

PEF Extract Plant Bioactive Components

Subjects: **Plant Sciences**

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Different parts of a plant (seeds, fruits, flower, leaves, stem, and roots) contain numerous biologically active compounds called “phytoconstituents” that consist of phenolics, minerals, amino acids, and vitamins. The conventional techniques applied to extract these phytoconstituents have several drawbacks including poor performance, low yields, more solvent use, long processing time, and thermally degrading by-products. In contrast, modern and advanced extraction nonthermal technologies such as pulsed electric field (PEF) assist in easier and efficient identification, characterization, and analysis of bioactive ingredients. Other advantages of PEF include cost-efficacy, less time, and solvent consumption with improved yields.

PEF

green extraction techniques

phytochemical extraction

oil extraction

food waste

1. Introduction

Plants with complex structures contain phenolic compounds in cell vacuoles and a lipoprotein bilayer. However, the membrane envelope controls intracellular movement within the intact cells ^{[1][2][3][4][5][6]}. From medieval cultures, natural products and food have been extracted as an essential source of spiritual, cosmetic, and nutritious ingredients. The Pharaonic civilization (more than 4000 years ago) was the first to use solid–liquid extraction to separate aromas, colors, and other materials from plants ^[7]. Soxhlet, heat reflux, soaking, water percolation, maceration, magnetic stirrer, boiling, and grinding are the traditional practices used to obtain phytoconstituents but involve high solvent consumption, a long processing time, poor or low extraction yields, and few have thermal degradation risks. To overcome the limitations of these traditional technologies, numerous innovative non-thermal extraction techniques have been evaluated ^{[8][9]}.

Pulsed electric field (PEF) has gained more attention in the past few years to extract beneficial materials from food waste/by-products through diffusion, osmosis, pressing, and drying ^[10]. It minimizes the deleterious effects of conventional heating processes ^{[11][12][13]}. PEF technology, as a promising alternative to other methods (boiling, microwave- and ultrasound-assisted extractions, etc.), has been used for the successful separation, intensification, stabilization, and dehydration of important compounds without affecting the nutritive properties ^{[14][15][16][17]}. Apart from improving the extraction, PEF has been recently proposed as a tool to induce stress in plant cells, thus stimulating the biosynthesis of active components ^[18].

Due to its ability to electroporate cell membrane, PEF is used as a pre-treatment to facilitate the recoveries of bioactive material followed by a subsequent traditional or novel extraction step ^[19]. The PEF method, when applied on water, showed a reduced temperature while consuming less solvent, and enhancing the rate at which the

constituents were extracted [20]. PEF has also been used for recycling food waste and by-products through extracting valuable ingredients [21]. It decreased energy costs, improved the extraction yield, lessened the degradation of heat-sensitive substances, and purified extraction with no environmental impact [15]. The current review confines the latest research on PEF assisted extraction focusing on its types, mechanism, and applications in the food industry.

2. Working Principle of PEF-Assisted Extraction

The PEF technique uses moderate to high electric field strength (EFS) ranging from 100–300 V/cm in batch mode and 20–80 kV/cm in continuous mode extraction. Two views are common among various hypotheses concerning the potential PEF mechanism. In the biological cell membrane, one is the speeding of chemical-based reactions from various compounds to enhance the solubility of solvent [22] and the other is the electroporation process. Electro-permeabilization or electroporation involves an external electrical force, which enhances the permeability of cell membranes [23]. Food or any other targeted materials are placed between the electrodes and a high-voltage electric field. The cell membrane is punctured by creating hydrophilic pores, which opened protein channels. The sample experiences a force per unit charge called the electric field when high-voltage electrical pulses are applied through the electrodes. The membrane loses its structural functionality, and the plant material is extracted [24]. This mechanism has been shown in **Figure 1**.

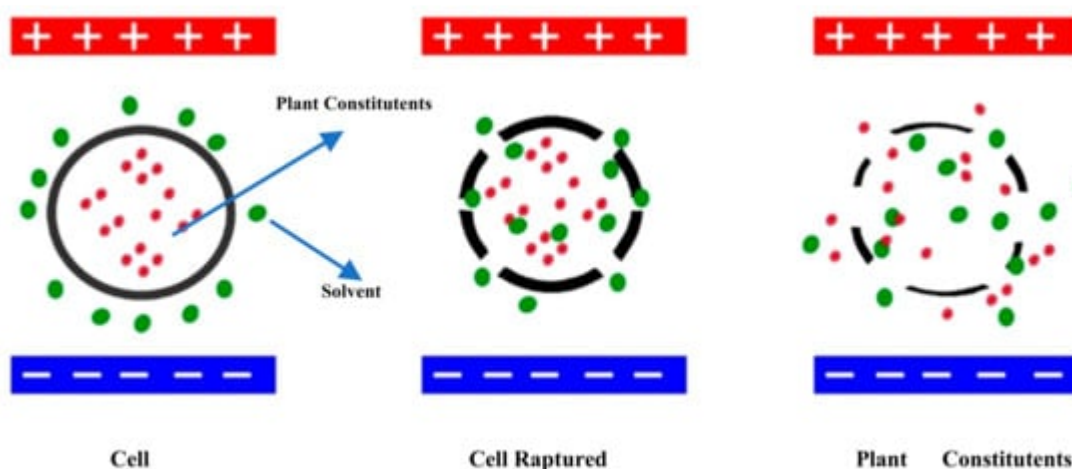


Figure 1. Electroporation mechanism for extraction.

An electric field can be applied either in exponentially decaying, oscillatory square waves, unipolar triangular, or bipolar pulses. The electroporation that occurs is either reversible or irreversible, but depending on the application, this effect can be controlled [25]. Generally, low specific energy (1–10 kJ/kg) and the time (nanoseconds to milliseconds) of the pulses cycle are efficient during the extraction process [26]. The electroporation of eukaryotic and prokaryotic cells and the formation of irreversible (permanent) and reversible (temporary) pores in their cell membranes were observed during the extraction process [27]. Irreversible electroporation increases the extraction process. However, EFS increases the cell membrane permeability, which depends on the size of the cell and cell geometry [28]. The strength of an electric field ranging from 0.1 to 10 kV/cm is enough for delicate plant tissues

(e.g., pericarp or mesocarp of few fruits); however, tough materials such as seeds need high intensities (i.e., 10 to 20 kV/cm) for efficient extraction [29]. This also provides some additional benefits such as preserving the nutritional, and sensory characteristics of liquid foods [30].

The cells subjected to an external electric field exhibited a high transmembrane potential due to the accumulation of charge on the surface of the membrane. To protect the membrane, a cell has an electrical endurance limit, i.e., cell membranes can endure a specific electrical field strength without considerable damage. The critical electric field is the smallest threshold value for the same cell type. This will facilitate the development of pores in vulnerable areas of the membrane that can be reversible or permanent, depending on the strength of the electric field. The EFS is directly proportional to the magnitude of the damage. Damage is reversible (temporary) when the intensity is low to moderate (not much greater than the critical value). An electric field of high intensity caused irreversible (permanent) damage to the cell membrane [31]. Accordingly, PEF enhances the mass transfer by demolishing the structure of the cell membrane [22]. The increased membrane permeability led to cell breakdown, while the increased rate of mass transfer assisted the discharge of intracellular substances [32]. The degree of electroporation depends on the EFS, type and number of pulses waveform, treatment time, and targeted plant material [33]. However, the specific EFS depends on the geometry and distance of the electrodes.

3. PEF-Assisted Extraction Equipment

A PEF unit consists of a high-voltage pulse generator, a treatment chamber with a fluid managing assembly and a monitoring and control system [34]. Additionally, PEF equipment has a charger (to convert AC in DC) and a device that stores energy in the generator. A high-voltage circuit is switched on and off to generate electric pulses. During the discharge of high electric energy, the high-voltage value and short time pulses make this process more complicated, while the capacitor is continuously monitored and stepped up if the voltage is interrupted [35]. The treatment chamber consists of two separate electrodes (one electrode attached to the high-voltage generator and the other to the ground) and a gap; filled with the targeted food. The different electric potential on either side of the membrane generates an electric field, which depends on the electrodes type and the distance among them and with the sample. Other factors include the electric pulses nature, the configuration of the treatment chamber, and the product's conductivity [28]. The PEF treatment parameters required to enhance polyphenols extraction, based on empirical experience, are categorized as high ($E > 1 \text{ kV cm}^{-1}$), medium ($E \approx 0.1\text{--}1 \text{ kV cm}^{-1}$), and low ($E < 0.1 \text{ kV cm}^{-1}$) electric fields [35]. Generally, the PEF extraction process can be grouped into the following two broad categories: one is batch, and the other is a continuous system of extraction concerning the way of operation [22].

3.1. PEF Batch Extraction

A standard batch extraction unit comprises a pretreatment chamber for PEF and essential equipment for solid–liquid extraction. The pretreatment unit of PEF contains a cylindrical vessel of polypropylene (30 mm diameter on the inner side), having two stainless steel electrodes arranged parallel to each other at a 10-millimeter distance. The processing parameters include electrical field strength, pulses width, shape, number, and frequency. Inside the cylindrical unit, the sample with a little solvent was first treated between two electrodes and connected to a PEF

generator. The treated sample was removed from the cylindrical unit and stirred at different velocities using a magnetic agitator to prevent solution evaporation [22].

Using a PEF batch extraction method, various intracellular compounds such as lipid [36], red beetroot pigment [37], anthocyanin [35], betanine [22], oil [38], polyphenols [31], and cellulose [39] have been extracted. **Figure 2** shows the PEF batch extraction system.

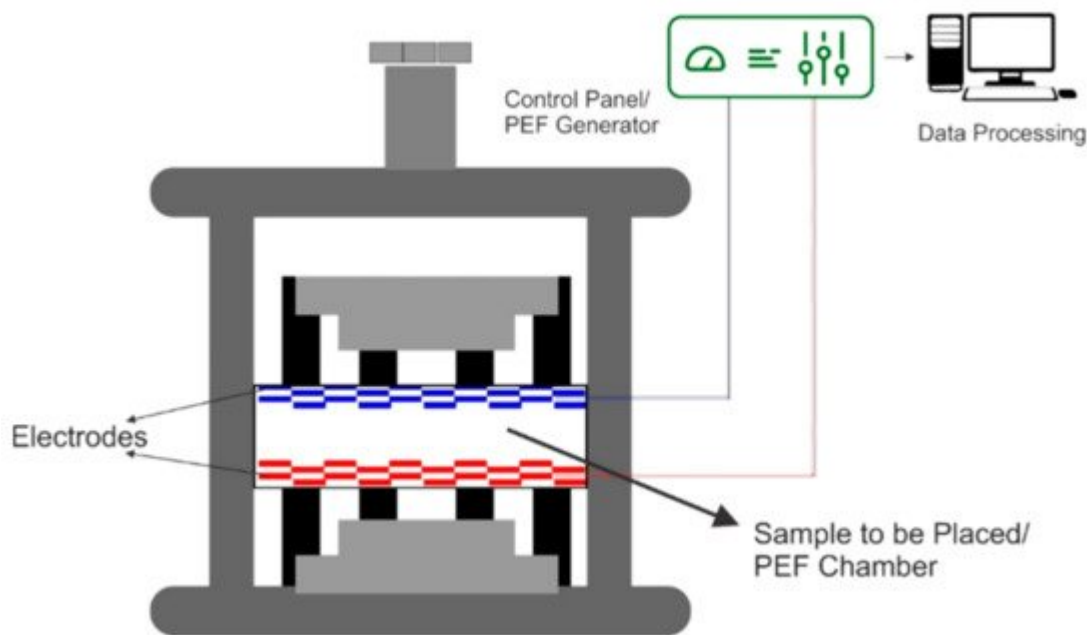


Figure 2. PEF (pulsed electric field) batch extraction system.

3.2. PEF Continuous Extraction

The batch extraction process of PEF gave promising results in terms of extraction; however, a significant increase in the operating time has been observed, which is due to the low capacity of batch mode systems. Therefore, it is crucial to devise a PEF extraction process to perform it in regular conditions at the industrial level. Since then, the PEF continuous extraction method has been in great focus. The continuous-flow treatment chamber was first successfully used by Yongguang et al. [35] for the extraction of polysaccharides from *Rana temporaria chensinensis* David in 2006 [40]. Their results depicted that the extraction yield was 55.59% with PEF (20 kV/cm, 0.5% of KOH) as compared to the traditional extraction technique. In the past few years, the continuous extraction PEF applied successfully in laboratories to obtain fishbone, broccoli juice, eggshell, tomato juice, etc. This technology has been applied in industries thus far.

A typical PEF continuous extraction system consists of a high-voltage pulse generator, a treatment chamber, a suitable product handling system, and a range of monitoring and controlling equipment. The oscilloscope can read the output voltage (up to 40 kV pulse voltage) directly, and the frequency is adjustable (40–3000 Hz). During continuous extraction PEF, the solvent mixture is pumped into the treatment chamber by a peristaltic pump at a constant fluid velocity. The cooling coil has a temperature of 25 °C in a water bath controlled by a thermostat during

the extraction procedure. Coaxial and the cofield continuous PEF treatment chambers are currently widely used due to their simple configurations [22]. **Figure 3** shows the PEF continuous extraction system.

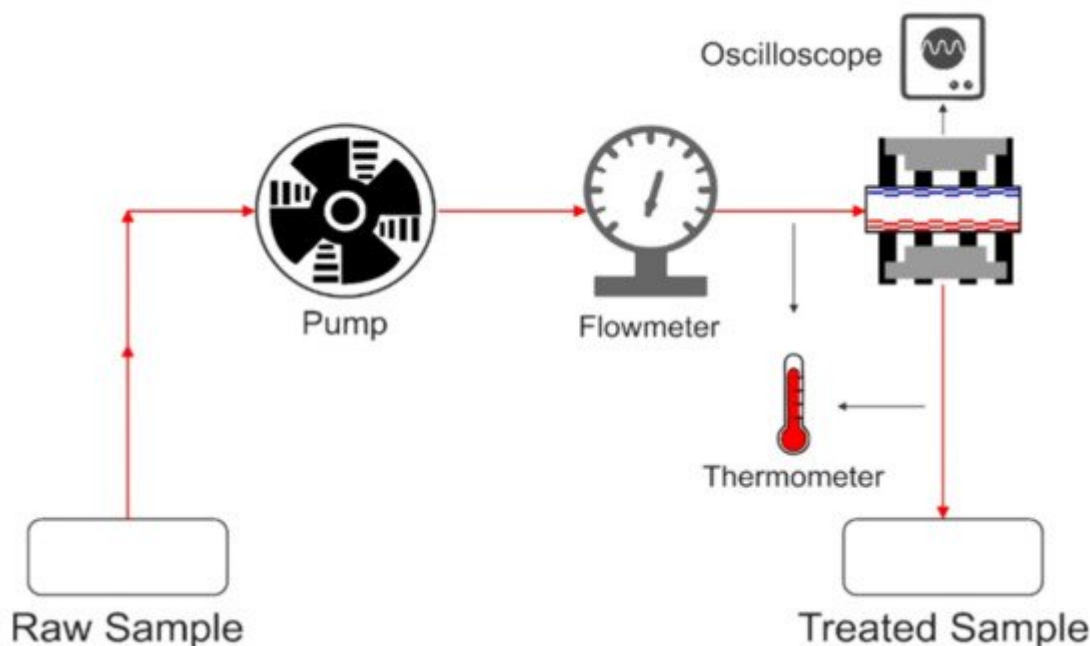


Figure 3. PEF (pulsed electric field) continuous extraction system.

4. Factors Influencing the PEF Extraction

The efficacy of the PEF extracting system depends not only on the processing parameters, but also on the solvent nature, sample composition (size, shape, pH, conductivity, etc.), and the extracted component's size and position in the cell cytoplasm or vacuoles. Moreover, the characteristics of the tissues and cells have a powerful impact on the efficiency of PEF-assisted extraction [28][41]. The extraction process improved in the low ionic strength. The ionic strength affects the cytoplasmic system in terms of the cell's compression and electroporation. However, the conductivity of the matrix affects the behavior of the electric field that passes through that matrix significantly [42].

EFS is a significant parameter in determining the degree of extraction as it affects the physical properties of the targeted compound such as diffusivity, surface tension, viscosity, and solubility [14]. The critical point is to assure the uniform distribution of electric fields across the treatment chamber. Electric field energy delivers through bipolar, exponential decaying, oscillatory, and square pulses. Among them, exponential square wave pulses are widely used in the PEF extraction process due to their high energy and lethal performance. Moreover, square waves are the most prevalent waveform used for the extraction process.

Generally, 12 to 45 kV/cm is enough to extract the valuable components from food; however, the PEF intensity depends on the food characteristics. Similarly, the extraction of target compounds is improved with an increase in the EFS due to high energy transfer in the food sample. Moreover, the PEF intensity (5–10 kV/cm) increased the hydrolysis rate and the total amino nitrogen content of abalone viscera protein. The results showed that the mild

PEF intensity caused the cell membrane to undergo reversible electroporation. Hence, mild PEF intensity strength reclosed the cell electric hole. However, a PEF intensity greater than 10 kV/cm exudated the intracellular material due to an irreversible breakdown of the cell membrane. According to the authors, 20 kV/cm was the optimal EFS for a better extraction of bioactive compounds [43]. Lin et al. [44] found a significant increase (5.042 ± 0.04 to 6.996 ± 0.03 mg/mL) in the calcium malate extraction from an eggshell using PEF at 0 to 10 kV/cm EFS. According to the authors, the PEF treatments accelerated the movements between ionic groups and electrons having more kinetic energy, which increased the extraction rate of calcium malate and malic acid. However, the higher EFS facilitated the dissolution of calcium ions. Alternatively, when the pulse time raised from 16 to 20 s, the content of dissoluble calcium malate was significantly reduced. They further explained that the narrow pulses excited the material to resonance vibration. It was also established that, the extreme high EFS, i.e., 35 kV/cm, caused the vitamin C degradation of broccoli juice [14].

Treatment temperature is another important factor affecting the PEF extraction process [45]. PEF extraction technology is a nonthermal process, thus it operates in room or near room temperature. Higher temperatures usually decrease the viscosity of liquid solvents, which is destructive for the extraction process. Treatment time (pulse numbers and width) is another parameter to measure PEF efficiency [42]. However, an increase in treatment time could raise the temperature of the product. According to the results, longer pulses maximized the extraction rate of polyphenols from fresh tea leaves at 0.9 kV/cm for 0.5 s and 1.1 kV/cm for 3 s [46]. Other than that, the selection of solvent is also crucial for better PEF extraction. It involves numerous factors such as the solubility, conductivity, and polarity of the solvent. The increased solvent conductivity enhanced the cell membrane electroporation that ultimately improved the extraction rate. Similarly, the high solubility of the extract in the solvent and the strong polarity of the solvent increased the mass transfer rate and extraction rate [47][48][49].

5. Applications

5.1. Fruits and Vegetables

Due to their nutritional value worldwide and the fact that they are rich sources of beneficial antioxidants, minerals, vitamins and fibers [50], fruits, and vegetables are the most common source of nutrition. A variety of antioxidant compounds can be present in fruits and vegetables, including phenolics, carotenoids, anthocyanins, and tocopherols [51]. PEF is a promising method to extract bioactive compounds (anthocyanins, betanines, carotenoids, etc.) from fruit and vegetables, as shown in **Table 1**. Several factors depend on the extraction of bioactive compounds, such as the process of extraction, the solvent used for extraction and raw materials [52]. Traditional extraction techniques such as Hydro distillation, maceration, and Soxhlet require agitation, high temperatures, and chemical or organic solvents [53]. Nevertheless, the Soxhlet method needs considerable time for extraction and significant quantities of the solvent [54].

Leong et al. [55] evaluated the extraction of anthocyanins from grape juices using PEF technology with 20 μ S pulse, 50 Hz frequency and 1.5 kV/cm electric field power. PEF-assisted extraction improved the release of vitamin C and anthocyanins as well as enhanced the antioxidant activity of grape juice as compared to untreated samples. The

resultant extract had good phytochemical composition and protected the cells from oxidation [55]. Similarly, the PEF treatment enhanced the antioxidant activity (1.03 times) and extraction of polyphenols (1.44 times) of green grape juice [56]. Moreover, the PEF pretreatment enhanced the recovery of red raspberry (*Rubus idaeus* L.) juice by 9–25% without affecting the total phenolic and anthocyanin content. Furthermore, it enhanced the red raspberry press-cake extract that involved a 20% increase in phenolic content and a 26% increase in anthocyanin content [57]. In another study, PEF (13.3 kV/cm, 0–564 kJ/kg) enhanced the extraction yields of fermented grape (*Dunkelfelder*) pomace up to 22 and 55%. The results showed more recovery of anthocyanins as compared to traditional grinding methods, ultrasound-assisted extraction, and high-voltage electrical discharge [58].

PEF treatment is the most suitable nonthermal technique to extract phenols and flavonoids from onions without significant quality losses. Plant tissues contain a cytomembrane, which affects the movement of intracellular substances between cells. PEF treatments altered the functionality (permeability) of cytomembrane by disintegration and improved the movement of mass through cells, providing greater yields. Compared to the control samples, the PEF treatment significantly enhanced the phenolic compounds (102.86 mg GAE/100 g) by 2.2 times and the flavonoid compounds (37.58 mg QE/100 g) by 2.7 times in onions [59]. According to Fincan [60], the disintegration index was 0.86 ± 0.02 at 99 pulses of 3 kV/cm (4102 ± 239 J/kg) in the PEF-treated sesame seed cake. The extraction yields (total phenolic, antioxidant power, and antioxidant activity) produced using microwave and heat technology was comparable with PEF-assisted extraction [60]. Further, Sarkis et al. [61] stated that the disintegration index and the proteins and polyphenols content increased until 83 kJ/kg energy inputs in the case of spearmint [61].

Xue et al. [62] showed that PEF treatment (38.4 kV/cm, 272 μ s) produced higher extraction yields of polysaccharides (97.7%), protein (48.9%), and polyphenolic compounds (50.9%) in white button mushrooms compared to traditional thermal treatment (95 °C for 1 h). The traditional technique has a prolonged treatment cycle as compared to the PEF system. However, PEF exhibited a fluid residence time of less than 2.6 min [62]. In the same context, Jeya et al. [25] investigated that PEF at 7 kV/cm and 2.5 kJ/kg resulted in a 4.2-fold enhancement in the yield of betanine as compared to untreated samples. According to the authors, 90% of betanine extraction was achieved in 35 min. Frontuto et al. [63] indicated that the PEF pretreatment of potato peel tissues greatly enhanced the extraction of phenolics. Accordingly, PEF pretreatment achieved the same amount (1062 mg GAE/kg) of phenolics in 144 min as compared to untreated samples (240 min) [63]. Similarly, PEF treatment (2 kV/cm, 11.225 kJ/kg) significantly enhanced the yield (13.3%) of phenolic content in olive paste [64].

Table 1. PEF Assisted extraction of Bioactive compounds.

Ref.	Raw Material	Extraction Technique	Pretreatment Condition	Extraction Condition	Solid-Solvent Ratio	Solvent	Yield	TPC	DPPH	FRAP	IC50	TFC	TAC	TCC
[65]	Cinnamon (<i>Cinnamomum verum</i>) powder	PEF-assisted extraction	Frequency: 1 Hz Voltage: 2–6 5.12 kV/cm No.	Temperature: ambient Time: 48 h	1:10 w/v	Ethanol 100 mL	5.06 %	505.9 mg GA/kg	91.7%	NR	NR	NR	NR	NR

Ref.	Raw Material	Extraction Technique	Pretreatment Condition	Extraction Condition	Solid-Solvent Ratio	Solvent	Yield	TPC	DPPH	FRAP	IC50	TFC	TAC	TCC
of pulses: 40–60 NR														
[66]	<i>Nepeta binaludensis</i>	PEF-assisted extraction	No. of pulses:60 Frequency: 1 Hz Voltage: 6 kV No. of pulses: 60	Temperature: ambient Time: 48 h	1:10 w/v	Ethanol 100 mL	11.36%	417.85 mg GA/g	74.8%	1688.53 μ mol Fe ²⁺ /g	0.32 mg/mL	NR	NR	NR
[24]	Thinned peach (<i>Prunus persica</i>)	PEF-assisted extraction	EFS: 0 kV/cm Specific energy: 0.61–9.98 kJ/kg Pulse frequency: 1 Hz No. of pulses per time: 30–150 μ s	Temperature: 35 °C Time: 10 h	NR	Methanol 80% 200 mL water–methanol solvent	NR	83.3 mg GAE/100 g	57.8 %	NR	NR	54.3 CE/100 g	NR	NR
[67]	Tomato (<i>Solanum lycopersicum</i>) peel	PEF-assisted extraction	EFS:5 kV/cm Total specific energy: 5 kJ/kg	-	1:40 g/mL	Acetone	NR	NR	4.2 \pm 0.4 mmol TE/100 g FW	NR	NR	NR	NR	80.4 \pm 2.2 mg/100 g FW
			-	Temperature: 50 °C Time: 4 h Speed: 160 rpm					5.2 \pm 0.4 mmolTE/100 g FW					84.0 \pm 8.3 mg/100 g FW
[68]	Sweet Cherries (<i>Prunus avium</i>)	PEF-assisted pressing	Variable field strength: 1 kV/cm Frequency: 5 Hz Pulse width: 20 μ s Total specific energy input: 10 kJ/kg	Pressure: 1.64 bar Time: 5 min	NR	NR	Juice yield40%	NR	NR	27.4%	NR	NR	29.2 \pm 1.1 mg/100 mL	NR
	Sweet cherries (<i>Prunus avium</i>) press cake	PEF-assisted extraction	Variable field strength: 0.5–1 kV/cm Frequency: 5 Hz	Time: 24 h Temperature: 25 °C	5:1 mL/g Solvent–cake ratio	Acidified aqueous ethanol (50% ethanol;	NR	NR	NR	21.0%	NR	NR	218.0 \pm 14.8 mg/100 mL	NR

Ref.	Raw Material	Extraction Technique	Pretreatment Condition	Extraction Condition	Solid-Solvent Ratio	Solvent	Yield	TPC	DPPH	FRAP	IC50	TFC	TAC	TCC
			Pulse width: 20 µs Total specific energy input: 10 kJ/kg			0.5% HCl, v/v)								
[15]	Moringa olifera dry leaves	PEF-assisted extraction	EFS: 7 kV/cm	Time: 40 min Temperature: ambient Pulse duration: 20 ms Pulse interval: 100 µs	NR	NR	NR	40.24 mg GAE/g of dry matter	98.31	108.22 µmol AAE/g dry matter	NR	NR	NR	NR
[63]	Potato (Solanum tuberosum) peels	PEF-assisted extraction	Pulse width: 3–25 µs Frequency: 1–450 Hz Electric field: 1 kV/cm Specific energy: 5 kJ/kg	Time: 30–240 min Temperature: 20–50 °C Speed: 160 rpm	1:20 g/mL	Water–ethanol mixture Ethanol concentration 50%	NR	1263.5 ± 43 mg GAE/kg FW PP	877.17/kg FWPP	NR	NR	NR	NR	NR
[69]	Date palm (Phoenix dactylifera)	PEF-assisted ethanolic extraction	EFS: 3 kV/cm Frequency: 10 Hz	Time: 6 h	4:1 v/v	Ethanol–water 300 mL	NR	67.35 mg GAE/100 g	50–72	NR	110 µL/mL	6.75 mg CE/100 g	2.08 mg/L	6.10 µg/mL
[70]	Tropical almond red leaves (Terminalia catappa)	PEF-assisted extraction	NR	Frequency: 1 Hz Electric field intensity: 0.75 kV/cm No. of pulses: 50 n	1:10	Water	74.6%	241.40 ± 2.15 mg GAE/g	93.40 ± 1.23%	NR	42 mg/mL	NR	NR	NR
[59]	Red onion (Allium cepa)	PEF-assisted water extraction	Pulse wide: 100 µs Frequency: 1 Hz Electric field intensity: 2.5 kV/cm No. of pulse: 90 Specific energy: 0.23–9.38 kJ/kg	Time: 2 h Shaking speed: 200 rpm/min Temperature: 42.5 °C	NR	Distilled water 50 mL	NR	102.86 mg GAE/100 g FW	262.39 %	[77]	NR	37.58 mg QE/100 g FW	NR	NR

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using PEF to recover valuable compounds that would contribute to the notion of zero waste.

Conventionally, these compounds are extracted by different treatments such as nanoemulsions used in the nutraceutical, cosmetics, and pharmaceutical industries [75]. The traditional solvent extraction techniques are time and energy consuming. For instance, the industrial batch extraction of polyphenols from grape peels is usually carried out at 50–60 °C for 20 h. Furthermore, the bioactive is enclosed in plant cell vacuoles and membrane bilayers (insoluble structures) that are not open to solvents. The usage of high temperature to enhance mass transfer and reduction in time may have serious drawbacks since high temperature (greater than 70 °C) caused rapid degradation of heat-sensitive components such as anthocyanins [78]. Kantar et al. [79] compared the total polyphenolic content of orange, grapefruit, and lemon in both juice extract and peel (flavedo/albedo) using PEF (3–10 kV/cm) as a pretreatment followed by the traditional extraction procedure (50% ethanol for 1h). The results showed an improved flavonoid content and total phenolic contents (2200 mg GAE/100 g) of fruit peels at 10 kV/cm [79].

5.3. Herbs and Spices

Herbs and spices have been widely used to strengthen or enhance the taste and preserve the quality of food along with their beneficial effects on human health. The plant essential oils contain 85% of polyphenols, terpenes, monoterpenes, and sesquiterpenes. However, essential oils (derived from herbs and spices) have 70 or more

Ref.	Raw Material	Extraction Technique	Pretreatment Condition	Extraction Condition	Solid-Solvent Ratio	Solvent	Yield	TPC	DPPH	FRAP	IC50	TFC	TAC	TCC
[71]	Wild blueberries (<i>Vaccinium myrtillus</i>)	PEF-assisted pressing	EFS:3 kV/cm Total specific energy: 10 kJ/kg Frequency: 10 Hz Pulse width: 20 µs	Time: 8 min Pressure: 1.32 bar	NR	NR	56.3	45.5%	NR	35.9 %	NR	NR	77.5%	NR
[72]	Sour cherries (<i>Prunus cerasus</i>)	PEF-assisted pressing	EFS: 5 kV/cm total specific input:10 kJ/kg constant frequency: 10 Hz pulse width:20 µs	Pressing time: 9 min	NR	NR	37.7 g 100/g1	133.90 ± 2.67 mg 100/mL Juice	NR	6.60 ± 0.13 µmol TE/mL	NR	NR	53.30 ± 0.97 mg 100/mL Juice	NR
[73]	Fresh pomelo fruits (Shantian Variety)	-	Number of pulses of 30 Electric field intensity: 4 kV/cm	Temperature: 40 °C	40:60, v/v	90 mL ethanol–water	16.19 mg/mL (naringin) increased by about 20%	NR	38.58% increased by 70%	NR	NR	NR	NR	NR
[74]	Thawed blueberries [85]	PEF-assisted pressing	EFS: 1, 3 kV/cm Total specific energy: 10 kJ/kg Constant frequency: 20 Hz Pulse width: 20 µs	Constant pressure: 1.32 bar Time: 8 min	NR	NR	36.3 ± 1%	8.0%	NR	NR	NR	NR	8.3% [11]	NR

Phyllanthus emblica L. (*Syn. Emblica officinalis*) is an ancient herb well known for its functional properties. PEF (18 to 24 kV/cm, 300 to 1000 µs) treated *Emblica* juice showed an improved (9 times) extraction of quercetin and ellagic acid than thermally processed juice. According to the authors, 22 kV/cm was the optimum electric field power for 0.79 disintegration index within 500 µs [85]. Similarly, PEF extraction at 20 kV/cm using 70% ethanol and water increased the yield (12.69 mg/g) of ginsenosides as compared to ultrasound assisted extraction, microwave assisted extraction, heat reflux extraction, and pressurized liquid extraction. The authors stated that it took less than 1 s to complete PEF extraction, which is much lower than the other methods tested [86].

5.4. Leaves

According to Segovia et al. [87], PEF treatment (300 Hz, 30 kV) increased the polyphenols from 1.3 to 6.6% and the Oxygen Radical Absorption Capacity from 2.0 to 13.7% in *Borago officinalis* L. leaves. Moreover, PEF-assisted extraction increased the antioxidant capacity of the extracts and reduced the extraction times [87]. Barba et al. [88] established that PEF extraction treatments (0 to 141 kJ/kg, using water as a solvent) disrupt the plant cells of *Stevia rebaudiana* Bertoni leaves. PEF extraction improved conductivity (≈25%) and soluble matter extraction yields (≈33%) as compared to diffusion. Furthermore, PEF enhanced the antioxidant activity (50%), total phenolic compounds activity (80%), chlorogenic acid (93%), caffeic acids (55%), ferulic acid (90%), and protocatechuic acids (45%) [88]. **Table 2** summarizes the range of parameters used in PEF-assisted extraction for the release of bioactive components from Agro-Industrial waste.

Table 2. The extraction of bioactive compounds assisted by PEF pretreatment.

Ref.	Raw Material	PEF-Pretreatment	Extraction Method	Extraction Conditions	Solid-Solvent Ratio	Solvent	Yield	TPC	AA	TAC	DPPH	FRAP	TFC
[89]	Thawed blackcurrant (<i>Ribes nigrum</i>)	EFS: 1318 kV/cm Pulses: 315	Cold pressing	Power: 150 W Velocity: 70 rpm Pressing time: 1.5 min	NR	NR	NR	3.8 ± 0.2 mg GA/g	1.88 ± 0.06 mg GA/g	NR	NR	NR	NR
[72]	Blueberry (<i>Vaccinium myrtillus</i>) press cake	EFS: 5 kV/cm	Solid–liquid extraction	Temperature: ambient Time: 24 h Shaking speed: 150 rpm	6:1 mL/g Solvent to press cake ratio	50% Ethanol; 0.5% HCl, v/v	NR	89.3%	NR	111%	NR	80%	NR
[90]	Fresh tea leaves (<i>Camellia sinensis</i>)	EFS: 1.00 kV/cm Pulses: 100 Energy: 22 kJ/kg Temperature: 1.5 °C	Organic solvent extraction	Time: 2 h Temperature: room Stirring speed: 250 rpm	1:1000 Biomass to solvent ratio	50% acetone/water (w/w) solution	NR	398 mg/L	NR	NR	NR	NR	NR
[72]	Sour cherry (<i>Prunus cerasus</i>) press cake	EFS: 5 kV/cm Specific energy input: 10 kJ/kg Frequency: 10 Hz Pulse width: 20 µs	Solvent extraction	Time: 24 h Temperature: ambient	10:1 v/w solvent to press cake	Acidified aqueous methanol 70% MeOH and 0.5% HCl, v/v	NR	407.90 ± 18.54 mg/100 g	NR	168.50 ± 3.99 mg/100 g	NR	56.10 ± 1.25 µmol TE/g	NR
[91]	Basil leaves (<i>Ocimum Americanum</i>)	EFS: 2–3 kV/cm Time: 1–2 min	Conventional method: maceration	Time: 3 h Temperature: room	NR	Distilled water 300 mL	33.15% ± 2.484%	115.203 ± 1.115 mg GAE/g extract	NR	NR	NR	NR	75.816 ± 0.723 mg QE/g
[19]	Rosmary (<i>Salvia rosmarinus</i> by)—product	Frequency: 10 Hz Pulse width: 30 µs Pulses: 167 EFS: 1.1 kV/cm Specific energy input 0.36/kg 24 g of 0.1%	Ultrasound assisted	Power: 200 W Temperature: 40 °C Time: 12.48 min	(1: 20 w/v)	100 mL of 55.19% aqueous EtOH	NR	297 mg GAE/100 g FW	NR	NR	593 mg TE/100 g FW	NR	NR

Ref.	Raw Material	PEF-Pretreatment	Extraction Method	Extraction Conditions	Solid-Solvent Ratio	Solvent	Yield	TPC	AA	TAC	DPPH	FRAP	TFC
[94][95]	Thyme (<i>Thymus vulgaris</i>) by-product	aqueous NaCl (1:1.4 w/v)						460 mg GAE/100 g FW	[93]		570 mg TE/100 g FW		
		Frequency: 10 Hz Pulse width: 30 µs Pulses:167 EFS: 1.1 kV/cm Specific energy input 0.46 kJ/kg 24 g of 0.1% aqueous NaCl 24 g of 0.1% aqueous NaCl (1:1.5 w/v)											
[57]	[15] Raspberry press cake	EFS: 1 kV/cm, 3 kV/cm Total specific energy: 1 kJ/kg, 6 kJ/kg Frequency: 20 Hz Pulse width: 20 µs	Solvent extraction	Temperature: ambient Time: 24 h Shaking speed: 150 rpm	Solvent to press cake 6:1 mL/g	50% ethanol, 0.5% HCl, v/v	NR	420.8 ± 39.33 mg GAE/100 g	NR	50.7 ± 3.38 mg/100 g	NR	24%	NR

nutritional value and freshness of the product [98].

Table 3. PEF assisted extraction of oils.

Ref.	Raw Material	PEF Pretreatment Conditions	Extraction Method	Extraction Equipment	Extraction Conditions	Solvent	Oil Yield
[99]	Sunflower seeds	EFS: 7.0 kV/cm Frequency: 0.5 Hz Solvent content: 50 wt.% Time: 90 s Pulse width: 30 µs	Solvent extraction	Shaker	Frequency: 400/min Time: 3 h Temperature: room	Bioethanol	55.9%
[100]	Damask rose flowers	EFS: 20 kV/cm Pulse number: 8	PEF-assisted Hydro distillation	Clevenger-type micro-apparatus	Distillation temperature: 100 °C Distillation time: 2 h	10% sodium chloride	yield of 0.105% with a 50% increase in essential oil
[101]	Sunflower seeds	EFS: 7 kV/cm Frequency: 1.5 Hz	Solvent extraction	Shaker	Frequency: 400/min Time: 3 h	Hexane 40 mL (50 wt.%)	48.24%

Ref.	Raw Material	PEF Pretreatment Conditions	Extraction Method	Extraction Equipment	Extraction Conditions	Solvent	Oil Yield
		Treatment time: 30 s Pulse width: 30 μs			Temperature: room		
[64]	Olive fruits (Arroniz variety)	NR	Pilot PEF-assisted extraction	Pilot PEF system	Electric fields: 2 kV/cm Frequency: 25 Hz flow rate: 520 kg/h Specific energy: 11.25 kJ/kg	NR	22.66 kg/100 kg
[102]	Olives (Nocellara del Belice variety)	EFS: 2 kV/cm Frequency: 25 Hz Pulse width: 50 μs Mass flow rate: 2300 kg/h Energy delivered per pulse: 210 J Specific energy: 7.83 kJ/kg	NR	Hammer crusher Malaxation machine	Malaxation time: 30 min Temperature: 27 °C	NR	85.5%

cell; therefore, it is essential to extract and purify them before use [103]. Microorganisms can carry out a broad range of reactions and are adaptable to a variety of conditions. They can be moved from nature into the laboratory to produce beneficial compounds on cheap sources such as carbon and nitrogen. Due to the biological activity of secondary metabolites originating from microbes, they are greatly beneficial for our nutrition and health. Research is progressed in the screening of naturally occurring microbial products, for the development of novel therapeutic agents. The search of novel chemicals is an important way forward to study the capacity of lesser-known or novel bacterial taxa [104].

Martínez et al. [105] used PEF (15–25 kV/cm, 60–150 μs, 10–40 °C) to enhance the extraction of a selective protein called phycocyanin (a water-soluble protein) from *Arthrospira platensis*’s fresh biomass. According to the authors, low molecular weight compounds move directly through the cytoplasmic membrane after electroporation, while high molecular weight compounds require that the pores produced by the PEF treatment expand over time. Hence, the resultant delay of 150 min was recorded during the extraction. [105]. Similarly, Jaeschke et al. [106] also succeeded in achieving a high protein and phycocyanin yield from *Arthrospira platensis* after a PEF treatment of 40 kV/cm utilizing pulses of 1 μs (112 kJ/kg) [106]. Martínez et al. [107] extracted another water-soluble protein (β-phycocerythrin) from *Porphyridium cruentum* by applying PEF for 24 h. However, this water-soluble phycobiliprotein

was unobservable in the untreated cells, even after extended periods of incubation. The results showed that the extraction of β -phycoerythrin involved not only the diffusion of the pigment through the cell membrane, but also the disassembly from the cell organization of the molecule. In this way, PEF released the *P. cruentum* organelles (hydrolytic enzymes) that broke the bonds between the pigment and other cell compounds; thus, the water- β -phycoerythrin complex diffused, carried by a concentration gradient, through the membrane. The enzymatic autolysis of microalgae caused by PEF must be further investigated for the industrial implementation of PEF extraction technology [108]. **Table 4** further confines some of the literature on PEF that involves the extraction of other phytoconstituents.

Table 4. The extraction of other phytoconstituents.

Phycocyanin						
References	Rawmaterial	PEFtreatment	Extractionmethod	Extraction conditions	Solvent	Yield
[105]	<i>Arthrospira platensis</i>	EFS: 25 kV/cm Treatment time: 150 μ s Temperature: 40 °C	PEF-assisted extraction	Temperature: 20 °C Time: 360 min	19 mL of distilled water	Extraction yield 151.94 \pm 14.22 mg/g
Total Amino Nitrogen Content (TANC)						
References	Raw material	PEF treatment	Extractionmethod	Extractionconditions	Solvent	Yield
[43]	Fresh abalone (<i>Haliotis Discus Hannai Ino</i>) viscera	NR	PEF-assisted enzymatic extraction	EFS: 20 kV/cm treatment times: 600 s	NR	42.35%, 175.20 mg/100 mL
Lipids						
References	Rawmaterial	PEF treatment	Extraction method	Extraction conditions	Solvent	Extractedlipid
[109]	fresh microalgae <i>Auxenochlorella protothecoides</i>	Flow rate: 0.1 mL/s Pulse duration 1 μ s EFS: 4 MV/m Specific energy: 150 kJ/L	NR	Time: overnight incubation conditions: Temperature: 25 °C Time: 20 h	Hexane–ethanol blend	97%
Pectin						

References	Raw material	extraction method	Extraction device	Extraction conditions	Powder–solvent ratio	Yield
[110]	Raw jackfruit (<i>Artocarpus heterophyllus</i>)	Combination of PEF- and microwave-assisted extraction	PEF generator microwave reactor	PEF conditions: Time: 5 min, EFS: 10 kV/cm. Microwave conditions: Time: 10 min, Power level of 650 W/g	1:4	18.3%
Cellulose						
References	Raw material	PEF treatment	Extraction method	Solvent		Yield
[39]	Mendong Fiber (<i>Fimbristylis globulosa</i>)	EFS: 1.3 kV/cm Frequency: 20 kHz Time 30 s	PEF-assisted alkali extraction	NaOH sol. (300 mL) 60% conc.		97.8%
Pigment						
References	Raw material	PEF treatment	Extraction method	Extraction conditions		Yield
[111]	Algae paste of <i>Nannochloropsis oceanica</i>	Constant flow rate: 20 mL/min Temperature: 25 °C Square wave: 5 µs EFS: 10 kV/cm Total specific energy inputs: 100 kJ/kg	PEF-assisted supercritical CO ₂ extraction	Pressure: 8, 14, and 20 MPa Fixed temperature: 35 °C CO ₂ /biomass ratio: 53.3 kgCO ₂ /kg DW Holding time: 7 min		Total carotenes: 36% Total chlorophyll: 52%

References

1. Yammine, S.; Brianceau, S.; Manteau, S.; Turk, M.; Ghidossi, R.; Vorobiev, E.; Mietton-Peuchot, M. Extraction and purification of high added value compounds from by-products of the winemaking chain using alternative/nonconventional processes/technologies. Crit. Rev. Food Sci. Nutr. 2018, 58, 1375–1390.

2. Ranjha, M.M.A.N.; Irfan, S.; Nadeem, M.; Mahmood, S. A Comprehensive Review on Nutritional Value, Medicinal Uses, and Processing of Banana. *Food Rev. Int.* 2020.
3. Ranjha, M.M.A.N.; Shafique, B.; Wang, L.; Irfan, S.; Safdar, M.N.; Murtaza, M.A.; Nadeem, M.; Mahmood, S.; Mueen-ud-Din, G.; Nadeem, H.R. A comprehensive review on phytochemistry, bioactivity and medicinal value of bioactive compounds of pomegranate (*Punica granatum*). *Adv. Tradit. Med.* 2021.
4. Nadeem, H.R.; Akhtar, S.; Ismail, T.; Sestili, P.; Lorenzo, J.M.; Ranjha, M.M.A.N.; Jooste, L.; Hano, C.; Aadil, R.M. Heterocyclic Aromatic Amines in Meat: Formation, Isolation, Risk Assessment, and Inhibitory Effect of Plant Extracts. *Foods* 2021, 10, 1466.
5. Ali Nawaz Ranjha, M.M. A Critical Review on Presence of Polyphenols in Commercial Varieties of Apple Peel, their Extraction and Health Benefits. *Open Access J. Biog. Sci. Res.* 2020, 6.
6. Mahmood, S. A Narrative Review on the Phytochemistry, Nutritional Profile and Properties of Prickly Pear Fruit. *Open Access J. Biog. Sci. Res.* 2021, 7.
7. Chemat, F.; Abert Vian, M.; Fabiano-Tixier, A.S.; Nutrizio, M.; Režek Jambrak, A.; Munekata, P.E.S.; Lorenzo, J.M.; Barba, F.J.; Binello, A.; Cravotto, G. A review of sustainable and intensified techniques for extraction of food and natural products. *Green Chem.* 2020, 22, 2325–2353.
8. Yan, L.G.; He, L.; Xi, J. High intensity pulsed electric field as an innovative technique for extraction of bioactive compounds—A review. *Crit. Rev. Food Sci. Nutr.* 2017, 57, 2877–2888.
9. Ranjha, M.M.A.N.; Amjad, S.; Ashraf, S.; Khawar, L.; Safdar, M.N.; Jabbar, S.; Nadeem, M.; Mahmood, S.; Murtaza, M.A. Extraction of Polyphenols from Apple and Pomegranate Peels Employing Different Extraction Techniques for the Development of Functional Date Bars. *Int. J. Fruit Sci.* 2020, 20, S1201–S1221.
10. Bansal, V.; Sharma, A.; Ghanshyam, C.; Singla, M.L.; Kim, K.H. Influence of pulsed electric field and heat treatment on *Emblica officinalis* juice inoculated with *Zygosaccharomyces bailii*. *Food Bioprod. Process.* 2015, 95, 146–154.
11. Gabrić, D.; Barba, F.; Roohinejad, S.; Gharibzahedi, S.M.T.; Radojčin, M.; Putnik, P.; Bursać Kovačević, D. Pulsed electric fields as an alternative to thermal processing for preservation of nutritive and physicochemical properties of beverages: A review. *J. Food Process. Eng.* 2018, 41, 12638.
12. Mohamad, A.; Shah, N.N.A.K.; Sulaiman, A.; Mohd Adzahan, N.; Aadil, R.M. Impact of the pulsed electric field on physicochemical properties, fatty acid profiling, and metal migration of goat milk. *J. Food Process. Preserv.* 2020, 44, 14940.
13. Aadil, R.M.; Zeng, X.A.; Sun, D.W.; Wang, M.S.; Liu, Z.W.; Zhang, Z.H. Combined effects of sonication and pulsed electric field on selected quality parameters of grapefruit juice. *LWT Food Sci. Technol.* 2015, 62, 890–893.

14. Sánchez-Vega, R.; Elez-Martínez, P.; Martín-Belloso, O. Influence of high-intensity pulsed electric field processing parameters on antioxidant compounds of broccoli juice. *Innov. Food Sci. Emerg. Technol.* 2015, 29, 70–77.
15. Bozinou, E.; Karageorgou, I.; Batra, G.; GDourtoglou, V.; ILalas, S. Pulsed Electric Field Extraction and Antioxidant Activity Determination of *Moringa oleifera* Dry Leaves: A Comparative Study with Other Extraction Techniques. *Beverages* 2019, 5, 8.
16. Evrendilek, G.A. Change regime of aroma active compounds in response to pulsed electric field treatment time, sour cherry juice apricot and peach nectars, and physical and sensory properties. *Innov. Food Sci. Emerg. Technol.* 2016, 33, 195–205.
17. Arshad, R.N.; Abdul-Malek, Z.; Munir, A.; Buntat, Z.; Ahmad, M.H.; Jusoh, Y.M.M.; Bekhit, A.E.D.; Roobab, U.; Manzoor, M.F.; Aadil, R.M. Electrical systems for pulsed electric field applications in the food industry: An engineering perspective. *Trends Food Sci. Technol.* 2020, 104, 1–13.
18. López-Gámez, G.; Elez-Martínez, P.; Martín-Belloso, O.; Soliva-Fortuny, R. Changes of carotenoid content in carrots after application of pulsed electric field treatments. *Lwt* 2021, 147, 111408.
19. Tzima, K.; Brunton, N.P.; Lyng, J.G.; Frontuto, D.; Rai, D.K. The effect of Pulsed Electric Field as a pre-treatment step in Ultrasound Assisted Extraction of phenolic compounds from fresh rosemary and thyme by-products. *Innov. Food Sci. Emerg. Technol.* 2021, 69, 102644.
20. Soquetta, M.B.; Terra, L.D.M.; Bastos, C.P. Green technologies for the extraction of bioactive compounds in fruits and vegetables. *CYTA J. Food* 2018, 16, 400–412.
21. Oroian, M.; Escriche, I. Antioxidants: Characterization, natural sources, extraction and analysis. *Food Res. Int.* 2015, 74, 10–36.
22. Xi, J.; Li, Z.; Fan, Y. Recent advances in continuous extraction of bioactive ingredients from food-processing wastes by pulsed electric fields. *Crit. Rev. Food Sci. Nutr.* 2021, 61, 1738–1750.
23. Panja, P. Green extraction methods of food polyphenols from vegetable materials. *Curr. Opin. Food Sci.* 2018, 23, 173–182.
24. Redondo, D.; Venturini, M.E.; Luengo, E.; Raso, J.; Arias, E. Pulsed electric fields as a green technology for the extraction of bioactive compounds from thinned peach by-products. *Innov. Food Sci. Emerg. Technol.* 2018, 45, 335–343.
25. Thulasidas, J.S.; Varadarajan, G.S.; Sundararajan, R. Pulsed Electric Field for Enhanced Extraction of Intracellular Bioactive Compounds from Plant Products: An Overview. *Nov. Approaches Drug Des. Dev.* 2019, 5.
26. Koubaa, M.; Roselló-Soto, E.; Šic Žlabur, J.; Režek Jambrak, A.; Brnčić, M.; Grimi, N.; Boussetta, N.; Barba, F.J. Current and New Insights in the Sustainable and Green Recovery of Nutritionally

- Valuable Compounds from *Stevia rebaudiana* Bertoni. *J. Agric. Food Chem.* 2015, 63, 6835–6846.
27. Barba, F.J.; Boussetta, N.; Vorobiev, E. Emerging technologies for the recovery of isothiocyanates, protein and phenolic compounds from rapeseed and rapeseed press-cake: Effect of high voltage electrical discharges. *Innov. Food Sci. Emerg. Technol.* 2015, 31, 67–72.
 28. Puértolas, E.; Koubaa, M.; Barba, F.J. An overview of the impact of electrotechnologies for the recovery of oil and high-value compounds from vegetable oil industry: Energy and economic cost implications. *Food Res. Int.* 2016, 80, 19–26.
 29. Sarkis, J.R.; Boussetta, N.; Tessaro, I.C.; Marczak, L.D.F.; Vorobiev, E. Application of pulsed electric fields and high voltage electrical discharges for oil extraction from sesame seeds. *J. Food Eng.* 2015, 153, 20–27.
 30. Barba, F.J.; Parniakov, O.; Koubaa, M.; Lebovka, N. Pulsed Electric Fields Assisted Extraction from Exotic Fruit Residues. In *Handbook of Electroporation*; Miklavcic, D., Ed.; Springer: Cham, Switzerland, 2016; pp. 1–18. ISBN 978-3-319-26779-1.
 31. Teh, S.S.; Niven, B.E.; Bekhit, A.E.D.A.; Carne, A.; Birch, E.J. Microwave and pulsed electric field assisted extractions of polyphenols from defatted canola seed cake. *Int. J. Food Sci. Technol.* 2015, 50, 1109–1115.
 32. Barba, F.J.; Parniakov, O.; Pereira, S.A.; Wiktor, A.; Grimi, N.; Boussetta, N.; Saraiva, J.A.; Raso, J.; Martin-Belloso, O.; Witrowa-Rajchert, D.; et al. Current applications and new opportunities for the use of pulsed electric fields in food science and industry. *Food Res. Int.* 2015, 77, 773–798.
 33. Moreira, S.A.; Alexandre, E.M.C.; Pintado, M.; Saraiva, J.A. Effect of emergent non-thermal extraction technologies on bioactive individual compounds profile from different plant materials. *Food Res. Int.* 2019, 115, 177–190.
 34. Alexandre, E.M.C.; Moreira, S.A.; Pintado, M.; Saraiva, J.A. Emergent extraction technologies to valorize fruit and vegetable residues. In *Agricultural Research Updates*; Gorawala, P., Mandhatri, S., Eds.; Nova Science: New York, NY, USA, 2017; Volume 17, pp. 37–79. ISBN 9781536109078.
 35. Ricci, A.; Parpinello, G.P.; Versari, A. Recent Advances and Applications of Pulsed Electric Fields (PEF) to Improve Polyphenol Extraction and Color Release during Red Winemaking. *Beverages* 2018, 4, 18.
 36. Papachristou, I.; Akaberi, S.; Silve, A.; Navarro-López, E.; Wüstner, R.; Leber, K.; Nazarova, N.; Müller, G.; Frey, W. Analysis of the lipid extraction performance in a cascade process for *Scenedesmus almeriensis* biorefinery. *Biotechnol. Biofuels* 2021, 14, 1–14.
 37. Nowacka, M.; Tappi, S.; Wiktor, A.; Rybak, K.; Miszczykowska, A.; Czyzewski, J.; Drozdal, K.; Witrowa-Rajchert, D.; Tylewicz, U. The impact of pulsed electric field on the extraction of bioactive compounds from beetroot. *Foods* 2019, 8, 244.

38. Moradi, N.; Rahimi, M. Effect of simultaneous ultrasound/pulsed electric field pretreatments on the oil extraction from sunflower seeds. *Sep. Sci. Technol.* 2018, 53, 2088–2099.
39. Suryanto, H.; Fikri, A.A.; Permanasari, A.A.; Yanuhar, U.; Sukardi, S. Pulsed Electric Field Assisted Extraction of Cellulose From Mendong Fiber (*Fimbristylis globulosa*) and its Characterization. *J. Nat. Fibers* 2018, 15, 406–415.
40. Yongguang, Y.; Yuzhu, H.; Yong, H. Pulsed electric field extraction of polysaccharide from *Rana temporaria chensinensis* David. *Int. J. Pharm.* 2006, 312, 33–36.
41. Arshad, R.N.; Abdul-Malek, Z.; Roobab, U.; Qureshi, M.I.; Khan, N.; Ahmad, M.H.; Liu, Z.W.; Aadil, R.M. Effective valorization of food wastes and by-products through pulsed electric field: A systematic review. *J. Food Process. Eng.* 2021, 44.
42. Zia, S.; Khan, M.R.; Shabbir, M.A.; Aslam Maan, A.; Khan, M.K.I.; Nadeem, M.; Khalil, A.A.; Din, A.; Aadil, R.M. An Inclusive Overview of Advanced Thermal and Nonthermal Extraction Techniques for Bioactive Compounds in Food and Food-related Matrices. *Food Rev. Int.* 2020.
43. Li, M.; Lin, J.; Chen, J.; Fang, T. Pulsed Electric Field-Assisted Enzymatic Extraction of Protein from Abalone (*Haliotis Discus Hannai* Ino) Viscera. *J. Food Process. Eng.* 2016, 39, 702–710.
44. Lin, S.; Wang, L.; Jones, G.; Trang, H.; Yin, Y.; Liu, J. Optimized extraction of calcium malate from eggshell treated by PEF and an absorption assessment in vitro. *Int. J. Biol. Macromol.* 2012, 50, 1327–1333.
45. Zhao, Y.; Zheng, Y.; He, H.; Sun, Z.; Li, A. Effective aluminum extraction using pressure leaching of bauxite reaction residue from coagulant industry and leaching kinetics study. *J. Environ. Chem. Eng.* 2021, 9, 104770.
46. Zderic, A.; Zondervan, E. Polyphenol extraction from fresh tea leaves by pulsed electric field: A study of mechanisms. *Chem. Eng. Res. Des.* 2016, 109, 586–592.
47. Kokosa, J.M. Selecting an extraction solvent for a greener liquid phase microextraction (LPME) mode-based analytical method. *TrAC Trends Anal. Chem.* 2019, 118, 238–247.
48. Efthymiopoulos, I.; Hellier, P.; Ladommatos, N.; Russo-Profil, A.; Eveleigh, A.; Aliev, A.; Kay, A.; Mills-Lamprey, B. Influence of solvent selection and extraction temperature on yield and composition of lipids extracted from spent coffee grounds. *Ind. Crops Prod.* 2018, 119, 49–56.
49. Pintać, D.; Majkić, T.; Torović, L.; Orčić, D.; Beara, I.; Simin, N.; Mimica-Dukić, N.; Lesjak, M. Solvent selection for efficient extraction of bioactive compounds from grape pomace. *Ind. Crop. Prod.* 2018, 111, 379–390.
50. Pintać, D.; Majkić, T.; Torović, L.; Orčić, D.; Beara, I.; Simin, N.; Mimica-Dukić, N.; Lesjak, M. Extraction of Bioactive Compound from Some Fruits and Vegetables (Pomegranate Peel, Carrot and Tomato). *Am. J. Food Nutr.* 2016, 4, 8–19.

51. Altemimi, A.; Lakhssassi, N.; Baharlouei, A.; Watson, D.G.; Lightfoot, D.A. Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. *Plants* 2017, 6, 42.
52. Tiwari, B.K. Ultrasound: A clean, green extraction technology. *TrAC-Trends Anal. Chem.* 2015, 71, 100–109.
53. Rodríguez-Pérez, C.; Quirantes-Piné, R.; Fernández-Gutiérrez, A.; Segura-Carretero, A. Optimization of extraction method to obtain a phenolic compounds-rich extract from *Moringa oleifera* Lam leaves. *Ind. Crops Prod.* 2015, 66, 246–254.
54. Heleno, S.A.; Diz, P.; Prieto, M.A.; Barros, L.; Rodrigues, A.; Barreiro, M.F.; Ferreira, I.C.F.R. Optimization of ultrasound-assisted extraction to obtain mycosterols from *Agaricus bisporus* L. by response surface methodology and comparison with conventional Soxhlet extraction. *Food Chem.* 2016, 197, 1054–1063.
55. Leong, S.Y.; Burritt, D.J.; Oey, I. Evaluation of the anthocyanin release and health-promoting properties of Pinot Noir grape juices after pulsed electric fields. *Food Chem.* 2016, 196, 833–841.
56. Shree, T.J.; Sree, V.G.; Sundararajan, R. Enhancement of Bioactive Compounds from Green Grapes Extract using Pulsed Electric Field treatment. *J. Cancer Prev. Curr. Res.* 2018, 9.
57. Lamanauskas, N.; Pataro, G.; Bobinas, Č.; Šatkauskas, S.; Viškelis, P.; Bobinaitė, R.; Ferrari, G. Impulsinio elektrinio lauko įtaka sulčių ir bioaktyvių medžiagų išgavimui iš aviečių ir jų produktų. *Zemdirbyste* 2016, 103, 83–90.
58. Barba, F.J.; Brianceau, S.; Turk, M.; Boussetta, N.; Vorobiev, E. Effect of Alternative Physical Treatments (Ultrasounds, Pulsed Electric Fields, and High-Voltage Electrical Discharges) on Selective Recovery of Bio-compounds from Fermented Grape Pomace. *Food Bioprocess. Technol.* 2015, 8, 1139–1148.
59. Liu, Z.W.; Zeng, X.A.; Ngadi, M. Enhanced extraction of phenolic compounds from onion by pulsed electric field (PEF). *J. Food Process. Preserv.* 2018, 42.
60. Fincan, M. Extractability of phenolics from spearmint treated with pulsed electric field. *J. Food Eng.* 2015, 162, 31–37.
61. Sarkis, J.R.; Boussetta, N.; Blouet, C.; Tessaro, I.C.; Marczak, L.D.F.; Vorobiev, E. Effect of pulsed electric fields and high voltage electrical discharges on polyphenol and protein extraction from sesame cake. *Innov. Food Sci. Emerg. Technol.* 2015, 29, 170–177.
62. Xue, D.; Farid, M.M. Pulsed electric field extraction of valuable compounds from white button mushroom (*Agaricus bisporus*). *Innov. Food Sci. Emerg. Technol.* 2015, 29, 178–186.
63. Frontuto, D.; Carullo, D.; Harrison, S.M.; Brunton, N.P.; Ferrari, G.; Lyng, J.G.; Pataro, G. Optimization of Pulsed Electric Fields-Assisted Extraction of Polyphenols from Potato Peels Using

- Response Surface Methodology. *Food Bioprocess. Technol.* 2019, 12, 1708–1720.
64. Puértolas, E.; Martínez De Marañón, I. Olive oil pilot-production assisted by pulsed electric field: Impact on extraction yield, chemical parameters and sensory properties. *Food Chem.* 2015, 167, 497–502.
 65. Pashazadeh, B.; Elhamirad, A.H.; Hajnajari, H.; Sharayei, P.; Armin, M. Optimization of the pulsed electric field -assisted extraction of functional compounds from cinnamon. *Biocatal. Agric. Biotechnol.* 2020, 23, 101461.
 66. Mahalleh, A.A.; Sharayei, P.; Mortazavi, S.A.; Azarpazhooh, E.; Niazmand, R. Optimization of the pulsed electric field-assisted extraction of functional compounds from nepeta binaludensis. *Agric. Eng. Int. CIGR J.* 2019, 21, 184–194.
 67. Pataro, G.; Carullo, D.; Ferrari, G. Effect of PEF pre-treatment and extraction temperature on the recovery of carotenoids from tomato wastes. *Chem. Eng. Trans.* 2019, 75, 139–144.
 68. Pataro, G.; Carullo, D.; Bobinaite, R.; Donsì, G.; Ferrari, G. Improving the extraction yield of juice and bioactive compounds from sweet cherries and their by-products by pulsed electric fields. *Chem. Eng. Trans.* 2017, 57, 1717–1722.
 69. Siddeeg, A.; Manzoor, M.F.; Ahmad, M.H.; Ahmad, N.; Ahmed, Z.; Khan, M.K.I.; Maan, A.A.; Mahr-Un-Nisa; Zeng, X.A.; Ammar, A.F. Pulsed electric field-assisted ethanolic extraction of date palm fruits: Bioactive compounds, antioxidant activity and physicochemical properties. *Processes* 2019, 7, 585.
 70. Moghaddam, T.N.; Elhamirad, A.H.; Saeidi Asl, M.R.; Shahidi Noghabi, M. Pulsed electric field-assisted extraction of phenolic antioxidants from tropical almond red leaves. *Chem. Pap.* 2020, 74, 3957–3961.
 71. Bobinaite, R.; Pataro, G.; Lamanaukas, N.; Šatkauskas, S.; Viškelis, P.; Ferrari, G. Application of pulsed electric field in the production of juice and extraction of bioactive compounds from blueberry fruits and their by-products. *J. Food Sci. Technol.* 2015, 52, 5898–5905.
 72. Bobinaite, R.; Pataro, G.; Visockis, M.; Bobinas, Č.; Ferrari, G.; Viškelis, P. Potential application of pulsed electric fields to improve the recovery of bioactive compounds from sour cherries and their by-products. In *Proceedings of the 11th Baltic Conference on Food Science and Technology*, Jelgava, Latvia, 27–28 April 2017; pp. 70–74.
 73. Niu, D.; Ren, E.F.; Li, J.; Zeng, X.A.; Li, S.L. Effects of pulsed electric field-assisted treatment on the extraction, antioxidant activity and structure of naringin. *Sep. Purif. Technol.* 2021, 265, 118480.
 74. Lamanaukas, N.; Bobinaite, R.; Šatkauskas, S.; Viškelis, P.; Pataro, G.; Ferrari, G. Sulčių spaudimas iš sušaldytų/atšildytų mėlynių taikant impulsinio elektrinio lauko metodą. *Zemdirbyste* 2015, 102, 59–66.

75. Saini, A.; Panesar, P.S.; Bera, M.B. Valorization of fruits and vegetables waste through green extraction of bioactive compounds and their nanoemulsions-based delivery system. *Bioresour. Bioprocess.* 2019, 6.
76. Rudra, S.G.; Nishad, J.; Jakhar, N.; Kaur, C. Food Industry Waste: Mine of Nutraceuticals. *Int. J. Sci. Enviro. Technol.* 2015, 4, 205–229.
77. Sagar, N.A.; Pareek, S.; Sharma, S.; Yahia, E.M.; Lobo, M.G. Fruit and Vegetable Waste: Bioactive Compounds, Their Extraction, and Possible Utilization. *Compr. Rev. Food Sci. Food Saf.* 2018, 17, 512–531.
78. Medina-Meza, I.G.; Barbosa-Cánovas, G.V. Assisted extraction of bioactive compounds from plum and grape peels by ultrasonics and pulsed electric fields. *J. Food Eng.* 2015, 166, 268–275.
79. El Kantar, S.; Boussetta, N.; Lebovka, N.; Foucart, F.; Rajha, H.N.; Maroun, R.G.; Louka, N.; Vorobiev, E. Pulsed electric field treatment of citrus fruits: Improvement of juice and polyphenols extraction. *Innov. Food Sci. Emerg. Technol.* 2018, 46, 153–161.
80. Irfan, S.; Ranjha, M.M.A.N.; Mahmood, S.; Saeed, W.; Alam, M.Q. Lemon Peel: A Natural Medicine. *Int. J. Biotechnol. Allied Fields* 2018, 7, 185–194.
81. Irfan, S.; Ranjha, M.M.A.N.; Mahmood, S.; Mueen-ud-Din, G.; Rehman, S.; Saeed, W.; Qamrosh Alam, M.; Mahvish Zahra, S.; Yousaf Quddoos, M.; Ramzan, I.; et al. A Critical Review on Pharmaceutical and Medicinal Importance of Ginger. *Acta Sci. Nutr. Health* 2019, 3, 78–82.
82. Faniyi, T.O.; Adewumi, M.K.; Jack, A.A.; Adegbeye, M.J.; Elghandour, M.M.M.Y.; Barbabosa-Pliego, A.; Salem, A.Z.M. Extracts of herbs and spices as feed additives mitigate ruminal methane production and improve fermentation characteristics in West African Dwarf sheep. *Trop. Anim. Health Prod.* 2021, 53, 312.
83. Valdés, A.; Mellinas, A.C.; Ramos, M.; Burgos, N.; Jiménez, A.; Garrigós, M.C. Use of herbs, spices and their bioactive compounds in active food packaging. *RSC Adv.* 2015, 5, 40324–40335.
84. Roselló-Soto, E.; Parniakov, O.; Deng, Q.; Patras, A.; Koubaa, M.; Grimi, N.; Boussetta, N.; Tiwari, B.K.; Vorobiev, E.; Lebovka, N.; et al. Application of Non-conventional Extraction Methods: Toward a Sustainable and Green Production of Valuable Compounds from Mushrooms. *Food Eng. Rev.* 2016, 8, 214–234.
85. Giacometti, J.; Bursać Kovačević, D.; Putnik, P.; Gabrić, D.; Bilušić, T.; Krešić, G.; Stulić, V.; Barba, F.J.; Chemat, F.; Barbosa-Cánovas, G.; et al. Extraction of bioactive compounds and essential oils from mediterranean herbs by conventional and green innovative techniques: A review. *Food Res. Int.* 2018, 113, 245–262.
86. Zhang, Q.W.; Lin, L.G.; Ye, W.C. Techniques for extraction and isolation of natural products: A comprehensive review. *Chin. Med.* 2018, 13.

87. Segovia, F.J.; Luengo, E.; Corral-Pérez, J.J.; Raso, J.; Almajano, M.P. Improvements in the aqueous extraction of polyphenols from borage (*Borago officinalis* L.) leaves by pulsed electric fields: Pulsed electric fields (PEF) applications. *Ind. Crop. Prod.* 2015, 65, 390–396.
88. Barba, F.J.; Grimi, N.; Vorobiev, E. Evaluating the potential of cell disruption technologies for green selective extraction of antioxidant compounds from *Stevia rebaudiana* Bertoni leaves. *J. Food Eng.* 2015, 149, 222–228.
89. Gagneten, M.; Leiva, G.; Salvatori, D.; Schebor, C.; Olaiz, N. Optimization of Pulsed Electric Field Treatment for the Extraction of Bioactive Compounds from Blackcurrant. *Food Bioprocess. Technol.* 2019, 12, 1102–1109.
90. Liu, Z.; Esveld, E.; Vincken, J.P.; Bruins, M.E. Pulsed Electric Field as an Alternative Pre-treatment for Drying to Enhance Polyphenol Extraction from Fresh Tea Leaves. *Food Bioprocess. Technol.* 2019, 12, 183–192.
91. Hendrawan, Y.; Sabrinauly, S.; Hawa, L.C.; Rachmawati, M.; Argo, B.D. Analysis of the phenol and flavonoid content from basil leaves (*Ocimum americanum* L) extract using pulsed electric field (PEF) pre-treatment. *Agric. Eng. Int. CIGR J.* 2019, 21, 149–158.
92. Mushtaq, A.; Roobab, U.; Denoya, G.I.; Inam-Ur-Raheem, M.; Gullón, B.; Lorenzo, J.M.; Barba, F.J.; Zeng, X.A.; Wali, A.; Aadil, R.M. Advances in green processing of seed oils using ultrasound-assisted extraction: A review. *J. Food Process. Preserv.* 2020, 44, e14740.
93. Yusuf, A.K. A Review of Methods Used for Seed Oil Extraction. *Int. J. Sci. Res.* 2017, 8, 1854–1861.
94. Haji-Moradkhani, A.; Rezaei, R.; Moghimi, M. Optimization of pulsed electric field-assisted oil extraction from cannabis seeds. *J. Food Process. Eng.* 2019, 42, e13028.
95. Batool, M.; Kauser, S.; Nadeem, H.R.; Perveen, R.; Irfan, S.; Siddiq, A.; Shafique, B.; Zahra, S.M.; Waseem, M.; Khalid, W.; et al. A Critical Review on Alpha Tocopherol: Sources, RDA and Health Benefits. *J. Appl. Pharm.* 2020, 12, 19–39.
96. Chemat, F.; Rombaut, N.; Sicaire, A.G.; Meullemiestre, A.; Fabiano-Tixier, A.S.; Abert-Vian, M. Ultrasound assisted extraction of food and natural products. Mechanisms, techniques, combinations, protocols and applications. A review. *Ultrason. Sonochem.* 2017, 34, 540–560.
97. Vorobiev, E.; Lebovka, N. Pulsed electric energy-assisted biorefinery of oil crops and residues. In *Handbook of Electroporation*; Miklavčič, D., Ed.; Springer: Cham, Switzerland, 2017; Volume 4, pp. 2863–2881. ISBN 9783319328867.
98. Sitzmann, W.; Vorobiev, E.; Lebovka, N. Applications of electricity and specifically pulsed electric fields in food processing: Historical backgrounds. *Innov. Food Sci. Emerg. Technol.* 2016, 37, 302–311.

99. Shorstkii, I.; Koshevoi, E. Extraction Kinetic of Sunflower Seeds Assisted by Pulsed Electric Fields. *Iran. J. Sci. Technol. Trans. A Sci.* 2019, 43, 813–817.
100. Zhou, Y.J.; Xue, C.M.; Zhang, S.S.; Yao, G.M.; Zhang, L.; Wang, S.J. Effects of high intensity pulsed electric fields on yield and chemical composition of rose essential oil. *Int. J. Agric. Biol. Eng.* 2017, 10, 295–301.
101. Shorstkii, I.; Mirshekarloo, M.S.; Koshevoi, E. Application of Pulsed Electric Field for Oil Extraction from Sunflower Seeds: Electrical Parameter Effects on Oil Yield. *J. Food Process. Eng.* 2017, 40.
102. Tamborrino, A.; Urbani, S.; Servili, M.; Romaniello, R.; Perone, C.; Leone, A. Pulsed electric fields for the treatment of olive pastes in the oil extraction process. *Appl. Sci.* 2020, 10, 114.
103. Martínez, J.M.; Delso, C.; Álvarez, I.; Raso, J. Pulsed electric field-assisted extraction of valuable compounds from microorganisms. *Compr. Rev. Food Sci. Food Saf.* 2020, 19, 530–552.
104. Yellamanda, B.; Vijayalakshmi, M.; Kavitha, A.; Reddy, D.K.; Venkateswarlu, Y. Extraction and bioactive profile of the compounds produced by *Rhodococcus* sp. VLD-10. *3 Biotech.* 2016, 6, 261.
105. Martínez, J.M.; Luengo, E.; Saldaña, G.; Álvarez, I.; Raso, J. C-phycocyanin extraction assisted by pulsed electric field from *Arthrospira platensis*. *Food Res. Int.* 2017, 99, 1042–1047.
106. Jaeschke, D.P.; Mercali, G.D.; Marczak, L.D.F.; Müller, G.; Frey, W.; Gusbeth, C. Extraction of valuable compounds from *Arthrospira platensis* using pulsed electric field treatment. *Bioresour. Technol.* 2019, 283, 207–212.
107. Martínez, J.M.; Delso, C.; Álvarez, I.; Raso, J. Pulsed electric field permeabilization and extraction of phycoerythrin from *Porphyridium cruentum*. *Algal Res.* 2019, 37, 51–56.
108. Carullo, D.; Abera, B.D.; Casazza, A.A.; Donsì, F.; Perego, P.; Ferrari, G.; Pataro, G. Effect of pulsed electric fields and high pressure homogenization on the aqueous extraction of intracellular compounds from the microalgae *Chlorella vulgaris*. *Algal Res.* 2018, 31, 60–69.
109. Silve, A.; Kian, C.B.; Papachristou, I.; Kubisch, C.; Nazarova, N.; Wüstner, R.; Leber, K.; Strässner, R.; Frey, W. Incubation time after pulsed electric field treatment of microalgae enhances the efficiency of extraction processes and enables the reduction of specific treatment energy. *Bioresour. Technol.* 2018, 269, 179–187.
110. Nandhu Lal, A.M.; Prince, M.V.; Sreeja, R. Studies on Characterisation of Combined Pulsed Electric Field and Microwave Extracted Pectin from Jack Fruit Rind and Core. *Int. J. Curr. Microbiol. Appl. Sci.* 2020, 9, 2371–2380.
111. Pataro, G.; Carullo, D.; Ferrari, G. PEF-assisted supercritical CO₂ extraction of pigments from microalgae *nannochloropsis oceanica* in a continuous flow system. *Chem. Eng. Trans.* 2019, 74, 97–102.

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