## Neuroinflammatory Pathways in the Amyotrophic Lateral Sclerosis-FTD Continuum

#### Subjects: Neurosciences

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Amyotrophic Lateral Sclerosis (ALS) and Frontotemporal dementia (FDT) are progressive neurodegenerative disorders that, in several cases, overlap in clinical presentation, and genetic and pathological disease mechanisms. About 10–15% of ALS cases and up to 40% of FTD are familial, usually with dominant traits. ALS and FTD, in several cases, share common gene mutations, such as in *C9ORF72*, *TARDBP*, *SQSTM-1*, *FUS*, *VCP*, *CHCHD10*, and *TBK-1*.

genetic

amyotrophic lateral sclerosis

frontotemporal dementia

immune system

neuroinflammation

### **1. Genetic in the Amyotrophic Lateral Sclerosis-Frontotemporal Dementia Continuum**

During the last few years, several mutations in genes related to amyotrophic lateral sclerosis (ALS) and Frontotemporal dementia (FTD) were discovered due to improvements in genetic sequencing analysis and their ease of use <sup>[1]</sup>.

Superoxide dismutase 1 (*SOD1*) is the primary gene-related only to ALS development <sup>[2]</sup>. It was discovered in 1993, with more than 200 different mutations described. Although SOD1 mutations are highly heterogeneous, symptoms are often similar to sporadic forms of ALS. In some cases, patients with *SOD1* mutations present a more extended disease duration and a pure motor phenotype, generally starting from the lower limbs <sup>[3]</sup>. Other genes implicated almost exclusively in ALS are the NIMA Related Kinase 1 (*NEK1*), vesicle-associated membrane protein-associated protein B (*VAPB*), angiogenin (*ANG*), senataxin (*SETX*), and alsin (*ALS2*) <sup>[4][5][6]</sup>.

Granulin (*GRN*) and microtubule-associated protein tau (*MAPT*) are the primary genes related to FTD <sup>[Z][8]</sup>. *GRN* gene mutations were first identified in families with FTD (5–20%), mainly as autosomal dominant forms associated with chromosome 17. Until now, more than 120 *GRN* mutations have been identified, excluding most missense variants of *GRN* that are risk factors for Alzheimer's disease (AD) rather than FTD <sup>[9]</sup>. Heterozygous mutations in *GRN* cause FTD, while homozygous mutations cause a lysosomal storage disorder named neuronal ceroid lipofuscinosis, suggesting that progranulin is involved in lysosomal biogenesis and homeostasis <sup>[10]</sup>.

The *MAPT* gene, identified more than 40 years ago, is located on chromosome 17q21 and encodes for tau protein, a protein expressed in neurons of several brain regions and less in glial cells <sup>[11]</sup>. Until now, more than 100 *MAPT* mutations have been identified <sup>[12]</sup>. From a pathological point of view, mutations in *MAPT* occur primarily in the microtubule-binding repeat domain, and consequently, tau has less affinity to microtubules <sup>[12][13]</sup>. Other minor gene mutations exclusively associated with FTD and not ALS were found on *CHMP2B, CCNF*, and *TIA1* <sup>[14][15]</sup>.

Some gene mutations are detectable in the ALS-FTD spectrum disorder and include *C9ORF72*, TAR DNA-binding protein (*TARDBP*), Sequestosome-1 (*SQSTM-1*), Fused in sarcoma (*FUS*), Valosin containing protein (*VCP*), Coiled-coil-helix-coiled-coil-helix domain containing 10 (*CHCHD10*), Optineurin (*OPTN*) and Tank-binding kinase 1 (*TBK-1*) <sup>[16][17][18][19][20][21]</sup>. The first genetic link between ALS and FTD was the discovery of the *TARDBP* mutation <sup>[22]</sup>. Mutations involving *TARDBP* act on TDP-43 protein, a DNA/RNA binding protein physiologically involved in RNA metabolism. For *TARDBP*, more than 70 different genetic variants have been described <sup>[18][20]</sup>. Mutations in this gene can act with two possible pathogenic mechanisms: the first is a depletion of this protein, leading to a loss of function, while the second is a gain of function of the aggregates in terms of toxicity <sup>[13]</sup>.

The hexanucleotide repeat expansion within *C9ORF72* on chromosome 9p21 was identified in 2011, and it is responsible for a high percentage of familial cases of ALS and FTD, especially when behavioral symptoms are present <sup>[23][24]</sup>. Discovered for the first time associated with ALS-FTD cases, it has also been associated with AD, Parkinson's Disease, atypical parkinsonism, and Huntington's disease. Although the exact function of C9Orf72 is yet to be defined, the pathological degeneration related to this gene is provoked by several mechanisms, such as accumulation of RNA containing GGGGCC repeat in the brain and spinal cord, direct haploinsufficiency of C9Orf72 protein, dipeptide repeat protein toxicity arising from repeat-associated non-AUG translation occurring off the expansion, and disruption of nucleocytoplasmic transport <sup>[17][25][26][27][28]</sup>.

*SQSTM1* gene is located on 5q35 and encodes p62, a multifunctional protein involved in various cellular functions related to neurodegenerative processes, including apoptosis, the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) signaling, ubiquitin-mediated autophagy, and transcription regulation. Defective p26 accumulates in aggregates, disturbing selective autophagy pathways <sup>[29]</sup>.

Among the most recently discovered genes, in 2015, *TBK1* was described as associated with ALS-FTD in roughly 1% of European cases. *TBK1* is a member of the IkB kinase family involved in autophagy, mitophagy, and innate immune signaling. Most mutations are loss of function due to deletion of the C-terminal domain leading to activation of autophagy and inflammatory pathways via interferon (IFN) type 1. Even with high variability in age at onset and disease duration, clinically, *TBK1* mutations are associated mainly with bulbar onset ALS and cognitive impairment <sup>[19][30][31][32]</sup>.

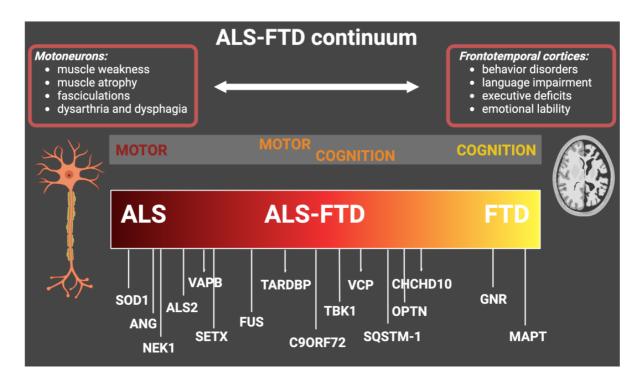
*VCP* represented another rare early example of gene overlapping; to date, more than 30 mutations have been reported <sup>[33]</sup>. Many of them are located on exon five within the N-terminal CDC48 domain, which is involved in ubiquitin-binding, meaning that mutations in this region may negatively affect the ubiquitin protein degradation pathway <sup>[34]</sup>. From a clinical point of view, mutations in *VCP* are associated with several disorders, Paget's disease

and inclusion body myositis among them <sup>[33]</sup>. Patients with ALS-FTD carriers of *VCP* mutations are phenotypically similar to sporadic forms <sup>[33]</sup>.

Mutations in *OPTN*, encoding optineurin, are rare and associated with both neurological and non-neurological conditions, including the ALS-FTD continuum <sup>[35]</sup>. *OPTN* was identified for the first time as an ALS causative gene in 2010, and since then, more than 20 mutations have been reported. The involvement of *OPTN* in FTD is less clear, although the case series reported a percentage of 5% of FTD patients carrying the *OPTN* mutation <sup>[36]</sup>. Optineurin is a protein involved in several cellular processes that act on several pathways. Importantly, *OPTN* influences the innate immune response by negatively regulating inflammatory pathways <sup>[37]</sup>.

Lastly, mutations in the *CHCHD10* gene encode for a small percentage of ALS-FTD patients. *CHCHD10* is a mitochondrial protein that regulates mitochondrial metabolism, synthesizes respiratory chain components, and modulates cell apoptosis <sup>[38]</sup>. At least 30 variants have since been reported, and they are concentrated on exon two of the gene encoding the non-structured N-terminal <sup>[38]</sup>. In addition to ALS-FTD, disorders related to *CHCHD10* mutations are several and varied, such as myopathy, Charcot-Marie Tooth neuropathy, and cerebellar ataxia <sup>[39][40]</sup>.

The involved genes along the ALS-FTD continuum are summarized in **Figure 1**.



**Figure 1.** Genes with genetic mutations that cause ALS, ALS-FTD, and FTD. The figure shows the main causative genes associated with the ALS-FTD continuum and the main related clinical features. ALS: amyotrophic lateral sclerosis; FTD: frontotemporal dementia.

# 2. Autoimmune Disorders and Systemic Inflammation in Genetic Amyotrophic Lateral Sclerosis- Frontotemporal

## Dementia

Several clinical studies focused on the epidemiology of sporadic ALS and FTD support the idea that patients affected by these disorders have a higher incidence of autoimmune diseases. In 2013, the group of Turner evaluated a large UK cohort with autoimmune diseases (e.g., asthma, celiac disease, young-onset diabetes, multiple sclerosis, and systemic lupus erythematosus), reporting in this group a higher percentage of ALS compared to the general population and hypothesizing the possibility of shared genetic and/or environmental risk factors <sup>[41]</sup>. Although the data is less evident in cohorts of sporadic FTD patients <sup>[42]</sup>, an interesting paper investigated the association between immune disorders and patients with mutations in the *GRN* gene, observing a significantly increased risk of autoimmune disorders clustered around inflammatory arthritis, cutaneous and gastrointestinal disorders in the PGNR cohort <sup>[43]</sup>.

Regarding other selected cohorts of genetic patients, more results regard the C9Orf72 patients. Firstly, in 2016, in a US cohort, it was reported a high prevalence of autoimmune diseases (similar to what was described for GRN patients) in C9Orf72 ALS/FTD patients compared to controls and other sporadic neurodegenerative patients <sup>[44]</sup>. A recent multicentric Italian cohort study described 150 ALS C9Orf72 patients, in which an increased incidence of autoimmune diseases was not observed, whereas the authors reported a longer survival in C9Orf72 patients with ALS and thyroid disorders <sup>[45]</sup>. Looking at the problem in reverse, genotyping a small cohort of patients with ALS and multiple sclerosis, 80% of patients with both neurological conditions carried a C9Orf72 mutation. In the same report, they described an imbalance in the immune pathway, with an NF-κB activation and a concomitant CXCL10 downregulation in the CSF <sup>[46]</sup>. The presented studies and the possible correlation between NDDs, genetics, and autoimmunity are supported by the recent detection of *TBK1* mutations in the ALS/FTD spectrum. In fact, as researchers have mentioned, *TBK1* plays a crucial role as the hub for many innate immune signaling pathways <sup>[47]</sup>.

Also, while systemic inflammation is overall present in ALS/FTD patients, it is not clear if these phenomena are a disease consequence or have a causative role. As global markers of inflammation and peripheral immune disbalance, several recent studies focused on alterations in peripheral markers, such as the neutrophil-to-lymphocyte ratio, in ALS and FTD. Alterations in this ratio, and others related, were associated with poorer prognosis and shorter survival by several groups <sup>[48][49][50]</sup>, but in no study, the genetic component is considered. Also, the studies that analyzed the C-reactive protein in ALS did not primarily focus on genetic subgroups of patients <sup>[51][52]</sup>.

### References

- 1. Guerreiro, R.; Brás, J.; Hardy, J. SnapShot: Genetics of ALS and FTD. Cell 2015, 160, 798.
- Rosen, D.R.; Siddique, T.; Patterson, D.; Figlewicz, D.A.; Sapp, P.; Hentati, A.; Donaldson, D.; Goto, J.; O'Regan, J.P.; Deng, H.-X. Mutations in Cu/Zn Superoxide Dismutase Gene Are Associated with Familial Amyotrophic Lateral Sclerosis. Nature 1993, 362, 59.

- Bali, T.; Self, W.; Liu, J.; Siddique, T.; Wang, L.H.; Bird, T.D.; Ratti, E.; Atassi, N.; Boylan, K.B.; Glass, J.D. Defining SOD1 ALS Natural History to Guide Therapeutic Clinical Trial Design. J. Neurol. Neurosurg. Psychiatry 2017, 88, 99–105.
- Grassano, M.; Calvo, A.; Moglia, C.; Sbaiz, L.; Brunetti, M.; Barberis, M.; Casale, F.; Manera, U.; Vasta, R.; Canosa, A. Systematic Evaluation of Genetic Mutations in ALS: A Population-Based Study. J. Neurol. Neurosurg. Psychiatry 2022, 93, 1190–1193.
- Riva, N.; Pozzi, L.; Russo, T.; Pipitone, G.B.; Schito, P.; Domi, T.; Agosta, F.; Quattrini, A.; Carrera, P.; Filippi, M. NEK1 Variants in a Cohort of Italian Patients with Amyotrophic Lateral Sclerosis. Front. Neurosci. 2022, 16, 833051.
- 6. Chia, R.; Chiò, A.; Traynor, B.J. Novel Genes Associated with Amyotrophic Lateral Sclerosis: Diagnostic and Clinical Implications. Lancet Neurol. 2018, 17, 94–102.
- 7. Van Swieten, J.C.; Heutink, P. Mutations in Progranulin (GRN) within the Spectrum of Clinical and Pathological Phenotypes of Frontotemporal Dementia. Lancet Neurol. 2008, 7, 965–974.
- B. Ghetti, B.; Oblak, A.L.; Boeve, B.F.; Johnson, K.A.; Dickerson, B.C.; Goedert, M. Invited Review: Frontotemporal Dementia Caused by Microtubule-associated Protein Tau Gene (MAPT) Mutations: A Chameleon for Neuropathology and Neuroimaging. Neuropathol. Appl. Neurobiol. 2015, 41, 24–46.
- 9. Wang, X.; Zeng, P.; Fang, Y.; Zhang, T.; Tian, Q. Progranulin in Neurodegenerative Dementia. J. Neurochem. 2021, 158, 119–137.
- 10. Simon, M.J.; Logan, T.; DeVos, S.L.; Di Paolo, G. Lysosomal Functions of Progranulin and Implications for Treatment of Frontotemporal Dementia. Trends Cell Biol. 2022, 33, 324–339.
- 11. Rademakers, R.; Cruts, M.; Van Broeckhoven, C. The Role of Tau (MAPT) in Frontotemporal Dementia and Related Tauopathies. Hum. Mutat. 2004, 24, 277–295.
- 12. Strang, K.H.; Golde, T.E.; Giasson, B.I. MAPT Mutations, Tauopathy, and Mechanisms of Neurodegeneration. Lab. Investig. 2019, 99, 912–928.
- 13. Virgilio, E.; De Marchi, F.; Contaldi, E.; Dianzani, U.; Cantello, R.; Mazzini, L.; Comi, C. The Role of Tau beyond Alzheimer's Disease: A Narrative Review. Biomedicines 2022, 10, 760.
- 14. Olszewska, D.A.; Lonergan, R.; Fallon, E.M.; Lynch, T. Genetics of Frontotemporal Dementia. Curr. Neurol. Neurosci. Rep. 2016, 16, 1–15.
- 15. Greaves, C.V.; Rohrer, J.D. An Update on Genetic Frontotemporal Dementia. J. Neurol. 2019, 266, 2075–2086.
- 16. Ferrari, R.; Kapogiannis, D.; D Huey, E.; Momeni, P. FTD and ALS: A Tale of Two Diseases. Curr. Alzheimer Res. 2011, 8, 273–294.

- 17. Balendra, R.; Isaacs, A.M. C9orf72-Mediated ALS and FTD: Multiple Pathways to Disease. Nat. Rev. Neurol. 2018, 14, 544–558.
- Kabashi, E.; Valdmanis, P.N.; Dion, P.; Spiegelman, D.; McConkey, B.J.; Velde, C.V.; Bouchard, J.-P.; Lacomblez, L.; Pochigaeva, K.; Salachas, F. TARDBP Mutations in Individuals with Sporadic and Familial Amyotrophic Lateral Sclerosis. Nat. Genet. 2008, 40, 572–574.
- Cui, R.; Tuo, M.; Li, P.; Zhou, C. Association between TBK1 Mutations and Risk of Amyotrophic Lateral Sclerosis/Frontotemporal Dementia Spectrum: A Meta-Analysis. Neurol. Sci. 2018, 39, 811–820.
- 20. Lattante, S.; Rouleau, G.A.; Kabashi, E. TARDBP and FUS Mutations Associated with Amyotrophic Lateral Sclerosis: Summary and Update. Hum. Mutat. 2013, 34, 812–826.
- Kwiatkowski, T.J.; Bosco, D.A.; Leclerc, A.L.; Tamrazian, E.; Vanderburg, C.R.; Russ, C.; Davis, A.; Gilchrist, J.; Kasarskis, E.J.; Munsat, T. Mutations in the FUS/TLS Gene on Chromosome 16 Cause Familial Amyotrophic Lateral Sclerosis. Science 2009, 323, 1205–1208.
- Rutherford, N.J.; Zhang, Y.-J.; Baker, M.; Gass, J.M.; Finch, N.A.; Xu, Y.-F.; Stewart, H.; Kelley, B.J.; Kuntz, K.; Crook, R.J.P. Novel Mutations in TARDBP (TDP-43) in Patients with Familial Amyotrophic Lateral Sclerosis. PLoS Genet. 2008, 4, e1000193.
- DeJesus-Hernandez, M.; Mackenzie, I.R.; Boeve, B.F.; Boxer, A.L.; Baker, M.; Rutherford, N.J.; Nicholson, A.M.; Finch, N.A.; Flynn, H.; Adamson, J. Expanded GGGGCC Hexanucleotide Repeat in Noncoding Region of C9ORF72 Causes Chromosome 9p-Linked FTD and ALS. Neuron 2011, 72, 245–256.
- Renton, A.E.; Majounie, E.; Waite, A.; Simón-Sánchez, J.; Rollinson, S.; Gibbs, J.R.; Schymick, J.C.; Laaksovirta, H.; Van Swieten, J.C.; Myllykangas, L. A Hexanucleotide Repeat Expansion in C9ORF72 Is the Cause of Chromosome 9p21-Linked ALS-FTD. Neuron 2011, 72, 257–268.
- 25. Vatsavayai, S.C.; Nana, A.L.; Yokoyama, J.S.; Seeley, W.W. C9orf72-FTD/ALS Pathogenesis: Evidence from Human Neuropathological Studies. Acta Neuropathol. 2019, 137, 1–26.
- 26. Smeyers, J.; Banchi, E.-G.; Latouche, M. C9ORF72: What It Is, What It Does, and Why It Matters. Front. Cell Neurosci. 2021, 15, 661447.
- Farg, M.A.; Sundaramoorthy, V.; Sultana, J.M.; Yang, S.; Atkinson, R.A.K.; Levina, V.; Halloran, M.A.; Gleeson, P.A.; Blair, I.P.; Soo, K.Y.; et al. C9ORF72, Implicated in Amytrophic Lateral Sclerosis and Frontotemporal Dementia, Regulates Endosomal Trafficking. Hum. Mol. Genet. 2014, 23, 3579–3595.
- 28. Gendron, T.F.; Petrucelli, L. Disease Mechanisms of C9ORF72 Repeat Expansions. Cold Spring Harb. Perspect. Med. 2018, 8, a024224.

- Le Ber, I.; Camuzat, A.; Guerreiro, R.; Bouya-Ahmed, K.; Bras, J.; Nicolas, G.; Gabelle, A.; Didic, M.; De Septenville, A.; Millecamps, S. SQSTM1 Mutations in French Patients with Frontotemporal Dementia or Frontotemporal Dementia with Amyotrophic Lateral Sclerosis. JAMA Neurol. 2013, 70, 1403–1410.
- Freischmidt, A.; Wieland, T.; Richter, B.; Ruf, W.; Schaeffer, V.; Müller, K.; Marroquin, N.; Nordin, F.; Hübers, A.; Weydt, P. Haploinsufficiency of TBK1 Causes Familial ALS and Fronto-Temporal Dementia. Nat. Neurosci. 2015, 18, 631–636.
- Gijselinck, I.; Van Mossevelde, S.; van der Zee, J.; Sieben, A.; Philtjens, S.; Heeman, B.; Engelborghs, S.; Vandenbulcke, M.; De Baets, G.; Bäumer, V.; et al. Loss of TBK1 Is a Frequent Cause of Frontotemporal Dementia in a Belgian Cohort. Neurology 2015, 85, 2116–2125.
- 32. Herhaus, L. TBK1 (TANK-Binding Kinase 1)-Mediated Regulation of Autophagy in Health and Disease. Matrix Biol. 2021, 100–101, 84–98.
- Koppers, M.; van Blitterswijk, M.M.; Vlam, L.; Rowicka, P.A.; van Vught, P.W.J.; Groen, E.J.N.; Spliet, W.G.M.; Engelen-Lee, J.; Schelhaas, H.J.; de Visser, M. VCP Mutations in Familial and Sporadic Amyotrophic Lateral Sclerosis. Neurobiol. Aging 2012, 33, 837.e7–837.e13.
- Ju, J.-S.; Fuentealba, R.A.; Miller, S.E.; Jackson, E.; Piwnica-Worms, D.; Baloh, R.H.; Weihl, C.C. Valosin-Containing Protein (VCP) Is Required for Autophagy and Is Disrupted in VCP Disease. J. Cell Biol. 2009, 187, 875–888.
- Maruyama, H.; Morino, H.; Ito, H.; Izumi, Y.; Kato, H.; Watanabe, Y.; Kinoshita, Y.; Kamada, M.; Nodera, H.; Suzuki, H. Mutations of Optineurin in Amyotrophic Lateral Sclerosis. Nature 2010, 465, 223–226.
- 36. Pottier, C.; Bieniek, K.F.; Finch, N.; van de Vorst, M.; Baker, M.; Perkersen, R.; Brown, P.; Ravenscroft, T.; van Blitterswijk, M.; Nicholson, A.M. Whole-Genome Sequencing Reveals Important Role for TBK1 and OPTN Mutations in Frontotemporal Lobar Degeneration without Motor Neuron Disease. Acta Neuropathol. 2015, 130, 77–92.
- 37. Shen, W.-C.; Li, H.-Y.; Chen, G.-C.; Chern, Y.; Tu, P. Mutations in the Ubiquitin-Binding Domain of OPTN/Optineurin Interfere with Autophagy-Mediated Degradation of Misfolded Proteins by a Dominant-Negative Mechanism. Autophagy 2015, 11, 685–700.
- Lehmer, C.; Schludi, M.H.; Ransom, L.; Greiling, J.; Junghänel, M.; Exner, N.; Riemenschneider, H.; van der Zee, J.; Van Broeckhoven, C.; Weydt, P.; et al. A Novel CHCHD10 Mutation Implicates a Mia40-Dependent Mitochondrial Import Deficit in ALS. EMBO Mol. Med. 2018, 10, e8558.
- Ronchi, D.; Riboldi, G.; Del Bo, R.; Ticozzi, N.; Scarlato, M.; Galimberti, D.; Corti, S.; Silani, V.; Bresolin, N.; Comi, G. Pietro CHCHD10 Mutations in Italian Patients with Sporadic Amyotrophic Lateral Sclerosis. Brain 2015, 138, e372.

- Chaussenot, A.; Le Ber, I.; Ait-El-Mkadem, S.; Camuzat, A.; de Septenville, A.; Bannwarth, S.; Genin, E.C.; Serre, V.; Augé, G.; Brice, A. Screening of CHCHD10 in a French Cohort Confirms the Involvement of This Gene in Frontotemporal Dementia with Amyotrophic Lateral Sclerosis Patients. Neurobiol. Aging 2014, 35, 2884.e1–2884.e4.
- 41. Turner, M.R.; Goldacre, R.; Ramagopalan, S.; Talbot, K.; Goldacre, M.J. Autoimmune Disease Preceding Amyotrophic Lateral Sclerosis: An Epidemiologic Study. Neurology 2013, 81, 1222– 1225.
- 42. Jin, T.; Huang, W.; Cao, F.; Yu, X.; Guo, S.; Ying, Z.; Xu, C. Causal Association between Systemic Lupus Erythematosus and the Risk of Dementia: A Mendelian Randomization Study. Front. Immunol. 2022, 13, 1063110.
- 43. Miller, Z.A.; Rankin, K.P.; Graff-Radford, N.R.; Takada, L.T.; Sturm, V.E.; Cleveland, C.M.; Criswell, L.A.; Jaeger, P.A.; Stan, T.; Heggeli, K.A. TDP-43 Frontotemporal Lobar Degeneration and Autoimmune Disease. J. Neurol. Neurosurg. Psychiatry 2013, 84, 956–962.
- 44. Miller, Z.A.; Sturm, V.E.; Camsari, G.B.; Karydas, A.; Yokoyama, J.S.; Grinberg, L.T.; Boxer, A.L.; Rosen, H.J.; Rankin, K.P.; Gorno-Tempini, M.L. Increased Prevalence of Autoimmune Disease within C9 and FTD/MND Cohorts: Completing the Picture. Neurol. -Neuroimmunol. Neuroinflammation 2016, 3, e301.
- Mandrioli, J.; Zucchi, E.; Martinelli, I.; Van der Most, L.; Gianferrari, G.; Moglia, C.; Manera, U.; Solero, L.; Vasta, R.; Canosa, A. Factors Predicting Disease Progression in C9ORF72 ALS Patients. J. Neurol. 2023, 270, 877–890.
- 46. Ismail, A.; Cooper-Knock, J.; Highley, J.R.; Milano, A.; Kirby, J.; Goodall, E.; Lowe, J.; Scott, I.; Constantinescu, C.S.; Walters, S.J. Concurrence of Multiple Sclerosis and Amyotrophic Lateral Sclerosis in Patients with Hexanucleotide Repeat Expansions of C9ORF72. J. Neurol. Neurosurg. Psychiatry 2013, 84, 79–87.
- 47. Xu, D.; Jin, T.; Zhu, H.; Chen, H.; Ofengeim, D.; Zou, C.; Mifflin, L.; Pan, L.; Amin, P.; Li, W. TBK1 Suppresses RIPK1-Driven Apoptosis and Inflammation during Development and in Aging. Cell 2018, 174, 1477–1491.
- 48. Choi, S.-J.; Hong, Y.-H.; Kim, S.-M.; Shin, J.-Y.; Suh, Y.J.; Sung, J.-J. High Neutrophil-to-Lymphocyte Ratio Predicts Short Survival Duration in Amyotrophic Lateral Sclerosis. Sci. Rep. 2020, 10, 428.
- 49. Wei, Q.-Q.; Hou, Y.-B.; Zhang, L.-Y.; Ou, R.-W.; Cao, B.; Chen, Y.-P.; Shang, H.-F. Neutrophil-to-Lymphocyte Ratio in Sporadic Amyotrophic Lateral Sclerosis. Neural Regen. Res. 2022, 17, 875.
- 50. Leone, M.A.; Mandrioli, J.; Russo, S.; Cucovici, A.; Gianferrari, G.; Lisnic, V.; Muresanu, D.F.; Giuliani, F.; Copetti, M.; Consortium, P.R.O.-A.A.L.S.C.T. Neutrophils-to-Lymphocyte Ratio Is

Associated with Progression and Overall Survival in Amyotrophic Lateral Sclerosis. Biomedicines 2022, 10, 354.

- 51. Lunetta, C.; Lizio, A.; Maestri, E.; Sansone, V.A.; Mora, G.; Miller, R.G.; Appel, S.H.; Chiò, A. Serum C-Reactive Protein as a Prognostic Biomarker in Amyotrophic Lateral Sclerosis. JAMA Neurol. 2017, 74, 660–667.
- Beers, D.R.; Zhao, W.; Neal, D.W.; Thonhoff, J.R.; Thome, A.D.; Faridar, A.; Wen, S.; Wang, J.; Appel, S.H. Elevated Acute Phase Proteins Reflect Peripheral Inflammation and Disease Severity in Patients with Amyotrophic Lateral Sclerosis. Sci. Rep. 2020, 10, 15295.

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