Multispectral Imaging for Quality Determinations of White Meat

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White meat is the nutritional term for lighter-colored meat that contains less myoglobin than red meat, which contains a great deal. White meat includes poultry (e.g., chicken, duck, goose and turkey), fish, reptiles (e.g., land snail), amphibians (e.g., frog), crustaceans (e.g., shrimp and crab) and bivalves (e.g., oyster and clam), but it excludes all mammal flesh such as beef, pork, and lamb. White meat has high nutritional value and plays an important role in human diet. The production and sale of white meat need to meet specific quality and safety standards. Fluorescence spectroscopy, color imaging and multispectral imaging (MSI) have emerged as effective analytical methods for the non-destructive detection of quality attributes of various white meat products such as fish, shrimp, chicken, duck and goose.

white meat multispectral imaging

fluorescence spectroscopy

convolutional neural network

quality detection

1. Introduction

As a global issue, food safety and quality are of increasing concern to companies and customers [1]. White meat is the nutritional term for lighter-colored meat that contains less myoglobin than red meat, which contains a great deal. Compared with white meat, the intake of red meat has a greater correlation with colorectal cancer (CRC), indicating that white meat intake is more beneficial to human health [2]. White meat includes poultry (e.g., chicken, duck, goose and turkey), fish, reptiles (e.g., land snail), amphibians (e.g., frog), crustaceans (e.g., shrimp and crab) and bivalves (e.g., oyster and clam), but it excludes all mammal flesh such as beef, pork, and lamb. White meat has high nutritional value and plays an important role in human diet. The production and sale of white meat need to meet specific quality and safety standards. The freshness of fish is one of the important indicators for evaluating its quality because of its high perishability ³. Moreover, poultry products are particularly susceptible to oxidation as this meat contains relatively high levels of unsaturated fatty acids and low levels of natural antioxidants, such as vitamin E. In addition, chemical residues in white meat may have an adverse effect on human health. For example, fluoroguinolone antibiotics are effective against a wide range of Gram-negative and positive bacteria, thus they are widely used in the medical and veterinary fields. However, their use in animals has raised concerns, as this practice may lead to an increase in microbial resistance [4]. Moreover, nitrofuran drugs (NFs), including furazolidone (FZD), nitrofurazone (NFZ), and furantazone (FTD) are broad-spectrum antimicrobials. The potential risk of these compounds to human health is of great concern because of their carcinogenic and mutagenic properties. It is therefore crucial to ensure the quality and safety of white meat.

Traditional methods for meat quality and safety evaluation, such as manual inspection, mechanical and chemical methods, are time-consuming and destructive, and cannot meet the requirements of rapid inspection ^[5]. For example, methods for freshness evaluation are based on human sensory qualities, such as appearance, taste and texture. However, human senses exhibit a very high degree of subjectivity and can therefore be questioned in certain situations ^[3]. Even if manual inspection could meet accuracy requirements, it is still a labor-intensive and time-consuming process. Recently, the meat industry has adopted the most advanced high-speed processing technologies, and meat products in order to achieve economic benefits. The requirement for real-time monitoring of food has encouraged the development of non-destructive measurement systems ^[5]. Optical technology is becoming increasingly important in research and industrial applications to measure the quality attributes of meat and meat products in order tory popular in recent years ^[8]. In addition, fluorescence spectroscopy and multispectral imaging (MSI) also show obvious advantages and capabilities in the non-destructive evaluation of white meat.

There have been several reviews of these new techniques of meat quality assessment. These papers show that these spectroscopic methods have been implemented as an alternative to traditional methods, but they mainly focus on one technique for quality detection of one specific category of meat, e.g., fish ^[3], shrimp ^[4], chicken ^[9], duck ^[10], or red meat ^[11]. As far as the researchers know, there is no literature review analyzing the application of various imaging techniques in the non-destructive quality inspection of various white meats. (The published reviews based on these three imaging techniques are tabulated in **Table 1**).

Table 1. Summary of reviews on fluorescence spectroscopy, RGB- and MSI techniques in food evaluation.

Technology	Product	Target Attributes	Reference
MSI	Meat	Adulteration	Ropodi et al. ^[12]
MSI, HSI	Meat	Defects	Feng et al. ^[13]
MSI	Food	Quality	Su and Sun ^[14]
MSI, IRS, SERS, LIBS and HSI	Food	Quality	Wang et al. ^[15]
MSI, HSI and VS	Food	Authenticity, quality and safety	Ropodi et al. ^[<u>16</u>]

Technology	Product	Target Attributes	Reference
Fluorescence spectroscopy	Food	Quality	Karoui and Blecker [<u>17</u>]
Fluorescence spectroscopy	Food	Quality	Strasburg and Ludescher ^[<u>18</u>]
Visible/Infrared, Raman and Fluorescence spectroscopy	Raw and processed food	Quality	He and Sun ^[19]
Fluorescence spectroscopy	Food	Quality	Ahmad et al. ^[20]
Fluorescence spectroscopy	Dairy products	Quality and safety	Shaikh and O'Donnell ^[21]
Fluorescence spectroscopy	Fresh and frozen- thawed muscle foods	Muscle classification	Hassoun ^[22]
RGB-Imaging	Meat	Quality and safety	Taheri-Garavand et al. ^[23]
RGB-Imaging	Fish	Quality	Dowlati et al. ^[24]
RGB-Imaging	Food	Quality	Gomes and Leta
RGB-Imaging	Food	Quality	Amani et al. ^[26]

J. Quality Evaluation of white meat

The application of fluorescence spectroscopy, RGB imaging and MSI for white meat quality inspection has been thoroughly and extensively researched as shown in **Table 2**. For MSI techniques, correlation coefficient (*R*) or **MSI ficiantisphetermination** (R^2) Histan-Hypertapicatatistical gengtricities as a spectroscopy indicate the management of the second statistical gengtricities between the second statistical gengtricities as a spectroscopy indicate the management of the second statistical gengtricities as a spectroscopy indicate the management of the second statistical gengtricities as a spectroscopy indicate statistical gengtricities as a spectroscopy in the spectroscopy

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White Meat	Module	Quality Parameters	Accuracy	Reference	sible light at higher
Fish	MSI	TVB-N, PPC	$R_p^2 = 0.862$ for TVB-N, $R_p^2 = 0.921$ for PPC	Khoshnoudi-Nia and Moosavi-Nasab ^[39] , Khoshnoudi-Nia and Moosavi-Nasab ^[40]	, with the nment, so ction and vels, low
Fish	MSI	TVC	$R^2 = 0.62$	Govari, et al. ^[41]	
Fish	MSI	TVC	$R^2 = 0.683$	Fengou, et al. ^[42]	
Fish	MSI	Astaxanthin concentration	<i>R</i> ² = 0.86	Dissing, et al. ^[43]	
Fish	MSI	TVB-N, TBARS, K	$R_{p}^{2} = 0.922$ for TVB-N, $R_{p}^{2} = 0.867$ for TBARS, $R_{p}^{2} = 0.936$ for K	Cheng, et al. ^[44]	dark state" ached state hardware scanners nd image jes in the
Fish	MSI	A 'standard freshness index' of K	R ² = 0.94,	Omwange, et al. ^[45]	mages of Ind visual , low cost
Fish	Fluorescence spectroscopy	A 'standard freshness index' of K	<i>R</i> ² = 0.92	Omwange, et al. ^[46]	s spatial dentifying has been

White Meat	Module	Quality Parameters	Accuracy	Reference	
Fish	Fluorescence spectroscopy	A 'standard freshness index' of K	<i>R</i> ² = 0.95	Liao, et al. ^[<u>47</u>]	
Fish	Fluorescence spectroscopy	AEC; NADH	$R^2 = 0.90$ for AEC, $R^2 = 0.85$ for NADH	Rahman, et al. ^[48]	
Fish	Fluorescence spectroscopy	NADH	90.5%	Hassoun and Karoui ^[49]	าclosure
Fish	RGB imaging	Classification performance	99.5%	Park, et al. ^[50]	
Fish	RGB imaging	Astaxanthin concentration	<i>R</i> ² = 0.66	Dissing et al. ^[43]	samples
Fish	RGB imaging	Freshness of tuna meat cuts	86.67%	Lugatiman, et al. ^[51]	r band of t spectral
Fish	RGB imaging	The main color of the sample	75%	Mateo, et al. ^[52]	n provide ide more cations in
Fish	RGB imaging	Texture features	86.3%	Gu, et al. ^[53]	i re 3 . The six bands,
Fish	RGB imaging	Color of Salmon Fillets	<i>R</i> = 0.95	Quevedo, et al. ^[54]	
Fish	RGB imaging	Gill and eye color changes in the sparus aurata	<i>R</i> ² = 0.994	Dowlati, et al. ^[55]	_
Fish	RGB imaging	Body color of carp	94.97%	Taheri-Garavand, et al. ^[56]	

White Meat	Module	Quality Parameters	Accuracy	Reference	
Fish	RGB imaging	Freshness	98.2%	Rocculi, et al. ^[57]	
Shrimp	Fluorescence spectroscopy	4-hexylresorcinol	81.6%	Jonker and Dekker ^[58]	_
Shrimp	Fluorescence spectroscopy	K, pH	<i>R</i> ² = 0.80	Rahman, et al. ^[59]	1
Shrimp	RGB imaging	рН	100%	Witjaksono, et al. ^[60]	
Shrimp	RGB imaging	Identification accuracy of the proposed ShrimpNet for shrimp	95.48%	Hu, et al. ^[61]]
Shrimp	RGB imaging	Shrimp dehydration levels	<i>R</i> = 0.86	Mohebbi, et al. ^[62]	JSA) and [<u>38</u>]
Shrimp	RGB imaging	Color changes in the head, legs and tail of pacific white shrimp (litopenaeus vannamei)	90%	Ghasemi-Varnamkhasti, et al. ^[63]	
Chicken	Fluorescence spectroscopy	Hydroxyproline concentration	$R^2 = 0.82$	Monago-Maraña, et al. ^[64]	_
Chicken	MSI	Skin tumors	86%	Chao, et al. ^[65]	_
Chicken	MSI	TVC	90.4%	Spyrelli, et al. ^[66]	
Chicken	MSI	pork-chicken adulteration	90.00% for fresh samples, 86.67%	Fengou, et al. ^[67]	

White Meat	Module	Quality Parameters	Accuracy	Reference
			for frozen-thawed samples	
Chicken	MSI	Sepsis in chickens	98.6% for septic chickens, 96.3% for healthy chickens	Yang, et al. ^[68]
Chicken	MSI	Contamination detection	96%	Park, et al. ^[69]
Chicken	MSI	Chicken heart disease characterization	100%	Chao, et al. ^[70]
Chicken	MSI; Fluorescence spectroscopy	Contamination detection	92.5%	Seo, et al. ^[71]
Chicken	Fluorescence spectroscopy	Lipid oxidation	R = 0.73	Gatellier, et al. ^[72]
Chicken	Fluorescence spectroscopy	P. aeruginosa concentration	96%	Abdel-Salam, et al. ^[73]
Chicken	Fluorescence spectroscopy	chicken meat tenderness	<i>R</i> = 0.870	Yu, et al. ^[74]
Chicken	Fluorescence spectroscopy	Contamination detection	96.6%	Cho, et al. ^[75]

White Meat	Module	Quality Parameters	Accuracy	Reference	_
Chicken	Fluorescence spectroscopy	Measurement of lipid oxidation	98%	Wold and Kvaal ^[76]	_
Chicken	RGB imaging	Avian flu infected chickens	97.43%	Cuan, et al. [77]	_
Chicken	RGB im-aging	Color	94%	Yumono, et al. ^[78]	_
Chicken	RGB im-aging	Freshness	R = 0.987	Taheri-Garavand, et al. ^[79]	A—Linear ≀S—Thio-
Duck	Fluorescence spectroscopy	Gentamicin Residual in Duck Meat	R = 0.996	Wang, et al. ^[80]	adenine
Duck	Fluorescence spectroscopy	Doxycycline content in duck meat	R = 0.998	Wang, et al. ^[81]	ז Nutr
Duck	Fluorescence spectroscopy	Carbaryl residue in duck meat	<i>R</i> = 0.976	Xiao et al. ^[10]	
Duck	Fluorescence spectroscopy	Tetracycline content	<i>R</i> = 0.952	Zhao, et al. ^[82]	lahos, with
Duck	Fluorescence spectroscopy	Triazophos content	$R_{p}^{2} = 0.974,$	Zhao, et al. ^[83]	J-
Duck	Fluorescence spectroscopy	Neomycin residue	R = 0.999	Jiang, et al. ^[84]	es for , 57,
Duck	Fluorescence	Carbofuran residue	$R_{p}^{2} = 0.999$	XIAO, et al. ^[85]	tion of m. Acta

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White Meat	Module	Quality Parameters	Accuracy	Reference	ication . Food
	spectroscopy				
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^{69, 225–234.}

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