

The Glutamatergic System

Subjects: **Neurosciences**

Contributor: Maria Meringolo , Paola Bonsi , Martina Montanari , Giuseppina Martella

Neurotransmitters play a critical role in developing both the peripheral and the central nervous systems. It is therefore conceivable that neurotransmitter dysfunctions may be involved in ASD pathophysiology. Among neurotransmitters, glutamate (Glu) is considered a good candidate as it is directly involved in brain development and synaptogenesis, memory, behavior, and motor activity regulation, and gastrointestinal functions.

glutamate receptors

brain development

1. Introduction

Autism Spectrum Disorder (ASD) is a mosaic of neurodevelopmental conditions, which show common deficits in two behavioral domains: (i) social interaction and communication difficulties, (ii) narrow interests, and repetitive and stereotyped behaviors (DSM-5) ^{[1][2]}. ASD can be associated with several co-occurring conditions, including seizures, attention-deficit/hyperactivity disorder (ADHD), and other cognitive impairments ^{[3][4]}. The estimated prevalence of ASD is about 1% of the human population ^[5], with males affected four times more frequently than females ^[6].

Although it has become clear that ASD has a complex and multifactorial etiopathogenesis, twin and family studies indicate a strong genetic background and high heritability, with a concordance rate of 60% to 95% between monozygotic twins versus 0% to 30% between dizygotic twins ^[7]. Despite the high heterogeneity between ASD cases, the occurrence of shared symptoms suggests common deficits in some neurodevelopmental pathways. Neurotransmitters play a critical role in developing both the peripheral and the central nervous systems. It is therefore conceivable that neurotransmitter dysfunctions may be involved in ASD pathophysiology. Among neurotransmitters, glutamate (Glu) is considered a good candidate ^{[8][9][10]} as it is directly involved in brain development and synaptogenesis ^{[11][12]}, memory, behavior, and motor activity regulation ^{[13][14][15]}, and gastrointestinal functions ^{[16][17]}. Studies on postmortem or body fluids samples provide important evidence about critical changes in Glu concentration in both pediatric and adult patients with ASD ^{[18][19]}. Furthermore, abnormalities in Glu receptors genes and deregulation of glutamatergic pathways have been reported both in ASD patients and animal models ^{[20][21][22][23]}. In general, excitotoxicity has already been connected with ASD. In addition, the increased prevalence of epilepsy in autistic patients compared to the general population ^{[24][25][26]} further strengthens the hypothesis of the dysfunction in excitatory and/or inhibitory network activity. In the past decades, two opposite theories about the role of Glu signaling in ASD have been proposed. In 1998, Carlsson suggested that ASD might result from a decrease in Glu signaling according to (i) overlapping between the ASD condition and the symptoms produced by Glu antagonist, (ii) the neuroanatomical and neuroimaging evidence of

glutamatergic areas impairment in ASD patients, (iii) the similarities with ASD symptoms in hypoglutamatergic animals treated with NMDA antagonists [27]. Conversely, Fatemi proposed an opposite theory, suggesting an hyperglutamatergic state in ASD, based on the observation of increased Glu levels in blood samples [19].

2. The Critical Role of the Glutamatergic System in Brain Development

ASD symptoms appear early in development, within the first three years of life, during the fundamental period of rapid synapse formation and maturation [28][29]. Brain development involves a number of processes, including synaptogenesis, axonal and dendritic arborization, migration, and synaptic plasticity. These functions have the overall aim of building a functional brain. They can, however, cause cellular, biochemical, and structural alterations in the neonatal brain [30]. During brain development, the role played by neurotransmitters and their receptors is of primary importance. The distribution and molecular characteristics of Glu receptors change considerably during brain development, making the brain vulnerable to changes in Glu neurotransmission during growth. Indeed, alterations in the expression and regulation of Glu receptors are known to be implicated in some neuropathological conditions such as neurological and psychiatric disorders, including Alzheimer's and Parkinson's diseases, schizophrenia, mood disorders, depression, epilepsy, anxiety, stress, and ASD [31][32][33][34]. Studies in animal models of neurologic disease suggest an altered expression of Glu proteins [12][35]. Knowledge and research on the ontogenetic alterations in Glu receptor function, subunit expression, and binding properties in pathological conditions are still very incomplete. However, in vivo and in vitro studies have provided information about the differences in both regional densities and the time-course of changes in the expression levels of the different Glu receptors subunits (**Figure 1**) [36].

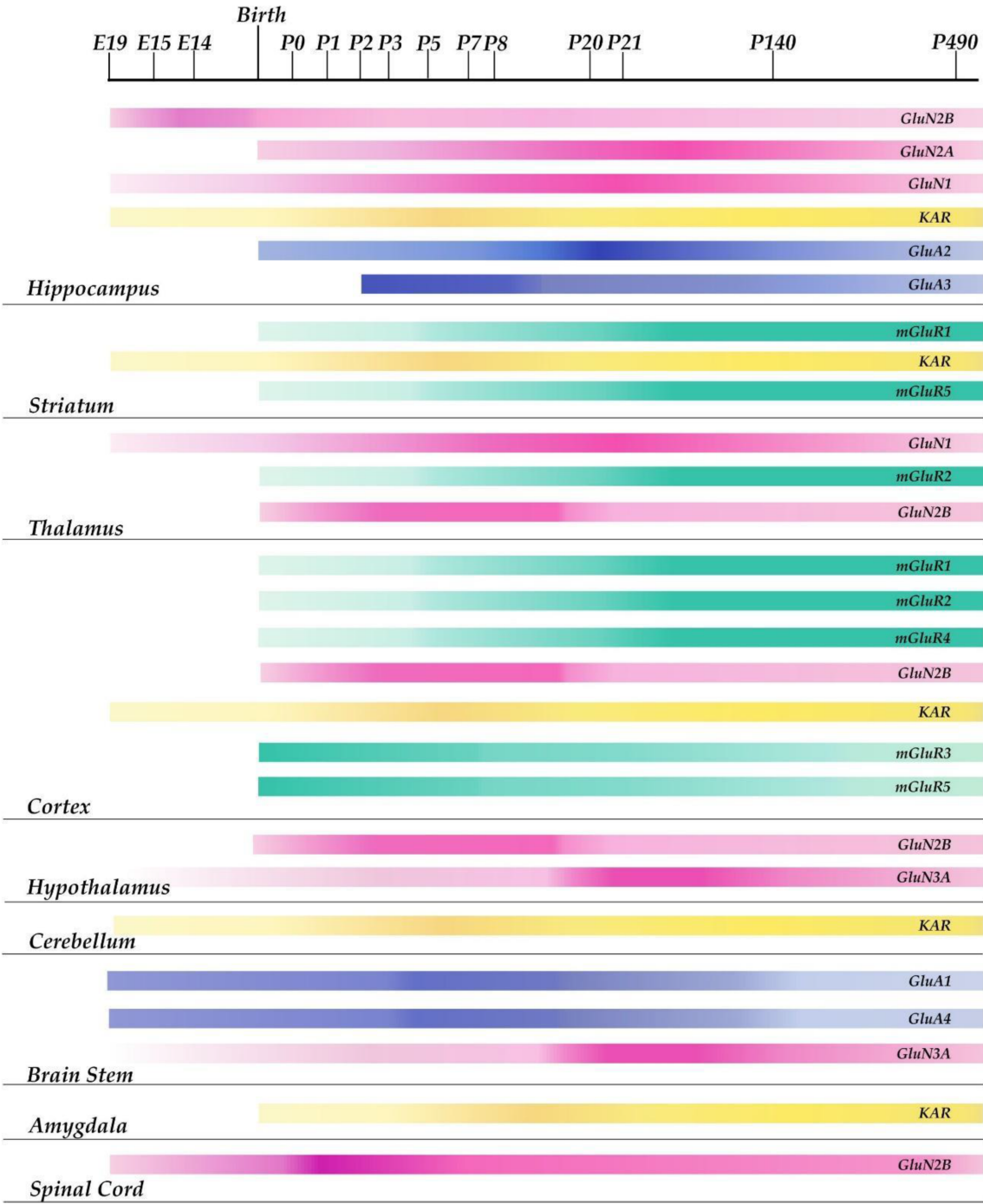


Figure 1. Schematic representation of the developmental time course of glutamatergic receptor subunits expression in different rodent brain areas. The variations in color intensity within the single bars represent changes along time in the expression levels of each receptor subunit in a particular area of the rodent brain. The midbrain, pons, and medulla are represented collectively as brain stem in the figure.

For example, the α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor (AMPA) is constituted by tetramers of different subunits (GluR1-4) [37]. The expression pattern of the GluR2 subunit during the period of postnatal development, maturation, and aging in male mice was investigated: it was observed that the expression of the GluR2 subunit is gradually upregulated in the hippocampus from postnatal day 0 (P0) to adult age (20

weeks) and subsequently down-regulated in 70 week-old male mice [38]. GluR1 and GluR4 subunits are expressed at significant levels in the midbrain from embryonic day 15 (E15), they remain constant until delivery, and then decrease immediately after birth, while GluR2 and GluR3 levels increase after early development in the hippocampus [34]. N-methyl-D-aspartate receptors (NMDAR) are expressed during the developmental periods of intense synaptic formation. Similar to AMPAR, the expression of the various NMDAR subunits varies over the course of development and in different regions of the brain [39]. Studies in rodents have shown that the GluN1 subunit is expressed before E14 in areas of the brain related to cognitive function, such as the cortex, hippocampus, and thalamus. Three weeks after birth, GluN1 levels reach their highest levels and then decrease [40]. The GluN2 subunits are expressed at variable levels at specific stages of neonatal development in rats. In the hippocampus, low levels of GluN2A are detected at P0, while a peak is observed at P21 [41]. As the brain develops, the protein expression of the GluN2A subunit gradually increases. GluN2B is the predominant NMDA receptor subunit located at immature synapses [36]. During prenatal development, it is expressed in the cortex, thalamus, and spinal cord, and its concentrations increase at birth. Lower amounts are expressed in the colliculi, hippocampus, and hypothalamus [34]. After birth, GluN2B levels reach a peak of expression in the cortex and hypothalamus, two important brain structures involved in cognition. The GluN3A subunit is involved in developing dendritic spines and synaptogenesis [34]. This subunit starts to be expressed in the medulla, pons, and hypothalamus at E15 [42][43]; its levels reach a peak at P8 and then decrease at P20. Kainate-2-carboxy-3-carboxy-methyl-4-isopropenylpyrrolidine receptors (KAR) are expressed at low amounts from E19, during embryonic development, in the cortex, hippocampus, cerebellum, and striatum [34]. Their activation regulates the network and synaptic activity in the neonatal hippocampus [44][45]. KAR are widely expressed in the amygdala within the first postnatal week, a period that coincides with the process of synaptogenesis, suggesting its involvement in the process of synapse formation [45]. Rodent studies have demonstrated a low expression at birth of type 1, 2, and 4 Glu metabotropic receptors (mGlu1, mGlu2, mGlu4) that increase during neonatal development [43]. Conversely, at P0, the levels of type 3 and 5 Glu metabotropic receptors, mGlu3 and mGlu5, are very high and then decrease during the maturation period. In fact, both receptors are involved in synaptogenesis, and GRM5 is involved in the proliferation and survival of neural progenitor cells, as well as in the migration of cortical neurons [46]. Another important aspect of the brain development process is the maintenance of the excitation-inhibition (E/I) balance between Glu and Gamma-Aminobutyric Acid (GABA) [47]. Opposite to Glu, GABA is involved in inhibitory neurotransmission [48]. However, at the beginning of development, GABAergic neurons form excitatory synapses, which become inhibitory only later during the maturation process [49]. For normal brain development and functioning, it is essential that the E/I balance is maintained stably in neuron's synapses and neural circuits [10][50]. Consequently, disturbances in the E/I balance have been implicated in neurodevelopmental disorders, including ASD [51][52].

3. Sex Differences in the Glutamatergic System

Accumulating evidence has revealed that gonadal steroids finely modulate neurotransmission mediated by amino acids, including Glu, in developing and adult brains [53]. Indeed, there appear to be functionally relevant baseline sex differences in neurotransmitter systems [11][54]. Clinical studies have highlighted increased Glu levels in frontal

grey matter and basal ganglia (BG) of females compared to males, while parietal grey matter (PGM) Glu concentration is higher in men than in women [55]. Further studies have identified in more detail sex differences in Glu concentration in specific brain regions. Females were found to exhibit higher Glu levels than males in the sensorimotor cortex, anterior cingulate cortex (ACC), striatum (STR), and cerebellum [56][57]. In contrast, prefrontal cortex (PFC) Glu concentration is higher in males than in females [58]. Along with the Glu concentration changes observed in the CNS, gender differences were also found in plasma Glu concentrations. Clinical studies reported higher Glu levels in males than females, which appear inversely correlated to estrogen and progesterone levels [59]. Gender differences in Glu concentration appear to become more conspicuous with age. Males, but not females, display an age-dependent decline in BG and PGM Glu levels [55], while females show a more pronounced age-dependent decrease in ACC Glu levels [60]. In addition, blood Glu levels exhibit an age-related increase in plasma in females but not in males [61]. Similar to what is seen in humans, preclinical studies have revealed significant sex-related differences in the Glu system of rodents. Increased Glu levels have been found in the lateral hypothalamus and habenula of male rats [62]. Additionally, rodents show a sex bias in Glu receptors distribution. For example, female rats show an enhanced expression of mGlu2/3, mGlu5, and the GluN1 and Glu2B subunits of the NMDAR in the hippocampus [63][64][65], along with greater mGlu5 expression in the PFC [66]. In line with the reported sex-related differences in the rodent Glu system, in female rats, AMPAR-dependent synaptic events are altered: synaptic transmission is enhanced, whereas the long-term potentiation (LTP) magnitude is reduced, the latter alteration likely due to reduced phosphorylation of the GluR1 subunit [67].

Sex-specific factors have been hypothesized to increase male's risk of ASD, or increase the protection of females from ASD, as more males than females are affected. The sex-related differences in the Glu system discussed above suggest the importance of elucidating the molecular mechanisms by which Glu dysfunction could differentially affect males and females.

References

1. Yin, H.H.; Knowlton, B.J. The role of the basal ganglia in habit formation. *Nat. Rev. Neurosci.* 2006, 7, 464–476.
2. American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders*, 5th ed.; American Psychiatric Association: Washington, DC, USA, 2013; ISBN 0-89042-555-8.
3. Fetit, R.; Hillary, R.F.; Price, D.J.; Lawrie, S.M. The neuropathology of autism: A systematic review of post-mortem studies of autism and related disorders. *Neurosci. Biobehav. Rev.* 2021, 129, 35–62.
4. Constantino, J.N.; Abbacchi, A.M.; Saulnier, C.; Klaiman, C.; Mandell, D.S.; Zhang, Y.; Hawks, Z.; Bates, J.; Klin, A.; Shattuck, P.; et al. Timing of the diagnosis of autism in African American children. *Pediatrics* 2020, 146, e20193629.

5. Marotta, R.; Risoleo, M.C.; Messina, G.; Parisi, L.; Carotenuto, M.; Vetri, L.; Roccella, M. The neurochemistry of autism. *Brain Sci.* 2020, 10, 163.
6. Pickles, A.; McCauley, J.B.; Pepa, L.A.; Huerta, M.; Lord, C. The adult outcome of children referred for autism: Typology and prediction from childhood. *J. Child Psychol. Psychiatry* 2020, 61, 760–767.
7. Carroll, L.; Braeutigam, S.; Dawes, J.M.; Krsnik, Z.; Kostovic, I.; Coutinho, E.; Dewing, J.M.; Horton, C.A.; Gomez-Nicola, D.; Menassa, D.A. Autism spectrum disorders: Multiple routes to, and multiple consequences of, abnormal synaptic function and connectivity. *Neuroscientist* 2021, 27, 10–29.
8. Rojas, D.C. The role of glutamate and its receptors in autism and the use of glutamate receptor antagonists in treatment. *J. Neural Transm.* 2014, 121, 891–905.
9. Carlson, G.C. Glutamate receptor dysfunction and drug targets across models of autism spectrum disorders. *Pharmacol. Biochem. Behav.* 2012, 100, 850–854.
10. Choudhury, P.R.; Lahiri, S.; Rajamma, U. Glutamate mediated signaling in the pathophysiology of autism spectrum disorders. *Pharmacol. Biochem. Behav.* 2012, 100, 841–849.
11. Wickens, M.M.; Bangasser, D.A.; Briand, L.A. Sex differences in psychiatric disease: A focus on the glutamate system. *Front. Mol. Neurosci.* 2018, 11, 197.
12. Basu, S.K.; Pradhan, S.; du Plessis, A.J.; Ben-Ari, Y.; Limperopoulos, C. GABA and glutamate in the preterm neonatal brain: In-vivo measurement by magnetic resonance spectroscopy. *Neuroimage* 2021, 238, 118215.
13. Guimaraes, I.M.; Carvalho, T.G.; Ferguson, S.S.; Pereira, G.S.; Ribeiro, F.M. The metabotropic glutamate receptor 5 role on motor behavior involves specific neural substrates. *Mol. Brain* 2015, 8, 24.
14. Gécz, J. Glutamate receptors and learning and memory. *Nat. Genet.* 2010, 42, 925–926.
15. Zoicas, I.; Kornhuber, J. The role of metabotropic glutamate receptors in social behavior in rodents. *Int. J. Mol. Sci.* 2019, 20, 1412.
16. Tomé, D. The roles of dietary glutamate in the intestine. *Ann. Nutr. Metab.* 2018, 73 (Suppl. 5), 15–20.
17. Kirchgessner, A.L. Glutamate in the enteric nervous system. *Curr. Opin. Pharmacol.* 2001, 1, 591–596.
18. Aldred, S.; Moore, K.M.; Fitzgerald, M.; Waring, R.H. Plasma amino acid levels in children with autism and their families. *J. Autism Dev. Disord.* 2003, 33, 93–97.

19. Fatemi, S.H. The hyperglutamatergic hypothesis of autism. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 2008, 32, 912–913.
20. Serajee, F.J.; Zhong, H.; Nabi, R.; Huq, A.H.M.M. The metabotropic glutamate receptor 8 gene at 7q31: Partial duplication and possible association with autism. *J. Med. Genet.* 2003, 40, e42.
21. Jamain, S.; Betancur, C.; Quach, H.; Philippe, A.; Fellous, M.; Giros, B.; Gillberg, C.; Leboyer, M.; Bourgeron, T. Paris Autism Research International Sibpair (PARIS) Study Linkage and association of the glutamate receptor 6 gene with autism. *Mol. Psychiatry* 2002, 7, 302–310.
22. Purcell, A.E.; Jeon, O.H.; Zimmerman, A.W.; Blue, M.E.; Pevsner, J. Postmortem brain abnormalities of the glutamate neurotransmitter system in autism. *Neurology* 2001, 57, 1618–1628.
23. Blandini, F. An update on the potential role of excitotoxicity in the pathogenesis of Parkinson's disease. *Funct. Neurol.* 2010, 25, 65–71.
24. Bozzi, Y.; Provenzano, G.; Casarosa, S. Neurobiological bases of autism-epilepsy comorbidity: A focus on excitation/inhibition imbalance. *Eur. J. Neurosci.* 2018, 47, 534–548.
25. Bolton, P.F.; Carcani-Rathwell, I.; Hutton, J.; Goode, S.; Howlin, P.; Rutter, M. Epilepsy in autism: Features and correlates. *Br. J. Psychiatry* 2011, 198, 289–294.
26. Lee, B.H.; Smith, T.; Paciorkowski, A.R. Autism spectrum disorder and epilepsy: Disorders with a shared biology. *Epilepsy Behav.* 2015, 47, 191–201.
27. Carlsson, M.L. Hypothesis: Is infantile autism a hypoglutamatergic disorder? Relevance of glutamate–serotonin interactions for pharmacotherapy. *J. Neural Transm.* 1998, 105, 525–535.
28. Comery, T.A.; Harris, J.B.; Willems, P.J.; Oostra, B.A.; Irwin, S.A.; Weiler, I.J.; Greenough, W.T. Abnormal dendritic spines in fragile X knockout mice: Maturation and pruning deficits. *Proc. Natl. Acad. Sci. USA* 1997, 94, 5401–5404.
29. Südhof, T.C.; Malenka, R.C. Understanding synapses: Past, present, and future. *Neuron* 2008, 60, 469–476.
30. Fedder, K.N.; Sabo, S.L. On the role of glutamate in presynaptic development: Possible contributions of presynaptic NMDA receptors. *Biomolecules* 2015, 5, 3448–3466.
31. Nicoletti, F.; Bockaert, J.; Collingridge, G.L.; Conn, P.J.; Ferraguti, F.; Schoepp, D.D.; Wroblewski, J.T.; Pin, J.P. Metabotropic glutamate receptors: From the workbench to the bedside. *Neuropharmacology* 2011, 60, 1017–1041.
32. Catania, M.V.; D'Antoni, S.; Bonaccorso, C.M.; Aronica, E.; Bear, M.F.; Nicoletti, F. Group I metabotropic glutamate receptors: A role in neurodevelopmental disorders? *Mol. Neurobiol.* 2007, 35, 298–307.

33. Dong, X.; Wang, Y.; Qin, Z. Molecular mechanisms of excitotoxicity and their relevance to pathogenesis of neurodegenerative diseases. *Acta Pharmacol. Sin.* 2009, 30, 379–387.
34. Egbenya, D.L.; Aidoo, E.; Kyei, G. Glutamate receptors in brain development. *Childs Nerv. Syst.* 2021, 37, 2753–2758.
35. Maragakis, N.J.; Rothstein, J.D. Glutamate transporters: Animal models to neurologic disease. *Neurobiol. Dis.* 2004, 15, 461–473.
36. Behuet, S.; Cremer, J.N.; Cremer, M.; Palomero-Gallagher, N.; Zilles, K.; Amunts, K. Developmental changes of glutamate and GABA receptor densities in wistar rats. *Front. Neuroanat.* 2019, 13, 100.
37. Pellegrini-Giampietro, D.E.; Gorter, J.A.; Bennett, M.V.; Zukin, R.S. The GluR2 (GluR-B) hypothesis: Ca(2+)-permeable AMPA receptors in neurological disorders. *Trends Neurosci.* 1997, 20, 464–470.
38. Pandey, S.P.; Rai, R.; Gaur, P.; Prasad, S. Development- and age-related alterations in the expression of AMPA receptor subunit GluR2 and its trafficking proteins in the hippocampus of male mouse brain. *Biogerontology* 2015, 16, 317–328.
39. Paupard, M.C.; Friedman, L.K.; Zukin, R.S. Developmental regulation and cell-specific expression of N-methyl-D-aspartate receptor splice variants in rat hippocampus. *Neuroscience* 1997, 79, 399–409.
40. Waters, K.A.; Machaalani, R. NMDA receptors in the developing brain and effects of noxious insults. *Neurosignals* 2004, 13, 162–174.
41. Szczurowska, E.; Mareš, P. NMDA and AMPA receptors: Development and status epilepticus. *Physiol. Res.* 2013, 62 (Suppl. 1), S21–S38.
42. Strunecka, A.; Blaylock, R.L.; Patocka, J.; Strunecky, O. Immunoexcitotoxicity as the central mechanism of etiopathology and treatment of autism spectrum disorders: A possible role of fluoride and aluminum. *Surg. Neurol. Int.* 2018, 9, 74.
43. Holopainen, I.E.; Laurén, H.B. Glutamate signaling in the pathophysiology and therapy of prenatal insults. *Pharmacol. Biochem. Behav.* 2012, 100, 825–834.
44. Bahn, S.; Volk, B.; Wisden, W. Kainate receptor gene expression in the developing rat brain. *J. Neurosci.* 1994, 14, 5525–5547.
45. Lauri, S.E.; Segerstråle, M.; Vesikansa, A.; Maingret, F.; Mulle, C.; Collingridge, G.L.; Isaac, J.T.R.; Taira, T. Endogenous activation of kainate receptors regulates glutamate release and network activity in the developing hippocampus. *J. Neurosci.* 2005, 25, 4473–4484.
46. Jansson, L.C.; Åkerman, K.E. The role of glutamate and its receptors in the proliferation, migration, differentiation and survival of neural progenitor cells. *J. Neural Transm.* 2014, 121,

819–836.

47. Chaudhury, S.; Sharma, V.; Kumar, V.; Nag, T.C.; Wadhwa, S. Activity-dependent synaptic plasticity modulates the critical phase of brain development. *Brain Dev.* 2016, 38, 355–363.
48. Holtmaat, A.; Svoboda, K. Experience-dependent structural synaptic plasticity in the mammalian brain. *Nat. Rev. Neurosci.* 2009, 10, 647–658.
49. Gaiarsa, J.-L.; Caillard, O.; Ben-Ari, Y. Long-term plasticity at GABAergic and glycinergic synapses: Mechanisms and functional significance. *Trends Neurosci.* 2002, 25, 564–570.
50. Bak, L.K.; Schousboe, A.; Waagepetersen, H.S. The glutamate/GABA-glutamine cycle: Aspects of transport, neurotransmitter homeostasis and ammonia transfer. *J. Neurochem.* 2006, 98, 641–653.
51. Kim, S.; Kwon, S.-H.; Kam, T.-I.; Panicker, N.; Karuppagounder, S.S.; Lee, S.; Lee, J.H.; Kim, W.R.; Kook, M.; Foss, C.A.; et al. Transneuronal Propagation of Pathologic α -Synuclein from the Gut to the Brain Models Parkinson's Disease. *Neuron* 2019, 103, 627–641.e7.
52. Kim, J.-W.; Park, K.; Kang, R.J.; Gonzales, E.L.T.; Kim, D.G.; Oh, H.A.; Seung, H.; Ko, M.J.; Kwon, K.J.; Kim, K.C.; et al. Pharmacological modulation of AMPA receptor rescues social impairments in animal models of autism. *Neuropsychopharmacology* 2019, 44, 314–323.
53. McCarthy, M.M.; Davis, A.M.; Mong, J.A. Excitatory neurotransmission and sexual differentiation of the brain. *Brain Res. Bull.* 1997, 44, 487–495.
54. Giacometti, L.L.; Barker, J.M. Sex differences in the glutamate system: Implications for addiction. *Neurosci. Biobehav. Rev.* 2020, 113, 157–168.
55. Sailasuta, N.; Ernst, T.; Chang, L. Regional variations and the effects of age and gender on glutamate concentrations in the human brain. *Magn. Reson. Imaging* 2008, 26, 667–675.
56. Grachev, I.D.; Apkarian, A.V. Chemical heterogeneity of the living human brain: A proton MR spectroscopy study on the effects of sex, age, and brain region. *Neuroimage* 2000, 11, 554–563.
57. Zahr, N.M.; Mayer, D.; Rohlfing, T.; Chanraud, S.; Gu, M.; Sullivan, E.V.; Pfefferbaum, A. In vivo glutamate measured with magnetic resonance spectroscopy: Behavioral correlates in aging. *Neurobiol. Aging* 2013, 34, 1265–1276.
58. O'Gorman, R.L.; Michels, L.; Edden, R.A.; Murdoch, J.B.; Martin, E. In vivo detection of GABA and glutamate with MEGA-PRESS: Reproducibility and gender effects. *J. Magn. Reson. Imaging* 2011, 33, 1262–1267.
59. Zlotnik, A.; Gruenbaum, B.F.; Mohar, B.; Kuts, R.; Gruenbaum, S.E.; Ohayon, S.; Boyko, M.; Klin, Y.; Sheiner, E.; Shaked, G.; et al. The effects of estrogen and progesterone on blood glutamate levels: Evidence from changes of blood glutamate levels during the menstrual cycle in women. *Biol. Reprod.* 2011, 84, 581–586.

60. Hädel, S.; Wirth, C.; Rapp, M.; Gallinat, J.; Schubert, F. Effects of age and sex on the concentrations of glutamate and glutamine in the human brain. *J. Magn. Reson. Imaging* 2013, 38, 1480–1487.
61. Kouchiwa, T.; Wada, K.; Uchiyama, M.; Kasezawa, N.; Niisato, M.; Murakami, H.; Fukuyama, K.; Yokogoshi, H. Age-related changes in serum amino acids concentrations in healthy individuals. *Clin. Chem. Lab. Med.* 2012, 50, 861–870.
62. Frankfurt, M.; Fuchs, E.; Wuttke, W. Sex differences in γ -aminobutyric acid and glutamate concentrations in discrete rat brain nuclei. *Neurosci. Lett.* 1984, 50, 245–250.
63. Zuena, A.R.; Mairesse, J.; Casolini, P.; Cinque, C.; Alemà, G.S.; Morley-Fletcher, S.; Chiodi, V.; Spagnoli, L.G.; Gradini, R.; Catalani, A.; et al. Prenatal restraint stress generates two distinct behavioral and neurochemical profiles in male and female rats. *PLoS ONE* 2008, 3, e2170.
64. Anderson, D.W.; Mettil, W.A.; Schneider, J.S. Rearing environment, sex and developmental lead exposure modify gene expression in the hippocampus of behaviorally naïve animals. *Neurochem. Int.* 2013, 62, 510–520.
65. Higuera-Matas, A.; Miguéns, M.; Coria, S.M.; Assis, M.A.; Borcel, E.; del Olmo, N.; Ambrosio, E. Sex-specific disturbances of the glutamate/GABA balance in the hippocampus of adult rats subjected to adolescent cannabinoid exposure. *Neuropharmacology* 2012, 62, 1975–1984.
66. Wang, Y.; Ma, Y.; Hu, J.; Cheng, W.; Jiang, H.; Zhang, X.; Li, M.; Ren, J.; Li, X. Prenatal chronic mild stress induces depression-like behavior and sex-specific changes in regional glutamate receptor expression patterns in adult rats. *Neuroscience* 2015, 301, 363–374.
67. Monfort, P.; Gomez-Gimenez, B.; Llansola, M.; Felipe, V. Gender differences in spatial learning, synaptic activity, and long-term potentiation in the hippocampus in rats: Molecular mechanisms. *ACS Chem. Neurosci.* 2015, 6, 1420–1427.

Retrieved from <https://encyclopedia.pub/entry/history/show/53500>