

# Insect-Specific Viruses (ISVs)

Subjects: Virology  
Contributor: Ahmed Mohamed

Unlike arboviruses, which have a dual-host tropism by cycling between vertebrate hosts and arthropod vectors, ISVs replicate exclusively in arthropod populations, causing a persistent viral infection, as such they are mainly maintained in nature by vertical transmission route. ISVs are also refereed to as mosquito-specific viruses as they are generally identified and discovered in mosquitoes. They are nonetheless an important part of the mosquito microbiome. The first ISV identified is cell-fusing agent virus (CFAV), which was isolated from an *Aedes aegypti* (*Ae. aegypti*) cell culture.

Keywords: insect-specific viruses ; Mosquito-specific viruses ; Flaviviruses ; Host-restriction ; Mosquitoes ; biocontrol agents

## 1. Introduction

Since the discovery of CFAV, wild mosquito populations have been shown to act as a reservoir for a wide variety of ISVs, which suggests a significant heterogeneity among these viruses. These viruses are characterized by their competency to replicate in their vectors, while their replication are restricted in vertebrate cells.<sup>[1]</sup> Thanks to the advent of next-generation sequencing applications and advanced bioinformatics tools, metagenomic studies in this scope have identified a large number of ISVs harboring wild-caught mosquitoes over wide geographical areas (reviewed in <sup>[2]</sup>). Phylogenetic analyses based on sequence identities between ISVs and other mosquito-borne viruses suggest strong evidence that ISVs may be ancestral to arboviruses. Thereby, ISVs could serve as a model to study arbovirus evolution and their transition from single- to dual-host identity.

## 2. Classification of ISVs

ISVs have been classified within multiple different taxa, mostly in the family Flaviviridae and the order Bunyavirales (Table 1). The Togaviridae, Rhabdoviridae, and Mesoniviridae also contain a smaller number of ISVs, as well as other taxa.<sup>[3]</sup>

Table 1. Classification of some insect-specific viruses.

Taxa	Genus	ISV	Host <sup>1</sup>	Reference
Flaviviridae	Flavivirus	Binjari virus	<i>Aedes normanensis</i>	<sup>[4]</sup>
		Cell fusing agent virus	<i>Aedes</i> spp.	<sup>[5]</sup>
		Kamiti River virus	<i>Aedes macintoshi</i>	<sup>[6]</sup>
		Niénokoué virus	<i>Culex</i> spp.	<sup>[7]</sup>
		Palm Creek virus	<i>Coquillettidia xanthogaster</i>	<sup>[8]</sup>
	Phasivirus	Parramatta River virus	<i>Aedes vigilax</i>	<sup>[9]</sup>
		Phasi Charoen-like phasivirus	<i>Aedes aegypti</i>	<sup>[10]</sup>
Bunyavirales	Orthoferavirus	Ferak orthoferavirus	<i>Culex decens</i>	<sup>[11]</sup>
	Goukovirus	Gouléako goukovirus	<i>Anopheles</i> spp. <i>Culex</i> spp. <i>Uranotaenia</i> spp.	<sup>[12]</sup>
	Herbevirus	Herbert herbevirus	<i>Culex nebulosus</i>	<sup>[13]</sup>
	Orthojonvirus	Jonchet orthojonvirus	<i>Culex</i> spp.	<sup>[11]</sup>
Birnaviridae	Entomobirnavirus	Espirito Santo virus	N/A	<sup>[14]</sup>

Taxa	Genus	ISV	Host <sup>1</sup>	Reference
Mesoniviridae	Alphamesonivirus 1	Cavally virus	Aedes spp. Anopheles spp. Culex spp. Uranotaenia spp.	[15]
		Nam Dinh virus	Culex spp. Aedes albopictus	[16][17]
		Dianke virus	Aedes spp. Anopheles spp. Culex spp. Mansonia spp. Uranotaenia spp. ceratopogonids	[18]
Reoviridae	Dinovernavirus	Fako virus	Aedes spp. Eretmapodites spp.	[19]
Rhabdoviridae	Almendravirus	Arboretum almindavirus	Ochlerotatusfulvus	[20]
	Mousrhavirus	Moussa Mousrhavirus	Culex decens	[21]
		Agua Salud alphavirus	Culex declarator	[22]
Togaviridae	Alphavirus	Eilat virus	Anopheles coustani	[23]
		Yada yada virus	N/A	[24]

<sup>1</sup> The host range of insect-specific viruses identified up to date. N/A the vector species of ISV are not determined yet.

### 3. ISV maintenance in nature

The vertical transmission routes (transovarial or transovum transmission) are considered the primary means by which ISVs are maintained and propagated in their vector populations.<sup>[26][27]</sup> They have also been shown to be venereally transmitted, yet to a lesser extent.<sup>[28]</sup> In addition, ISVs such as Eilat virus (EILV) and Negev virus could be experimentally transmitted to adult mosquitoes via a high-titer of an infectious blood meal.<sup>[29][30]</sup>

### 4. Host Restriction of ISVs

Viral tropism is fundamental to consider with regards to host restriction of ISVs. It is defined as the ability of a particular virus to productively infect and replicate in a specific cell type, tissue, or species.<sup>[25]</sup> Viral tropism is determined by host cell susceptibility and permissiveness. For an efficient viral replication cycle in the host cell, the virus should be able to interact with multiple host factors at each step of its replicative cycle and to antagonize the host immune response that hinders its replication. Consequently, ISVs must overcome several integrated bottlenecks present at different levels to potentially emerge as a new dual-host virus: their genetic determinants, the vertebrate host factors, and the host microenvironment needed for efficient replication (Figure 1) (reviewed in <sup>[1]</sup>).

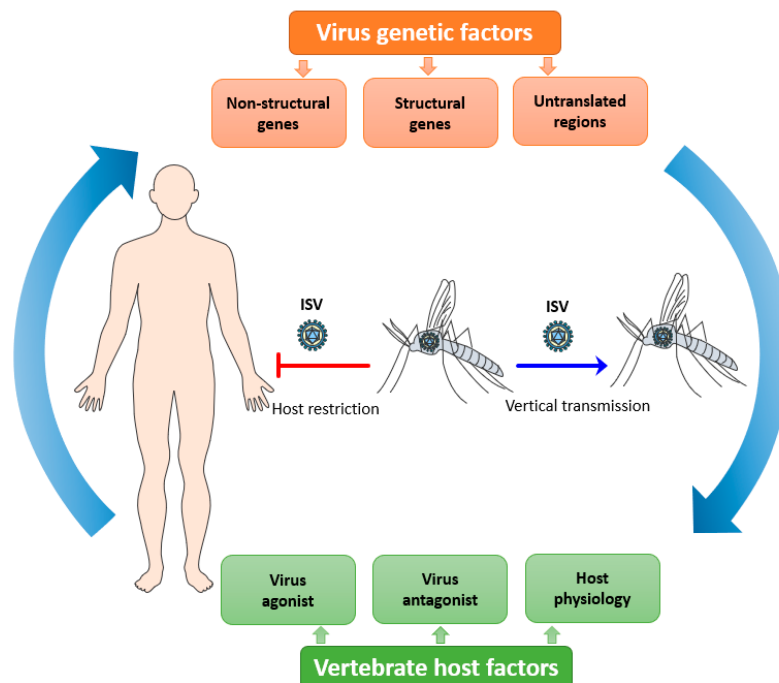


Figure 1. Putative overview of the host tropism of insect-specific viruses (ISVs). ISVs are maintained in mosquito populations by a vertical route of transmission. The infection and replication of ISVs are restricted in vertebrate hosts due to the complex interplay between multiple viral, host, and microenvironmental factors.<sup>[1]</sup>

## 5. Potential applications of ISVs

### 5.1. Novel biocontrol agents

Since ISV discovery in natural mosquito populations as circulating viruses more attention has been given to their role in modulating pathogenic arbovirus transmission. Recently, different studies have focused on ISV interactions with dual-host arboviruses by investigating the phenomenon of superinfection exclusion, reviewed in <sup>[2][31][32]</sup>. Accordingly, ISVs can modulate the replication or dissemination of dual-host viruses in their competent vectors.<sup>[33]</sup>

### 5.2. Vaccine and diagnostic platforms

Thanks to ISV host-restriction in vertebrate cells, ISVs have been manipulated as vaccine and diagnostic platforms. Additionally, the similarity between ISVs and medically-important arboviruses in gene structure and order have paved the way to generate chimeras that could harbor structural proteins of these arboviruses. Consequently, the dual-host structural proteins could maintain the native structure and conformation required for vaccine and diagnostic platforms. In this regard, Eilat virus and Binjari virus have been utilized as recombinant platforms for vaccine and diagnostic development.<sup>[34][35][36]</sup>

## References

1. Ahmed Me Elrefaey; Rana Abdelnabi; Ana Lucia Rosales Rosas; Lanjiao Wang; Sanjay Basu; Leen Delang; Understanding the Mechanisms Underlying Host Restriction of Insect-Specific Viruses. *Viruses* **2020**, *12*, 964, [10.3390/v12090964](https://doi.org/10.3390/v12090964).
2. Eric Agboli; Mayke Leggewie; Mine Altinli; Esther Schnettler; Mosquito-Specific Viruses—Transmission and Interaction. *Viruses* **2019**, *11*, 873, [10.3390/v11090873](https://doi.org/10.3390/v11090873).
3. Charles H. Calisher; Stephen Higgs; The Discovery of Arthropod-Specific Viruses in Hematophagous Arthropods: An Open Door to Understanding the Mechanisms of Arbovirus and Arthropod Evolution?. *Annual Review of Entomology* **2018**, *63*, 87-103, [10.1146/annurev-ento-020117-043033](https://doi.org/10.1146/annurev-ento-020117-043033).
4. Jessica J. Harrison; Jody Hobson-Peters; Agathe M. G. Colmant; Joanna Koh; Natalee D. Newton; David Warrilow; Hel le Bielefeldt-Ohmann; Thisun B. H. Piyasena; Caitlin A. O'Brien; Laura J. Vet; et al. Antigenic Characterization of New Lineage II Insect-Specific Flaviviruses in Australian Mosquitoes and Identification of Host Restriction Factors. *mSphere* **2020**, *5*, 1, [10.1128/msphere.00095-20](https://doi.org/10.1128/msphere.00095-20).

5. Victor Stollar; Virginia L. Thomas; An agent in the *Aedes aegypti* cell line (Peleg) which causes fusion of *Aedes albopictus* cells. *Virology* **1975**, 64, 367-377, [10.1016/0042-6822\(75\)90113-0](https://doi.org/10.1016/0042-6822(75)90113-0).
6. M. B. Crabtree; R. C. Sang; V. Stollar; L. M. Dunster; B. R. Miller; Genetic and phenotypic characterization of the newly described insect flavivirus, Kamiti River virus. *Archives of Virology* **2003**, 148, 1095-1118, [10.1007/s00705-003-0019-7](https://doi.org/10.1007/s00705-003-0019-7).
7. Rebecca Halbach; Sandra Junglen; Ronald P. Van Rij; Mosquito-specific and mosquito-borne viruses: evolution, infection, and host defense. *Current Opinion in Insect Science* **2017**, 22, 16-27, [10.1016/j.cois.2017.05.004](https://doi.org/10.1016/j.cois.2017.05.004).
8. Jody Hobson-Peters; Alice Wei Yee Yam; Jennifer Lu; Yin Xiang Setoh; Fiona May; Nina Kurucz; Susan Walsh; Natalie A. Prow; Steven S. Davis; Richard Weir; et al. A New Insect-Specific Flavivirus from Northern Australia Suppresses Replication of West Nile Virus and Murray Valley Encephalitis Virus in Co-infected Mosquito Cells. *PLOS ONE* **2013**, 8, e56534, [10.1371/journal.pone.0056534](https://doi.org/10.1371/journal.pone.0056534).
9. Breeanna J. McLean; Jody Hobson-Peters; Cameron E. Webb; Daniel Watterson; Natalie A. Prow; Hong Duyen Nguyen; Sonja Hall-Mendelin; David Warrilow; Cheryl A. Johansen; Cassie C. Jansen; et al. A novel insect-specific flavivirus replicates only in *Aedes*-derived cells and persists at high prevalence in wild *Aedes vigilax* populations in Sydney, Australia. *Virology* **2015**, 486, 272-283, [10.1016/j.virol.2015.07.021](https://doi.org/10.1016/j.virol.2015.07.021).
10. Xiaomin Zhang; Suibin Huang; Tao Jin; Peng Lin; Yalan Huang; Chunli Wu; Bo Peng; Lan Wei; Hin Chu; Miao Wang; et al. Discovery and high prevalence of Phasi Charoen-like virus in field-captured *Aedes aegypti* in South China. *Virology* **2018**, 523, 35-40, [10.1016/j.virol.2018.07.021](https://doi.org/10.1016/j.virol.2018.07.021).
11. Marco Marklewitz; Florian Zirkel; Andreas Kurth; Christian Drosten; Sandra Junglen; Evolutionary and phenotypic analysis of live virus isolates suggests arthropod origin of a pathogenic RNA virus family. *Proceedings of the National Academy of Sciences* **2015**, 112, 7536-7541, [10.1073/pnas.1502036112](https://doi.org/10.1073/pnas.1502036112).
12. Marco Marklewitz; S. Handrick; W. Grasse; A. Kurth; Alexander N. Lukashev; C. Drosten; H. Ellerbrok; F. H. Leendertz; G. Pauli; Sandra Junglen; et al. Gouleako Virus Isolated from West African Mosquitoes Constitutes a Proposed Novel Genus in the Family Bunyaviridae. *Journal of Virology* **2011**, 85, 9227-9234, [10.1128/jvi.00230-11](https://doi.org/10.1128/jvi.00230-11).
13. Marco Marklewitz; Florian Zirkel; Innocent B. Rwego; Hanna Heidemann; Pascal Trippner; Andreas Kurth; René Kallies; Thomas Briesse; W. Ian Lipkin; Christian Drosten; et al. Discovery of a Unique Novel Clade of Mosquito-Associated Bunyaviruses. *Journal of Virology* **2013**, 87, 12850-12865, [10.1128/jvi.01862-13](https://doi.org/10.1128/jvi.01862-13).
14. Ricardo Vancini; Angel Paredes; Mariana Ribeiro; Kevin Blackburn; Davis Ferreira; Joseph P. Kononchik; Raquel Hernandez; Dennis T. Brown; Espirito Santo Virus: a New Birnavirus That Replicates in Insect Cells. *Journal of Virology* **2011**, 86, 2390-2399, [10.1128/jvi.06614-11](https://doi.org/10.1128/jvi.06614-11).
15. Florian Zirkel; Andreas Kurth; Phenix-Lan Quan; Thomas Briesse; Heinz Ellerbrok; Georg Pauli; Fabian H. Leendertz; W. Ian Lipkin; John Ziebuhr; Christian Drosten; et al. An Insect Nidovirus Emerging from a Primary Tropical Rainforest. *mBio* **2011**, 2, 1, [10.1128/mbio.00077-11](https://doi.org/10.1128/mbio.00077-11).
16. [1371/journal.ppat.1002215](https://doi.org/10.1371/journal.ppat.1002215)
17. [1186/1743-422X-11-97](https://doi.org/10.1186/1743-422X-11-97)
18. Moussa Moïse Diagne; Alioune Gaye; Marie Henriette Dior Ndione; Martin Faye; Gamou Fall; Idrissa Dieng; Steven G. Widen; Thomas G. Wood; Vsevolod Popov; Hilda Guzman; et al. Dianke virus: A new mesonivirus species isolated from mosquitoes in Eastern Senegal. *Virus Research* **2019**, 275, 197802, [10.1016/j.virusres.2019.197802](https://doi.org/10.1016/j.virusres.2019.197802).
19. Albert J. Augustine; Jason T. Kaelber; Eric B. Fokam; Hilda Guzman; Christine V. F. Carrington; Jesse H. Erasmus; Basile Kamgang; Vsevolod L. Popov; Joanita Jakana; Xiangnan Liu; et al. A Newly Isolated Reovirus Has the Simplest Genomic and Structural Organization of Any Reovirus. *Journal of Virology* **2014**, 89, 676-687, [10.1128/jvi.02264-14](https://doi.org/10.1128/jvi.02264-14).
20. Nikos Vasilakis; Fanny Castro-Llanos; Steven G. Widen; Patricia V. Aguilar; Hilda Guzman; Carolina Guevara; Roberto Fernandez; Albert J. Augustine; Thomas G. Wood; Vsevolod Popov; et al. Arboretum and Puerto Almendras viruses: two novel rhabdoviruses isolated from mosquitoes in Peru. *Journal of General Virology* **2014**, 95, 787-792, [10.1099/vir.0.058685-0](https://doi.org/10.1099/vir.0.058685-0).
21. Phenix-Lan Quan; S. Junglen; Alla Tashmukhamedova; Sean Conlan; Stephen K. Hutchison; Andreas Kurth; Heinz Ellerbrok; Michael Egholm; Thomas Briesse; Fabian H. Leendertz; et al. Moussa virus: A new member of the Rhabdoviridae family isolated from *Culex decens* mosquitoes in Côte d'Ivoire. *Virus Research* **2010**, 147, 17-24, [10.1016/j.virusres.2009.09.013](https://doi.org/10.1016/j.virusres.2009.09.013).
22. Kyra Hermanns; Marco Marklewitz; Florian Zirkel; Gijs J. Overheul; Rachel A. Page; Jose R. Loaiza; Christian Drosten; Ronald P. Van Rij; Sandra Junglen; Agua Salud alphavirus defines a novel lineage of insect-specific alphaviruses discovered in the New World. *Journal of General Virology* **2020**, 101, 96-104, [10.1099/jgv.0.001344](https://doi.org/10.1099/jgv.0.001344).
23. Farooq Nasar; Gustavo Palacios; Rodion V. Gorchakov; Hilda Guzman; Amelia P. Travassos Da Rosa; Nazir Savji; Vsevolod L. Popov; Michael B. Sherman; W. Ian Lipkin; Robert B. Tesh; et al. Eilat virus, a unique alphavirus with host range

- e restricted to insects by RNA replication. *Proceedings of the National Academy of Sciences* **2012**, 109, 14622-14627, [10.1073/pnas.1204787109](https://doi.org/10.1073/pnas.1204787109).
24. Jana Batovska; Jan P. Buchmann; Edward C. Holmes; Stacey E. Lynch; Coding-Complete Genome Sequence of Yada Yada Virus, a Novel Alphavirus Detected in Australian Mosquitoes. *Microbiology Resource Announcements* **2020**, 9, 1, [10.1128/mra.01476-19](https://doi.org/10.1128/mra.01476-19).
  25. Grant McFadden; Mohamed R. Mohamed; Masmudur M. Rahman; Eric Barte; Cytokine determinants of viral tropism. *Nature Reviews Immunology* **2009**, 9, 645-655, [10.1038/nri2623](https://doi.org/10.1038/nri2623).
  26. Maria Angelica Contreras-Gutierrez; Hilda Guzman; Saravanan Thangamani; Nikos Vasilakis; Robert B. Tesh; Experimental Infection with and Maintenance of Cell Fusing Agent Virus (Flavivirus) in *Aedes aegypti*. *The American Journal of Tropical Medicine and Hygiene* **2017**, 97, 299-304, [10.4269/ajtmh.16-0987](https://doi.org/10.4269/ajtmh.16-0987).
  27. Joel J. L. Lutomiah; Charles Mwandawiro; Japhet Magambo; Rosemary Sang; Infection and Vertical Transmission of Karamiti River Virus in Laboratory Bred *Aedes aegypti* Mosquitoes. *Journal of Insect Science* **2007**, 7, 1-7, [10.1673/031.007.5501](https://doi.org/10.1673/031.007.5501).
  28. Bethany G. Bolling; Francisco J. Olea-Popelka; Lars Eisen; Chester G. Moore; Carol D. Blair; Transmission dynamics of an insect-specific flavivirus in a naturally infected *Culex pipiens* laboratory colony and effects of co-infection on vector competence for West Nile virus. *Virology* **2012**, 427, 90-97, [10.1016/j.virol.2012.02.016](https://doi.org/10.1016/j.virol.2012.02.016).
  29. Farooq Nasar; Andrew D. Haddow; Robert B. Tesh; Scott C Weaver; Eilat virus displays a narrow mosquito vector range. *Parasites & Vectors* **2014**, 7, 595, [10.1186/s13071-014-0595-2](https://doi.org/10.1186/s13071-014-0595-2).
  30. Nikos Vasilakis; Naomi L. Forrester; Gustavo Palacios; Farooq Nasar; Nazir Savji; Shannan L. Rossi; Hilda Guzman; Thomas G. Wood; Vsevolod Popov; Rodion Gorchakov; et al. Negevirus: a Proposed New Taxon of Insect-Specific Viruses with Wide Geographic Distribution. *Journal of Virology* **2012**, 87, 2475-2488, [10.1128/jvi.00776-12](https://doi.org/10.1128/jvi.00776-12).
  31. Pontus Öhlund; Hanna Lundén; Anne-Lie Blomström; Insect-specific virus evolution and potential effects on vector competence. *Virus Genes* **2019**, 55, 127-137, [10.1007/s11262-018-01629-9](https://doi.org/10.1007/s11262-018-01629-9).
  32. Edward I Patterson; Jandouwe Villinger; Joseph N Muthoni; Lucien Dobel-Ober; Grant L Hughes; Exploiting insect-specific viruses as a novel strategy to control vector-borne disease. *Current Opinion in Insect Science* **2020**, 39, 50-56, [10.1016/j.cois.2020.02.005](https://doi.org/10.1016/j.cois.2020.02.005).
  33. Artem Baidaliuk; Elliott F. Miot; Sebastian Lequime; Isabelle Moltini-Conclois; Fanny Delaigue; Stéphanie Dabo; Laura B. Dickson; Fabien Aubry; Sarah H. Merkl; Van-Mai Cao-Lormeau; et al. Cell-Fusing Agent Virus Reduces Arbovirus Dissemination in *Aedes aegypti* Mosquitoes In Vivo. *Journal of Virology* **2019**, 93, e00705-19, [10.1128/jvi.00705-19](https://doi.org/10.1128/jvi.00705-19).
  34. Jesse H. Erasmus; Albert J. Augustine; Jason T. Kaelber; Huanle Luo; Shannan L. Rossi; Karla Fenton; Grace Leal; Dal Young Kim; Wah Chiu; Tian Wang; et al. A chikungunya fever vaccine utilizing an insect-specific virus platform. *Nature Medicine* **2016**, 23, 192-199, [10.1038/nm.4253](https://doi.org/10.1038/nm.4253).
  35. Jesse H. Erasmus; James Needham; Syamal Raychaudhuri; Michael S. Diamond; David W. C. Beasley; Stan Morkowski; Henrik Salje; Ildelfonso Fernandez Salas; Dal Young Kim; Ilya Frolov; et al. Utilization of an Eilat Virus-Based Chimera for Serological Detection of Chikungunya Infection. *PLOS Neglected Tropical Diseases* **2015**, 9, e0004119, [10.1371/journal.pntd.0004119](https://doi.org/10.1371/journal.pntd.0004119).
  36. Jesse H. Erasmus; Robert L. Seymour; Jason T. Kaelber; Dal Young Kim; Grace Leal; Michael B. Sherman; Ilya Frolov; Wah Chiu; Scott C Weaver; Farooq Nasar; et al. Novel insect-specific Eilat virus-based chimeric vaccine candidates provide durable, mono- and multi-valent, single dose protection against lethal alphavirus challenge. *Journal of Virology* **2017**, 92, JVI.01274-17, [10.1128/jvi.01274-17](https://doi.org/10.1128/jvi.01274-17).