done for all articles. Data extraction was performed by the same authors through a format prepared on a Microsoft Excel workbook.

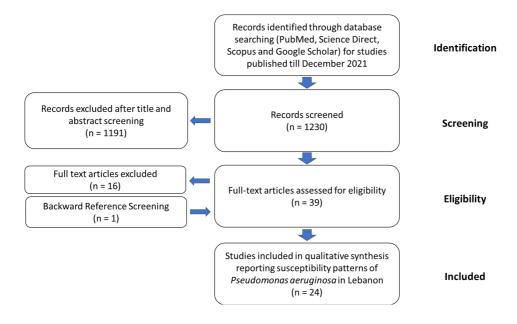


Figure 2. A Flow diagram describing the selection of the studies and the inclusion/exclusion process according to PRISMA guidelines.

The search strategy initially resulted in 1230 studies. Subsequently, a total of 40 manuscripts were screened in the full-text review and 24 studies describing the epidemiology and susceptibility profiles of P. aeruginosa isolates in Lebanon were identified as eligible according to the inclusion criteria. The number of studies excluded or included at each stage is summarized in **Figure 2**. There were three nationwide studies identified; however, most of the included studies were conducted in Beirut (12 of 21), particularly at the American University of Beirut Medical Center (AUBMC) (N = 5). Additionally, eight studies were performed in the North and Akkar governorates (North Lebanon region). Furthermore, almost all reports (23 of 24) studied human samples, particularly in hospital settings. Only one paper investigated the susceptibility patterns of P. aeruginosa in animals, reporting the emergence of P. aeruginosa producing VIM-2 carbapenemase in Lebanese livestock [38].

The first data on AMR profiles of *P. aeruginosa* in Lebanon were generated at the AUBMC and Makassed general hospital located in Beirut [39][40]. At that time, 11% of the circulating clinical *P. aeruginosa* isolates were resistant to ceftazidime [39] [40] and 8% were resistant to imipenem [40]. In the first decade of the 21st century, it appears that the susceptibility of strains to ceftazidime and imipenem significantly decreased. Specifically, in a 11-year retrospective study at the AUBMC, the prevalence of resistance against both antimicrobials reached 17% and 19%, respectively [41]. However, another large scale study that included 5090 clinical samples from patients suffering from healthcare- or community-acquired infections at the Hôtel-Dieu de France Hospital (Beirut) showed a higher prevalence of resistance to ceftazidime (34.7%) and imipenem (41.1%) [42]. Similar findings were obtained in other Lebanese geographical regions such as Tripoli, the North governorate of Lebanon [43]. Recent data from three studies conducted between 2014 and 2018 in local tertiary care centers in Beirut, North, and Akkar governorates were alarming, showing that 40-97.1% of P. aeruginosa infections were due to carbapenem-resistant isolates [44][45][46]. Furthermore, resistance against other antimicrobials has been reported in the Lebanese clinical settings. For example, 26.4% and 36.2% of the isolates were resistant to amikacin and levofloxacin between 2005 and 2009 at Hôtel-Dieu de France Hospital, respectively [42]. Yet again, the susceptibility of circulating P. aeruginosa against different antimicrobials appears to have continued to decrease over time. Although P. aeruginosa susceptibility patterns have been reported in several studies in Lebanon during the last three decades, most reports investigated a limited number of isolates (<150 isolates in 16 studies) in monocentric study locations. Therefore, like other clinically important pathogens, the full burden of AMR of P. aeruginosa in Lebanon remains unclear due to the lack of national surveillance data, a limited number of well-designed national studies, weak epidemiological tracking, and the absence of adequate funding, infrastructure, and oversight among other factors [9]. Nevertheless, the three nationwide retrospective investigations based on aggregated institutional antimicrobial susceptibility testing data from tertiary care centers located in different Lebanese districts have confirmed the relatively high level of resistance to ceftazidime, imipenem, amikacin, and levofloxacin. This was corroborated by the results observed in local studies conducted in Lebanon [47][48][49]. Additionally, a nationwide study reported for the first time the emergence of colistin-resistant P. aeruginosa isolates in Lebanon [47]. This was followed by another more recent and geographically-constrained study that also corroborated the emergence of colistin-resistant *P. aeruginosa* isolates in Lebanese clinical settings [44]. Taken together, these findings highlight a worrying trend that has been developing in Lebanon in recent decades and perhaps reflect the inappropriate use and/or over-reliance on carbapenems and colistin in the treatment of infections [I][9].

Unfortunately, only a few studies have addressed the occurrence of resistance genes in P. aeruginosa (**Figure 3**). Nevertheless, the limited data showed that carbapenem-resistant P. aeruginosa in Lebanon encompassed (1) carbapenem-hydrolyzing enzymes (including bla_{VIM-2} , bla_{GES-6} , bla_{IMP-1} , bla_{IMP-2} , and bla_{IMP-15}), (2) non-enzymatic mechanisms (alteration of the outer membrane porin protein OprD, overexpression of efflux pumps), and (3) a combination of reduced membrane permeability and/or drug efflux pumps with enzyme inactivation mechanisms such as Class C β -lactamase hyperproduction (e.g., PDC-13, AmpC) $\frac{49[50][51][52][53]}{152[53]}$. It should be noted that a fingerprint analysis of strains isolated from various Lebanese hospitals indicated that the VIM-2 occurrence in P. aeruginosa was primarily due to clonal dissemination $\frac{49}{9}$. These results corroborated previous reports in the Middle East and North Africa (MENA) region that showed that different types of carbapenemases (VIM-2 was predominant) have been described in P. aeruginosa isolates in the countries surrounding Lebanon $\frac{53}{9}$. Recently, the emergence of carbapenemase-producing P aeruginosa harboring bla_{VIM-2} has also been reported in livestock in Lebanon, potentially suggesting a zoo-anthropogenic transmission of VIM-2 producing P. aeruginosa and raising further concerns about the dissemination of MDR P aeruginosa in animals and via zoonosis $\frac{54}{9}$.

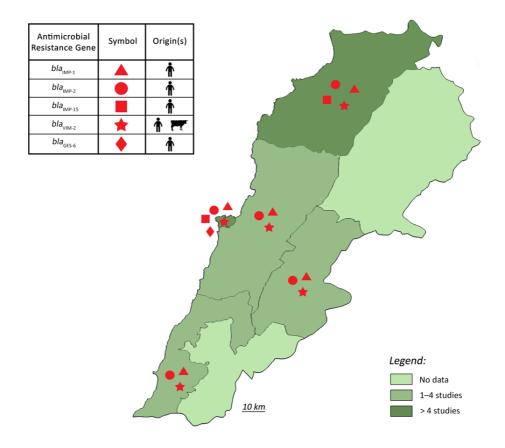


Figure 3. The geographical distribution of β-lactamase genes in *Pseudomonas aeruginosa* isolated from humans and animals in Lebanon.

Due to the emergence and spread of resistance to traditional antimicrobial agents, healthcare professionals have been using the U.S. Food and Drug Administration (FDA) approved antipseudomonal beta-lactam drugs, ceftolozane/tazobactam and ceftazidime/avibactam [55]. These two new combinations of β -lactam/ β -lactamase inhibitor antibiotics were recently registered at the Lebanese Ministry of Public Health as agents active against many MDR isolates of *P. aeruginosa* [55][56]. Regarding ceftolozane/tazobactam use in Lebanon, a study performed at AUBMC has shown a high susceptibility (96%) against non-MDR *P. aeruginosa* isolates but a low susceptibility (42%) against MDR isolates [56]. To date, national data on the resistance of *P. aeruginosa* to ceftazidime/avibactam are not available. These observations further highlight the urgency needed to tackle the glaring gaps in knowledge about MDR *P. aeruginosa* infections and associated controls and treatments in Lebanon.

4. Conclusions

Although Lebanon joined the World Health Organization's (WHO) Global Antimicrobial Resistance Surveillance System (GLASS) in 2017, antimicrobial stewardship is still underdeveloped across the country. This situation has likely resulted in MDR *P. aeruginosa* to be prevalent in Lebanese hospitals and precipitated the emergence of carbapenem resistance that is associated predominantly with VIM-2 production. In conclusion, there is a critical need to establish robust monitoring and AMR stewardship programs and to devise interventions at the policy level that will bolster a national strategic plan to combat AMR in Lebanon. Otherwise, the country will face undesirable public health problems. AMR stewardship programs and extensive awareness campaigns must be integrated in the Lebanese vulnerable health system which has been impeded by a plethora of challenges. These interventions are required to curb mortality and morbidity due to AMR in the Lebanese population as well as in the large refugee population that is currently hosted in Lebanon. Given the proximity of Lebanon to many European, Middle Eastern and African countries and the mobility of the Lebanese and refugee populations, there is a risk that MDR can spill across the Lebanese borders, affecting other countries in the region and beyond.

References

- 1. Horcajada, J.P.; Montero, M.; Oliver, A.; Sorlí, L.; Luque, S.; Gómez-Zorrilla, S.; Benito, N.; Grau, S. Epidemiology and Treatment of Multidrug-Resistant and Extensively Drug-Resistant Pseudomonas aeruginosa Infections. Clin. Microbiol. Rev. 2019, 32, e00031-19.
- 2. Filho, F.; Nascimento, A.P.; Costa, M.; Thiago, M.; Menezes, M.; Marisa, N.; Trindade dos Santos, M.; Carvalho-Assef, A.P.; da Silva, F. A systematic strategy to find potential therapeutic targets for Pseudomonas aeruginosa using integrated computational models. Front. Mol. Biosc. 2021, 8, 728129.
- 3. Abbara, A.; Rawson, T.M.; Karah, N.; El-Amin, W.; Hatcher, J.; Tajaldin, B.; Dar, O.; Dewachi, O.; Abu Sitta, G.; Uhlin, B.E.; et al. Antimicrobial resistance in the context of the Syrian conflict: Drivers before and after the onset of conflict and key recommendations. Int. J. Infect. Dis. 2018, 73, 1–6.
- 4. Osman, M.; Halimeh, F.B.; Rafei, R.; Mallat, H.; Tom, J.E.; BouRaad, E.; Diene, S.D.; Jamal, S.; Al Atrouni, A.; Dabboussi, F.; et al. Investigation of an XDR-Acinetobacter baumannii ST2 outbreak in an intensive care unit of a Lebanese tertiary care hospital. Future Microbiol. 2020, 15, 1535–1542.
- 5. Hmede, Z.; Kassem, I.I. The Colistin Resistance Gene mcr-1 Is Prevalent in Commensal Escherichia coli Isolated from Preharvest Poultry in Lebanon. Antimicrob. Agents Chemother. 2018, 62, e01304-18.
- Hmede, Z.; Sulaiman, A.A.A.; Jaafar, H.; Kassem, I.I. Emergence of plasmid-borne colistin resistance gene mcr-1 in multidrug-resistant Escherichia coli isolated from irrigation water in Lebanon. Int. J. Antimicrob. Agents 2019, 54, 102– 104
- 7. Al-Omari, S.; Al Mir, H.; Wrayde, S.; Merhabi, S.; Dhaybi, I.; Jamal, S.; Chahine, M.; Bayaa, R.; Tourba, F.; Tantawi, H.; et al. First Lebanese Antibiotic Awareness Week campaign: Knowledge, attitudes and practices towards antibiotics. J. Hosp. Infect. 2019, 101, 475–479.
- 8. Samer, S.; Ghaddar, A.; Hamam, B.; Sheet, I. Antibiotic use and resistance: An unprecedented assessment of university students' knowledge, attitude and practices (KAP) in Lebanon. BMC Public Health 2020, 20, 535.
- 9. Osman, M.; Kasir, D.; Kassem, I.I.; Hamze, M. Shortage of appropriate diagnostics for antimicrobial resistance in Lebanese clinical settings: A crisis amplified by COVID-19 and economic collapse. J. Glob. Antimicrob. Resist. 2021, 27, 72–74.
- 10. ESCWA. Escwa Warns: More Than Half of Lebanon's Population Trapped in Poverty 2020. Available online: https://www.unescwa.org/news/lebanon-population-trapped-poverty (accessed on 6 May 2022).
- 11. Kassem, I.I.; Osman, M. A brewing storm: The impact of economic collapse on the access to antimicrobials in Lebanon. J. Glob. Antimicrob. Resist. 2022; ahead of print.
- 12. Dadouch, S.; Durgham, N. Lebanon was Famed for Its Medical Care. Now, Doctors and Nurses are Fleeing in Droves. The Washington Post 2021. Available online: https://www.washingtonpost.com/world/middle_east/lebanon-crisis-healthcare-doctors-nurses/2021/11/12/6bf79674-3e33-11ec-bd6f-da376f47304e_story.html (accessed on 6 May 2022).
- 13. Dagher, L.A.; Hassan, J.; Kharroubi, S.; Jaafar, H.; Kassem, I.I. Nationwide Assessment of Water Quality in Rivers across Lebanon by Quantifying Fecal Indicators Densities and Profiling Antibiotic Resistance of Escherichia coli. Antibiotics 2021, 10, 883.
- 14. Hassan, J.; Zein Eddine, R.; Mann, D.; Li, S.; Deng, X.; Saoud, I.P.; Kassem, I.I. The mobile colistin resistance gene, mcr-1.1, is carried on lncx4 plasmids in multidrug resistant E. coli isolated from rainbow trout aquaculture. Microorganisms 2020, 8, 1636.
- 15. Sourenian, T.; Mann, D.; Li, S.; Deng, X.; Jaafar, H.; Kassem, I.I. Dissemination of multidrug-resistant Escherichia coli harboring the mobile colistin resistance gene mcr-1.1 on transmissible plasmids in the Mediterranean Sea. J. Glob. Antimicrob. Resist. 2020, 22, 84–86.
- 16. Al-Mir, H.; Osman, M.; Drapeau, A.; Hamze, M.; Madec, J.-Y.; Haenni, M. Spread of ESC-, carbapenem- and colistin-resistant Escherichia coli clones and plasmids within and between food workers in Lebanon. J. Antimicrob. Chemother. 2021, 76, 3135–3143.
- 17. Al-Mir, H.; Osman, M.; Drapeau, A.; Hamze, M.; Madec, J.-Y.; Haenni, M. WGS Analysis of Clonal and Plasmidic Epidemiology of Colistin-Resistance Mediated by mcr Genes in the Poultry Sector in Lebanon. Front. Microbiol. 2021, 12, 624194.
- 18. Osman, M.; Al Mir, H.; Rafei, R.; Dabboussi, F.; Madec, J.-Y.; Haenni, M.; Hamze, M. Epidemiology of antimicrobial resistance in Lebanese extra-hospital settings: An overview. J. Glob. Antimicrob. Resist. 2019, 17, 123–129.
- 19. Kassem, I.I.; Hijazi, M.A.; Saab, R. On a collision course: The availability and use of colistin-containing drugs in human therapeutics and food-animal farming in Lebanon. J. Glob. Antimicrob. Resist. 2019, 16, 162–164.

- 20. Kassem, I.I.; Nasser, N.A.; Salibi, J. Prevalence and Loads of Fecal Pollution Indicators and the Antibiotic Resistance Phenotypes of Escherichia coli in Raw Minced Beef in Lebanon. Foods 2020, 9, 1543.
- 21. Blair, J.M.A.; Webber, M.A.; Baylay, A.J.; Ogbolu, D.O.; Piddock, L.J.V. Molecular mechanisms of antibiotic resistance. Nat. Rev. Microbiol. 2015, 13, 42–51.
- 22. Pang, Z.; Raudonis, R.; Glick, B.R.; Lin, T.-J.; Cheng, Z. Antibiotic resistance in Pseudomonas aeruginosa: Mechanisms and alternative therapeutic strategies. Biotechnol. Adv. 2019, 37, 177–192.
- 23. Hirsch, E.B.; Brigman, H.V.; Zucchi, P.C.; Chen, A.; Anderson, J.C.; Eliopoulos, G.M.; Cheung, N.; Gilbertsen, A.; Hunter, R.C.; Emery, C.L.; et al. Ceftolozane-tazobactam and ceftazidime-avibactam activity against b-lactam-resistant Pseudomonas aeruginosa and extended-spectrum b-lactamase-producing enterobacterales clinical isolates from U.S. medical centres. J. Glob. Antimicrob. Resist. 2020, 22, 689–694.
- 24. Cultrera, R.; Libanore, M.; Barozzi, A.; D'Anchera, E.; Romanini, L.; Fabbian, F.; De Motoli, F.; Quarta, B.; Stefanati, A.; Bolognesi, N.; et al. Ceftolozane/Tazobactam and Ceftazidime/Avibactam for Multidrug-Resistant Gram-Negative Infections in Immunocompetent Patients: A Single-Center Retrospective Study. Antibiotics 2020, 9, 640.
- 25. Zamudio, R.; Hijazi, K.; Joshi, C.; Aitken, E.; Oggioni, M.R.; Gould, I.M. Phylogenetic analysis of resistance to ceftazidime/avibactam, ceftolozane/tazobactam and carbapenems in piperacillin/tazobactam-resistant Pseudomonas aeruginosa from cystic fibrosis patients. Int. J. Antimicrob. Agents 2019, 53, 774–780.
- 26. Ramirez, M.S.; Tolmasky, M.E. Aminoglycoside modifying enzymes. Drug Resist. Updat. 2010, 13, 151-171.
- 27. Zhao, L.; Wang, S.; Li, X.; He, X.; Jian, L. Development of in vitro resistance to fluoroquinolones in Pseudomonas aeruginosa. Antimicrob. Resist. Infect. Control. 2020, 9, 124.
- 28. Walters, M.S.; Grass, J.E.; Bulens, S.N.; Hancock, E.B.; Phipps, E.C.; Muleta, D.; Mounsey, J.; Kainer, M.A.; Concannon, C.; Dumyati, G.; et al. Carbapenem-resistant Pseudomonas aeruginosa at US emerging infections program sites, 2015. Emerg. Infect. Dis. 2019, 25, 1281–1288.
- 29. Hamel, M.; Rolain, J.-M.; Baron, S. The History of Colistin Resistance Mechanisms in Bacteria: Progress and Challenges. Microorganisms 2021, 9, 442.
- 30. Fernández, L.; Álvarez-Ortega, C.; Wiegand, I.; Olivares, J.; Kocíncová, D.; Lam, J.S.; Martínez, J.L.; Hancock, R.E.W. Characterization of the Polymyxin B Resistome of Pseudomonas aeruginosa. Antimicrob. Agents Chemother. 2013, 57, 110–119.
- 31. Pathak, A.; Singh, S.; Kumar, A.; Prasad, K. Emergence of chromosome borne colistin resistance gene, mcr-1 in clinical isolates of Pseudomonas aeruginosa. Int. J. Infect. Dis. 2020, 101, 22.
- 32. El-Baky, R.M.A.; Masoud, S.M.; Mohamed, D.S.; Waly, N.G.; Shafik, E.A.; Mohareb, D.A.; Elkady, A.; Elbadr, M.M.; Hetta, H.F. Prevalence and Some Possible Mechanisms of Colistin Resistance Among Multidrug-Resistant and Extensively Drug-Resistant Pseudomonas aeruginosa. Infect. Drug Resist. 2020, 13, 323–332.
- 33. Liu, Y.-Y.; Chandler, C.E.; Leung, L.M.; McElheny, C.L.; Mettus, R.T.; Shanks, R.M.Q.; Liu, J.-H.; Goodlett, D.R.; Ernst, R.K.; Doi, Y. Structural Modification of Lipopolysaccharide Conferred by mcr-1 in Gram-Negative ESKAPE Pathogens. Antimicrob. Agents Chemother. 2017, 61, e00580-17.
- 34. Snesrud, E.; Maybank, R.; Kwak, Y.I.; Jones, A.R.; Hinkle, M.K.; McGann, P. Chromosomally Encoded mcr-5 in Colistin-Nonsusceptible Pseudomonas aeruginosa. Antimicrob. Agents Chemother. 2018, 62, e00679-18.
- 35. Abbott, I.J.; Van Gorp, E.; Wijma, R.A.; Dekker, J.; Croughs, P.D.; Meletiadis, J.; Mouton, J.W.; Peleg, A.Y. Efficacy of single and multiple oral doses of fosfomycin against Pseudomonas aeruginosa urinary tract infections in a dynamic in vitro bladder infection model. J. Antimicrob. Chemother. 2020, 75, 1879–1888.
- 36. Ito, R.; Mustapha, M.M.; Tomich, A.D.; Callaghan, J.D.; McElheny, C.L.; Mettus, R.T.; Shanks, R.M.Q.; Sluis-Cremer, N.; Doi, Y. Widespread Fosfomycin Resistance in Gram-Negative Bacteria Attributable to the Chromosomal fosA Gene. MBio 2017, 8, e00749-17.
- 37. Díez-Aguilar, M.; Morosini, M.I.; Tedim, A.P.; Rodríguez, I.; Aktaş, Z.; Cantón, R. Antimicrobial Activity of Fosfomycin-Tobramycin Combination against Pseudomonas aeruginosa Isolates Assessed by Time-Kill Assays and Mutant Prevention Concentrations. Antimicrob. Agents Chemother. 2015, 59, 6039–6045.
- 38. Al Bayssari, C.; Dabboussi, F.; Hamze, M.; Rolain, J.-M. Emergence of carbapenemase-producing Pseudomonas aeruginosa and Acinetobacter baumannii in livestock animals in Lebanon. J. Antimicrob. Chemother. 2015, 70, 950–951.
- 39. Shaar, T.; Al-Hajjar, R. Antimicrobial susceptibility patterns of bacteria at the Makassed General Hospital in Lebanon. Int. J. Antimicrob. Agents 2000, 14, 161–164.

- 40. Araj, G.F.; Uwaydah, M.M.; Alami, S.Y. Antimicrobial susceptibility patterns of bacterial isolates at the american university medical center in Lebanon. Diagn. Microbiol. Infect. Dis. 1994, 20, 151–158.
- 41. Araj, G.F.; Avedissian, A.Z.; Ayyash, N.S.; Bey, H.A.; El Asmar, R.G.; Hammoud, R.Z.; Itani, L.Y.; Malak, M.R.; Sabai, S.A. A reflection on bacterial resistance to antimicrobial agents at a major tertiary care center in Lebanon over a decade. J. Med. Liban. 2012, 60, 125–135.
- 42. Hamouche, E.; Sarkis, D.K. Evolution of susceptibility to antibiotics of Escherichia coli, Klebsiella pneumoniae, Pseudomonas aeruginosa and Acinetobacter baumanii, in a university hospital center of Beirut between 2005 and 2009. Pathol. Biol. 2012, 60, e15–e20.
- 43. Hamze, M.; Dabboussi, F.; Izard, D. A 4-year study of Pseudomonas aeruginosa susceptibility to antibiotics (1998–2001) in northern Lebanon. Med. Mal. Infect. 2004, 34, 321–324.
- 44. Bourgi, J.; Said, J.M.; Yaakoub, C.; Atallah, B.; Al Akkary, N.; Sleiman, Z.; Ghanimé, G. Bacterial infection profile and predictors among patients Aadmitted to a burn care center: A retrospective study. Burns 2020, 46, 1968–1976.
- 45. Hamze, M.; Osman, M.; Mallat, H.; Achkar, M. Prevalence and antibiotic susceptibility of ear pathogens isolated from patients in Tripoli, north of Lebanon. Int. Arab. J. Antimicrob. Agents 2017, 7, 1–10.
- 46. Osman, M.; Mallat, H.; Hamze, M.; Bou Raad, E. Prevalence and antibiotic susceptibility patterns of bacteria causing urinary uract infections in Youssef hospital center: First report from akkar governorate, north Lebanon. Int. Arab. J. Antimicrob. Agents 2017, 7, 2–4.
- 47. Moghnieh, R.; Araj, G.F.; Awad, L.; Daoud, Z.; Mokhbat, J.E.; Jisr, T.; Abdallah, D.; Azar, N.; Irani-Hakimeh, N.; Balkis, M.M.; et al. A compilation of antimicrobial susceptibility data from a network of 13 Lebanese hospitals reflecting the national situation during 2015–2016. Antimicrob. Resist. Infect. Control. 2019, 8, 41.
- 48. Chamoun, K.; Farah, M.; Araj, G.; Daoud, Z.; Moghnieh, R.; Salameh, P.; Saade, D.; Mokhbat, J.; Abboud, E.; Hamze, M.; et al. Surveillance of antimicrobial resistance in Lebanese hospitals: Retrospective nationwide compiled data. Int. J. Infect. Dis. 2016, 46, 64–70.
- 49. Halat, D.H.; Moubareck, C.A.; Sarkis, D.K. Heterogeneity of Carbapenem Resistance Mechanisms Among Gram-Negative Pathogens in Lebanon: Results of the First Cross-Sectional Countrywide Study. Microb. Drug Resist. 2017, 23, 733–743.
- 50. Hammoudi, D.; Moubareck, C.A.; Kanso, A.; Nordmann, P.; Sarkis, D.K. Surveillance of carbapenem non-susceptible Gram negative strains and characterization of carbapenemases of classes A, B, and D in a Lebanese hospital. J. Med. Liban. 2015, 63, 66–73.
- 51. Yaghi, J.; Fattouh, N.; Akkawi, C.; El Chamy, L.; Maroun, R.G.; Khalil, G. Unusually High Prevalence of Cosecretion of Ambler Class A and B Carbapenemases and Nonenzymatic Mechanisms in Multidrug-Resistant Clinical Isolates of Pseudomonas aeruginosa in Lebanon. Microb. Drug Resist. 2020, 26, 150–159.
- 52. Dagher, T.N.; Al-Bayssari, C.; Diene, S.; Azar, E.; Rolain, J.-M. Emergence of plasmid-encoded VIM-2–producing Pseudomonas aeruginosa isolated from clinical samples in Lebanon. New Microbes New Infect. 2019, 29, 100521.
- 53. Al-Bayssari, C.; Dagher, T.N.; El Hamoui, S.; Fenianos, F.; Makdissy, N.; Rolain, J.-M.; Nasreddine, N. Carbapenem and colistin-resistant bacteria in North Lebanon: Coexistence of MCR-1 and NDM-4 genes in Escherichia coli. J. Infect. Dev. Ctries. 2021, 15, 934-342.
- 54. Al Bayssari, C.; Diene, S.M.; Loucif, L.; Gupta, S.K.; Dabboussi, F.; Mallat, H.; Hamze, M.; Rolain, J.-M. Emergence of VIM-2 and IMP-15 Carbapenemases and Inactivation of oprD Gene in Carbapenem-Resistant Pseudomonas aeruginosa Clinical Isolates from Lebanon. Antimicrob. Agents Chemother. 2014, 58, 4966–4970.
- 55. van Duin, D.; Bonomo, R.A. Ceftazidime/avibactam and ceftolozane/tazobactam: Second-generation b-lactam/b-lactamase inhibitor combinations. Clin. Infect. Dis. 2016, 63, 234–241.
- 56. Araj, G.F.; Berjawi, D.M.; Musharrafieh, U.; El Beayni, N.K. Activity of ceftolozane/tazobactam against commonly encountered antimicrobial resistant Gram-negative bacteria in Lebanon. J. Infect. Dev. Ctries. 2020, 14, 559–564.