

# Atypical Femoral Fractures Related to Bisphosphonate Treatment

Subjects: [Medicine, Research & Experimental](#) | [Genetics & Heredity](#)

Contributor: Natalia Garcia-Giralt

Atypical femoral fractures (AFF) are rare fragility fractures in the subtrochanteric or diaphysis femoral region associated with long-term bisphosphonate (BP) treatment. The etiology of AFF is still unclear even though a genetic basis is suggested.

atypical femoral fractures

bisphosphonates

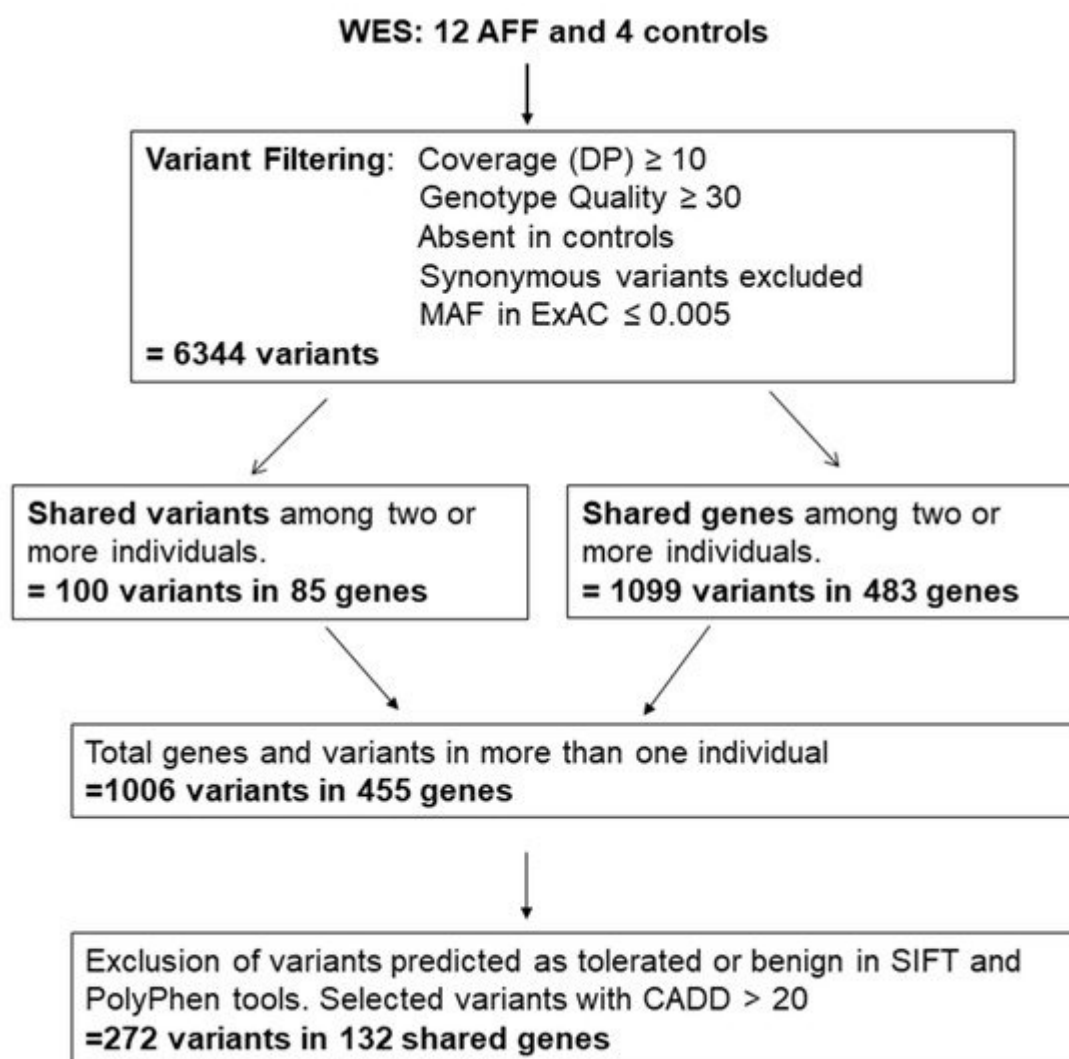
## 1. Introduction

Atypical femoral fractures (AFF) are a very rare type of bone fractures associated mainly with bisphosphonates (BP) and very rarely also with denosumab use [\[1\]\[2\]\[3\]](#). Genetic factors have been suggested as a possible explanation for both the higher risk of AFF in Asian populations and the low proportion of BP users that develop AFF [\[4\]](#).

Many attempts have been made to identify these genetic factors that may predispose some BP users to sustain AFF. Among them, a few studies have revealed that genetic variants in genes implicated in the mevalonate pathway, which is targeted by BP, may affect bone mineral density, bone turnover, and predispose to AFF, in response to BP treatment [\[5\]\[6\]\[7\]\[8\]](#). However, a recent genome-wide association study (GWAS) and candidate gene study comparing 51 AFF cases to 324 BP-treated controls was unable to find evidence of common genetic variants for BP-associated AFF [\[9\]](#). Hence, the authors proposed to perform GWAS with a larger sample size as well as whole-exome or whole-genome sequencing studies. This combination of studies would help to uncover the genetic background associated with BP-related AFF, which has a high genetic heterogeneity, sometimes associated with monogenic disorders [\[10\]\[11\]](#) or otherwise with a polygenic etiology and large variability among individuals [\[12\]\[13\]](#).

## 2. Variant Selection

In order to identify genes putatively involved in AFF, authors first removed all variants identified in the four control samples, and then selected those genes harboring rare genetic variants (ExAC and CSVS < 0.005) shared in at least two patients (**Figure 1**). Authors identified 100 rare variants in 85 genes that were shared by at least two patients. In addition, 483 genes presented a rare variant in at least 2 patients with AFF (same gene, different variant). In total, 1006 variants in 455 genes were identified.



**Figure 1.** Pipeline of selected variants obtained by whole exome sequencing of 12 patients with BP-related AFF and 4 controls (individuals with long-term BP treatment without AFF). Only variants or genes mutated in at least two patients were considered for further analysis.

Variants were then prioritized based on functional prediction (excluding variants with CADD score < 20, and those considered tolerated or benign by SIFT or PolyPhen\_humDiv, respectively). Considering only genes with at least two carriers of a rare variant, a total 272 variants in 132 genes remained (**Figure 1** and **Table 1**).

**Table 1.** Genes with at least two individuals carrying a rare variant; Variants were prioritized based on functional prediction (excluding variants with CADD score <20, and those considered tolerated or benign by SIFT or PolyPhen\_humDiv, respectively).

Genes with Rare Variants in Two AFF Cases				Genes with Rare Variants in More Than two AFF Cases	
Two Different Variants		One Variant			
Gene Name	Gene Name	Gene Name	Gene Name	Gene Name	Number of Variants and (Carriers)
AASS	DNAH10	PSD3	ACADL	C8orf46	1 (3)

Genes with Rare Variants in Two AFF Cases				Genes with Rare Variants in More Than two AFF Cases	
Two Different Variants		One Variant			
Gene Name	Gene Name	Gene Name	Gene Name	Gene Name	Number of Variants and (Carriers)
ABCA10	DNAH12	PTH1R	C1orf87	CHRNA1	3 (3)
ABCA4	DNAH6	PYHIN1	CD1A	DAAM2	3 (3, one homoz)
ABL2	DYSF	R3HDML	CITED4	DNAH14	4 (4)
ADAMTS12	EFHB	RET	GBA	DNAH2	3 (3)
ANAPC11	EP400	RMDN1	IQSEC3	DNAH9	3 (3)
ANK3	ERCC5	RNF157	NSMAF	FSIP2	3 (3)
ANKRD40	FAT4	RNF34	PPP2R1B	HLA-DRB1	2 (4)
ARHGEF18	FBLN7	RTEL1	SERPINB2	HRASLS	1 (3)
ARID1B	FLJ00418	SCN9A	SPTBN1	IGFLR1	2 (2, one homoz)
ASH1L	GBP3	5-Sep	SYDE1	KRT10	1 (5)
ATAD2	GPX4	SH3BP2	TNFRSF25	LAMA1	3 (3)
ATP10B	HK3	SHROOM4	TRAPPC2L	LRP5	4 (3)
BIN1	HPS6	SIRT5	TRIM32	MRPS12	1 (3)
C10orf54	IGFN1	SLC26A9		NEB	4 (4)
C12orf42	IGSF10	SLC2A7		OBSCN	5 (5)
C14orf159	IGSF22	SLC34A3		TCOF1	3 (4)
C17orf107	KLHL33	SLC52A2		TNXB	3 (3)
C6	LLGL1	SPTBN5		TTN	8 (8)
C9orf84	MEX3D	SRCAP		UTRN	3 (3)
CA9	MKS1	TAF15		VEGFB	1 (3)
CDC42BPG	MMP20	TENM4		ZC3H3	3 (3)
CERKL	MSLN	TJP3			
CHAMP1	NOD2	TMEM143			
CLCN2	NUP153	TNRC6B			

Genes with Rare Variants in Two AFF Cases			Genes with Rare Variants in More Than two AFF Cases	
Two Different Variants		One Variant		
Gene Name	Gene Name	Gene Name	Gene Name	Number of Variants and (Carriers)
<i>CRYBA1</i>	<i>OPLAH</i>	<i>TOPORS</i>		
<i>CTSE</i>	<i>PACSIN2</i>	<i>TSFM</i>		
<i>CUL7</i>	<i>PARD6B</i>	<i>TTC14</i>		
<i>CYYR1</i>	<i>PCDHAC1</i>	<i>ZNF34</i>		
<i>DAB2IP</i>	<i>PDE4DIP</i>	<i>ZNF646</i>		
<i>DAW1</i>	<i>PISD</i>	<i>ZNF729</i>		
<i>DHX34</i>	<i>PLA2G4D</i>	<i>ZSCAN32</i>		

Function enrichment analysis using the BinGO and GeneMANIA app in Cytoscape yielded adjusted significant scores for dynein complex, contractile fiber, microtubule motor activity, ciliary transition zone, actin cytoskeleton organization and pyrophosphatase activity.

Afterwards, authors intersected this list with previously described genes involved in bone metabolism and/or AFF [13][14][15][16]. Twelve genes were identified and selected as candidate genes (**Table 2**) for further in silico analyses using the AFFNET tool. Half of the AFF patients were carriers of variants in one Wnt signaling gene: *DAAM2* (3 carriers, one each for p.(P555L) (homozygous), p.(P582H) and p.(R989L), and a fourth with a variant predicted as tolerated by SIFT (p.(K776T))) and *LRP5* (3 carriers, one each for p.(R258C) and p.(P1504L) and one carrying two variants (p.(R1036Q) and p.(S1482L)), suggesting a role of this pathway in AFF triggering.

**Table 2.** Genes involved in bone metabolism and/or AFF containing deleterious rare variants in at least two AFF patients of this study.

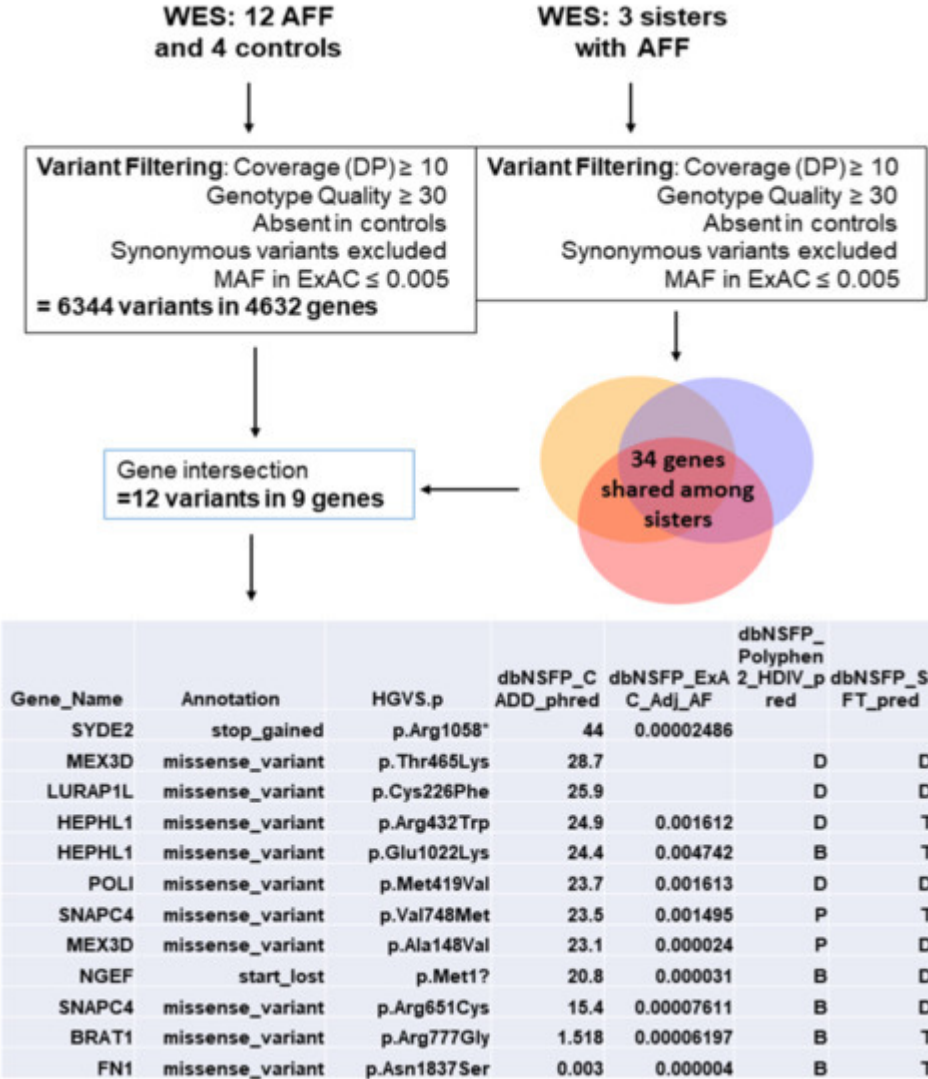
Gene ID	Number of Carriers	Function	Bone Association	Bibliography Source
<i>CUL7</i>	2	A core component of the 3 M complex required to regulate microtubule dynamics and genome integrity	Mutations in this gene produce the 3 m syndrome, which causes skeletal abnormalities	Genecards
<i>DAAM2</i>	3	Involved in the canonical Wnt signaling, a pathway critical for bone formation and repair	SNPs in this gene are associated with estimated bone mineral density (eBMD). <i>Daam2</i> knockout mouse showed decreased bone strength	Musculoskeletal Knowledge Portal, Morris et al., 2019 [17]

Gene ID	Number of Carriers	Function	Bone Association	Bibliography Source
<i>DNAH10</i>	2	Found in cilia and flagella; ATPase activity and microtubule motor activity	SNPs in this gene are associated with waist-hip ratio and eBMD.	Musculoskeletal Knowledge Portal
<i>DNAH12</i>	2	ATPase activity and microtubule motor activity	SNPs in this gene are associated with waist-hip ratio and eBMD	Musculoskeletal Knowledge Portal
<i>LAMA1</i>	3	A major component of the basal membrane which has been implicated in a wide variety of biological processes including cell adhesion, differentiation, migration, and signaling	Binding to cells via a high affinity receptor, laminin is thought to mediate the attachment, migration and organization of cells into tissues during embryonic development by interacting with other extracellular matrix components.	Genecards
<i>LRP5</i>	4	A co-receptor with Frizzled protein family members for transducing signals by Wnt proteins	It plays a key role in skeletal homeostasis and many bone density related diseases are caused by mutations in this gene	Genecards
<i>MEX3D</i>	2	RNA binding protein, may be involved in post-transcriptional regulatory mechanisms	Found mutated in three sisters with AFF	Roca-Ayats N, et al. 2018 <a href="#">[12]</a>
<i>PTH1R</i>	2	A receptor for parathyroid hormone (PTH) and for parathyroid hormone-like hormone (PTHrH).	Involved in the Hedgehog and PTH signaling pathways in bone and cartilage development	Genecards
<i>SLC34A3</i>	2	Involved in the transporting phosphate into cells via sodium cotransport in the renal brush border membrane, and contributes to the maintenance of inorganic phosphate concentration in the kidney	Mutations in this gene are associated with hereditary hypophosphatemic rickets with hypercalciuria.	Genecards
<i>SPTBN1</i>	2	Spectrin is an actin crosslinking and molecular scaffold protein that links the plasma membrane to the actin cytoskeleton, and functions in the determination of cell shape, arrangement of	SNPs in this gene are associated with eBMD and total body BMD	Musculoskeletal Knowledge Portal

Gene ID	Number of Carriers	Function	Bone Association	Bibliography Source
		transmembrane proteins, and organization of organelles		
<i>TNRC6B</i>	2	Involved in cellular senescence, innate or adaptive immune system, Wnt signaling, and calcium modulating pathways	SNPs in this gene are mainly associated with lean mass. One SNP was also associated with lower lumbar spine BMD and increased risk of fractures	Karasik D, et al. 2019 <a href="#">[18]</a>
<i>TNXB</i>	3	A member of the tenascin family of extracellular matrix glycoproteins	Mutations in this gene are associated with the Ehlers-Danlos Syndrome	Genecards

Genecards: <https://www.genecards.org/>, accessed on 9 December 2021. Musculoskeletal Knowledge Portal (MSK portal): <https://msk.hugeamp.org/>, accessed on 9 December 2021.

In parallel, authors compared all genes carrying rare variants in this study with previous results obtained from a WES in three sisters with BP-related AFF [\[12\]](#). A total of 9 genes were found overlapping both studies (**Figure 2**). Four of them carried damaging rare variants: *LURAP1L*, *MEX3D*, *POLI*, and *SYDE2*. These genes were also considered candidate genes for the network analysis.

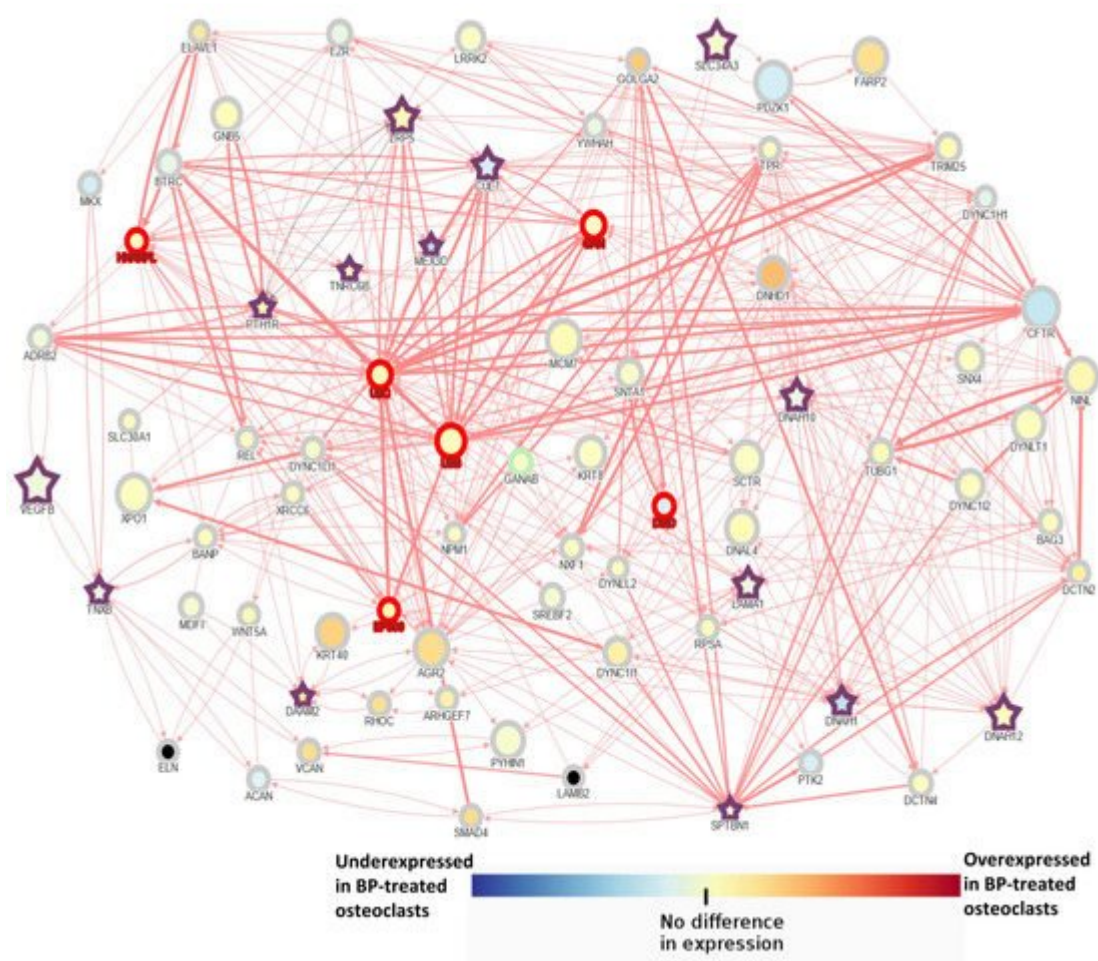


**Figure 2.** All mutated genes from the WES were intersected with genes also mutated in a previous study with 3 sisters who sustained AFF [12].

### 3. AFF Network Analysis with Candidate Genes

Interactions among identified genes were explored using the AFFNET tool. In order to simplify the network display, the shortest path interactions among bone-related genes (described in Table 2) were explored. Therefore, only direct interactions between candidate genes are displayed. Candidate genes were interconnected with each other, even though in some cases through other intermediate genes (Figure 3).





**Figure 3.** Network analysis of bone-related genes mutated in at least 2 AFF patients using the AFFNET tool. The shortest path interactions among candidate genes were displayed. Purple stars indicate candidate genes. The size of the nodes is determined by observed/expected loss of function score in the gnomAD database. This score is the ratio of the observed and expected loss of function variants in a particular gene. This score provides insight into how tolerant a gene is to loss of function variation. The red border tags genes that are outlier in the constrain metrics for LoF or missense variants according to gnomAD. Node color represents their expression depending on the expression data obtained in the GSE63009: Osteoclastic precursor cells treated or not with bisphosphonates (alendronate or risedronate) during their differentiation into mature osteoclasts.

Besides, a very complex network with multiple interactions among candidate genes and with the other mutated genes (**Table 1**) is displayed when all possible interactions were included in the analysis. Finally, interactions of mutated genes overlapped between the present study and the previous study with 3 sisters with AFF (**Figure 2**) were also explored. No direct interactions were found among these genes even though they were interconnected through one intermediate gene. One gene, *LURAP1L*, had no interactions with other genes. The *MEX3D* gene, which is mutated in 2 patients plus the 3 sisters, is under expressed in BP-treated osteoclasts.

The main conclusion of this study is that AFF may present a multigenic background, specific to each patient, in which an accumulation of susceptibility variants may lead to a predisposition to BP-related AFF. Our analysis suggested that Wnt signaling may play a relevant role in the BP-related AFFs as half of the patients had mutations in a gene of this pathway. In silico analysis suggested a complex interaction network among the different mutated



genes as well as a biological enrichment for cytoskeleton and cilium organization. WES analysis provided evidence to support the hypothesis that several genes and their interactions may be involved in the development of AFF, and, along with BP treatment and, in some cases, glucocorticoids, they may trigger the perfect storm.

## References

1. Shane, E.; Burr, D.; Abrahamsen, B.; Adler, R.A.; Brown, T.D.; Cheung, A.M.; Cosman, F.; Curtis, J.R.; Dell, R.; Dempster, D.W.; et al. Atypical subtrochanteric and diaphyseal femoral fractures: Second report of a task force of the American Society for Bone and Mineral Research. *J. Bone Miner. Res.* 2014, 29, 1–23.
2. Lo, J.C.; Grimsrud, C.D.; Ott, S.M.; Chandra, M.; Hui, R.L.; Ettinger, B. Atypical femur fracture incidence in women increases with duration of bisphosphonate exposure. *Osteoporos. Int.* 2019, 30, 2515–2520.
3. Starr, J.; Tay, Y.K.D.; Shane, E. Current Understanding of Epidemiology, Pathophysiology, and Management of Atypical Femur Fractures. *Curr. Osteoporos Rep.* 2018, 16, 519–529.
4. Lo, J.C.; Hui, R.L.; Grimsrud, C.D.; Chandra, M.; Neugebauer, R.S.; Gonzalez, J.R.; Budayr, A.; Lau, G.; Ettinger, B. The association of race/ethnicity and risk of atypical femur fracture among older women receiving oral bisphosphonate therapy. *Bone* 2016, 85, 142–147.
5. Roca-Ayats, N.; Balcells, S.; Garcia-Giralt, N.; Falco-Mascaro, M.; Martinez-Gil, N.; Abril, J.F.; Urreiziti, R.; Dopazo, J.; Quesada-Gomez, J.M.; Nogues, X.; et al. GGPS1 Mutation and Atypical Femoral Fractures with Bisphosphonates. *N. Engl. J. Med.* 2017, 376, 1794–1795.
6. Marini, F.; Falchetti, A.; Silvestri, S.; Bagger, Y.; Luzi, E.; Tanini, A.; Christiansen, C.; Brandi, M.L. Modulatory effect of farnesyl pyrophosphate synthase (FDPS) rs2297480 polymorphism on the response to long-term amino-bisphosphonate treatment in postmenopausal osteoporosis. *Curr. Med. Res. Opin.* 2008, 24, 2609–2615.
7. Choi, H.J.; Choi, J.Y.; Cho, S.W.; Kang, D.; Han, K.O.; Kim, S.W.; Kim, S.Y.; Chung, Y.S.; Shin, C.S. Genetic polymorphism of geranylgeranyl diphosphate synthase (GGSP1) predicts bone density response to bisphosphonate therapy in Korean women. *Yonsei Med. J.* 2010, 51, 231–238.
8. Olmos, J.M.; Zarrabeitia, M.T.; Hernandez, J.L.; Sanudo, C.; Gonzalez-Macias, J.; Riancho, J.A. Common allelic variants of the farnesyl diphosphate synthase gene influence the response of osteoporotic women to bisphosphonates. *Pharm. J.* 2012, 12, 227–232.
9. Kharazmi, M.; Michaelsson, K.; Schilcher, J.; Eriksson, N.; Melhus, H.; Wadelius, M.; Hallberg, P. A Genome-Wide Association Study of Bisphosphonate-Associated Atypical Femoral Fracture. *Calcif. Tissue Int.* 2019, 105, 51–67.

10. Marini, F.; Brandi, M.L. Atypical femur fractures: A distinctive tract of adult hypophosphatasia. *Clin. Cases Miner. Bone Metab.* 2017, 14, 324–328.
11. Tournis, S.; Dede, A.D. Osteogenesis imperfecta—A clinical update. *Metabolism* 2018, 80, 27–37.
12. Roca-Ayats, N.; Ng, P.Y.; Garcia-Giralt, N.; Falco-Mascaro, M.; Cozar, M.; Abril, J.F.; Quesada Gomez, J.M.; Prieto-Alhambra, D.; Nogues, X.; Dunford, J.E.; et al. Functional Characterization of a GGPPS Variant Identified in Atypical Femoral Fracture Patients and Delineation of the Role of GGPPS in Bone-Relevant Cell Types. *J. Bone Miner. Res.* 2018, 33, 2091–2098.
13. Perez-Nunez, I.; Perez-Castrillon, J.L.; Zarrabeitia, M.T.; Garcia-Ibarbia, C.; Martinez-Calvo, L.; Olmos, J.M.; Briongos, L.S.; Riancho, J.; Camarero, V.; Munoz Vives, J.M.; et al. Exon array analysis reveals genetic heterogeneity in atypical femoral fractures. A pilot study. *Mol. Cell Biochem.* 2015, 409, 45–50.
14. Morris, J.A.; Kemp, J.P.; Youlten, S.E.; Laurent, L.; Logan, J.G.; Chai, R.C.; Vulpescu, N.A.; Forgetta, V.; Kleinman, A.; Mohanty, S.T.; et al. An atlas of genetic influences on osteoporosis in humans and mice. *Nat. Genet.* 2019, 51, 258–266.
15. Nguyen, H.H.; van de Laarschot, D.M.; Verkerk, A.; Milat, F.; Zillikens, M.C.; Ebeling, P.R. Genetic Risk Factors for Atypical Femoral Fractures (AFFs): A Systematic Review. *JBMR Plus* 2018, 2, 1–11.
16. Koromani, F.; Trajanoska, K.; Rivadeneira, F.; Oei, L. Recent Advances in the Genetics of Fractures in Osteoporosis. *Front. Endocrinol* 2019, 10, 337.
17. Karasik, D.; Zillikens, M.C.; Hsu, Y.H.; Aghdassi, A.; Akesson, K.; Amin, N.; Barroso, I.; Bennett, D.A.; Bertram, L.; Bochud, M.; et al. Disentangling the genetics of lean mass. *Am. J. Clin. Nutr.* 2019, 109, 276–287.

---

Retrieved from <https://encyclopedia.pub/entry/history/show/45199>