# **Asthma Pathogenesis**

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Asthma is a widespread chronic disease of the bronchopulmonary system with a heterogeneous course due to the complex etiopathogenesis. Natural-climatic and anthropogenic factors play an important role in the development and progression of this pathology.

asthma temperature thermosensory transient receptor potential ion channels genes

## 1. Introduction

Asthma is a widespread, chronic heterogeneous disease with a complex etiopathogenesis and constantly improving approaches to therapy. Nowadays, over 350 million people of all ages worldwide suffer from asthma, and about 350 thousand people per year die from the disease. The growing prevalence of this pathology among people of different age groups is of particular concern. Uncontrolled asthma that is diagnosed in 45% of patients with a high frequency of exacerbations and hospitalizations remains an important global health problem <sup>[1]</sup>. In Europe, about 15% of school-age children suffer from asthma; 5% of them have difficult-to-control asthma. According to the Global Burden of Disease Study, asthma occupies the 14th ranking place among the leading causes of disability. The global burden of asthma is associated with direct economic costs and indirect social and economic consequences <sup>[2]</sup>. Obviously, achieving asthma control and reducing exacerbation frequency in patients with the pathology are among the most pressing problems of our time.

According to current research, bronchospasm and asthma-like symptoms in asthma patients can occur under the influence of trigger factors, such as the combination of cold temperatures and high/low air humidity, as well as hot temperatures and high air humidity <sup>[3][4][5]</sup>. The contribution of high and low temperatures in airway inflammation in asthma as well as the main molecular mechanism underlying the disease have not yet been fully understood <sup>[5][6]</sup>. The high sensitivity of the bronchi to environmental factors is caused by impaired regulation of afferent nerves of the respiratory tract and chronic neurogenic inflammation <sup>[2]</sup>. Vagal sensory nerves, of which 75% are non-myelinated nociceptive C-fibers of the lungs, innervate the respiratory tract. In addition, nociceptor neurons are involved in the activation of the bronchoconstrictor mechanism in airway inflammation. The sensitivity of lung nociceptors is mediated by thermosensory channels, members of a subfamily of the transient receptor potential (TRP) ion channels <sup>[8]</sup>. The channels are considered as main sensors of physicochemical environmental stimuli involved in the regulation of body temperature <sup>[5]</sup>. The channels are activated by temperature changes <sup>[9]</sup>. TRP channels are expressed by neuronal cells and cells of the respiratory tract (bronchial epithelium and endothelium, smooth muscle cells, unmyelinated nociceptive C-fibers of the lungs) interves of the lungs) <sup>[10][11][12][13]</sup>. TRP channel activation in the nerve endings of the respiratory tract leads to the stimulation of protective reflexes, but under certain conditions, it

can be the pathogenetic basis of asthma <sup>[12][13][14][15]</sup>. The role of TRP channels in asthma pathogenesis has been demonstrated <sup>[13][16][17][18][19][20][21]</sup>, which makes them promising targets for disease therapy <sup>[12][13]</sup>.

Asthma is initiated not only by the influence of natural and climatic factors but also by the interaction between these factors and the genetic component of the body. However, not all asthma patients react to changes in the temperature and humidity of inhaled air, which may be related to genetic factors <sup>[22]</sup>. Recently, there has been considerable interest in the genetic contribution to asthma pathogenesis <sup>[23][24][25][26][27][28][29][30][31][32]</sup>. Of particular interest is the identification of SNPs of TRP genes and their interaction with physicochemical environmental factors.

#### 2. The Role of Natural-Climatic Factors in Asthma Pathogenesis

In recent years, a number of studies have shown that high and low air temperatures impact on the development and exacerbation of asthma <sup>[6][33][34]</sup>. In addition, the temperature effect is modulated by other environmental conditions (humidity, visibility, cloud cover, air pressure, wind speed, air pollution, etc.). For example, simultaneous exposure to low temperature and poor air quality is an important factor for the appearance of asthma symptoms <sup>[35]</sup>. Cold moist air induces more respiratory symptoms in asthma patients than cold dry air. Therefore, people suffering from this pathology should avoid adverse environmental conditions and should limit their outdoor activities during periods of extreme temperatures, as well as during periods of high humidity combined with low/high temperature and low humidity combined with low temperature <sup>[33]</sup>.

Under normal conditions, nasal breathing partially compensates for the cold air exposure, and low airway symptoms at rest or during light exercise do not occur. Intensive exercise in cold weather generates only a short-term acute bronchospasm and cough symptoms in healthy people <sup>[36]</sup>. Nevertheless, the cold air is an important trigger factor for severe asthma in patients with cold airway hyperresponsiveness. Asthma patients exhibit an increase in the frequency of cold-air-induced symptoms by 50% compared with healthy subjects. The risk of developing respiratory symptoms provoked by cold air (shortness of breath, wheezing, sputum discharge) is more elevated in patients with asthma coexisting with allergic rhinitis <sup>[4]</sup>. M. D'Amato et al. have revealed that breathing of +20 °C air at 15 L/min reduces the temperature of the trachea proximal sections to 34 °C, while breathing the same air at 100 L/min decreases this temperature to 31 °C <sup>[37]</sup>. In addition to bronchoconstriction, cold air hyperventilation also brings about coughing. Cough and bronchospasm are independent reactions since pretreatment with salbutamol blocks bronchoconstriction but does not affect cold-air-induced cough. Additionally, cold air is dry, and consequently, cold air hyperventilation causes airway dehydration, leading to the release of mediators initiating bronchospasm <sup>[5]</sup>.

The effect of conditioned cold air is also injurious to asthma patients. In particular, quick cooling of the indoor air without gradual adaptation to a temperature 2–3 °C lower than the outdoor temperature—and especially with humidity ranging between 40% and 60%—may give rise to asthma exacerbation over several hours or days <sup>[37]</sup>.

There are few data on the negative impact of high ambient temperature on the respiratory tract <sup>[5][38][39][40]</sup>. Additionally, increased metabolic rate (physical activity) and difficult heat dissipation (for example, in a warm environment) are common causes of hyperthermia <sup>[40]</sup>. Junior M.A.V.C. et al. have found that exercise-induced bronchospasm is diagnosed in most asthmatic patients living in a hot dry climate, while only 10% of patients with rhinitis and 5% of healthy people living under the same conditions developed exercise-induced bronchospasm <sup>[38]</sup>. Non-myelinated nociceptive C-fibers of the lungs have been demonstrated to be activated when the intrathoracic temperature was elevated to 39.2 °C. Hot air hyperventilation induces the development of bronchoconstriction in asthma patients, and pretreatment with ipratropium bromide aerosol completely prevented bronchospasm in these patients <sup>[40]</sup>. Furthermore, hot air breathing provokes cough in these patients, which indicates damage to the respiratory nerves. Thus, hyperventilation with hot humid air raises the temperature of the airways and triggers bronchoconstriction in asthma patients by activating pulmonary C-fibers <sup>[40]</sup>.

The association between asthma hospitalizations and air temperature is of interest. It has been established that in the cold season, temperature is inversely correlated with asthma hospital admissions <sup>[33]</sup>. Patients with poorly controlled asthma are more likely to exhibit cold-weather-related respiratory symptoms <sup>[35]</sup>. It is worth noting that the number of hospital admissions among adults depends not only on low temperature but also on high temperature, while this relationship is not observed among children under five years of age. In the hot season, a minimum number of asthma hospitalizations is observed at an ambient temperature of 27 °C; this parameter reaches the maximum at 30 °C and achieves a plateau in the temperature range 30–32 °C. High temperature can have a protective effect for adults but is dangerous for non-adults <sup>[41]</sup>. The amount of evidence suggesting a nonlinear association between air temperature and the number of asthma hospitalizations of asthma temperature and the number of asthma hospitalizations demonstrated high sensitivity to both hot temperature in summer and low temperature in winter <sup>[34]</sup>.

Thus, the combination of cold temperatures and high/low air humidity as well as the combination of hot temperatures and high air humidity lead to bronchospastic reactions and asthma-like symptoms (coughing, wheezing, shortness of breath) in asthma patients, thereby increasing the exacerbation frequency and reducing the control of the disease. It is known that thermosensory TRP channels are responsible for the reception of physicochemical environmental stimuli and the regulation of body temperature <sup>[5][8][9][42][43][44]</sup>. The impairment of their functioning plays an important role in asthma pathogenesis <sup>[16][17][18][19][20][21][45]</sup>, which makes these channels promising targets for disease treatment <sup>[12][46]</sup>.

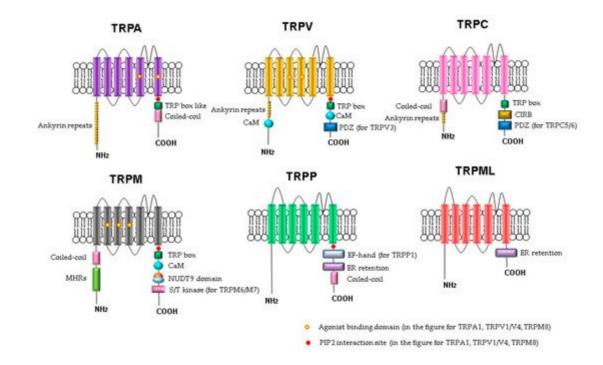
### 3. Transient Receptor Potential Ion Channels

TRP channels belong to the voltage-gated ion channel superfamily (voltage-gated K<sup>+</sup>/Na<sup>+</sup>/Ca<sup>2+</sup> channels, cyclic nucleotide-gated channels) localized mainly on the plasma membrane of cells <sup>[47]</sup>. The main difference between the TRP channel superfamily and other ion channel families is their activation in response to exogenous stimuli, such as temperature, chemicals, light, and sound <sup>[48][49]</sup>. The TRP superfamily consists of 28 channels grouped into

7 subfamilies, in each of which several subunits are distinguished <sup>[50][51]</sup>. The eighth subfamily was recently identified in yeast and named TRPY (**Table 1**).

TRP Channel Subfamily	Subfamily Subunits
canonical receptors	TRPC 1–7
vanilloid receptors	TRPV1–6
polycysteine receptors	TRPP1–3
mucolipin receptors	TRPML1-3
ankyrin-like receptors	TRPA1
melastatin receptors	TRPM1-8
Drosophila NOMPC in mammals	TRPN

The molecular understanding of TRP channels has been improved by structural biology methods <sup>[51]</sup>. Monomers of TRP channels consist of a six-pass transmembrane protein (S1–S6) and a re-entrant loop between S5 and S6, forming the pore or ion conduction pathway (**Figure 1**). The C- and N-termini of the TRP channels are in the cytoplasm. Some of the TRP channels contain ankyrin repeat domains in the N-terminus and a TRP-domain in the C-terminus. The number of ankyrin repeats in the ankyrin domains of the TRP subfamilies differs. For example, TRPC channels contain 3–4 repeats, TRPV channels have 6, TRPA has 14–18 repeats, and TRPN has 29. It is believed that the role of the ankyrin domain is in protein–protein interactions: channel tetramerization and binding to ligands and partner proteins. The ankyrin domains TRPV1, TRPV3, and TRPV4 bind ATP, which competes for binding with calmodulin, a protein that inhibits sensitization of TRPV receptors. The coiled-coil domain is another common motif in the proteins of the TRP family. It can be located both at the C-terminus and at the N-terminus and is involved in cytosolic interactions and binding of polyphosphates.



**Figure 1.** Schematic structure of TRP channels. Monomers of TRP channels consist of six transmembrane segments and a pore-forming return loop between S5 and S6. The intracellular N- and C-termini are distinguished in the length and domains. The TRP box is similar for TRPC, TRPV, TRPM, and a TRP box-like region for the TRPA1 channel. Coiled-coil domains (CC) are located in the C-terminus of TRPA and in the N-terminus of TRPC and TRPM. TRPM share melastatine high homology regions in their N-terminus, (MHRs). The TRPC subfamily of TRP channels has binding domains for calmodulin (CaM), inositol triphosphate receptor binding site (CIRB), and the PDZ-binding specific motif (PDZ). Substrates of NUDT9 are the adenosine 5'-diphosphoribose (ADPR) or the ADPR-2'-phosphate (ADPRP). NUDT9 domain is specific for TRPM2. TRPM6 and TRPM7 contain a serine/threonine kinase domain (S/T kinase). TRPP1 presents a calcium-binding motif (EF-hand), while TRPP and TRPML exhibit an endoplasmic reticulum retention (ER retention) domain. TRP channels are modulated by phosphatidylinositol-4,5-bisphosphate (PIP2).

The operating principle of these receptors is the activation of cation influx in response to stimuli <sup>[47]</sup>. These channels are involved in the processes of apoptosis and proliferation, and they affect vascular tone. TRP channels are expressed primarily by neuronal cells. At the same time, the channels were also found on other cells, particularly in the respiratory tract (bronchial epithelium and endothelium, smooth muscle cells, non-myelinated nociceptive C-fibers of the lungs) <sup>[11][16][18][19][20][43][50][52][53][54][55][56][57][58][59] (**Table 2**).</sup>

Table 2. Tissue or cells expre	essing thermosensory TRP channels.
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TRP Channels	Tissue or Cells	References
TRPA1	T cells	[ <u>11][43][48][49]</u>
	B cells	
	peptidergic and non-peptidergic neurons	
	myelinated Aβ-fibers	
	epithelial cells	

RP Channels	Tissue or Cells	References
	melanocytes	
	mast cells	
	fibroblasts	
	enterochromaffin cells	
	lung cells	
	epithelial cells	
	melanocytes	
	mast cells	
	fibroblasts	
	enterochromaffin cells	
	lung cells	
TRPV1	airway-specific neurons	[ <u>52][53][54]</u>
	C fibres	
	Aδ-fibres	
	central nervous system (neurons, glial cells, astrocytes)	
	pulmonary arteries	
	aorta	
	gastrointestinal tract	
	neuronal and non-neuronal tissues	
TRPV2	pulmonary arteries	[52]
IRPV2	aorta	
	gastrointestinal tract	
	epithelial cells of skin	
	oral cavity	
	gastrointestinal tract	
	tongue	
TRPV3	dorsal root ganglion	[ <u>52</u> ][ <u>55</u> ]
IRPVS	trigeminal ganglion	
	spinal cordbrain	
	lung cells	
	neuronspulmonary arteries	
	aorta	
TRPV4	gastrointestinal tract	[16][47][52][56]
	prostate	
	smooth muscle cells	
	vascular endothelium	
	pulmonary arteries	
	aorta	
	epidermal keratinocyte cells	
	fallopian tubes	
	epithelial cells of the human cornea	
	insulin secreting $\beta$ cells of the pancreas urothelial cells in the renal pelvis	
	insulin secreting picels of the particleas trothelial cells in the renai pelvis	
	ureters	
	ureters	
	ureters urethra	

TRP Channels	Tissue or Cells	References
	macrophages	
	submucosal glands	
	lung cells	
	central nervous system (neurons, glial cells, astrocytes)	
	peripheral nervous systems	
TRPM3	lung cells	
	pulmonary arteries	
	aorta	[50][57]
	cancerous tissues	
	dorsal root ganglia	
	cardiomyocytes	
	insulin secreting $\beta$ cells of the pancreas	
TRPM8	lung cells	
	pulmonary arteries	
	aorta	
	afferent neurons	[ <u>19][20][44][50][58</u> ]
	in cold-sensitive afferents expressed in the upper and lower airways	
	nasal trigeminal neurons	
	retrogradely labelled jugular neurons	
	cancerous tissues	

#### References

- 1. Global Strategy for Asthma Management and Prevention Global Initiative for Asthma (GINA). 2017. Available online: (accessed on 20 August 2018).
- 2. Ferrante, G.; La Grutta, S. The Burden of Pediatric Asthma. Front. Pediatr. 2018, 6, 186.
- 3. Kytikova, O.Y.; Gvozdenko, T.A.; Antonyuk, M.V. Modern aspects of prevalence of chronic bronchopulmonary diseases. Bull. Physiol. Pathol. Respir. 2017, 64, 94–100. (In Russian)
- Hyrkäs, H.; Jaakkola, M.S.; Ikäheimo, T.M.; Hugg, T.T.; Jaakkola, J.J. Asthma and allergic rhinitis increase respiratory symptoms in cold weather among young adults. Respir. Med. 2014, 108, 63– 70.
- 5. Millqvist, E. TRP channels and temperature in airway disease-clinical significance. Temperature 2015, 2, 172–177.
- 6. Xu, Z.; Crooks, J.L.; Davies, J.M.; Khan, A.F.; Hu, W.; Tong, S. The association between ambient temperature and childhood asthma, a systematic review. Int. J. Biometeorol. 2018, 62, 471–481.
- Kytikova, O.Y.; Novgorodtseva, T.P.; Antonyuk, M.V.; Gvozdenko, T.A. The role of regulatory neuropeptides and neurotrophic factors in asthma pathophysiology. Russ. Open Med. J. Ther. 2019, 8, 4.

- 8. De Logu, F.; Patacchini, R.; Fontana, G.; Geppetti, P. TRP functions in the broncho-pulmonary system. Semin. Immunopathol. 2016, 38, 321–329.
- López-Romero, A.E.; Hernández-Araiza, I.; Torres-Quiroz, F.; Tovar-Y-Romo, L.B.D.; Islas, L.; Rosenbaum, T. TRP ion channels, Proteins with conformational flexibility. Channels 2019, 13, 207–226.
- 10. Xia, Y.; Xia, L.; Lou, L.; Jin, R.; Shen, H.; Li, W. Transient receptor potential channels and chronic airway inflammatory diseases, A comprehensive review. Lungs 2018, 196, 505–516.
- 11. Marsakova, L.; Barvik, I.; Zima, V.; Zimova, L.; Vlachova, V. The First Extracellular Linker Is Important for Several Aspects of the Gating Mechanism of Human TRPA1. Channel. Front. Mol. Neurosci. 2017, 10, 16.
- 12. Dietrich, A. Modulators of Transient Receptor Potential (TRP) Channels as Therapeutic Options in Lung Disease. Pharmaceutics 2019, 12, 23.
- 13. Dietrich, A.; Steinritz, D.; Gudermann, T. Transient receptor potential (TRP) channels as molecular targets in lung toxicology and associated diseases. Cell Calcium 2017, 67, 123–137.
- Poveda, J.A.; Giudici, A.M.; Renart, M.L.; Morales, A.; Gonzalez-Ros, J.M. Towards Understanding the Molecular Basis of Ion Channel Modulation by Lipids, Mechanistic Models and Current Paradigms. Biochim. Biophys. Acta Biomembr. 2017, 1859, 1507–1516.
- 15. Taylor-Clark, T.E. Role of reactive oxygen species and TRP channels in the cough reflex. Cell Calcium 2016, 60, 155–162.
- Palaniyandi, S.; Rajendrakumar, A.M.; Periasamy, S.; Goswami, R.; Tuo, W.; Zhu, X.; Rahaman, S.O. TRPV4 is dispensable for the development of airway allergic asthma. Lab. Investig. 2020, 100, 265–273.
- 17. Belvisi, M.G.; Birrell, M.A. The emerging role of transient receptor potential channels in chronic lung disease. Eur. Respir. J. 2017, 50.
- 18. Li, N.; He, Y.; Yang, G.; Yu, Q.; Li, M. Role of TRPC1 channels in pressure-mediated activation of airway remodeling. Respir. Res. 2019, 20, 91.
- 19. Liu, H.; Liu, Q.; Hua, L.; Pan, J. Inhibition of transient receptor potential melastatin 8 alleviates airway inflammation and remodeling in a murine model of asthma with cold air stimulus. Acta Biochim. Biophys. Sin. 2018, 50, 499–506.
- 20. Cheon, D.Y.; Kim, J.H.; Jang, Y.S.; Hwang, Y.I.; Park, S.; Kim, D.G.; Jang, S.H.; Jung, K.S. The activation of transient receptor potential melastatin 8 (TRPM8) receptors of bronchial epithelial cells induces airway inflammation in bronchial asthma. Eur. Respir. J. 2016, 48, PA3997.
- 21. McGarvey, L.P.; Butler, C.A.; Stokesberry, S.; Polley, L.; McQuaid, S.; Abdullah, H.; Heaney, L.G. Increased expression of bronchial epithelial transient receptor potential vanilloid 1 channels in

patients with severe asthma. J. Allergy Clin. Immunol. 2014, 133, 704–712.e4.

- 22. Naumov, D.E.; Perelman, J.M.; Kolosov, V.P.; Potapova, T.A.; Maksimov, V.N.; Zhou, X. Transient receptor potential melastatin 8 gene polymorphism is associated with cold-induced airway hyperresponsiveness in bronchial asthma. Respirol. Ther. 2015, 20, 1192–1197.
- 23. Hernandez-Pacheco, N.; Pino-Yanes, M.; Flores, C. Genomic Predictors of Asthma Phenotypes and Treatment Response. Front. Pediatr. 2019, 7, 6.
- 24. Vicente, C.T.; Revez, J.A.; Ferreira, M.A.R. Lessons from ten years of genome-wide association studies of asthma. Clin. Transl. Immunol. 2017, 6, 165.
- 25. Pickrell, J.K.; Berisa, T.; Liu, J.Z.; Segurel, L.; Tung, J.Y.; Hinds, D.A. Detection and interpretation of shared genetic influences on 42 human traits. Nat. Genet. 2016, 48, 709–717.
- 26. Li, M.; Fan, X.; Ji, L.; Fan, Y.; Xu, L. Exacerbating effects of trimellitic anhydride in ovalbumininduced asthmatic mice and the gene and protein expressions of TRPA1; TRPV1; TRPV2 in lung tissue. Int. Immunopharmacol. 2019, 69, 159–168.
- 27. Gallo, V.; Dijk, F.N.; Holloway, J.W.; Ring, S.M.; Koppelman, G.H.; Postma, D.S.; Strachan, D.P.; Granell, R.; de Jongste, J.C. TRPA1 gene polymorphisms and childhood asthma. Pediatr. Allergy Immunol. 2017, 28, 191–198.
- 28. Bønnelykke, K.; Ober, C. Leveraging gene-environment interactions and endotypes for asthma gene discovery. J. Allergy Clin. Immunol. 2016, 13, 667–679.
- 29. Ober, C. Asthma Genetics in the Post-GWAS Era. Ann. Am. Thorac. Soc. 2016, 13, S85–90.
- 30. Willis-Owen, S.A.G.; Cookson, W.O.C.; Moffatt, M.F. The genetics and genomics of asthma. Ann. Rev. Genom. Hum. Genet. 2018, 19, 223–246.
- 31. Moheimani, F.; Hsu, A.C.; Reid, A.T.; Williams, T.; Kicic, A.; Stick, S.M.; Knight, D.A. The genetic and epigenetic landscapes of the epithelium in asthma. Respir. Res. 2016, 17, 119.
- 32. Moffatt, M.F.; Gut, I.G.; Demenais, F.; Strachan, D.P.; Bouzigon, E.; Heath, S.; von Mutius, E.; Farrall, M.; Lathrop, M.; Cookson, W.; et al. A large-scale; consortium-based genomewide association study of asthma. N. Engl. J. Med. 2010, 363, 1211–1221.
- Lam, H.C.; Li, A.M.; Chan, E.Y.; Goggins, W.B., 3rd. The short-term association between asthma hospitalisations; ambient temperature; other meteorological factors and air pollutants in Hong Kong, a time-series study. Thorax 2016, 71, 1097–1109.
- Lam, H.C.Y.; Hajat, S.; Chan, E.Y.Y.; Goggins, W.B. Different sensitivities to ambient temperature between first- and re-admission childhood asthma cases in Hong Kong-A time series study. Environ. Res. 2019, 170, 487–492.

- 35. Hyrkäs, H.; Ikäheimo, T.M.; Jaakkola, J.J.; Jaakkola, M.S. Asthma control and cold weatherrelated respiratory symptoms. Respir. Med. 2016, 113, 1–7.
- 36. Frischhut, C.; Kennedy, M.D.; Niedermeier, M.; Faulhaber, M. Effects of a heat and moisture exchanger on respiratory function and symptoms post-cold air exercise. Scand. J. Med. Sci. Sports 2019.
- 37. D'Amato, M.; Molino, A.; Calabrese, G.; Cecchi, L.; Annesi-Maesano, I.; D'Amato, G. The impact of cold on the respiratory tract and its consequences to respiratory health. Clin. Transl. Allergy 2018, 8, 20.
- 38. Correia Junior, M.A.V.; Costa, E.C.; Sarinho, S.W.; Rizzo, J.Â.; Sarinho, E.S.C. Exercise-induced bronchospasm in a hot and dry region, study of asthmatic; rhinitistic and asymptomatic adolescents. Expert Rev. Respir. Med. 2017, 11, 1013–1019.
- 39. Bullone, M.; Murcia, R.Y.; Lavoie, J.P. Environmental heat and airborne pollen concentration are associated with increased asthma severity in horses. Equine Vet. J. 2016, 48, 479–484.
- 40. Don Hayes, J.R.; Paul, B.C.; Mehdi, K.; Ruei-Lung, L.; Lu-Yuan, L. Bronchoconstriction Triggered by Breathing Hot Humid Air in Patients with Asthma. Am. J. Respir. Crit. Care Med. 2012, 185, 1190–1196.
- Chan, T.C.; Hu, T.H.; Chu, Y.H.; Hwang, J.S. Assessing effects of personal behaviors and environmental exposure on asthma episodes, a diary-based approach. BMC Pulm. Med. 2019, 19, 231.
- 42. Li, L.; Hongmei, Y.; Jing, T.; Wei, L.; Xinxin, Y.; Fang, Y.; Qiaoli, C.; Jiaxing, X.; Nanshan, Z.; Kian, F.C.; et al. Heterogeneity of cough hypersensitivity mediated by TRPV1 and TRPA1 in patients with chronic refractory cough. Respir. Res. 2019, 20, 112.
- Shapiro, D.; Deering-Rice, C.E.; Romero, E.G.; Hughen, R.W.; Light, A.R.; Veranth, J.M.; Bevans, T.S.; Phan, Q.M.; Reilly, C.A. Activation of transient receptor potential ankyrin-1 (TRPA1) in lung cells by wood smoke particulate material. Chem. Res. Toxicol. 2013, 26, 750–758.
- 44. Liu, H.; Hua, L.; Liu, Q.; Pan, J.; Bao, Y. Cold Stimuli Facilitate Inflammatory Responses through Transient Receptor Potential Melastatin 8 (TRPM8) in Primary Airway Epithelial Cells of Asthmatic Mice. Inflamm. Ther. 2018, 41, 1266–1275.
- 45. Deng, L.; Ma, P.; Wu, Y.; Ma, Y.; Yang, X.; Li, Y.; Deng, Q. High and low temperatures aggravate airway inflammation of asthma, Evidence in a mouse model. Environ. Pollut. 2020, 256, 113433.
- 46. Moran, M.M. TRP Channels as Potential Drug Targets. Annu. Rev. Pharmacol. Toxicol. 2018, 58, 309–330.
- 47. Wilkes, M.; Madej, M.G.; Kreuter, L.; Rhinow, D.; Heinz, V.; De Sanctis, S.; Ruppel, S.; Richter, R.M.; Joos, F.; Grieben, M.; et al. Molecular insights into lipid-assisted Ca2+ regulation of the TRP

channel Polycystin-2. Nat. Struct. Mol. Biol. 2017, 24, 123–130.

- 48. Fine, M.; Li, X.; Dang, S. Structural insights into group II TRP channels. Cell Calcium 2020, 86, 102107.
- 49. Steinritz, D.; Stenger, B.; Dietrich, A.; Gudermann, T.; Popp, T. TRPs in Tox, Involvement of Transient Receptor Potential-Channels in Chemical-Induced Organ Toxicity-A Structured Review. Cells 2018, 7, 98.
- 50. Huang, Y.; Fliegert, R.; Guse, A.H.; Lü, W.; Du, J. A structural overview of the ion channels of the TRPM family. Cell Calcium 2020, 85, 102111.
- 51. Madej, M.G.; Ziegler, C.M. Dawning of a new era in TRP channel structural biology by cryoelectron microscopy. Pflügers Arch. Eur. J. Physiol. 2018, 470, 213–225.
- Toledo Mauriño, J.J.; Fonseca-Camarillo, G.; Furuzawa-Carballeda, J.; Barreto-Zuñiga, R.; Martínez Benítez, B.; Granados, J.; Yamamoto-Furusho, J.K. TRPV Subfamily (TRPV2, TRPV3, TRPV4, TRPV5, and TRPV6) Gene and Protein Expression in Patients with Ulcerative Colitis. J. Immunol. Res. 2020, 2906845.
- 53. Gao, Y.; Cao, E.; Julius, D.; Cheng, Y. TRPV1 Structures in Nanodiscs Reveal Mechanisms of Ligand and Lipid Action. Nature 2016, 534, 347–351.
- 54. Joanna, K.; Bujak, D.; Kosmala, I.M.; Szopa, K.; Majchrzak, P.B. Inflammation; Cancer and Immunity—Implication of TRPV1 Channel. Front. Oncol. 2019, 9, 1087.
- 55. Singh, A.K.; McGoldrick, L.L.; Sobolevsky, A.I. Structure and gating mechanism of the transient receptor potential channel TRPV3. Nat. Struct. Mol. Biol. 2018, 25, 805–813.
- 56. Suresh, K.; Servinsky, L.; Jiang, H.; Bigham, Z.; Yun, X.; Kliment, C.; Huetsch, J.; Damarla, M.; Shimoda, L.A. Reactive oxygen species induced Ca2+ influx via TRPV4 and microvascular endothelial dysfunction in the SU5416/hypoxia model of pulmonary arterial hypertension. Am. J. Physiol. Lung Cell. Mol. Physiol. 2018, 314, L893–L907.
- 57. Webster, C.M.; Tworig, J.; Caval-Holme, F.; Morgans, C.W.; Feller, M.B. The Impact of Steroid Activation of TRPM3 on Spontaneous Activity in the Developing Retina. Eneuro 2020, 7.
- 58. Yin, Y. Structural basis of cooling agent and lipid sensing by the cold-activated TRPM8 channel. Science 2019, 363, 6430.
- 59. Yin, Y.; Wu, M.; Zubcevic, L.; Borschel, W.F.; Lander, G.C.; Lee, S.Y. Structure of the cold- and menthol-sensing ion channel TRPM8. Sci. Ther. 2018, 359, 237–241.

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