# **Tongue and Systemic Connections Microbiota**

Subjects: Dentistry, Oral Surgery & Medicine

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The tongue is able to quickly reflect the state of health or disease of the human body. Tongue inspection is an important diagnostic approach. It is a unique method that allows to explore the pathogenesis of diseases based on the guiding principles of the holistic concept that involves the observation of changes in the lining of the tongue in order to understand the physiological functions and pathological changes of the body. It is a potential method of screening and early detection of cancer.

Keywords: tongue microbiota ; host–microbiota symbiosis ; tongue diagnosis ; systems correlations ; minimally invasive ; hygiene

### 1. Introduction

For thousands of years, doctors have diagnosed a patient's health by inspecting the tongue, especially the back of the tongue. Inspection of the tongue as a method of clinical diagnosis is a unique feature of Traditional Chinese Medicine (TCM).

The tongue inspection is a procedure that has been reported from the Shang Dynasty (from 1600 BC to 1000 BC) that should be performed by the clinical observation the tongue body to evaluate its morphological and color homogeneity. In accordance to the Traditional Chinese Medicine, the tongue is able to absolve a key role of a biosensor, while the different tongue regions are able to provide information to check the diagnosis the organ systems health  $\frac{|\mathbf{1}||\mathbf{2}|}{|\mathbf{1}||\mathbf{2}|}$ .

In accordance to Traditional Chinese Medicine, "A pathology arises associated to a tongue surface coating appearance. The surface coating represents the exterior manifestation, which corresponds to the sustaining disease, and is the primary indication for the diagnosis making". In this way, the appropriate knowledge of the molecular aspects of tongue response is a key factor to comprehend this semeiotic medical practice [1][3][4]. Several studies on the microbiology of the healthy human tongue, through the sequencing of 16S rRNA, which is applied for the detection, categorization, and evaluation of the microbic charge within the biological mixtures, have shown that the most abundant phyla are Fusobacteria, Actinobacteria, Firmicutes, Proteobacteria, and Bacteroidetes [1][4][5]. The variation in the microbiota of the tongue lining recorded by metagenomic sequencing identifies potential biomarkers for several diseases, including the precancerous cascade [1][6][7]. The thin white or reddish tongue coating with 0/1 patina is a symbol of good health (Figure 1 and Figure 2), while the "white-fat" and "yellow-dense" coating (Figure 3 and Figure 4) may be an indication of inflammation, infection, and stress, as well as of immune and endocrine disorders (Figure 5) [8][9]. Thick coat and tongue moisture are increased in tumors. The relationships between tongue diagnosis and diseases, including rheumatoid arthritis, gastritis and precancerous lesions, liver, pancreatic, and lung cancer, were explored [1][10][11][12][13][14][15]. Tongue-lining microbiota biomarkers could provide a complement for the diagnosis of several diseases from the noninvasive, individualized, and long-term monitoring aspects [1][16]. Tongue diagnosis is a simple, noninvasive method of assessing physiological conditions by observing the thickness and color of the coating [6]. The tongue coating is categorized into three different classes in accordance to their color typologies: white coating (W), yellow coating (Y), and black and gray coating (Table 1). A yellow coat is considered an indication of water retention and heat [3]. The most common syndromes are the following four: external cold syndrome, internal cold syndrome, external heat syndrome, and internal heat syndrome, and are associated with thin white, thick white, thin yellow, and thick yellow tongue coatings [1]. Cold and hot syndromes are two contrary but internally related conditions [8]. According to the TCM theory, the tongue is an external extension of the spleen and stomach [17][18]. Western medicine refers to the indexes of the thickness of the coat on the back of the tongue, classifying it in four degrees [19] (Table 2).



Figure 1. Strawberry tongue health symbol with thin coat (TCM), grade 0/1 according to Miyazaki.



Figure 2. Thin white tongue (TCM), grade 2 according to Miyazaki.

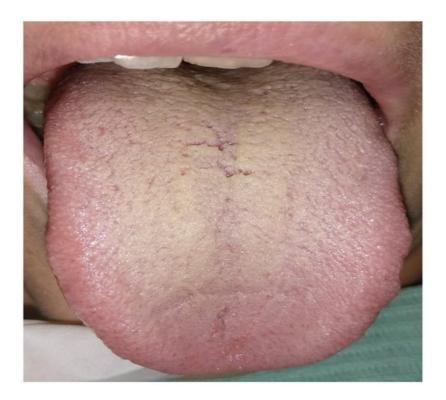


Figure 3. Thick white and yellow tongue (TCM), grade 3 according to Miyazaki.



Figure 4. Yellow fat tongue (TCM), grade 2 according to Miyazaki.



Figure 5. Desquamated coat of tongue related to severe hypovitaminosis.

 Table 1. Classification of the lingual patina according to the TCM standard.

standard.

Description	
Thin white, thick white, and fat white tongue coating	
Tongue coating thin yellow, thick yellow, and fat yellow	
Thin black and gray tongue coating and thick gray black	

**Table 2.** Classification of the lingual patina according to the Miyazaki standard.

Value	Description
0	No visible patina on the back of the tongue
1	Patina present only on the posterior III of the tongue
2	Patina that completely covers the dorsal surface of the tongue but does not mask the underlying mucosa
3	Very thick patina covering the entire dorsal surface of the tongue

Several studies have shown that *Lautropia* was significantly increased in patients with gingivitis, oral lichen planus, and chronic periodontitis [20][21][22][23][24]. The level of *Capnocytophaga* was higher in the saliva of patients with oral cancer [25] [26]. These results indicated that the tongue thin coating (W) is an important reference of the oral microbiota. The salivary *Megasphaera* was observed more abundant in lung cancer patients, associated to an increased rate of white coated tongue [5], fecal *Selenomonas* was increased in colon cancer patients, and the data suggested that (W) thick coat may be

correlated with the risk of intestinal tumors [1][27][28]. *Prevotella* has been implicated with periodontal infection [29]. Perimplant *prevotella maculosa* was increased in patients with smoking mucositis but decreased in patients with nonsmoking mucositis, indicating that thick coating (W) is associated with periodontal disease [28][29]. As for the yellow tongue coating, few potential microbiota have been observed, particularly opportunistic bacteria [20]. Researchers have shown that the oral microbiota is associated with many diseases, such as pancreatic disease, pediatric inflammatory bowel disease, obesity, coronary heart disease, rheumatoid arthritis, gastrointestinal cancer, liver cirrhosis, cardiovascular disease, and pneumonia. The relationship between the tongue lining microbiome and diagnosis was also reported to be relevant for the differentiation of these syndromes [3][9][10][16][17][30]. The mouth environment represents one of the widest microbial reservoirs, and a large quantity of bacterial species are able to provide a complex and stable community. *Mutans streptococci* and red complex bacterial group, including *Porphyromonas gingivalis*, *Tannerella forsythia*, and *Treponema denticola*, represent species strongly correlated to a high clinical risk for tooth decay and periodontal pathologies [2][31][32]. In addition, the existence of specific oral bacterial species related to systemic diseases has been reported in recent years [33]. For this reason, oral dysbiosis has attracted attention as an etiology of oral and systemic diseases

After the birth, the oral environment is exposed to a wide variety of microbes, while the human microbiome develops with the organism growth. In this phase, the facultative anaerobic bacterial species such as streptococcus and actinomycetes are considered the pioneering colonizers of the mouth environment, while the Streptococcus salivarius is considered the predominant colonizer species [35]. Although the Firmicutes phylum predominates in both the oral microbiota and the gut, the genus Streptococcus within the Firmicutes phylum is rarely detected in the gut microbiota [36]. Initially, a Streptococcus species predominance is primarily associated to the first phases of the consolidation of the mouth microbiome. It has also been reported that the formation of the oral indigenous microbiota begins within the first 6 weeks of life, and Streptococcus rapidly dominates the oral cavity during this phase [37]. The microbiome quality of the tongue tissues during the transition phase, between 80 and 120 weeks, is more similar to the adults' microbiota composition. Colonies corresponding to Streptococcus peroris and Streptococcus lactarius decrease exponentially immediately after 30–49 weeks, while Granulicatella Adjacens, Actinomyces odontolyticus, and Fusobacterium periodonticum increase over the same period Streptococcus S

The posterior part of the tongue has a large surface with papillary structures [39], which can retain numerous aerobic and anaerobic microorganisms [40], fungi, metabolites in the blood, saliva, and exfoliated keratinized epithelium. Filiform papillae are a specific structure that involves the formation of the lining of the tongue. This structure causes many cracks and folds that increase the surface of the tongue, and the warm, moist, and nourishing environment provides a suitable medium for colonization, growth, and proliferation of microorganisms [36][41][42][43][44][45][46][47]. The mucous membrane of the tongue accumulates more microbes than other parts of the oral cavity do, boasting a relatively complete and independent microecosystem that can be easily harvested from saliva [48][49]. These features imply that more attention should be paid to the tongue microbiota in older adults susceptible to swallowing problems [50]. An increase in tongue lining in edentulous older adults has been associated with aspiration pneumonia and fever  $\frac{[51][52]}{}$ . Usually, the detached microbes ingested with the salivary fluids move through the esophagus tract to the stomach region, where they go to be inactivated by the action of the gastric acid and proteolytic enzymes. However, alterations in swallowing with aging allow for aspiration into the lower respiratory tract and subsequent lung infection [53]. Older adults with fewer teeth, poor oral hygiene, and more dental caries consistently experience dysbiosis, with a greater relative abundance of Prevotella, Veillonella, and Streptococcus species and a greater number of fungal species. Particular attention should be paid to the state of the tongue by implementing correct oral hygiene habits with particular attention to the back of the tongue. Specifically, mechanical oral hygiene to lower the oral microbial load is recognized as an effective approach to reduce the death rate from aspiration pneumonia, so much so that in some countries, the figure of the oral health professional is guaranteed as standard assistance for the treatment of the oral cavity hygiene in frail elderly in hospitals and nursing homes [54]. Although aspirated saliva contains microorganisms that colonize various oral sites, the bacterial composition indicates that the dominant source is the tongue microbiota [36][41][55].

Some studies have shown that bacteria implicated in periodontal disease, such as *porphyromonas gingivalis*, *tannerella forsythia*, and *treponema denticola*, are a risk factor for atherosclerotic vascular disease, type 2 diabetes, and nonalcoholic fatty liver disease (**Figure 6**) [56][57][58]. Furthermore, a repeated oral ingestion of *Porphyromonas gingivalis* promotes systemic inflammation [59].



Figure 6. Thick white fatty coat of tongue related to type 2 diabetes, hypertension, and nonalcoholic fatty liver disease.

Since we ingest approximately 600 mL of saliva per day containing up to  $10^9$  bacteria/10 mL, it is reasonable to suspect that some oral bacteria induce intestinal microbiota disturbances  $\frac{[60]}{}$ .

## 2. Current Insights on Tongue and Systemic Connections Microbiota

The oral cavity represents a key ecosystem for microbial proliferation, while in healthy subjects, the mouth could present over ten billion bacteria. Bacteria are localized in tongue crypts and they could invade into the vascular system through the rich vascularized tissue components of this region [18].

Several studies have suggested a link between the development of the human microbiome and rheumatoid arthritis (RA). *Porphyromonas gingivalis* appears to be involved in the initiation and progression of RA, supported by the high presence of periodontitis [61][62]. In a case-control study, Ceccarelli F., Orrù G., and Pilloni A. et al. analyzed the presence and quantification of *P. gingivalis* in a large healthy cohort, and there was a significant association between the percentage of *P. gingivalis* on the total tongue biofilm and disease activity in patients with RA, suggesting that the microbiological status of the oral cavity could play a role in the mechanisms of inflammation. In this study, the percentage of *P. gingivalis* on the total language biofilm was analyzed for the first time; using this new measurement, an association was identified between this value and disease activity, new information on the bacterium's influence on RA. The authors hypothesized that the presence of *P. gingivalis* can chronically stimulate the immune system, regardless of the presence of periodontitis, leading to a state of chronic systemic inflammation [63].

#### 3. Conclusions

A small organ such as the tongue can be a very effective visual detector of apparently nonvisible pathologies and can be considered as a potential method of rapid cancer screening and diagnosis.

Many cancers are diagnosed at an advanced stage, mainly because most patients are asymptomatic in the early stage. Diagnose in the early stage could increase the number of curable cancers and improve survival. Evidently, it is necessary to develop new strategies in oral medicine for the early diagnosis of diseases, and the diagnosis of the tongue as a minimally invasive method is certainly one of them.

#### References

- 1. Sun, B.; Zhou, D.; Tu, J.; Lu, Z. Evaluation of the Bacterial Diversity in the Human Tongue Coating Based on Genus-Sp ecific Primers for 16S rRNA Sequencing. BioMed Res. Int. 2017, 2017, 1–12.
- 2. Anastasi, J.K.; Currie, L.M.; Kim, G.H. Understanding diagnostic reasoning in TCM practice: Tongue diagnosis. Altern. Ther. Heal. Med. 2009, 15, 18–28.

- 3. Ye, J.; Cai, X.; Yang, J.; Sun, X.; Hu, C.; Xia, J.; Shen, J.; Su, K.; Yan, H.; Xu, Y.; et al. Bacillus as a potential diagnostic marker for yellow tongue coating. Sci. Rep. 2016, 6, 32496.
- 4. Asakawa, M.; Takeshita, T.; Furuta, M.; Kageyama, S.; Takeuchi, K.; Hata, J.; Ninomiya, T.; Yamashita, Y. Tongue Micro biota and Oral Health Status in Community-Dwelling Elderly Adults. mSphere 2018, 3, e00332-18.
- 5. Welch, J.L.M.; Utter, D.; Rossetti, B.J.; Welch, D.B.M.; Eren, A.M.; Borisy, G.G. Dynamics of tongue microbial communi ties with single-nucleotide resolution using oligotyping. Front. Microbiol. 2014, 5, 568.
- 6. Cui, J.; Cui, H.; Yang, M.; Du, S.; Li, J.; Li, Y.; Liu, L.; Zhang, X.; Li, S. Tongue coating microbiome as a potential biomar ker for gastritis including precancerous cascade. Protein Cell 2019, 10, 496–509.
- 7. Kageyama, S.; Asakawa, M.; Takeshita, T.; Ihara, Y.; Kanno, S.; Hara, T.; Takahashi, I.; Yamashita, Y. Transition of Bact erial Diversity and Composition in Tongue Microbiota during the First Two Years of Life. mSphere 2019, 4, e00187-19.
- 8. Jiang, B.; Liang, X.; Chen, Y.; Ma, T.; Liu, L.; Li, J.; Jiang, R.; Chen, T.; Zhang, X.; Li, S. Integrating next-generation seq uencing and traditional tongue diagnosis to determine tongue coating microbiome. Sci. Rep. 2012, 2, srep00936.
- 9. Hsu, C.-H.; Yu, M.-C.; Lee, C.-H.; Lee, T.-C.; Yang, S.-Y. High Eosinophil Cationic Protein Level in Asthmatic Patients w ith "Heat" Zheng. Am. J. Chin. Med. 2003, 31, 277–283.
- 10. Zhao, Y.; Mao, Y.-F.; Tang, Y.-S.; Ni, M.-Z.; Liu, Q.-H.; Wang, Y.; Feng, Q.; Peng, J.-H.; Hu, Y.-Y. Altered oral microbiota in chronic hepatitis B patients with different tongue coatings. World J. Gastroenterol. 2018, 24, 3448–3461.
- 11. Farrell, J.J.; Zhang, L.; Zhou, H.; Chia, D.; Elashoff, D.; Akin, D.; Paster, B.J.; Joshipura, K.; Wong, D.T.W. Variations of oral microbiota are associated with pancreatic diseases including pancreatic cancer. Gut 2011, 61, 582–588.
- 12. Mitsuhashi, K.; Nosho, K.; Sukawa, Y.; Matsunaga, Y.; Ito, M.; Kurihara, H.; Kanno, S.; Igarashi, H.; Naito, T.; Adachi, Y.; et al. Association of Fusobacterium species in pancreatic cancer tissues with molecular features and prognosis. Onco target 2015, 6, 7209–7220.
- 13. Ballini, A.; Di Palma, G.; Isacco, C.G.; Boccellino, M.; Di Domenico, M.; Santacroce, L.; Nguyễn, K.C.; Scacco, S.; Calvani, M.; Boddi, A.; et al. Oral Microbiota and Immune System Crosstalk: A Translational Research. Biology 2020, 9, 13 1.
- 14. Ramírez, J.H.; Parra, B.; Gutiérrez, S.; Arce, R.M.; Jaramillo, A.; Ariza, Y.; Contreras, A. Biomarkers of cardiovascular d isease are increased in untreated chronic periodontitis: A case control study. Aust. Dent. J. 2014, 59, 29–36.
- 15. Fava, F.; Danese, S. Intestinal microbiota in inflammatory bowel disease: Friend of foe? World J. Gastroenterol. 2011, 17, 557–566.
- 16. Ahn, J.; Chen, C.Y.; Hayes, R.B. Oral microbiome and oral and gastrointestinal cancer risk. Cancer Causes Control. 20 12, 23, 399–404.
- 17. Besnard, P.; Christensen, J.E.; Brignot, H.; Bernard, A.; Passilly-Degrace, P.; Nicklaus, S.; De Barros, J.-P.P.; Collet, X.; Lelouvier, B.; Servant, F.; et al. Author Correction: Obese Subjects With Specific Gustatory Papillae Microbiota and Sali vary Cues Display an Impairment to Sense Lipids. Sci. Rep. 2018, 8, 9773.
- 18. Chen, L.Y.; Lu, G.Z. Analysis of Tongue Images in 114 Patients with Gastric Cancer. Zhong Yi Za Zhi 2011, 52, 1935–1 938.
- 19. Miyazaki, H.; Sakao, S.; Katoh, Y.; Takehara, T. Correlation Between Volatile Sulphur Compounds and Certain Oral He alth Measurements in the General Population. J. Periodontol. 1995, 66, 679–684.
- 20. Xu, J.; Xiang, C.; Zhang, C.; Xu, B.; Wu, J.; Wang, R.; Yang, Y.; Shi, L.; Zhang, J.; Zhan, Z. Microbial biomarkers of common tongue coatings in patients with gastric cancer. Microb. Pathog. 2019, 127, 97–105.
- 21. Huang, S.; Yang, F.; Zeng, X.; Chen, J.; Li, R.; Wen, T.; Li, C.; Wei, W.; Liu, J.; Chen, L.; et al. Preliminary characterizati on of the oral microbiota of Chinese adults with and without gingivitis. BMC Oral Health 2011, 11, 33.
- 22. Kistler, J.O.; Booth, V.; Bradshaw, D.J.; Wade, W.G. Bacterial Community Development in Experimental Gingivitis. PLo S ONE 2013, 8, e71227.
- 23. He, Y.; Gong, D.; Shi, C.; Shao, F.; Shi, J.; Fei, J. Dysbiosis of oral buccal mucosa microbiota in patients with oral liche n planus. Oral Dis. 2017, 23, 674–682.
- 24. Cao, Y.; Qiao, M.; Tian, Z.; Yu, Y.; Xu, B.; Lao, W.; Ma, X.; Li, W. Comparative Analyses of Subgingival Microbiome in C hronic Periodontitis Patients with and Without IgA Nephropathy by High Throughput 16S rRNA Sequencing. Cell. Physi ol. Biochem. 2018, 47, 774–783.
- 25. Healy, C.; Moran, G.P. The microbiome and oral cancer: More questions than answers. Oral Oncol. 2019, 89, 30–33.
- 26. Mager, D.L.; Haffajee, A.D.; Devlin, P.M.; Norris, C.M.; Posner, M.R.; Goodson, J.M. The salivary microbiota as a diagn ostic indicator of oral cancer: A descriptive, non-randomized study of cancer-free and oral squamous cell carcinoma su

- bjects. J. Transl. Med. 2005, 3, 27.
- 27. Allali, I.; Boukhatem, N.; Bouguenouch, L.; Hardi, H.; Boudouaya, H.A.; Cadenas, M.B.; Ouldim, K.; Amzazi, S.; Azcarat e-Peril, M.A.; Ghazal, H. Gut microbiome of Moroccan colorectal cancer patients. Med Microbiol. Immunol. 2018, 207, 211–225.
- 28. Downes, J.; Sutcliffe, I.C.; Booth, V.; Wade, W.G. Prevotella maculosa sp. nov., isolated from the human oral cavity. Int. J. Syst. Evol. Microbiol. 2007, 57, 2936–2939.
- 29. Tsigarida, A.; Dabdoub, S.; Nagaraja, H.; Kumar, P. The Influence of Smoking on the Peri-Implant Microbiome. J. Dent. Res. 2015, 94, 1202–1217.
- 30. Ehrenfest, D.M.D.; Del Corso, M.; Inchingolo, F.; Charrier, J.-B. Selecting a relevant in vitro cell model for testing and c omparing the effects of a Choukroun's platelet-rich fibrin (PRF) membrane and a platelet-rich plasma (PRP) gel: Tricks and traps. Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endodontol. 2010, 110, 409–411.
- 31. Aas, J.A.; Griffen, A.L.; Dardis, S.R.; Lee, A.M.; Olsen, I.; Dewhirst, F.E.; Leys, E.J.; Paster, B.J. Bacteria of Dental Cari es in Primary and Permanent Teeth in Children and Young Adults. J. Clin. Microbiol. 2008, 46, 1407–1417.
- 32. Takahashi, N.; Nyvad, B. The Role of Bacteria in the Caries Process: Ecological perspectives. J. Dent. Res. 2010, 90, 2 94–303.
- 33. Goodson, J.; Groppo, D.; Halem, S.; Carpino, E. Is Obesity an Oral Bacterial Disease? J. Dent. Res. 2009, 88, 519–52 3.
- 34. Takeshita, T.; Kageyama, S.; Furuta, M.; Tsuboi, H.; Takeuchi, K.; Shibata, Y.; Shimazaki, Y.; Akifusa, S.; Ninomiya, T.; Kiyohara, Y.; et al. Bacterial diversity in saliva and oral health-related conditions: The Hisayama Study. Sci. Rep. 2016, 6, 22164.
- 35. Carlsson, J.; Grahnén, H.; Jonsson, G.; Wikner, S. Early Establishment of Streptococcus salivarius in the Mouths of Infants. J. Dent. Res. 1970, 49, 415–418.
- 36. Segata, N.; Haake, S.K.; Mannon, P.; Lemon, K.P.; Waldron, L.; Gevers, D.; Huttenhower, C.; Izard, J. Composition of t he adult digestive tract bacterial microbiome based on seven mouth surfaces, tonsils, throat and stool samples. Genom e Biol. 2012, 13, R42.
- 37. Chu, D.M.; Ma, J.; Prince, A.L.; Antony, K.M.; Seferovic, M.D.; Aagaard, K.M. Maturation of the infant microbiome community structure and function across multiple body sites and in relation to mode of delivery. Nat. Med. 2017, 23, 314–32 6.
- 38. Mason, M.R.; Chambers, S.; Dabdoub, S.; Thikkurissy, S.; Kumar, P.S. Characterizing oral microbial communities acros s dentition states and colonization niches. Microbiome 2018, 6, 1–10.
- 39. Seerangaiyan, K.; Jüch, F.; Winkel, E.G. Tongue coating: Its characteristics and role in intra-oral halitosis and general h ealth—A review. J. Breath Res. 2017, 12, 034001.
- 40. Loesche, W.J. Microbiology and treatment of halitosis. Curr. Infect. Dis. Rep. 2003, 5, 220-226.
- 41. NISC Comparative Sequencing Program; Oh, J.; Byrd, A.L.; Deming, C.; Conlan, S.; Kong, H.H.; Segre, J.A. Biogeogra phy and individuality shape function in the human skin metagenome. Nature 2014, 514, 59–64.
- 42. Ballini, A.; Gnoni, A.; De Vito, D.; DiPalma, G.; Cantore, S.; Isacco, C.G.; Saini, R.; Santacroce, L.; Topi, S.; Scarano, A.; et al. Effect of probiotics on the occurrence of nutrition absorption capacities in healthy children: A randomized doubl e-blinded placebo-controlled pilot study. Eur. Rev. Med. Pharmacol. Sci. 2019, 23, 8645–8657.
- 43. Inchingolo, F.; Martelli, F.S.; Isacco, C.G.; Borsani, E.; Cantore, S.; Corcioli, F.; Boddi, A.; Nguyễn, K.C.; De Vito, D.; Ait yan, S.K.; et al. Chronic Periodontitis and Immunity, Towards the Implementation of a Personalized Medicine: A Transla tional Research on Gene Single Nucleotide Polymorphisms (SNPs) Linked to Chronic Oral Dysbiosis in 96 Caucasian Patients. Biomedicines 2020, 8, 115.
- 44. Cantore, S.; Ballini, A.; De Vito, D.; Abbinante, A.; Altini, V.; DiPalma, G.; Inchingolo, F.; Saini, R. Clinical results of improvement in periodontal condition by administration of oral probiotics. J. Biol. Regul. Homeost. Agents 2018, 32, 1329–1 334
- 45. Inchingolo, F.; Santacroce, L.; Ballini, A.; Topi, S.; DiPalma, G.; Haxhirexha, K.; Bottalico, L.; Charitos, I.A. Oral Cancer: A Historical Review. Int. J. Environ. Res. Public Health 2020, 17, 3168.
- 46. Topi, S.; Santacroce, L.; Bottalico, L.; Ballini, A.; Inchingolo, A.D.; Dipalma, G.; Charitos, I.A.; Inchingolo, F. Gastric Cancer in History: A Perspective Interdisciplinary Study. Cancers 2020, 12, 264.
- 47. Boccellino, M.; Di Stasio, D.; Di Palma, G.; Cantore, S.; Ambrosio, P.; Coppola, M.; Quagliuolo, L.; Scarano, A.; Malcan gi, G.; Borsani, E.; et al. Steroids and growth factors in oral squamous cell carcinoma: Useful source of dental-derived s

- tem cells to develop a steroidogenic model in new clinical strategies. Eur. Rev. Med. Pharmacol. Sci. 2019, 23, 8730–8 740.
- 48. Yang, I.; Nell, S.; Suerbaum, S. Survival in hostile territory: The microbiota of the stomach. FEMS Microbiol. Rev. 2013, 37, 736–761.
- 49. Gibbons, R.J.; Houte, J.V. Bacterial Adherence in Oral Microbial Ecology. Annu. Rev. Microbiol. 1975, 29, 19-42.
- 50. Cabre, M.; Serra-Prat, M.; Palomera, E.; Almirall, J.; Pallares, R.; Clavé, P. Prevalence and prognostic implications of d ysphagia in elderly patients with pneumonia. Age Ageing 2010, 39, 39–45.
- 51. Abe, S.; Ishihara, K.; Adachi, M.; Okuda, K. Tongue-coating as risk indicator for aspiration pneumonia in edentate elderly. Arch. Gerontol. Geriatr. 2008, 47, 267–275.
- 52. Shimazaki, Y.; Tomioka, M.; Saito, T.; Nabeshima, F.; Ikematsu, H.; Koyano, K.; Yamashita, Y. Influence of oral health on febrile status in long-term hospitalized elderly patients. Arch. Gerontol. Geriatr. 2009, 48, 411–414.
- 53. Marik, P.E.; Kaplan, D. Aspiration Pneumonia and Dysphagia in the Elderly. Chest 2003, 124, 328-336.
- 54. Sjögren, P.; Nilsson, E.; Forsell, M.; Johansson, O.; Hoogstraate, J.; Sjögren, P. A Systematic Review of the Preventive Effect of Oral Hygiene on Pneumonia and Respiratory Tract Infection in Elderly People in Hospitals and Nursing Home s: Effect Estimates and Methodological Quality of Randomized Controlled Trials: Oral hygiene and pneumonia in elderly. J. Am. Geriatr. Soc. 2008, 56, 2124–2130.
- 55. Zhou, Y.; Gao, H.; Mihindukulasuriya, A.K.; La Rosa, P.S.; Wylie, K.M.; Vishnivetskaya, T.; Podar, M.; Warner, B.; Tarr, I. P.; Nelson, E.D.; et al. Biogeography of the ecosystems of the healthy human body. Genome Biol. 2013, 14, R1.
- 56. Humphrey, L.L.; Fu, R.; Buckley, D.I.; Freeman, M.; Helfand, M. Periodontal Disease and Coronary Heart Disease Incid ence: A Systematic Review and Meta-analysis. J. Gen. Intern. Med. 2008, 23, 2079–2086.
- 57. Ide, R.; Hoshuyama, T.; Wilson, D.; Takahashi, K.; Higashi, T. Periodontal Disease and Incident Diabetes: A Seven-Yea r Study. J. Dent. Res. 2011, 90, 41–46.
- 58. Yoneda, M.; Naka, S.; Nakano, K.; Wada, K.; Endo, H.; Mawatari, H.; Imajo, K.; Nomura, R.; Hokamura, K.; Ono, M.; et al. Involvement of a periodontal pathogen, Porphyromonas gingivalis on the pathogenesis of non-alcoholic fatty liver di sease. BMC Gastroenterol. 2012, 12, 16.
- 59. Iwauchi, M.; Horigome, A.; Ishikawa, K.; Mikuni, A.; Nakano, M.; Xiao, J.; Odamaki, T.; Hironaka, S. Relationship betwe en oral and gut microbiota in elderly people. Immun. Inflamm. Dis. 2019, 7, 229–236.
- 60. Dawes, C. Salivary flow patterns and the health of hard and soft oral tissues. J. Am. Dent. Assoc. 2008, 139, 18S-24S.
- 61. Isacco, C.G.; Ballini, A.; De Vito, D.; Nguyen, K.C.D.; Cantore, S.; Bottalico, L.; Quagliuolo, L.; Boccellino, M.; Di Dome nico, M.; Santacroce, L. Rebalance the Oral Microbiota as Efficacy Tool in Endocrine, Metabolic, and Immune Disorder s. Endocr. Metab. Immune Dis. Drug Targets 2021, 21, 777–784.
- 62. Hajishengallis, G.; Darveau, R.P.; Curtis, M.A. The keystone-pathogen hypothesis. Nat. Rev. Microbiol. 2012, 10, 717–725.
- 63. Ceccarelli, F.; Orrù, G.; Pilloni, A.; Bartosiewicz, I.; Perricone, C.; Martino, E.; Lucchetti, R.; Fais, S.; Vomero, M.; Olivier i, M.; et al. Porphyromonas gingivalisin the tongue biofilm is associated with clinical outcome in rheumatoid arthritis pati ents. Clin. Exp. Immunol. 2018, 194, 244–252.

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