Microalgae-Based Biorefineries: Challenges and Future Trends

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Microalgae-based biorefineries allow the simultaneous production of microalgae biomass enriched in a particular macromolecule and high-added and low-value products if a proper selection of the microalgae species and the cultivation conditions are adequate for the purpose.

third-generation biorefineries multi-product biorefineries

circular economy

1. Introduction

Microalgae have been recognized as a very important group of photosynthetic microorganisms for many of their attributes and the roles that they play in nature. Their importance as one of the major groups for CO₂ sequestration and for releasing oxygen either in marine environments $\begin{bmatrix} 1 \\ 2 \end{bmatrix}$, in freshwater bodies $\begin{bmatrix} 2 \\ 2 \end{bmatrix}$, and even in arid and semiarid environments [2], has attracted the attention of several research groups for decades. On the other hand, microalgae have been identified as a very versatile and diverse group of organisms with a very high potential for application in biotechnology [1][4] due to their high capacity to produce and accumulate various macromolecules (proteins, carbohydrates, and lipids) with several uses in the food and feed industry, and more recently, as a source of biofuels [5]6]. Furthermore, microalgae are a very important source of several high-added value products such as pigments, PUFAs (poly-unsaturated fatty acids), peptides, exo-polysaccharides, among others, with a clear benefit to human health and nutrition [7][8] (Figure 1).

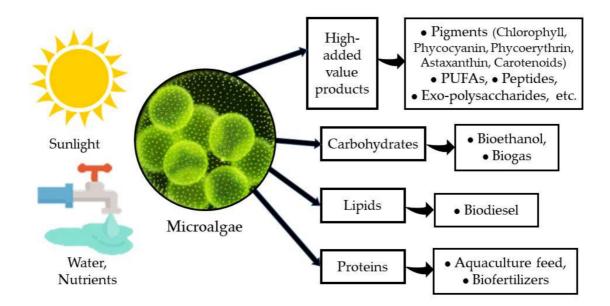


Figure 1. Microalgae and their products.

Despite all of these attributes that indicate that microalgae are a very useful feedstock for biofuels ^[9], food and feed supplements ^[10], and high-added value products ^[11], it has been recognized that several constraints still have to be overcome to make their large-scale production economically feasible, such as low productivity, risk of contamination in outdoor cultures, and high harvesting and downstream processing costs ^[12]. Thus, one of the strategies to overcome such drawbacks is their production following a "Biorefinery" concept.

A widely used Biorefinery definition is the one launched by the IEA (International Energy Agency) Bioenergy Task 42: "Biorefinery is the sustainable processing of biomass into a spectrum of marketable products (food, feed, materials, chemicals) and energy (fuels, power, heat)" ^[13], and it has been applied in recent decades to the processing of biomass for production of biofuels and chemical products with the expectation of reduced environmental impact ^{[14][15]}. It has drawn the attention of several research groups following different approaches. With the purpose of optimizing the environmental and economic performance of a biorefinery, a useful tool that combines the economic value and the environmental impact (EVEI) has been developed ^[16].

On the other hand, microalgae have been used for treating wastewater since many decades ago, when the pioneer group led by Prof. William Oswald initiated their use on a large scale in California ^[17]. Following this approach, research has been oriented toward the design and establishment of integrated systems in which microalgae were used for treating wastewater and producing valuable biomass ^{[18][19][20][21]}. Currently, microalgae-based biorefineries have been investigated by several research groups ^{[22][23][24]}, and in some cases, such a type of biorefinery has integrated the treatment of wastewater and used some of the treated effluents or digestates as a source of nutrients ^{[25][26][27]}.

It is important to mention that both the integrated systems for treating wastewater with microalgae and the microalgae-based biorefineries treating and using wastewater follow the circular economy concept (Figure 2). In

fact, the term circular economy has been described as the efficient utilization of biomass, including waste, for the sustainable production of high-added and low-value products (e.g., food, feed, biomaterials, biofuels, biofertilizers, and biostimulants) with the benefit of lowering the carbon footprint and the valorization of waste materials ^[28].

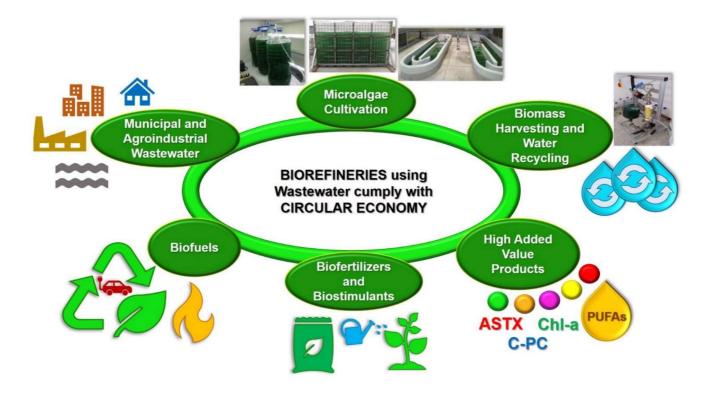


Figure 2. Microalgae-based biorefineries using and treating wastewater, follow the Circular Economy concept.

2. Photoautotrophic Production of High-Added Value Products

Despite the large potential of several microalgae for producing high-added value products, only a few species are produced at a large scale under photoautotrophic conditions: *Arthrospira* sp., *Chlorella* sp., *Dunaliella* sp., *Haematococcus* sp., and *Nannochloropsis* sp. ^{[29][30][31][32]}

The production of food supplements or specific bioactive compounds gained interest for large-scale commercialization ^[34], mainly due to the demand in high-income countries. The genus *Arthrospira* sp. is the one which is being produced in the largest quantity (tons/year) and in many countries due to its high content of protein and other well-known high added-value compounds ^[35]. Earthrise Nutraceuticals[®] started production of this valuable cyanobacteria in California and Hawaii more than 40 years ago. *Arthrospira* is currently one of the most widely commercially cultivated microalgae in open raceway ponds worldwide, with China and the United States leading the global production, followed by Japan, Taiwan, Thailand, India, and Australia. According to a recent publication from Meticulous Research, the *Arthrospira* market is expected to record a compounded annual growth rate (CAGR) of 13.2% from 2021 to 2028 to reach USD 968.6 million dollars by 2028. In terms of volume,

the *Arthrospira* market is expected to register a CAGR of 18.1% from 2021 to 2028 to reach 98,768.5 tons by 2028 [36][37].

The second genus produced ina very large quantity is *Chlorella*, also in various countries. *Dunaliella* sp. is being produced either as whole dried biomass or for the direct production of β -carotene. *Haematococcus* sp. has also been cultivated outdoors for the production of asthaxanthin, a very valuable pigment ^[35].

On the other hand, the production of pigments is still an attractive field of research, and there are several recent interesting reports dealing with different operational strategies (batch and semi-continuous cultures) and showing variable yield and productivity, depending on each strategy used and the strain of microalgae utilized. It is important to note that new information regarding new strains or new types of processes might lead to new commercial large-scale production worldwide.

3. Mixotrophic Production of High-Added Value and Low-Value Products Using Wastewater

Following the concept of circular economy and sustainable development, microalgae-based biorefineries treating and using wastewater have also been an important field of research in recent decades ^[5]. The initial trend was to produce biomass for biofuels production ^{[25][26][27][38]}, and more recently, the trend is to produce biomass for biofuels and high-added value products ^{[39][40][41]}. Within this context, *Arthrospira platensis*, *Haematococcus pluvialis*, *Chlorella sorokiniana*, *Dunaliella* sp., and *Nannochloropsis limnética*, *Nannochloropsis gaditana*, among others, have been cultivated in municipal wastewater ^{[42][43][44][45]}, industrial wastewater ^{[46][47][48]}, and agroindustrial wastewater ^{[49][50][51]} for the production of some of the most demanded pigments (astaxanthin and β-carotene). There have been estimations that by using this type of mixotrophic culture, the cost of biomass could be less than 5 EUR/kg of biomass ^{[52][53]}.

4. Production of Microalgae Biomass Enriched in Carbohydrates

The photosynthetic metabolism of microalgae, as well as the metabolism of carbohydrates and lipids, are similar to those of terrestrial plants. However, the photosynthetic efficiency of microalgae is in the range of 6–10%, exceeding the 1–2% that of terrestrial plants ^[54]. Some authors mention that in most microalgae, starch accumulates as a primary source of energy and carbon reserve, while lipids serve as secondary storage ^[55]. Carbohydrate productivity in microalgae is usually higher than the lipid one, as the accumulation of lipids requires intense stress, whereas carbohydrate production is "easily" achieved by photosynthesis through the Calvin cycle and is markedly increased by nitrogen deficiency ^[56].

The contents of lipids, carbohydrates, and proteins depend on the type of microalgae and the environmental and nutritional conditions applied to the culture ^{[57][58][59]}. The ability of microalgae to produce energy-rich compounds,

such as lipids and carbohydrates, has led them to be considered one of the most important raw materials used in the production of third-generation biofuels ^[57].

5. Polysaccharides from Cyanobacteria for Developing New Bioactive Products

Many cyanobacteria are characterized by the presence, outside their outer cell membrane, of additional external structures of a polysaccharidic nature classified, according to their characteristics, as sheaths, capsules, and slimes. The sheath is a well-defined electron-dense thin layer that surrounds single cells, single filaments, or cell groups, reflecting their shape. The capsule is a gelatinous layer that is intimately associated with the cell surface and capable of absorbing large quantities of water. The capsule is characterized by sharp outlines and is structurally coherent to exclude particles. The slime is an amorphous, mucilaginous shroud, only loosely associated with the cell surface and capable of enclosing a large number of cells or filaments that may or may not be also surrounded by capsules. In the three cases, most exopolysaccharide (EPS)-producing cyanobacteria release into the culture medium during the growth or at the end of the linear phase of the growth, and large amounts of water-soluble polymeric material that was named released polysaccharides (RPS) ^[60]. Most of these polymers are characterized by an anionic nature, owing to the presence of uronic acids and/or other charged groups such as the sulphate or ketal-linked pyruvate groups ^[61].

Owing to the peculiar features of the polysaccharides produced by these microorganisms, EPS-producing cyanobacteria have been considered a promising resource of biopolymers of applied interest, and their exploitation to produce macromolecules suitable for specific industrial applications has been proposed in many studies. The specific features that have been considered of particular interest for the exploitation of cyanobacterial EPSs for developing new industrial products for biomedical applications are the following ^[61]:

- (i) Most of the polymers so far studied (up to now close to 200) are characterized by the presence in the macromolecule of a large number of different types of monosaccharides (in about 75% of the cases ranging from 6 up to 15), a feature rarely found in the EPS of other microorganisms. This feature makes it possible to have polymers with similar composition but organized in various structures and conformational forms, enlarging the chances of finding new products with useful properties;
- (ii) In most of the polymers are present uronic acids and sulphate groups, which are rare in other bacterial EPS and confer an anionic charge to the polymers. In addition to that, the presence of sulphate groups has been considered relevant for conferring antiviral properties to the polymers ^[62];
- (iii)The presence of peptides and deoxysugars, which confer hydrophobicity of some areas of the polymers;
- (iv)The frequent presence of uncommon sugars, such as acetylated or amine sugars, which are considered possibly involved in polymers' biological activity.

Owing to these specific features, the industrial interest in EPSs of cyanobacterial origin has been rising, and a number of studies showed that some of the polymers so far characterized possess interesting biological activities.

A part of the international research has focused on the therapeutical applications of cyanobacterial polysaccharides, spanning a wide range of biological activities, namely antiviral, antibacterial, antioxidant, immunostimulatory, anti-inflammatory, antitumor, and also wound-healing (stimulating collagen synthesis) activities. A brief summary of these reported activities is given here, focusing on the literature specifically related to cyanobacterial EPS.

It was shown that the presence of sulphate groups confers antiviral properties to the sulfated exopolysaccharide (TK V3) produced by *Arthrospira platensis*, which was found to inhibit orthopoxvirus and other enveloped viruses ^[63]. Antiviral properties have also been shown for Nostoflan, an EPS produced by the terrestrial cyanobacterium *Nostoc flagelliforme* ^[64], as well as for the EPSs produced by other cyanobacteria ^[65].

Antibacterial activity is much more controversial, as studies conducted on *A. platensis* reported a different activity depending on the extracting solvent: ethanolic EPS extract showed no significant effect, while methanolic extracts showed bacteriostatic effects ^[66].

Regarding the use of EPS as antioxidants, the just mentioned *A. platensis* EPS ethanolic extracts showed a low antioxidant capacity, while methanolic extracts resulted in possessing a significantly higher antioxidant capacity ^[66]. It has to be stressed that most of the research about the antioxidant activity of EPS has been conducted on microalgal polysaccharides ^[67]. However, cyanobacterial EPS presents very interesting chemical properties which are generally considered relevant for radical scavenging, as the negative charges given and the complex tridimensional structure ^[68], this specific activity has not been well characterized yet.

Very recently, the highly sulfated (\approx 13% w/w) EPS produced by *Phormidium* sp. ETS05 showed to exert antiinflammatory and pro-resolution activities in chemical and injury-induced zebrafish inflammation models, downregulating NF- κ B signaling and reducing neutrophil recruitment, thus accelerating the clearance of these cells to recuperate tissue homeostasis ^[69]. The immunomodulation effect of sulphated EPSs produced by *Cyanothece* sp. PE 14 ^[70] or by *Cyanobacterium aponinum* ^[71] was also demonstrated in animal and human cell lines. *N. commune* extract resulted in being effective in downregulating IL-6 ^[72] and is, therefore, potentially useful for anti-allergic and wound-healing therapeutic purposes ^[73].

More recently, a number of studies showed interesting antitumor activities of the EPSs of cyanobacterial origin. Li et al. ^[74] reported the antitumor activity of the EPS produced by *Nostoc sphaeroides*. Ou et al. ^[75] showed that the EPS produced by the cyanobacterium *Aphanothece halophytica* (EPSAH) was capable of inducing apoptosis in HeLa cells. More recently, Flores et al. ^[76] showed that the EPS produced by a *Synechocystis* $\Delta sigF$ mutant is capable of decreasing the viability of melanoma, thyroid, and ovary carcinoma cells by inducing high levels of apoptosis.

The results reported above point out the potential of some of the EPSs produced by cyanobacteria for developing new products for biomedical application, but at the same time show the need for further studies for unveiling the molecular mechanisms of action of these macromolecules as well as their possible negative effects on healthy animal or human cells.

This brief overview of the research in the field of cyanobacterial EPS bioactivity puts into light the numerous potential applications of these polymers. However, their use in animal and clinical trials are yet to be explored as the high molecular weight of these molecules makes them complex to handle, which is also related to their rheological behavior ^[77].

6. Microalgae-Bioactive Compounds, A Natural Source of Potential Therapeutical and Health Promoting Agents: Insights from Innovative In Vivo Functional Studies

Microalgae are light energy-based biofactories that synthesize high-value bioactive compounds, being the more representative examples with health relevance: proteins, pigments, polyphenols, antioxidants, polyunsaturated fatty acids, vitamins, minerals, sterols, and polysaccharides. These bioactive compounds have multiple health properties as antiviral, anticancer, antioxidant, antidiabetic, antibacterial, antifungal, anti-inflammatory, neuro, cardiorespiratory and hepatoprotective, among others ^{[7][10][78][79]}. Within this frame, herein, recent functional studies in animal of microalgae-bioactive compounds with their remarkable health-promoting biological activities are summarized. These innovative microalgae bioactivities revisited here support the relevance that algal biomass has gained and are nowadays considered a valuable source of health-promoting agents with potential nutritional, biomedical, and therapeutical applications.

Furthermore, microalgae-bioactive compounds are potential health-promoting agents for the prevention and therapeutic application of relevant infectious, chronic, and degenerative diseases such as viral, fungal, bacterial infections, diabetes, metabolic, cardiorespiratory, cancer, inflammatory, neurodegenerative, among other pathologies.

Algal Health-Promoting Agents

Microalgae-bioactive compounds have shown very important therapeutical bioactivities for the prevention or treatment of diseases as proven by functional studies in animals:

• Potential skin healing/antifibrotic agents:

A study performed with a rat model proved that when *Arthrospira platensis* was applied over skin wounds of Wistar rats, this microalga helped in the healing process. In addition to skin reparation, molecular expression patterns as indicators of the upregulation of angiogenic genes and downregulation of fibrotic genes were also observed ^[80].

Another study conducted with a rat model showed increased skin wound healing due to a diet with docosahexaenoic acid (DHA) from *Schizochytrium* sp., demonstrating its immunostimulant effect ^[81].

• Promising liver health-promoters:

The administration of *Dunaliella salina* showed to revert liver disfunction, decreasing inflammation and oxidative stress by its anti-inflammatory and antioxidant properties, as it was observed in thioacetamide- (TAA-) induced hepatic encephalopathy rat model ^[82].

• Natural immunostimulators:

Exopolysaccharides from *Porphyridium cruentum* (purpureum) administrated to shrimps *Litopenaeus vannamei* resulted in functioning as immunostimulators elevating the immune response of shrimps and protecting them from Vibrio infection ^[83]. Another functional study performed with Senegalese sole larvae treated with microalgae showed that *Nannochloropsis gaditana* and *Phaeodactylum tricornutum* could induce immune response ^[84]. In an intestinal inflammation murine model, it has been proven that *Arthrospira platensis* induced immunomodulatory effects ^[85]. A study realized with a murine model indicated that fucoxanthin could be used as a natural health-promoting agent as it showed an anti-inflammatory effect, facilitating the recovery of dextran sulfate sodium (DSS)-induced colitis mice ^[86].

• Non-toxic anticancer agents:

An engineered *Chlorella vulgaris* was injected into a tumor-bearing mouse model showing to have antitumoral and anti-metastasis effects ^[87]. An in vivo rodent model showed that astaxanthin is safe and can be used as a nutraceutical and as an anticancer agent as it inhibits lung metastasis in mice ^[88].

• Improvement of nutrients deficiency:

A functional study with a murine model indicated that *Nannochloropsis oceanica* could be considered an algal nutraceutical as its consumption increased hemoglobin values in anemic mice ^[89].

• Prevention of type 2 Diabetes Mellitus development:

Microalgae antidiabetic agents are gaining relevance in the prevention and treatment of diabetes mellitus type 2 ^[90], as this disorder is considered to be the ninth worldwide leading cause of mortality, as 1 million deaths per year have been attributed to this metabolic disorder ^[91]. In a study performed with streptozotocin-induced diabetic rats, it was found that polysaccharides from *Porphyridium cruentum* had antihyperglycemic activity representing a potential natural antidiabetic agent ^[92]. In a murine model, it was demonstrated that consumption of n-3 fatty acids from microalgae could function as natural antidiabetic agents as an increment in the antioxidant capacity in adipose tissue of diabetic mice was observed ^[93].

• Natural antioxidant agents:

These health-promoting agents could decrease tissue oxidation and cell damage, as demonstrated by a functional study performed with a rat model, which showed that the consumption of *Chlorella vulgaris* acted as an antioxidant agent that decreased skeletal muscle oxidative stress preventing cell damage ^[94]. A mice model indicated that carotenoids from *Scenedesmus obliquus* demonstrated that these pigments could be used as antioxidant agents, considering that the endogenous antioxidant defense system of mice was increased ^[95].

Herein, based on functional bioactivities, it has been illustrated how natural bioactive compounds considered highadded value products derived from microalgal biomass are gaining more impact as natural therapeutic agents useful for the prevention or treatment of diseases. Furthermore, their relevance as a valuable source of innovative natural agents with potential biomedical and therapeutical applications is highlighted.

References

- 1. Hamed, I. The evolution and versatility of microalgal biotechnology: A review. Compr. Rev. Food. Sci. Saf. 2016, 15, 1104–1123.
- 2. Abdel-Raouf, N.; Al-Homaidan, A.A.; Ibraheem, I.B.M. Microalgae and wastewater treatment. Saudi J. Biol. Sci. 2012, 19, 257–275.
- Rossi, F.; Olguín, E.J.; Diels, L.; De Philippis, R. Microbial fixation of CO2 in water bodies and in drylands to combat climate change, soil loss and desertification. New Biotechnol. 2015, 32, 109– 120.
- Ho, S.H.; Kondo, A.; Hasunuma, T.; Chang, J.S. Engineering strategies for improving the CO fixation and carbohydrate productivity of Scenedesmus obliquus CNW-N used for bioethanol fermentation. Bioresour. Technol. 2013, 143, 163–171.
- 5. Alam, M.A.; Wang, Z.M. Microalgae Biotechnology for Development of Biofuel and Wastewater Treatment; Springer Nature: Singapore, 2019; p. 655.
- Patel, A.K.; Choi, Y.Y.; Sim, S.J. Emerging prospects of mixotrophic microalgae: Way forward to sustainable bioprocess for environmental remediation and cost-effective biofuels. Bioresour. Technol. 2020, 300, 122741.
- 7. Barkia, I.; Saari, N.; Manning, S.R. Microalgae for high-value products towards human health and nutrition. Mar. Drugs 2019, 17, 304.
- 8. Alam, M.A.; Xu, J.L.; Wang, Z. Microalgae Biotechnology for Food, Health and High Value Products; Springer Nature: Singapore, 2020; p. 483.

- Borowitzka, M.A. Energy from microalgae: A short history. In Algae for Biofuels and Energy. Developments in Applied Phycology; Borowitzka, M., Moheimani, N., Eds.; Springer Science+Business Media: Dordrecht, The Netherlands, 2013; Volume 5, pp. 1–15.
- Dolganyuk, V.; Belova, D.; Babich, O.; Prosekov, A.; Ivanova, S.; Katserov, D.; Patyukov, N.; Sukhikh, S. Microalgae: A promising source of valuable bioproducts. Biomolecules 2020, 10, 1153.
- 11. Dourou, M.; Dritsas, P.; Baeshen, M.N.; Elazzazy, A.; Al-Farga, A.; Aggelis, G. High-added value products from microalgae and prospects of aquaculture wastewaters as microalgae growth media. FEMS Microbiol. Lett. 2020, 367, fnaa081.
- 12. Chen, Y.; Xu, C.; Vaidyanathan, S. Microalgae: A robust "green bio-bridge" between energy and environment. Crit. Rev. Biotechnol. 2018, 38, 351–368.
- International Energy Agency (IEA). IEA BIOENERGY Task42 BIOREFINING. In Sustainable and Synergetic Processing of Biomass into Marketable Food & Feed Ingredients, Chemicals, Materials and Energy; van Ree, R., van Zeeland, A., Eds.; IEA: Wageningen, The Netherlands, 2014; Available online: http://www.ieabioenergy.com/wp-content/uploads/2014/09/IEA-Bioenergy-Task42-Biorefining-Brochure-SEP2014_LR.pdf (accessed on 3 May 2022).
- 14. Kamm, B.; Kamm, M. Principles of biorefineries. Appl. Microbiol. Biotechnol. 2005, 64, 137–145.
- Sadhukhan, J.; Martinez-Hernandez, E.; Amezcua-Allieri, M.A.; Aburto, J.; Honorato, J.A. Economic and environmental impact evaluation of various biomass feedstock for bioethanol production and correlations to lignocellulosic composition. Bioresour. Technol. Rep. 2019, 7, 100230.
- 16. Martinez-Hernandez, E.; Campbell, G.; Sadhukhan, J. Economic value and environmental impact (EVEI) analysis of biorefinery systems. Chem. Eng. Res. Des. 2013, 91, 1418–1426.
- Posadas, E.; Alcántara, C.; García-Encina, P.A.; Gouveia, L.; Guieysse, B.; Norvill, Z.; Acién, F.G.; Markou, G.; Congestri, R.; Koreiviene, J.; et al. 3-Microalgae cultivation in wastewater. In Microalgae-Based Biofuels and Bioproducts. From Feedstock Cultivation to End-Products; Woodhead Publishing Series in Energy: Kidlington, UK, 2017; pp. 67–91.
- Olguín, E.J.; Hernández, B.; Araus, A.; Camacho, R.; González, R.; Ramírez, M.E.; Galicia, S.; Mercado, G. Simultaneous high-biomass protein-production and nutrient removal using Spirulina maxima in sea-water supplemented with anaerobic effluents. World J. Microbiol. Biotechnol. 1994, 10, 576–578.
- Olguín, E.J.; Galicia, S.; Mercado, G.; Pérez, T.J. Annual productivity of Spirulina (Arthrospira) and nutrient removal in a pig wastewater recycling process under tropical conditions. J. Appl. Phycol. 2003, 15, 249–257.

- Flesch, A.; Beer, T.; Campbell, P.K.; Batten, D.; Grant, T. Greenhouse gas balance and algaebased biodiesel. In Algae for Biofuels and Energy. Developments in Applied Phycology; Borowitzka, M., Moheimani, N., Eds.; Springer Science+Business Media: Dordrecht, The Netherlands, 2013; Volume 5, pp. 233–254.
- 21. Jebali, A.; Acién, F.G.; Barradas, E.R.; Olguín, E.J.; Sayadi, S.; Molina-Grima, E. Pilot-scale outdoor production of Scenedesmus sp. in raceways using flue gases and centrate from anaerobic digestion as the sole culture medium. Bioresour. Technol. 2018, 262, 1–8.
- 22. Li, S.H.; Zhao, S.; Yan, S.L.; Qiu, Y.T.; Song, C.F.; Li, Y.; Kitamura, Y. Food processing wastewater purification by microalgae cultivation associated with high value-added compounds production—A review. Chin. J. Chem. Eng. 2019, 27, 2845–2856.
- 23. Khoo, C.G.; Dasan, Y.K.; Lam, M.K.; Lee, K.T. Algae biorefinery: Review on a broad spectrum of downstream processes and products. Bioresour. Technol. 2019, 292, 121964.
- 24. Figueroa-Torres, G.M.; Pittman, J.K.; Theodoropoulos, C. Optimisation of microalgal cultivation via nutrient-enhanced strategies: The biorefinery paradigm. Biotechnol. Biofuels 2021, 14, 64.
- 25. Olguín, E.J. Dual purpose microalgae–bacteria-based systems that treat wastewater and produce biodiesel and chemical products within a biorefinery. Biotechnol. Adv. 2012, 30, 1031–1046.
- Olguín, E.J.; Dorantes, E.; Castillo, O.; Hernández-Landa, V.J. Anaerobic digestates from vinasse promote growth and lipid enrichment in Neochloris oleoabundans cultures. J. Appl. Phycol. 2015, 27, 1813–1822.
- 27. Olguín, E.J.; Castillo, O.S.; Mendoza, A.; Tapia, K.; González-Portela, R.E.; Hernández, V.J. Dual purpose system that treats anaerobic effluents from pig waste and produce Neochloris oleoabundans as lipid rich biomass. New Biotechnol. 2015, 32, 387–395.
- Leong, H.Y.; Chang, C.K.; Khoo, K.S.; Chew, K.W.; Chia, S.R.; Lim, J.W.; Chang, J.S.; Show, P.L. Waste biorefinery towards a sustainable circular bioeconomy: A solution to global issues. Biotechnol. Biofuels 2021, 14, 87.
- 29. Araújo, R.; Calderón, F.V.; López, J.S.; Azevedo, I.C.; Bruhn, A.; Fluch, S.; Tasende, M.G.; Ghaderiardakani, F.; Ilmjärv, T.; Laurans, M.; et al. Current status of the algae production industry in Europe: An emerging sector of the Blue Bioeconomy. Front. Mar. Sci. 2021, 7, 626389.
- Herrador, M.; The Microalgae/Biomass Industry in Japan. An Assessment of Cooperation and Business Potential with European Companies. EU-Japan Centre for Industrial Cooperation. Minerva EU-Japan Fellowship. 2016. Available online: https://www.eujapan.eu/publications/microalgae (accessed on 5 May 2022).
- Novoveská, L.; Ross, M.E.; Stanley, M.S.; Pradelles, R.; Wasiolek, V.; Sassi, J.-F. Microalgal carotenoids: A review of production, current markets, regulations, and future direction. Mar. Drugs 2019, 17, 640.

- 32. Meticulous Blog. TOP 10 COMPANIES IN SPIRULINA MARKET. Available online: https://meticulousblog.org/top-10-companies-in-spirulina-market/ (accessed on 20 June 2022).
- Meticulous Blog. TOP 10 COMPANIES IN THE CHLORELLA MARKET. Available online: https://meticulousblog.org/top-10-companies-in-the-chlorella-market/ (accessed on 20 June 2022).
- 34. Borowitzka, M.A. High-value products from microalgae-their development and commercialization. J. Appl. Phycol. 2013, 25, 743–756.
- 35. Belay, A. Biology and Industrial Production of Arthrospira (Spirulina). In Handbook of Microalgal Culture: Applied Phycology and Biotechnology, 2nd ed.; Richmond, A., Hu, Q., Eds.; John Wiley & Sons-Blackwell Publishing Ltd.: Hoboken, NJ, USA, 2013; pp. 339–358.
- 36. Nguyen, D.B.; Doan, T.T.Y.; Phi, T.C.M.; Ngo, T.A.; Vu, L.D.H.; Dang, D.K. Arthrospira production in Vietnam: Current status and prospects. Bioresour. Technol. Rep. 2021, 15, 100803.
- 37. Meticulous Research. Spirulina Market. Available online: https://www.meticulousresearch.com/product/spirulina-market-5070 (accessed on 19 June 2022).
- Montero, E.; Olguín, E.J.; De Philippis, R.; Reverchon, F. Mixotrophic cultivation of Chlorococcum sp. under non-controlled conditions using a digestate from pig manure within a biorefinery. J. Appl. Phycol. 2018, 30, 2847–2857.
- Javed, F.; Aslam, M.; Rashid, N.; Shamir, Z.; Khan, A.L.; Yasin, M.; Fazal, T.; Hafeez, A.; Rehman, F.; Rehman, M.S.U.; et al. Microalgae-based biofuels, resource recovery and wastewater treatment: A pathway towards sustainable biorefinery. Fuel 2019, 255, 115826.
- 40. Nagarajan, D.; Lee, D.-J.; Chen, C.Y.; Chang, J.-S. Resource recovery from wastewaters using microalgae-based approaches: A circular bioeconomy perspective. Bioresour. Technol. 2020, 302, 122817.
- López-Pacheco, I.Y.; Silva-Núñez, A.; García-Pérez, J.S.; Carrillo-Nieves, D.; Salinas-Salazar, C.; Castillo-Zacarías, C.; Afewerki, S.; Barceló, D.; Iqbal, H.N.M.; Parra-Saldívar, R. Phycoremediation of swine wastewater as a sustainable model based on circular economy. J. Environ. Manag. 2021, 278, 111534.
- Sukumaran, P.; Nulit, R.; Zulkifly, S.; Halimoon, N.; Omar, H.; Ismail, A. Potential of fresh POME as a growth medium in mass production of Arthrospira platensis. Int. J. Curr. Microbiol. App. Sci. 2014, 3, 235–250. Available online: https://www.ijcmas.com/vol-3-4/Puganeswary%20Sukumaran,%20et%20al.pdf (accessed on 9 May 2022).
- 43. Arone Soul Raj, G.P.; Elumalai, S.; Sangeetha, T.; Roop Singh, D. Botryococcus braunii as a phycoremediation tool for the domestic waste water recycling from Cooum River, Chennai, India. J. Bioremed. Biodeg. 2015, 6, 294.

- 44. Basily, H.S.; Nassar, M.M.; El Diwani, G.I.; El-Enin, S.A.A. Exploration of using the algal bioactive compounds for cosmeceuticals and pharmaceutical applications. Egypt. Pharmaceut. J. 2018, 17, 109–120.
- 45. De Francisci, D.; Su, Y.; Iital, A.; Angelidaki, I. Evaluation of microalgae production coupled with wastewater treatment. Environ. Technol. 2018, 39, 581–592.
- 46. Safafar, H.; Van Wagenen, J.; Moller, P.; Jacobsen, C. Carotenoids, phenolic compounds and tocopherols contribute to the antioxidative properties of some microalgae species grown on industrial wastewater. Mar. Drugs 2015, 13, 7339–7356.
- Ji, M.K.; Yun, H.S.; Hwang, B.S.; Kabra, A.N.; Jeon, B.H.; Choi, J. Mixotrophic cultivation of Nephroselmis sp. using industrial wastewater for enhanced microalgal biomass production. Ecol. Eng. 2016, 95, 527–533.
- 48. Pan, M.M.; Zhu, X.Y.; Pan, G.; Angelidak, I. Integrated valorization system for simultaneous high strength organic wastewater treatment and astaxanthin production from Haematococcus pluvialis. Bioresour. Technol. 2021, 326, 124761.
- Han, T.; Lu, H.; Zhao, Y.; Xu, H.; Zhang, Y.; Li, B. Two-step strategy for obtaining Dunaliella sp. biomass and β-carotene from anaerobically digested poultry litter wastewater. Int. Biodeterior. Biodegrad. 2019, 143, 104714.
- 50. Arashiro, L.T.; Boto-Ordoñez, M.; Van Hulle, S.W.H.; Ferrer, I.; Garfi, M.; Rousseau, D.P.L. Natural pigments from microalgae grown in industrial wastewater. Bioresour. Technol. 2020, 303, 122894.
- 51. Jiménez-Veuthey, M.; Morillas-España, A.; Sánchez-Zurano, A.; Navarro-López, E.; Acién, G.; López-Segura, J.G.; Lafarga, T. Production of the marine microalga Nannochloropsis gaditana in pilot-scale thin-layer cascade photobioreactors using fresh pig slurry diluted with sea water. J. Water Process Eng. 2022, 48, 102869.
- 52. Ferreira, A.; Melkonyan, L.; Carapinha, S.; Ribeiro, B.; Figueiredo, D.; Avetisova, G.; Gouveia, L. Bioestimulant and biopesticide potential of microalgae growing in piggery wastewater. Environ. Adv. 2021, 4, 100062.
- 53. Kalra, R.; Gaur, S.; Goel, M. Microalgae bioremediation: A perspective towards wastewater treatment along with industrial carotenoids production. J. Water Process Eng. 2021, 40, 101794.
- 54. González-Fernández, C.; Ballesteros, M. Linking microalgae and cyanobacteria culture conditions and key-enzymes for carbohydrate accumulation. Biotechnol. Adv. 2012, 30, 1655–1661.
- 55. Cheng, D.; Li, D.; Yuan, Y.; Zhou, L.; Li, X.; Wu, T.; Wang, L.; Zhao, Q.; Wei, W.; Sun, Y. Improving carbohydrate and starch accumulation in Chlorella sp. AE10 by a novel two-stage process with cell dilution. Biotechnol. Biofuels 2017, 10, 75.

- 56. Chen, C.Y.; Zhao, X.Q.; Yen, H.W.; Ho, S.H.; Cheng, C.L.; Lee, D.J.; Bai, F.W.; Chang, J.S. Microalgae-based carbohydrates for biofuel production. Biochem. Eng. J. 2013, 78, 110.
- 57. Bastos, R.G. Biofuels from microalgae: Bioethanol. In Energy from Microalgae. Green Energy and Technology; Jacob-Lopes, E., Queiroz Zepka, L., Queiroz, M., Eds.; Springer: Cham, Switzerland, 2018; pp. 229–246.
- Ho, S.H.; Chen, C.Y.; Chang, J.S. Effect of light intensity and nitrogen starvation on CO2 fixation and lipid/carbohydrate production of an indigenous microalga Scenedesmus obliquus CNW-N. Bioresour. Technol. 2012, 113, 244–252.
- 59. da Silva-Braga, V.; da Silveira-Mastrantonio, D.J.; Vieira-Costa, J.A.; Greque de Morais, M. Cultivation strategy to stimulate high carbohydrate content in Spirulina biomass. Bioresour. Technol. 2018, 269, 221–226.
- De Philippis, R.; Vincenzini, M. Exocellular polysaccharides from cyanobacteria and their possible applications. FEMS Microbiol. Rev. 1998, 22, 151–175. Available online: https://academic.oup.com/femsre/article/22/3/151/643945 (accessed on 20 June 2022).
- Pereira, S.; Zille, A.; Micheletti, E.; Moradas-Ferreira, P.; De Philippis, R.; Tamagnini, P. Complexity of cyanobacterial exopolysaccharides: Composition, structures, inducing factors and putative genes involved in their biosynthesis and assembly. FEMS Microbiol. Rev. 2009, 33, 917– 941.
- 62. Ghosh, T.; Chattopadhyay, K.; Marschall, M.; Karmakar, P.; Mandal, P.; Ray, B. Focus on antivirally active sulfated polysaccharides: From structure–activity analysis to clinical evaluation. Glycobiology 2009, 19, 2–15.
- Radonic, A.; Thulke, S.; Achenbach, J.; Kurth, A.; Vreemann, A.; König, T.; Walter, C.; Possinger, K.; Nitsche, A. Anionic polysaccharides from phototrophic microorganisms exhibit antiviral activities to vaccinia virus. J. Antivir. Antiretrovir. 2010, 2, 27–35.
- Kanekiyo, K.; Lee, J.B.; Hayashi, K.; Takenaka, H.; Hayakawa, Y.; Endo, S.; Hayashi, T. Isolation of an antiviral polysaccharide, Nostoflan, from a terrestrial cyanobacterium, Nostoc flagelliforme. J. Nat. Prod. 2005, 68, 1037–1041.
- 65. Colica, G.; De Philippis, R. Exopolysaccharides from cyanobacteria and their possible industrial applications. In Cyanobacteria: An Economic Perspective; Sharma, N.K., Rai, A.K., Stal, L.J., Eds.; Wiley Blackwell: West Sussex, UK, 2014; pp. 197–207.
- 66. Challouf, R.; Trabelsi, L.; Dhieb, R.B.; El Abed, O.; Yahia, A.; Ghozzi, K.; Ammar, J.B.; Omran, H.; Ouada, H.B. Evaluation of Cytotoxicity and Biological Activities in Extracellular Polysaccharides Released by Cyanobacterium Arthrospira platensis. Braz. Arch. Biol. Technol. 2011, 54, 831–838.
- 67. de Jesus Raposo, M.F.; de Morais, A.M.B.; de Morais, R.M.S.C. Marine Polysaccharides from Algae with Potential Biomedical Applications. Mar. Drugs 2015, 13, 2967–3028.

- 68. Sun, L.; Wang, C.; Shi, Q.; Ma, C. Preparation of different molecular weight polysaccharides from Porphyridium cruentum and their antioxidant activities. Int. J. Biol. Macromol. 2009, 45, 42–47.
- 69. Zampieri, R.M.; Adessi, A.; Caldara, F.; Codato, A.; Furlan, M.; Rampazzo, C.; De Philippis, R.; La Rocca, N.; Dalla Valle, L. Anti-inflammatory activity of exopolysaccharides from Phormidium sp. ETS05, the most abundant cyanobacterium of the therapeutic Euganean thermal muds, using the zebrafish model. Biomolecules 2020, 10, 582.
- Campa-Córdova, A.I.; Hernández-Saavedra, N.Y.; De Philippis, R.; Ascencio, F. Generation of superoxide anion and SOD activity in haemocytes and muscle of American white shrimp (Litopenaeus vannamei) as a response to β-glucan and sulphated polysaccharide. Fish Shellfish Immunol. 2002, 12, 353–366.
- 71. Gudmundsdottir, A.B.; Omarsdottir, S.; Brynjolfsdottir, A.; Paulsen, B.S.; Olafsdottir, E.S.; Freysdottir, J. Exopolysaccharides from Cyanobacterium aponinum from the blue lagoon in Iceland increase IL-10 secretion by human dendritic cells and their ability to reduce the IL 17+RORyt+/IL-10+ FoxP3+ ratio in CD4+ T cells. Immunol. Lett. 2015, 163, 157–162.
- Liao, H.F.; Wu, T.J.; Tai, J.L.; Chi, M.C.; Lin, L.L. Immunomodulatory Potential of the Polysaccharide-Rich Extract from Edible Cyanobacterium Nostoc commune. Med. Sci. 2015, 3, 112–123.
- 73. Tseng, C.C.; Yeh, H.Y.; Liao, Z.H.; Hung, S.W.; Chen, B.; Lee, P.T.; Nan, F.H.; Shih, W.L.; Chang, C.C.; Lee, M.C. An in vitro study shows the potential of Nostoc commune (Cyanobacteria) polysaccharides extract for wound-healing and anti-allergic use in the cosmetics industry. J. Funct. Foods 2021, 87, 104754.
- 74. Li, H.F.; Su, L.N.; Chen, S.; Zhao, L.B.; Wang, H.Y.; Ding, F.; Chen, H.; Shi, R.; Wang, Y.; Huang, Z. Physicochemical characterization and functional analysis of the polysaccharide from the edible microalga Nostoc sphaeroides. Molecules 2018, 23, 508.
- 75. Ou, Y.; Xu, S.; Zhu, D.; Yang, X. Molecular mechanisms of exopolysaccharide from Aphanothece halophytica (EPSAH) induced apoptosis in HeLa cells. PLoS ONE 2014, 9, e87223.
- 76. Flores, C.; Lima, R.T.; Adessi, A.; Sousa, A.; Pereira, S.B.; Granja, P.L.; De Philippis, R.; Soares, P.; Tamagnini, P. Characterization and antitumor activity of the extracellular carbohydrate polymer from the cyanobacterium Synechosystis ΔsigF mutant. Int. J. Biol. Macromol. 2019, 136, 1219–1227.
- 77. de Jesus Raposo, M.F.; de Morais, A.M.M.B.; de Morais, R.M.S.C. Bioactivity and Applications of Polysaccharides from Marine Microalgae. In Polysaccharides; Ramawat, K., Mérillon, J.M., Eds.; Springer: Cham, Switzerland, 2014; pp. 1–38.
- 78. Alsenani, F.; Tupally, K.R.; Chua, E.T.; Eltanahy, E.; Alsufyani, H.; Parekh, H.S.; Schenk, P.M. Evaluation of microalgae and cyanobacteria as potential sources of antimicrobial compounds.

Saudi Pharm. J. 2020, 28, 1834–1841.

- Stirk, W.A.; Bálint, P.; Vambe, M.; Lovász, C.; Molnár, Z.; van Staden, J.; Ördög, V. Effect of cell disruption methods on the extraction of bioactive metabolites from microalgal biomass. J. Biotechnol. 2020, 307, 35–43.
- 80. Elbialy, Z.I.; Assar, D.H.; Abdelnaby, A.; Asa, S.A.; Abdelhiee, E.Y.; Ibrahim, S.S.; Abdel-Daim, M.M.; Almeer, R.; Atiba, A. Healing potential of Spirulina platensis for skin wounds by modulating bFGF, VEGF, TGF-ß1 and α-SMA genes expression targeting angiogenesis and scar tissue formation in the rat model. Biomed. Pharmacother. 2021, 137, 111349.
- 81. Komprda, T.; Sladek, Z.; Sevcikova, Z.; Svehlova, V.; Wijacki, J.; Guran, R.; Do, T.; Lackova, Z.; Polanska, H.; Vrlikova, L.; et al. Comparison of dietary oils with different polyunsaturated fatty acid n-3 and n-6 content in the rat model of cutaneous wound healing. Int. J. Mol. Sci. 2020, 21, 7911.
- 82. El-Baz, F.K.; Elgohary, R.; Salama, A. Amelioration of hepatic encephalopathy using Dunaliella salina microalgae in rats: Modulation of hyperammonemia/TLR4. BioMed Res. Int. 2021, 2021, 8843218.
- Risjani, Y.; Mutmainnah, N.; Manurung, P.; Wulan, S.N. Yunianta Exopolysaccharide from Porphyridium cruentum (purpureum) is not toxic and stimulates immune response against vibriosis: The assessment using zebrafish and white shrimp Litopenaeus vannamei. Mar. Drugs 2021, 19, 133.
- Carballo, C.; Mateus, A.P.; Maya, C.; Mantecón, L.; Power, D.M.; Manchado, M. Microalgal extracts induce larval programming and modify growth and the immune response to bioactive treatments and LCDV in Senegalese sole post-larvae. Fish Shellfish Immunol. 2020, 106, 263– 272.
- 85. de Oliveira-Garcia, F.A.; Sales-Campos, H.; Yuen, V.G.; Machado, J.R.; de Barros-Viana, G.S.; Freire-Oliveira, C.J.; McNeill, J.H. Arthrospira (Spirulina) platensis attenuates dextran sulfate sodium-induced colitis in mice by suppressing key pro-inflammatory cytokines. Korean J. Gastroenterol. 2020, 76, 150–158.
- Yang, Y.P.; Tong, Q.Y.; Zheng, S.H.; Zhou, M.D.; Zeng, Y.M.; Zhou, T.T. Anti-inflammatory effect of fucoxanthin on dextran sulfate sodium-induced colitis in mice. Nat. Prod. Res. 2020, 34, 1791– 1795.
- 87. Zhong, D.; Li, W.L.; Hua, S.Y.; Qi, Y.C.; Xie, T.T.; Qiao, Y.; Zhou, M. Calcium phosphate engineered photosynthetic microalgae to combat hypoxic-tumor by in-situ modulating hypoxia and cascade radio-phototherapy. Theranostics 2021, 11, 3580–3594.
- Tseng, C.C.; Lin, Y.J.; Liu, W.; Lin, H.Y.; Chou, H.Y.; Thia, C.; Wu, J.H.; Chang, J.S.; Wen, Z.H.; Chang, J.J.; et al. Metabolic engineering probiotic yeast produces 3S, 3'S-astaxanthin to inhibit B16F10 metastasis. Food Chem. Toxicol. 2020, 135, 110993.

- 89. Bhatnagar, R.S.; Miller, D.D.; Padilla-Zakour, O.I.; Lei, X.G. Supplemental microalgal iron helps replete blood hemoglobin in moderately anemic mice fed a rice-based diet. Nutrients 2020, 12, 2239.
- Popović-Djordjević, J.B.; Katanić-Stanković, J.S.; Mihailović, V.; Pereira, A.G.; Garcia-Oliveira, P.; Prieto, M.A.; Simal-Gandara, J. Algae as a source of bioactive compounds to prevent the development of type 2 Diabetes Mellitus. Curr. Med. Chem. 2021, 28, 4592–4615.
- Khan, M.A.B.; Hashim, M.J.; King, J.K.; Govender, R.D.; Mustafa, H.; Al-Kaabi, J. Epidemiology of type 2 Diabetes—Global burden of disease and forecasted trends. J. Epidemiol. Glob. Health 2020, 10, 107–111.
- 92. Setyaningsih, I.; Prasetyo, H.; Agungpriyono, D.R.; Tarman, K. Antihyperglycemic activity of Porphyridium cruentum biomass and extra-cellular polysaccharide in streptozotocin-induced diabetic rats. Int. J. Biol. Macromol. 2020, 156, 1381–1386.
- Gutiérrez-Pliego, L.E.; Martínez-Carrillo, B.E.; Reséndiz-Albor, A.A.; Valdés-Ramos, R. Effect on adipose tissue of diabetic mice supplemented with n-3 fatty acids extracted from microalgae. Endocr. Metab. Immune Disord. Drug Targets 2020, 20, 728–735.
- 94. Zainul-Azlan, N.; Mohd-Yusof, Y.A.; Makpol, S. Chlorella vulgaris ameliorates oxidative stress and improves the muscle regenerative capacity of young and old Sprague-Dawley rats. Nutrients 2020, 12, 3752.
- do Nascimento, T.C.; Cazarin, C.B.B.; Maróstica, M.R.J.; Zerlotti-Mercadante, A.; Jacob-Lopes, E.; Zepka, L.Q. Microalgae carotenoids intake: Influence on cholesterol levels, lipid peroxidation and antioxidant enzymes. Food Res. Int. 2020, 128, 108770.

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