

Recent Advances of Wearable Sweat-Sensing Devices

Subjects: [Engineering, Biomedical](#) | [Engineering, Electrical & Electronic](#)

Contributor: Nur Fatin Adini Ibrahim , Norhayati Sabani , Shazlina Johari , Asrulnizam Abd Manaf , Asnida Abdul Wahab , Zulkarnay Zakaria , Anas Mohd Noor

A sweat-sensing device requires a wearable device for the temporary attachment of its main components, including sensors, sweat collection devices, and electronic devices to the body's skin region.

sweat-sensing applications

wearable device

real-time measurement

1. Sweat-Sensing Device (SSD)

A sweat-sensing device requires a wearable device for the temporary attachment of its main components, including sensors, sweat collection devices, and electronic devices to the body's skin region. There are three primary types of wearable devices that are most commonly found in sweat applications: sweatbands [\[1\]](#), epidermal patches [\[2\]](#), and textiles [\[3\]](#). Several factors to be considered in selecting which types of wearable devices are best suited for use in SSDs include the skin surface on a body location, sample collection techniques, and environmental conditions, whether on dry land or in aquatic exercise. For example, a sweatband is appropriate for wearing on specific body parts such as the wrist [\[4\]](#), arm [\[5\]](#), back [\[6\]](#), or forehead [\[7\]](#), where the bands can be tightened, as shown in **Figure 1a**. In addition, most of the literature prefers wearable sweatbands on the arm for cystic fibrosis tests during the conducting of conventional pilocarpine iontophoresis and sweat collection samples [\[8\]](#)[\[9\]](#). The band can be worn repeatedly with reusable electrodes during pilocarpine iontophoresis to trigger perspiration at any time due to the ease of detachment and reattachment of the electrodes from the band.

An epidermal patch is characterized by a disposable style of adhesive skin tape. It is low-cost, making it a practical SSD in a disposable format [\[10\]](#). Wearable epidermal patches in SSDs typically come in various forms, such as skin patches [\[11\]](#), tattoos [\[12\]](#)[\[13\]](#), and bandages [\[14\]](#)[\[15\]](#), as shown in **Figure 1b–d**. The elements of substrate epidermal patches can promote strong skin adhesion, high mechanical strength, stretchability, and resilience in water conditions to manipulate physical skin performance. Moreover, an epidermal patch has a high flexibility for wearing on any part of the body's skin and a good adaptability in high-intensity exercise. Furthermore, they are versatile enough to be worn for use in water sports such as swimming [\[16\]](#). Reeder et al. proposed an epidermal patch for colorimetric sensing integrated with microfluidic and water-proof electronic systems that can perform real-time physiological measurements on swimmers and dryland athletes [\[17\]](#). In particular, most colorimetry sensing applies epidermal patches as wearable devices, as this device can be used for the single-shot measurement of sweat biomarkers once they change color, as reported by previous studies [\[18\]](#)[\[19\]](#).

A textile-based sensor has advantages over sweatbands and epidermal patches in terms of substrate washability when exposed to humidity and dirt. Moreover, the textile can serve either as a sweat collection device by absorption or as a wearable device. Recent textile-based sensing devices have a fitted sensor on regular cloth such as a t-shirt [\[20\]](#)[\[21\]](#), as shown in **Figure 1e**. Thus, they offer the advantages of being comfortable and allowing users to wear in any regular clothes. Recently, some SSDs have been woven with sensor and electronic components into highly stretchable fibers, making them suitable for applications requiring high-motion applications. The detection of motion and physiological signals by a fitted

sensor into a t-shirt also enable the easy examination of numerous physiological valuable data, including a person's movement for the detection of Parkinson's disease and stress levels [22][23][24]. Recently, several technologies and materials used in textile-based sensing devices have been improved to ensure they can operate under intense mechanical tension during regular activities and be reused without interfering with the analytical performance of a sensor after washing. Wicaksono et al. [20] and Martinez-Estrada et al. [25] developed a highly robust sensor-based textile that integrates with electronic component reusability after washing. A smart textile comprises water-resistant and detachable electronics for the convenience of washing, as well as a comfortable fabric sensor to prevent skin irritation for long-term monitoring [26].

An SSD can be composed of a single sensor or a combination of sensors with certain types of sweat collection devices to perform sweat analysis. A single sensor in an SSD is designed for direct skin contact to detect and measure sweat biomarkers such as metabolites that quickly degrade over time [27]. Thus, adopting a sweat collector, particularly a serpentine microfluidic device for analyzing proteins, is undesirable because sweat flow into a microchannel is time-consuming [28]. However, a sensor that has direct contact with the skin can irritate due to a rough sensor surface and contamination by perspiration from nearby areas. Combining a sensor with certain types of sweat collection devices can overcome these limitations. For example, utilizing paper-microfluidic integration with a sensor can prevent skin inflammation and contamination [29]. Moreover, the addition of paper to a microfluidic device is able to increase the flow rate of the transportation of sweat analytes into a sensor surface by absorption [30]. Therefore, sweat collection devices are essential components that can be added to an SSD. In addition, human sweat glands have small duct diameters, which are 5–40 μm for secretory coils and 10–20 μm for dermal ducts and upper coiled ducts [31], that limit the volume of sweat secretion. As a result, a sweat duct secretes a tiny portion of sweat from the bottom duct into the upper coiled duct region, with a microliter volume of total sweat being released at the skin's surface.

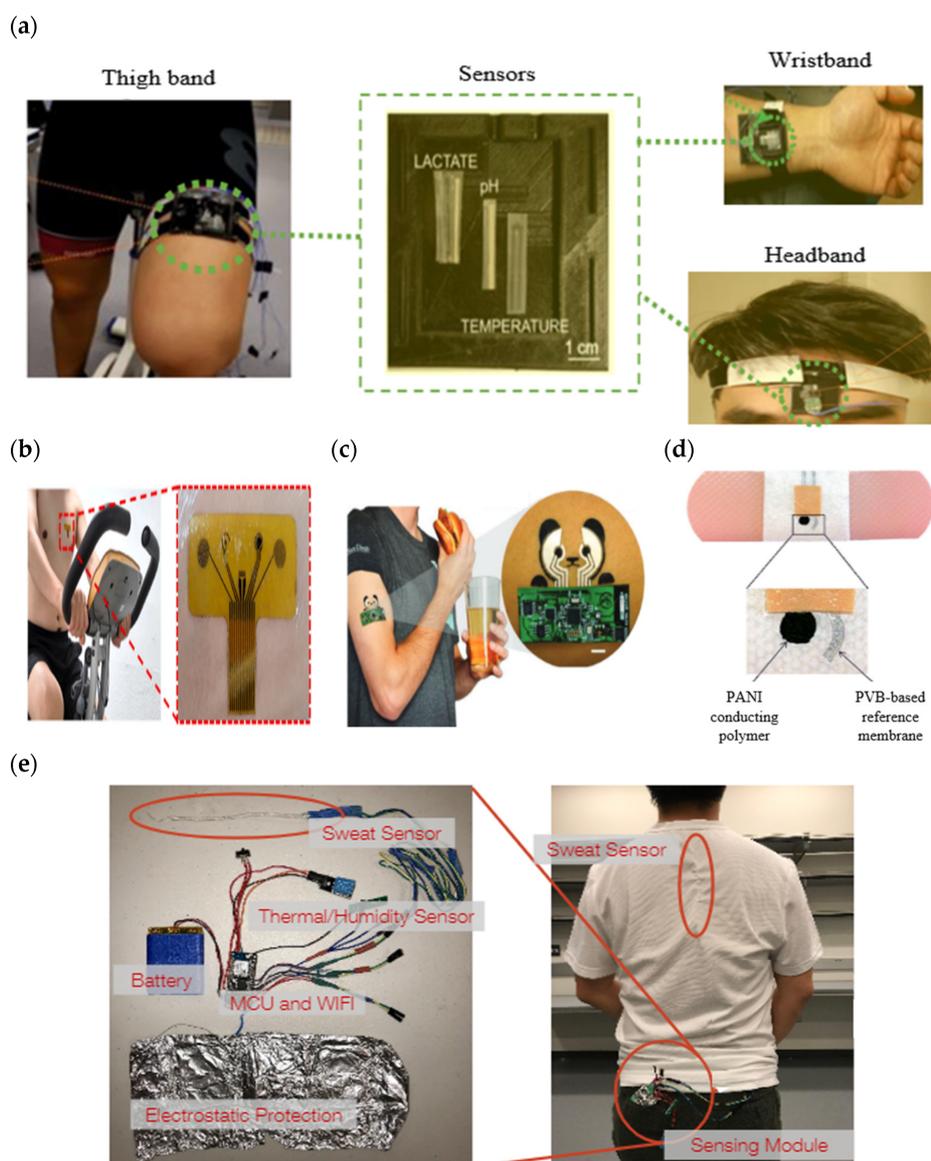


Figure 1. Types of wearable devices in sweat-sensing devices. (a) Sweatband [32]; (b) skin patch; reprinted from [11], copyright (2022), with permission from Elsevier. (c) Tattoo patch [13]; (d) bandage of epidermal patch [33] (Reused with the permission of copyright 2014, John Wiley, and Sons). (e) Textile [21].

A microfluidic device basically has a channel dimension of tens to hundreds of micrometers to reduce sample consumption and miniaturize microscale instruments for portability [34]. It largely uses body fluid samples such as sweat for the point-of-care diagnosis of diseases and certain laboratory tests. A microfluidic device can be sorted into the mechanism of active micropumps [35][36] and passive micropumps [37][38][39][40][41]. An active micropump requires an external power source to guide a continuous fluid flow with an adjustable flow rate into a microchannel. Typically, an electronic pump is used to deliver a solution efficiently with a setup of a steady flow rate at an inlet cavity that is similar to the natural average velocity [35], pressure, and mass flow rates of a biofluid. It is mainly used to enhance the smooth movement of liquid in a hydrophobic based material channel. Hydrophobic surface will increase resistance of capillary action to the fluid flow during passive flow. Its low surface energy makes it hard to wet on the wall surface, resulting in time-consuming sweat collection on a sensor surface [42].

The combination of a microfluidic device, absorbent-based materials, and microneedle injection demonstrates the best improvements in sweat collection devices, which has a high potential for efficiently transporting a solution in a short time while sharing its combined advantages and overcoming each other's limitations. **Figure 2** shows a summary of various SSD structures, including the types of wearable devices, categories of sweat collection devices, and sweat-sensing devices. There are two main groups for classifying sweat-sensing devices: continuous flow (CF) and non-continuous flow (NCF), based on the presence or absence of an integrated device outlet and real-time measurement. A microfluidic device consists of an outlet, allowing fresh sweat to continuously pass through a sensing area, providing the capability of performing continuous real-time analysis [43][44]. Real-time computation is essential for evaluating precise and valid current sweat analyte concentration withdrawal at a particular time, especially for sweat analysis over a longer period of time [16]. A sensor is commonly used to measure target sweat analytes in real time. CF and NCF SSDs can also be varied in terms of the types of wearable devices and sweat collection devices.

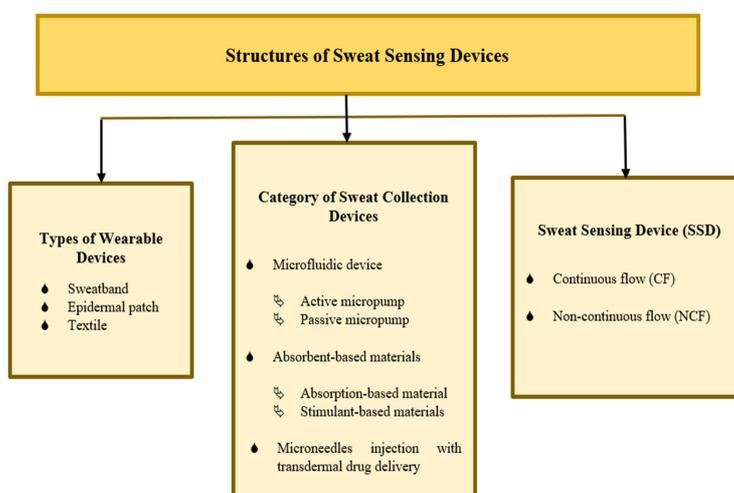


Figure 2. Summary of sweat-sensing device structures.

2. Recent Advances of SSDs: Optimal in Designs, Functionalities, and Performance

2.1. Sweat Collection Device

Adopting pharmacologic agents of sweat-stimulation such as hydrogel and pilocarpine can induce the skin to generate local sweat without requiring physical activity. The iontophoresis of pilocarpine can produce a sufficiently small amount of perspiration within the range of 15–100 μL [45]. Typically, the minimum volume capacity that a chemical sensor can react to and measure is at least 10 to 100 microliters [46]. In addition, this sweat stimulation can also be used to increase the biofluid for an individual that produces a low volume of sweat secretion during exercise. Moreover, the average sweat rate is approximately 1–15 $\mu\text{L min}^{-1}$ during active secretion [47][48], which slows fluid generation. Therefore, the use of pilocarpine/hydrogel is advocated for stimulating eccrine glands to elevate perspiration levels quickly for exercise and rest. However, most conventional pilocarpine iontophoresis methods are commonly utilized separate processes for collecting sweat samples that are inapplicable outside of the laboratory. Moreover, this process exposes electrical power that can cause skin burning, relying on expensive laboratory equipment and both bulky electrodes and benchtop analyzers [6][9]. Thus, adding a sensor while using this method can provide the dynamic real-time monitoring of target biomarkers on the spot. Kim et al.

developed an SSD that can simultaneously implement the iontophoresis of pilocarpine and the real-time measurement of the continuous monitoring of sweat glucose and alcohol [13].

Employing multiple sweat collection devices in a single SSD can efficiently accumulate a sufficient volume of sweat in a short duration and is able to provide fast hydration monitoring. Gunatilake et al. demonstrated a signal detection time readout of 4 min for lactate and 6 min for glucose using microfluidic paper-based analytical devices by incorporating a hydrogel of alginate-based materials and a colorimetric biosensor [49]. In addition, Alizadeh took 7–10 min for a hydration monitoring application using hydrogel that induced more sweat secretion while utilizing hydrophilic glass wicking in a microfluidic device [50]. In contrast, Nyein et al. only applied a single sweat collector in a microfluidic device which took 30–45 min [36]. However, the period for fitness performance analysis does not mostly depend on the number of sweat collection devices used because it can be flexible when continuously measuring at the user's preferred target time for estimating the loss of the electrolyte composition. The assembly of various sweat collection tools is primarily concentrated on supporting and maximizing the functions of each other. Incorporating a microfluidic device can hinder the problem of contamination, fast evaporation that reduces the sample collection, and an inevitable blend of renewal sweat concentration [51][52]. In addition, the introduction of a hydrophilic channel can promote capillary pressure, wall adhesion, and hydrostatic forces for the sweat flowing optimally in and out of the chamber. Ideally, a paper-based microfluidic patch could be a viable option to eliminate the direct contact with sensors that causes abrasion, avoid blocking the breathability of glands on the skin, and avoid backflow by increasing the pressure at the inlet during long-time monitoring [53]. Moreover, it facilitates continuous flow collection by transporting the sweat composition in liquid-filled paper channels, and it can also reduce the mixing of old and new sweat [54].

Microneedle transdermal injections can introduce new microbiomes from the surroundings into the skin's micropores. Although research has demonstrated that skin barrier function can recover the micropores within a few hours [55], utilizing wearable microneedles increases the risk of infection because bacteria can circulate through open micropores on the skin [56]. Moreover, hollow microneedles may fail and shatter due to additional compression, shear stress, excessive motion, the inherent discrepancy in complexion, or any other related pressures [57]. So, standard operating protocols for the use of microneedles are necessary for implementing proper clinical practice. Several mechanical and biological factors of evaluation approaches could be performed in vitro and in vivo to precisely assess possible risks and examine the safety of skin contact with innovative devices, especially when microneedles are used [58][59].

The repeatability of a microfluidic device is vital to ensure it can be frequently reusable, more practicable, prevent the waste of equipment, and avoid relying on disposable products even if they are the cheapest. Yang et al. introduced a reusable microfluidic device that integrated a high selectivity of concurrently detecting lactate and uric acid [60]. Their microfluidic device is portable, small in size, and has lower sample consumption, showing a good design for application prospects in clinical testing and personalized healthcare. Another study developed a microfluidic device that can be repeatedly reused by requiring its chips to be flushed with deionized water to remove the previous biofluid-measured ionic background [61]. Their microfluidic device collects the sweat volume with a small depth size to reduce time-consuming fluid flow. Their wearable microfluidic device is integrated with optimally modified sodium sensors that can measure several applications, such as fitness performance and diagnose cystic fibrosis.

Adding multiple inlet openings to a microfluidic device increases the exposed skin region that releases sweat into the channels while reducing time-consuming sweat secretion that depends on the accumulation in one hole. Xu et al. designed a microfluidic device consisting of eight inlets to reach a reservoir channel at the surface contact of a uric acid electrode sensing interface through the capillary effect [47]. During off-body test, they tested the performance of the microfluidic device by inserting a dye solution with a flow rate of 15 $\mu\text{L min}^{-1}$ at the inlet. The time required to fill the microfluidic reservoir

completely was 166 s. They switched the concentration of the solution between 0–80 μM with a flow rate of 15 $\mu\text{L min}^{-1}$, while the well-mixed renewal sweat could be used to determine the renewal time, which came out at 4.68 min. Meanwhile, adding multiple outlet openings in a microchannel could also improve passive pump performance through the spontaneous evaporation effect. Cheng et al. developed a wearable sweat sensor consisting of three micropores of the outlet with an inlet cavity [35]. The old sweat solution was quickly transported out by the fluid filling induced by capillary force and followed the spontaneous evaporation effect of micropores at three outlet cavities.

A simulation must be conducted for the fluid flow rate in a microfluidic device before developing its prototype. This is to determine the future channel size and what material properties of the wall are used to provide sufficient pressure without applying external energy. Moreover, a fluid simulation can also be used to mimic a real experiment for the validation of the channel performance of a microfluidic device by defining the average velocity of biological sweat secretion at the inlet. In the experiment, a flow rate sensor must be integrated with a microfluidic device to accurately measure the fluid flow rate. A high correspondence of a good flow rate and short-time results between the fluid simulation and experimental results exhibit the best verification of the effective working of microfluidic device channels. Nyein et al. [36] and the iGEM teams [44] realized both tests, varying the flow rates and solution concentrations. Hence, they had the advantage of exploring and gaining more information about the performance and capability of their device through a variety of changing channel sizes, shapes, and materials via theoretical simulation while saving time and money.

Table 1 shows other more recently developed SSDs with some features related to sweat collection device design. Most of them are CF SSDs that are integrated with sensors for real-time measurement and consist of an outlet in a microfluidic device. In addition, their developed microfluidic device promotes repeatability even though the wearable device is a single-use, disposable epidermal patch. This patch could be replaced with a new patch to allow the sweat collection device to be reused. Moreover, several of them incorporated microchannels with three, six, and eight inlets. Suction pumps and valves were also added to their microfluidic systems to control and increase fluid flow movement. Furthermore, the proposed microfluidic device's validation method was tested, including at least off-body test (calibration) and on-body tests to compare both evaluation performances. In addition, a few of them included a simulation to test the mechanical performance [62][63] as well as the performance in terms of fluid flow [64][65]. Mechanical testing is required to establish the level of flexibility and robustness of the microfluidic device in terms of bending, stretching, and twisting to identify which types of extreme activities they are appropriate for use in. For textile SSDs, it is essential to undergo a process of cleaning and drying for reusability. However, its design should be unaffected by the process, allowing it to retain its functionality.

Table 1. A summary of the latest SSDs' sweat collection device features advancements in 2022.

Categories of Sweat Collection Devices	SSD	Wearable Devices	Dimensions/Depth of Channel	Flow Rate and Time to Fill Channel	Reusable/Disposable	Additional Features	Validation Method	Mechanical Testing	References
Microfluidic device, portable iontophoresis of pilocarpine, adding hydrogel	CF	Epidermal patch	5 mm (Outer diameter), 1 mm (Inner diameter)	N/A 15 min (Total volume 32 μL)	Reusability	Multiple inlets (n = 3)	On-body test	Bending, stretching, twisting	[62]

Categories of Sweat Collection Devices	SSD	Wearable Devices	Dimensions/Depth of Channel	Flow Rate and Time to Fill Channel	Reusable/Disposable	Additional Features	Validation Method	Mechanical Testing	References
Modification of hydrophobic microfluidic device to a hydrophilic surface	CF	Epidermal patch	10 mm (diameter), 1 mm (thickness)	0.05–0.5 m/s (Total volume 200µL)	Reusability	Tesla valves	Simulation, off-body test, on-body test	N/A	[66]
Microfluidic device	CF	Epidermal patch	N/A	3–12 mm/s 13 min for indoor exercise, 20 min for outdoor exercise	Reusability	Multiple inlets (n = 3)	Off-body test, on-body test	Bending, stretching, twisting, tensile	[63]
Modification of hydrophobic microfluidic device to a hydrophilic surface	CF	Epidermal patch	1.5 mm (Inlet diameter), 4 mm (Reservoir diameter), 200 µm (thickness)	0.14 µL/min (each inlet), 0.84 µL/min (total) 14 min (Total volume 11.8 µL)	N/A	Multiple inlets (n = 6)	Simulation, off-body test, on-body test	Bending, pressing	[64]
Microfluidic device	CF	Epidermal patch	1 mm (diameter), 330 µm (thickness)	174.6 µL/min (Total volume 20µL)	N/A	Capillary bursting valves, colorimetric, multiple inlets (n = 8)	Simulation, off-body test, on-body test	Bending, stretching, twisting	[65]
Microfluidic device, absorptive pad	CF	Epidermal patch	N/A	5 µL/min (Total volume 10µL)	Reusability	Suction pump reset after filling the channel	Off-body test, on-body test	N/A	[67]
Textile	NCF	Stitched fabric with three button joints	N/A	N/A	Reusability	Washability	On-body test	Washing, drying (thermal)	[68]

able Platform for

Real-time monitoring of sodium in sweat. *Sensory Systems* 2019, 19, 1004–1009.

- He, W.; Wang, C.; Wang, H.; Jian, M.; Lu, W.; Liang, X.; Zhang, X.; Yang, F.; Zhang, Y. Integrated textile sensor patch for real-time and multiplex sweat analysis. *Sci. Adv.* 2019, 5, eaax0649.
- Mostafalu, P.; Akbari, M.; Alberti, K.A.; Xu, Q.; Khademhosseini, A.; Sonkusale, S.R. A toolkit of thread-based microfluidics, sensors, and electronics for 3D tissue embedding for medical diagnostics. *Microsyst. Nanoeng.* 2016, 2, 16039.
- Khemtonglang, K.; Chaiyaphet, N.; Kumsaen, T.; Chaiyachati, C.; Chuchuen, O. A Smart Wristband Integrated with an IoT-Based Alarming System for Real-Time Sweat Alcohol Monitoring. *Sensors* 2022, 22, 6435.
- Pirovano, P.; Dorrian, M.; Shinde, A.; Donohoe, A.; Brady, A.J.; Moyna, N.M.; Wallace, G.; Diamond, D.; McCaul, M. A wearable sensor for the detection of sodium and potassium in human sweat during exercise. *Talanta* 2020, 219, 121145.
- Schazmann, B.; Morris, D.; Slater, C.; Beirne, S.; Fay, C.; Reuveny, R.; Moyna, N.; Diamond, D. A wearable electrochemical sensor for the real-time measurement of sweat sodium concentration. *Anal. Methods* 2010,

2.2. Sensor

2. Wang, S.; Wu, Y.; Gu, Y.; Li, T.; Luo, H.; Li, L.H.; Bai, Y.; Li, L.; Liu, L.; Cao, Y.; et al. Wearable Sweatband Movement. Thus, a great resiliency and robustness of the sensor during extreme physical exercise are essential to enhance the reliability of sweat measurement with minimized dynamic motion artifacts [69]. These mechanical characteristics can maintain a good stability of the sensor's potential, current, or impedance of readable signals for prolonged monitoring even in fluctuating ion concentrations of analytes. The optimization of a surface electrode with polyurethane [70], carbon nanotubes [72], and poly(3,4-ethylenedioxythiophene) [71] of the solid contact coating has commonly maintained the contact of electrical conductivity and further high resistance to mechanical stress to ensure strong adherence to conventional substrates. Furthermore, these materials' properties generally include metallic conductivity, high tensile strength, high elasticity, thermal stability, high chemical inertness, and small sizes that are favorable for electrochemical sensor performance [78].
3. Choh, J.; Ghaffari, R.; Baker, L.B.; Rogers, J.A. Skin-interfaced systems for sweat collection and analytics. *Sci. Adv.* 2018, 4, eaar3921.
4. Gonzalez-Ruiz, J.; Mas, B.; de Haro, C.; Gabruja, E.; Camero, B.; Alonso-Lomillo, M.A.; Muñoz, F.J. Early determination of cystic fibrosis by electrochemical chloride quantification in sweat. *Biosens. Bioelectron.* 2009, 24, 1788–1791.
5. McLister, A.; McHugh, J.; Cundell, J.; Davis, J. New Developments in Smart Bandage Technologies for Wound Diagnostics. *Adv. Mater.* 2016, 28, 5732–5737.
6. Kim, J.; Yoon, S.; Yoon, H.; Zayed, M.A.; Park, C.; Kim, D.; Park, D. Multifunctional hydrogel skin patch for wearable smart healthcare applications. *Biosens. Bioelectron.* 2022, 196, 114665.
7. Bhandodkar, A.J.; Molinnus, D.; Mirza, O.; Guinovart, T.; Windmiller, J.R.; Valdés-Ramírez, G.; Andrade, F.J.; Schöning, M.J.; Wang, J. Epidermal tattoo potentiometric sodium sensors with wireless signal transduction for continuous non-invasive sweat monitoring. *Biosens. Bioelectron.* 2014, 54, 603–609.
8. Kim, C.; Sempier, S.; Molina, J.R.; Anania, S.; Harte, M.C.; Badiouin, A.; Tang, C.; Campbell, A.S.; McCreary, P.P. Wearable, self-powered, simultaneous monitoring of sweat and heart rate of the skin using a single wearable electrode. *Biosens. Bioelectron.* 2018, 5, 1800380.
9. Terse-Thakoor, T.; Puniya, M.; Matharu, Z.; Lyu, B.; Ahmad, M.; Giles, G.E.; Owyung, R.; Alaimo, F.; Shojaei Baghini, M.; Brunye, T.T.; et al. Thread-based multiplexed sensor patch for real-time sweat monitoring. *Npj Flex. Electron.* 2020, 4, 18.
10. Jiang, X.; Lillehoj, P.B. Microneedle-based skin patch for blood-free rapid diagnostic testing. *Microsyst. Nanosyst.* 2020, 6, 86.
11. Seshadri, D.R.; Li, R.T.; Voo, J.E.; Rowbottom, J.R.; Ailes, C.M.; Zorman, C.A.; Drummond, C.K. Wearable sensors for monitoring the physiological and biochemical profile of the athlete. *Npj Digit. Med.* 2019, 2, 72.
12. Reeder, J.T.; Choi, J.; Xue, Y.; Gutruf, P.; Hanson, J.; Liu, M.; Ray, T.; Bhandodkar, A.J.; Avila, R.; Xia, W.; et al. Waterproof electronics-enabled epidermal microfluidic devices for sweat collection, biomarker analysis, and thermography in aquatic settings. *Sci. Adv.* 2019, 5, eaau6356.
13. Wang, L.; Xu, T.; He, X.; Zhang, X. Flexible, self-healable, adhesive and wearable hydrogel patch for continuous sweat detection. *J. Mater. Chem. C* 2021, 9, 14938–14945.
14. Kim, J.; Wu, Y.; Luan, H.; Yang, D.S.; Cho, D.; Kwak, S.S.; Liu, S.; Ryu, H.; Ghaffari, R.; Rogers, J.A.; et al. A Skin-Interfaced, Miniaturized Microfluidic Analysis and Delivery System for Colorimetric Measurements of Nutrients in Sweat and Supply of Vitamins Through the Skin. *Adv. Sci.* 2021, 9, 2103331.
15. Weaks, I.; Puck, C.; Sun, F.; Guerrero, C.A.; Liu, C.; Woo, W.M.; Pence, E.J.; Dagnan, R.A. Tailored electronic textile conformable sensor for large-scale spatially resolved physiological sensing on a large area. *Npj Flex. Electron.* 2020, 4, 5.

37. Zhu, Y.; Grubisic, S. Rate-Dependent Slip of Newtonian Liquids at Smooth Surfaces. *Phys. Rev. Lett.* **2001**, *87*, 096105.
38. Choi, C.H.; Westin, K.J.A.; Breuer, K.S. Apparent slip flows in hydrophilic and hydrophobic microchannels. *Phys. Fluids* **2003**, *15*, 2897.
39. Zhang, Y.; Chen, Y.; Huang, J.; Liu, Y.; Peng, J.; Chen, S.; Song, K.; Ouyang, X.; Cheng, H.; Wang, X. Skin-interfaced microfluidic devices with one opening chambers and hydrophobic valves for sweat collection and analysis. *Lab. Chip* **2020**, *20*, 2635–2645.
40. White, T.; Kaya, T. Evaluation of Hydrophilic Properties of Polydimethylsiloxane for Possible Microfluidic Sweat Sensor Applications. **2016**. Available online: http://people.se.cmcich.edu/yelam1k/asee/proceedings/2016/student_regular_papers/2016_asee_ncs_paper_19.pdf (accessed on 26 February 2022).
41. Escobedo, P.; Ramos-Lorente, C.E.; Martínez-Olmos, A.; Cavajal, M.A.; Ortega-Muñoz, M.; de Orbe-Payá, I.; Hernández-Mateo, F.; Santoyo-González, F.; Capitán-Vallvey, J.F.; Palma, A.J.; et al. Wireless wearable wristband for continuous sweat pH monitoring. *Sens. Actuators B Chem.* **2021**, *327*, 128948.
42. Design engineer considerations when selecting materials and designing microfluidic devices. *Adv. Healthc. Mater.* **2018**, *6*, 1601405.
43. Curto, V.F.; Fay, C.; Coyle, S.; Byrne, R.; O'Toole, C.; Barry, C.; Hughes, S.; Moyna, N.; Diamond, D.; Benito-Lopez, F. Real-time sweat pH monitoring based on a wearable chemical barcode micro-fluidic platform incorporating ionic liquids. *Sens. Actuators B Chem.* **2012**, *171–172*, 1327–1334.
44. The International Council of Engineered Machine (ICEM) Hardware (Microfluidic Bio-sensors and Electronic Device) Report Final. **2021**. Available online: <https://www.icem.org/Team/Report/Hardware/#/tree/2> (accessed on 26 February 2022).
45. Li, S.; Hart, K.; Norton, N.; Ryan, C.A.; Gugliani, L.; Prausnitz, M.R. Administration of pilocarpine by microneedle patch as a novel method for cystic fibrosis sweat testing. *Bioeng. Transl. Med.* **2021**, *6*, e10222.
46. Kim, J.; de Araujo, W.P.; Samek, J.A.; Bando, A.; Jia, W.; Brunetti, B.; Rairao, T.R.; Wang, J. Wearable temporary tattoo sensor for real-time trace metal monitoring in human sweat. *Electrochem. Commun.* **2015**, *51*, 41–45.
47. Xu, Z.; Song, J.; Liu, B.; Lv, S.; Gao, F.; Luo, X.; Wang, P. A conducting polymer PEDOT:PSS hydrogel based wearable sensing sensor integrated with a microfluidic device to provide the multimodal analysis of glucose, lactate, chloride, pH, and sweat rate. *Sens. Actuators B Chem.* **2021**, *348*, 130674.
48. Lin, H.; Tan, J.; Zhu, J.; Lin, S.; Zhao, Y.; Yu, W.; Hojajii, H.; Wang, B.; Yang, S.; Cheng, X.; et al. A programmable epidermal microfluidic valving system for wearable biofluid management and contextual biomarker analysis. *Nat. Commun.* **2020**, *11*, 4405.
49. Gunatillake, U.B.; Garcia-Rey, S.; Ojeda, E.; Basabe-Desmonts, L.; Benito-Lopez, F. H₂O₂ Nanotubes Alginate Hydrogel Scaffold for Rapid Sensing of Sweat Biomarkers: Lactate and Glucose. *ACS Appl. Mater. Interfaces* **2021**, *13*, 37734–37745.

50. Alizadeh, A.; Borras-Alcalá, I.; R. Settings, R.; Ashby, J.; Porter, A.; McCann, M.; Barnett, R.; Diamond, D.; White, P.S. A wearable patch for continuous monitoring of sweat electrolytes during exercise. *Chin Chem Lett* 2018, 18, 2637–2641. Electrolyte sensors in a single SSD provides a better fitness performance analysis, which contributes to accurate measurements.
51. Martin, A.; Kim, J.; Kurniawan, J.F.; Sempionatto, J.R.; Moreto, J.R.; Tang, G.; Campbell, A.S.; Shin, A.; Lee, M.Y.; Liu, X.; et al. Epidermal Microfluidic Electrochemical Detection System: Enhanced Sweat Sampling and Metabolite Detection. *ACS Sens.* 2017, 2, 1860–1868. Additionally, an SSD that integrates with various chemical sensors can also be featured in multitasking sweat applications such as medical diagnostics and the analysis of fitness performance, to reduce time consumption by simultaneously analyzing
52. Chiriac, P.; Xue, S.; Xia, W.; Reas, T.R.; Reddy, J.; Bando, A.; Velkovic, D.; Xu, S.; Huang, Y.; Rogers, J.A. Soft, skin-attached microfluidic systems for measuring secretory fluidic pressures generated at the surfaces of the skin by eccrine sweat glands. *Lab Chip* 2017, 17, 2570–2580. Furthermore, Nyein et al. developed multiplex electrochemical sensors comprising pH, chloride ion, and levodopa composition sensing, which were used to
53. Liang, B.; Wei, J.; Tu, T.; Cao, Q.; Mao, X.; Fang, L.; Ye, X. A Smartwatch Integrated with a Paper-based Microfluidic Patch for Sweat Electrolytes Monitoring material science View project Two Dimensional Carbon Materials Based Gas Sensor Development View project A Smartwatch Integrated with a Paper-based Microfluidic Electroanalysis 2020, 33, 643–651. When at rest, sweating more or less may be an additional sign of autonomic dysfunction, diabetes, cerebrovascular disease, Parkinson's disease, and chronic psychological stress such as anxiety or pain. Sweat pH can indicate acid–base imbalances, whereas chloride levels are helpful in testing for cystic fibrosis, electrolyte balance, and hydration status
54. Lan, W.J.; Maxwell, F.J.; Parolo, C.; Bwambok, D.K.; Subramaniam, A.B.; Whitesides, G.M. Paper-based electroanalytical devices with an integrated, stable reference electrode. *Lab Chip* 2013, 13, 4103–4108. Sweat-based levodopa screening could contribute to precision therapy for Parkinson's patients. They optimally maximized multifunction sweat applications, including physical hydration, neurological afflictions, and mental condition.
55. Kalluri, H.; Kolli, C.S.; Banga, A.K. Characterization of microchannels created by metal microneedles: Formation and closure. *AAPS J.* 2011, 13, 473–481. In sweat analysis, adding physical/physiological signal sensors (e.g., blood pressure, heart rate, and electrocardiograms) that integrate with a chemical sensor can provide a sufficient overview of a patient's health condition, the body's response, and physiological changes to daily activities. Sempionatto et al. constructed an SSD that could monitor vital signs (blood pressure and heart rate), the concentration of metabolites (glucose and lactate), and exogenous chemical levels (caffeine and alcohol) by analyzing sweat extracted through exercise, iontophoresis, and external stimuli (intake of food, caffeine, and alcohol)
56. McGonville, A.; Hegarty, C.; Davis, J. Mini-Review: Assessing the Potential Impact of Microneedle Technologies on Home Healthcare Applications. *Medicines* 2018, 5, 50. These biomarker sensors were proposed to have a correlation with the health self-monitoring of common daily activities such as eating, drinking, and exercising. Their SSD was optimized to provide good mechanical resilience and a dependable substrate on microneedle array penetration into skin. *J. Pharm. Sci.* 2019, 102, 4100–4108.
57. Kochmar, J.S.; Soon, W.J.; Choi, J.; Zou, S.; Kang, L. Effect of microneedle geometry and supporting substrate on microneedle array penetration into skin. *J. Pharm. Sci.* 2019, 102, 4100–4108. These biomarker sensors were proposed to have a correlation with the health self-monitoring of common daily activities such as eating, drinking, and exercising. Their SSD was optimized to provide good mechanical resilience and a dependable substrate on microneedle array penetration into skin. *J. Pharm. Sci.* 2019, 102, 4100–4108.
58. Park, J.H.; Allen, M.G.; Prausnitz, M.R. Biodegradable polymer microneedles: Fabrication, mechanics and transdermal drug delivery. *J. Control. Release* 2005, 104, 51–66. In addition, Imani et al. proposed an epidermal patch that could continually track sweat lactate levels and ECG signals concurrently to assess a wearer's physical strength, glucose detection in sweat without crosstalk between the different sensors.
59. Bai, S.M.; Oussif, I.S.; Pavlyushin, B.; Bouwstra, J.A. A possible assessment of the safety and function of arrays using electrostimulation. *Electrochim. Acta* 2008, 55, 193–202. This device offers a practical approach to researching and developing
60. Yang, S.; Lu, F.; Liu, Y.; Ning, Y.; Tian, S.; Zuo, P.; Ji, X.; He, Z. Quantum dots-based hydrogel microspheres for visual determination of lactate and simultaneous detection coupled with microfluidic physiology comprehensively. *Microchem. J.* 2021, 171, 106801. This device offers a practical approach to researching and developing multimodal wearable sensors that incorporate physical, electrophysiological, and chemical sensors to further measure human microspheres for visual determination of lactate and simultaneous detection coupled with microfluidic physiology comprehensively.
61. Wang, Z.; Guo, C.; De Quigley, C.; McNamee, S.; Zulfari, C.; Payne, C.; Slendon, T.; Diamond, D. A microfluidic device for assessing and wireless communication platform for sensing sodium in sweat. *Anal. Methods* 2015, 7, 041a. These features are essential for determining whether the newest SSDs can be used
62. Bolat, G.; De la Paz, E.; Azeredo, N.F.; Kartolo, M.; Kim, J.; de Loyola e Silva, A.N.; Rueda, R.; Brown, C.; Angnes, L.; Wang, J.; et al. Wearable soft electrochemical microfluidic device integrated with iontophoresis sensors, biophysical sensing, and multiple analyte measurement. The life span of the developed sensors was kept stable, with minimal drift of measurement values in current or voltage. In addition, blood testing, together with sweat analysis, can diagnose tissue oxygenation and pressure ischemia. This device offers a practical approach to researching and developing
63. Wei, L.; Fang, C.; Kuang, Z.; Cheng, H.; Wu, H.; Guo, D.; Liu, A. 3D printed low cost fabrication and facile integration of flexible epidermal microfluidics platform. *Sens. Actuators B Chem.* 2022, 353, 131085. Sweat has remained comparatively unexplored in comparison to standard biofluids, such as blood, despite its great promise for noninvasive physiological monitoring.
64. He, X.; Fan, C.; Luo, Y.; Xu, L.; Zhang, X. Flexible microfluidic nanoplasmonic sensors for refreshable and portable recognition of sweat biochemical fingerprint. *Npj Flex. Electron.* 2022, 6, 60. In addition, blood testing, together with sweat analysis, can diagnose tissue oxygenation and pressure ischemia. This device offers a practical approach to researching and developing

65. Liu, S.; Som Yang, T.; Wang, S.; Wang, S.; Yuan, H.; Li, S.; Seok, S.; Yee, M.; Liu, B.; Aranyosi, A.; Joo, J.; Jafari, R.; Rogers, J.A.; John Rogers, C.A.; et al. Soft, environmentally degradable microfluidic devices for measurement of

Analytes Detection	Solid Contact Materials	Types of Sensors	Physical/Physiological Signal Sensor	Sample	Correlate with Blood Test	Techniques Measurements	Repeatability and Life Span	References
Levodopa	Zeolitic imidazolate framework/graphene oxide (ZIF-8/GO)	Electrochemical	N/A	Sweat	Yes	Chronoamperometry, cyclic voltammetry	N/A 7 days	[122]
Glucose	PB	Colorimetry, electrochemical	N/A	Sweat, blood	Yes	Amperometry	N/A	[62]
Lactate	N/A	Colorimetric	Temperature	Artificial sweat, human sweat	Yes	Convolutional neural networks (CNNs)	N/A	[123]
Glucose, pH	PANI, reduced glucose oxidase (GOx)/ PtNPs/ Gold (Au)	Electrochemical	ECG, temperature, heart rate	Artificial sweat, human sweat, blood	Yes	Amperometry, potentiometry	Repeatability 1 week	[11]
Cl ⁻ , pH	N/A	Colorimetric	N/A	Artificial sweat, human sweat	N/A	Color intensity changing (absorbance, wavelength)	N/A	[65]
Glucose	PB-PEDOT-N	Electrochemical	N/A	Sweat, blood	Yes	Chronoamperometry	Repeatability 1 month	[75]
Na ⁺ , K ⁺	PEDOT/PSS	Electrochemical	N/A	Sweat	N/A	Chronoamperometry, potentiometry	Repeatability -	[81]
Na ⁺ , K ⁺ , pH	PEDOT/PSS, PANI	Electrochemical	N/A	Sweat	N/A	Chronoamperometry, potentiometry	Repeatability 30 days	[124]
Na ⁺ , K ⁺ , Pb ²⁺ , Li ⁺	Platinum nanoparticles (PtNPs)	Electrochemical	Temperature	NaCl, KCl, LiCl, Pb(NO ₃) ₂	Yes	Potentiometry	N/A	[125]
Glucose, lactate	PB	Electrochemical	Heart rate	Sweat	Yes	Amperometry	Disposable	[126]
Uric acid	Metal azolate framework-7 (MAF-7)	Electrochemical	N/A	Artificial sweat, human sweat	N/A	Amperometry, cyclic voltammetry	N/A	[127]
Creatinine	N/A	Fluorescence	N/A	Sweat, urine	Yes	Color intensity changing (absorbance, wavelength)	N/A	[128]

74. Yoon, J.H.; Kim, S.M.; Eom, Y.; Koo, J.M.; Cho, H.W.; Lee, T.J.; Lee, K.G.; Park, H.J.; Kim, Y.K.; Yoo, H.J.; et al. Extremely Fast Self-Healable Bio-Based Supramolecular Polymer for Wearable Real-Time Sweat-Monitoring Sensor. *ACS Appl. Mater. Interfaces* 2019, 11, 46165–46175.

2.3. Electronic Device

75. Lin, P.H.; Sheu, S.C.; Chen, C.W.; Huang, S.C.; Li, B.R. Wearable hydrogel patch with noninvasive, miniaturized electronic SSDs in the form of a simple and fashionable daily-worn accessory, such as a smartwatch, can be the ideal choice for a sweat monitoring platform routine. Cao et al. introduced a smartwatch integrated with a paper-based

76. Zhai, Q.; Yao, D.; Wang, R.; Gong, S.; Guo, Z.; Liu, Y.; Liu, Q.; Wang, J.; Simon, G.; Cheng, W. Microfluidic platform for sweat monitoring providing a digital display of real-time detection results of potassium and sodium ions in sweat. *ACS Applied Materials and Interfaces* 2020, 12, 123187.

77. Veric, A.; Aligned Gold Nanowires as Stretchable and Wearable Epidermal Ion-Selective Electrode for Noninvasive Multiplexed Sweat Analysis. *Anal. Chem.* 2020, 92, 4647–4655.

78. Lim, H.R.; Kim, Y.S.; Kwon, S.; Mahmood, M.; Kwon, Y.T.; Lee, Y.; Lee, S.M.; Yeo, W.H. Wireless, Flexible, Ion-Selective Electrode System for Selective and Repeatable Detection of Sodium. *Sensors* 2020, 20, 3297.

79. Bardackar, A.S.; Jeeran, I.; You, J.M.; Nunez-Flores, R.; Wang, J. Highly Stretchable Fully Printed CNT-Based Electrochemical Sensors and Biofuel Cells: Combining Intrinsic and Design-Induced Stretchability.

94. Olorunleke, M.; Calvo, S.; Lomonaco, T.; Ghimertevs, S.; Salvadori, S.; Serpelloni, F.; Branigan, E. Potentiometric Besides glucose and lactate, is sweat suitable for the determination of a urea? *Anal. Chim. Acta* 2017, **969**, 80–87. [151](#)
95. Furlan De Oliveira, R.; Montes-García, V.; Livio, P.A.; Begona González-García, M.; Fanjul-Bolado, P.; Casalini, S.; Samori, P.; Furlan De Oliveira, R.; Montes-García, V.; Livio, P.A.; et al. Selective Ion Sensing harvesting from sweat, an important guideline for controlling alcohol ingestion for heavy drinkers. Sadly, enzymatic fuel cells cannot generate electricity for an extended period of time because many oxidation–reduction enzymes degrade rapidly with low interactions, limiting their biocatalytic activity, stability, endurance, and energy capacity. Interestingly, Ryu et al. pioneered high robustness and long-lasting perspiration-based electricity production via a wearable paper-based microbial fuel cell (MFC) made by using a spore-forming skin bacterium, *Bacillus Subtilis* (BS) [150](#). In an extreme condition that led to the limitation of sweat, the BS formed endospores. When sweat was introduced, the BS repeatedly germinated spores to prevent their denaturation or degradation. This circumstance enabled the electrogenic capability of the BS to generate a sufficiently high-power density for a small-scale battery. Moreover, sensors powered by bacteria could be used to monitor human skin health and possibly release antibiotics, making BS a more appropriate source of energy when wound-healing devices or microneedles with transdermal drug delivery systems are present.
96. Zhu, Z.; Zhong, W.; Zhang, Y.; Dong, P.; Sun, S.; Zhang, Y.; Li, X. Elucidating electrochemical intercalation mechanisms of biomass-derived hard carbon in sodium/potassium ion batteries. *Carbon Energy* 2021, **3**, 541–553. [152](#)
97. Mahmood, A.; Li, S.; Ali, Z.; Tabassum, H.; Zhu, B.; Liang, Z.; Meng, W.; Akab, W.; Guo, W.; Zhang, H.; et al. Ultrafast Sodium/Potassium Ion Intercalation into Hierarchically Porous Thin Carbon Sheets. *Adv. Mater.* 2019, **31**, 1805430. [153](#)
98. Alvin, S.; Cahyadi, H.S.; Hwang, J.; Chang, W.; Kwak, S.K.; Kim, J. Revealing the Intercalation Mechanisms of Lithium, Sodium and Potassium in Hard Carbon. *Adv. Energy Mater.* 2020, **10**, 2000283. [154](#)
99. Liu, Y.; Zhang, Y.; Xu, L.; Zhong, L.; Sun, Z.; Ma, Y.; Bao, Y.; Gan, S.; Niu, L. Solid Contact Ion Sensing without Using an Ion Selective Membrane through Classic Li-Ion Battery Materials. *Anal. Chem.* 2021, **93**, 7538–7539. [155](#)
100. Sekine, Y.; Kim, S.B.; Zhang, Y.; Bandodkar, A.J.; Xu, S.; Choi, J.; Irie, M.; Ray, T.R.; Kohli, P.; Kozai, N.; et al. A fluorometric skin-interfaced microfluidic device and smartphone imaging module for in situ quantitative analysis of sweat chemistry. *Lab Chip* 2018, **18**, 2178–2186. [156](#)
101. Bandodkar, A.J.; Wit, G.; Pao, C.H.; Lee, P.; Belin, D.; Reber, D.; Poon, J.; Wang, W.; Arayonchi, A.; Lee, S.P.; et al. Battery-free, skin-interfaced microfluidic electronic systems for perspiration analysis for security purposes, chemical, and clinical safety and volitional analysis of sweat. *Sci. Adv.* 2019, **5**, eaah3294. [157](#)
102. Xiao, J.; Liu, Y.; Su, L.; Zhao, D.; Zhao, L.; Zhang, X. Microfluidic Chip-Based Wearable Colorimetric Sensor for Simple and Facile Detection of Sweat Glucose. *Anal. Chem.* 2019, **91**, 14803–14807. [158](#)
103. Kim, S.; Lee, B.; Reader, J.T.; Seo, S.H.; Lee, S.H.; Houria-Fargette, A.; Shin, J.; Sekine, Y.; Jeong, H. On-Passive Soft Skin-Interfaced Microfluidic Systems with Integrated Immunoassays, Fluorometric Sensors, and Impedance Measurement Capabilities. *Proc. Natl. Acad. Sci. USA* 2020, **117**, 27906–27915. [159](#)
104. Jiang, N.; Yetisen, A.K.; Linhart, N.; Flisikowski, K.; Dong, J.; Dong, X.; Butt, H.; Jakobi, M.; Schnieke, A.; Koch, A.W. Fluorescent dermal tattoo biosensors for electrolyte analysis. *Sens. Actuators B Chem.* 2020, intelligent digital networks with wide-ranging applications in fields including healthcare monitoring, disease detection, and personalized medical treatment. An ML algorithm offers advanced tools for processing and analyzing a large volume and complexity of raw data from SDS to improve the performance and quality of their system for personalized perspiration analysis. *Sens. Actuators B Chem.* 2020, 320, 128378. [160](#)
105. Andari, S.; Huseinifard, M.; Yosuf, M.; Gornhamadi, D. Towards smart personalized perspiration analysis: An IoT-integrated cellulose-based microfluidic wearable patch for smartphone fluorimetric multi-sensing of sweat biomarkers. *Biosens. Bioelectron.* 2020, **168**, 112459. [161](#)
106. Chung, M.; Fortunato, G.; Radacs, N. Wearable flexible sweat sensors for healthcare monitoring: A review. *J. R. Soc. Interface* 2019, **16**, 20190217. [162](#)
107. Califf, R.M. Biomarker definitions and their applications. *Exp. Biol. Med.* 2018, **243**, 213. [163](#)

108. Blawieńska, Ź.; Michalska, A.; Maksymiuk, K. Optimization of capacitance of conducting polymer solid state anion-selective electrodes. *Electrochim. Acta* 2016, 187, 397–405.

109. Nicholson, R.S. Theory and Application of Cyclic Voltammetry for Measurement of Electrode Reaction Kinetics. *Anal. Chem.* 2002, 37, 1351–1355.

110. Lo, T.W.B.; Aldous, J.; Ompton, R.C. The use of nano-carbon as an alternative to multi-walled carbon nanotubes in modified electrodes for adsorptive stripping voltammetry. *Sens. Actuators B Chem.* 2012, 162, 361–368.

111. Rose, D.P.; Ratterman, M.E.; Griffin, D.K.; Hou, L.; Kelley-Loughnane, N.; Nak, R.R.; Hager, J.A.; Papautsky, I.; Heikenfeld, J.C. Adhesive RFID Sensor Patch for Monitoring of Sweat Electrolytes. *IEEE Trans. Biomed. Eng.* 2015, 62, 1457–1465.

112. Zhang, Y.; Guo, H.; Kim, S.B.; Wu, Y.; Osteich, D.; Park, S.H.; Wang, X.; Weng, Z.; Li, B.; Bandodkar, A.J.; et al. Passive sweat collection and colorimetric analysis of biomarkers relevant to kidney disorders using a

Table 3: A summary of the latest SSDs' electronic device features advancements in 2022.

SSD Forms	Types of Power Source	Sensor Involved	Wireless Communication	Commercial Product	Machine Learning	Applications	References
Smartwatch	Rechargeable LiPo battery	Temperature sensor, relative humidity sensor, glucose sensor	Bluetooth	N/A	Decision tree regression algorithm	Continuous glucose monitoring	[160]
Adhesive tape	TENG	Acoustic sensors, epidermal sensor, triboelectric sensor, heart rate sensor	Internet-of-Things (IoT)	N/A	Deep learning algorithms	Human activity monitoring, cardiovascular monitoring, acoustic-biometric applications	[161]
Smart necklace	BFC	Sodium, hydrogen, potassium, glucose sensor	Vector network analyzer (VNA)	N/A	A low-pass fast Fourier transform algorithm	Detect sweat electrolytes and glucose	[162]
Hexagonal bounding shape of microfluidic patch	N/A	Colorimetric, sodium sensor, chloride sensors	Image capture from microfluidic patch sweat metrics using smartphone	N/A	Canny edge detection algorithm, image analysis algorithms, multiple regressions	Sweating rate, total sweat loss, sweat electrolyte concentration loss	[163]
A nano-porous polyamide substrate	Battery	Cytokine sensor	N/A	N/A	Supervised discriminant factor analysis	Detect of Interleukin-31 (IL-31),	[164]

121. Imani, S.; Bandodkar, A.J.; Mohan, A.M.; Kumar, R.; Yu, S.; Wang, J.; Mercier, P.P. A wearable chemical-electrophysiological hybrid biosensing system for real-time health and fitness monitoring. *Nat. Commun.* 2016, 7, 11650.

122. Xiao, J.; Fan, C.; Xu, T.; Su, L.; Zhang, X. An electrochemical wearable sensor for levodopa quantification in sweat based on a metal–Organic framework/graphene oxide composite with integrated enzymes. *Sens. Actuators B Chem.* 2022, 359, 131586.

SSD Forms	Types of Power Source	Sensor Involved	Wireless Communication	Commercial Product	Machine Learning	Applications	References
along with serpentine gold electrodes					(DFA) linear regression model of a binary classifier	chronic skin disease	highly accurate 132489.
Wristband	3.7 V LiPo battery (168 h on single charge)	Interferon-inducible protein (IP-10), tumor necrosis factor-related apoptosis-inducing ligand (TRAIL), and C-reactive protein (CRP) sensors	Bluetooth (Smartphone app)	SWEATSENSOR Dx-EnLiSense	N/A	Detect simultaneously and continuously specific IP-10, TRAIL, CRP	et al. Human wearable Chem. 2021, fabricated 2022, 368, works integrated mechanisms of 0, 045003.
Smartwatch	110 mAh Li-ion battery	Cortisol sensor	Bluetooth	Aptamer-FET biosensing smartwatch	N/A	Track stress level	[166] Paper-based
Wristband	N/A	IL-6 sensor, pH sensor	Bluetooth	WRRIST	N/A	Detect IL-6 levels (Inflammatory biomarkers)	[167] by iontophoretic delivery of the slowly-metabolized cholinergic agent carbachol. <i>J. Dermatol. Sci.</i> 2016, 69, 40–51.

131. Francis, J.; Stamper, I.; Heikenfeld, J.; Gomez, E.F. Digital nanoliter to milliliter flow rate sensor with in vivo demonstration for continuous sweat rate measurement. *Lab Chip* 2018, 19, 178–185.

132. Zhao, J.; Lin, Y.; Wu, J.; Nyein, H.Y.Y.; Bariya, M.; Tai, L.C.; Chao, M.; Ji, W.; Zhang, G.; Fan, Z.; et al. A Fully Integrated and Self-Powered Smartwatch for Continuous Sweat Glucose Monitoring. *ACS Sens.* 2019, 4, 1925–1933.

133. Song, Y.; Wang, H.; Cheng, X.; Li, G.; Chen, X.; Chen, H.; Miao, L.; Zhang, X.; Zhang, H. High-efficiency self-charging smart bracelet for portable electronics. *Nano Energy* 2019, 55, 29–36.

134. Yu, Y.; Nassar, J.; Xu, C.; Min, J.; Yang, Y.; Dai, A.; Doshi, R.; Huang, A.; Song, Y.; Gehlhar, R.; et al. Biofuel-powered soft electronic skin with multiplexed and wireless sensing for human-machine interfaces. *Sci. Robot.* 2020, 5, eaaz7946.

135. Tai, L.C.; Gao, W.; Chao, M.; Bariya, M.; Ngo, Q.P.; Shahpar, Z.; Nyein, H.Y.Y.; Park, H.; Sun, J.; Jung, Y.; et al. Methylxanthine Drug Monitoring with Wearable Sweat Sensors. *Adv. Mater.* 2018, 30, 1707442.

136. Zhao, J.; Zha, J.; Zeng, Z.; Tan, C. Recent advances in wearable self-powered energy systems based on flexible energy storage devices integrated with flexible solar cells. *J. Mater. Chem. A* 2021, 9, 18887–18905.

137. Dong, P.; Rodrigues, M.T.F.; Zhang, J.; Borges, R.S.; Kalaga, K.; Reddy, A.L.; Silva, G.G.; Ajayan, P.M.; Lou, J. A flexible solar cell/supercapacitor integrated energy device. *Nano Energy* 2017, 42, 181–186.

138. Siddiqui, S.; Lee, H.B.; Kim, D.I.; Duy, L.T.; Hanif, A.; Lee, N.E. An Omnidirectionally Stretchable Piezoelectric Nanogenerator Based on Hybrid Nanofibers and Carbon Electrodes for Multimodal Straining

- and Human Kinematics Energy Harvesting. *Adv. Energy Mater.* 2018, 8, 1701520.
139. Zhao, T.; Fu, Y.; Sun, C.; Zhao, X.; Jiao, C.; Du, A.; Wang, Q.; Mao, Y.; Liu, B. Wearable biosensors for real-time sweat analysis and body motion capture based on stretchable fiber-based triboelectric nanogenerators. *Biosens. Bioelectron.* 2022, 205, 114115.
140. Zhang, Q.; Jin, T.; Cai, J.; Xu, L.; He, T.; Wang, T.; Tian, Y.; Li, L.; Peng, Y.; Lee, C. Wearable Triboelectric Sensors Enabled Gait Analysis and Waist Motion Capture for IoT-Based Smart Healthcare Applications. *Adv. Sci.* 2022, 9, 2103694.
141. Wang, J.; Li, S.; Yi, F.; Zi, Y.; Lin, J.; Wang, X.; Xu, Y.; Wang, Z.L. Sustainably powering wearable electronics solely by biomechanical energy. *Nat. Commun.* 2016, 7, 12744.
142. Liu, D.; Yin, X.; Guo, H.; Zhou, L.; Li, X.; Zhang, C.; Wang, J.; Wang, Z.L. A constant current triboelectric nanogenerator arising from electrostatic breakdown. *Sci. Adv.* 2019, 5, eaav6437.
143. Chen, X.; Song, Y.; Su, Z.; Chen, H.; Cheng, X.; Zhang, J.; Han, M.; Zhang, H. Flexible fiber-based hybrid nanogenerator for biomechanical energy harvesting and physiological monitoring. *Nano Energy* 2017, 38, 43–50.
144. Kwak, S.S.; Yoon, H.J.; Kim, S.W. Textile-Based Triboelectric Nanogenerators for Self-Powered Wearable Electronics. *Adv. Funct. Mater.* 2019, 29, 1804533.
145. Yi, Q.; Pei, X.; Das, P.; Qin, H.; Lee, S.W.; Esfandyarpour, R. A self-powered triboelectric MXene-based 3D-printed wearable physiological biosignal sensing system for on-demand, wireless, and real-time health monitoring. *Nano Energy* 2022, 101, 107511.
146. Falk, M.; Pankratov, D.; Lindh, L.; Arnebrant, T.; Shleev, S. Miniature Direct Electron Transfer Based Enzymatic Fuel Cell Operating in Human Sweat and Saliva. *Fuel Cells* 2014, 14, 1050–1056.
147. Wang, C.; Shim, E.; Chang, H.K.; Lee, N.; Kim, H.R.; Park, J. Sustainable and high-power wearable glucose biofuel cell using long-term and high-speed flow in sportswear fabrics. *Biosens. Bioelectron.* 2020, 169.
148. Jia, W.; Valdés-Ramírez, G.; Bandodkar, A.J.; Windmiller, J.R.; Wang, J. Epidermal Biofuel Cells: Energy Harvesting from Human Perspiration. *Angew. Chem. Int. Ed.* 2013, 52, 7233–7236.
149. Garcia, S.O.; Ulyanova, Y.V.; Figueroa-Teran, R.; Bhatt, K.H.; Singhal, S.; Atanassov, P. Wearable Sensor System Powered by a Biofuel Cell for Detection of Lactate Levels in Sweat. *ECS J. Solid State Sci. Technol.* 2016, 5, M3075.
150. Koushanpour, A.; Gamella, M.; Katz, E. A Biofuel Cell Based on Biocatalytic Reactions of Lactate on Both Anode and Cathode Electrodes—Extracting Electrical Power from Human Sweat. *Electroanalysis* 2017, 29, 1602–1611.
151. Sun, M.; Gu, Y.; Pei, X.; Wang, J.; Liu, J.; Ma, C.; Bai, J.; Zhou, M. A flexible and wearable epidermal ethanol biofuel cell for on-body and real-time bioenergy harvesting from human sweat. *Nano Energy* 2021, 86, 106061.
152. Cederbaum, A.I. Alcohol metabolism. *Clin. Liver Dis.* 2012, 16, 667–685.

153. Ryu, J.; Landers, M.; Choi, S. A sweat-activated, wearable microbial fuel cell for long-term, on-demand power generation. *Biosens. Bioelectron.* 2022, 205, 114128.
154. Foster, K.R. Is Wi-Fi a Health Threat in Schools? *Educ. Next* 2019, 19, 28–37.
155. Ulrich, C.M.; Demiris, G.; Kennedy, R.; Rothwell, E. The ethics of sensor technology use in clinical research. *Nurs. Outlook* 2020, 68, 720–726.
156. Abdulla, R.; Sathish Kumar Selvaperumal, A.; Bathich, A.; Ali Khan, H.; Kumar Selvaperumal, S. IoT based on secure personal healthcare using RFID technology and steganography Wireless power transfer using conical and spiral coils View project IoT based on secure personal healthcare using RFID technology and steganography. *Artic. Int. J. Electr. Comput. Eng.* 2021, 11, 3300–3309.
157. Hureib, E.; Gutub, A.; Bin Hureib, E.S.; Gutub, A.A. Enhancing Medical Data Security via Combining Elliptic Curve Cryptography and Image Steganography. *Int. J. Comput. Sci. Netw. Secur.* 2020, 20, 1–8.
158. Baik, S.; Lee, J.; Jeon, E.J.; Park, B.Y.; Kim, D.W.; Song, J.H.; Lee, H.J.; Han, S.Y.; Cho, S.W.; Pang, C. Diving beetle-like miniaturized plungers with reversible, rapid biofluid capturing for machine learning-based care of skin disease. *Sci. Adv.* 2021, 7, 5695–5711.
159. Lin, S.; Hu, S.; Song, W.; Gu, M.; Liu, J.; Song, J.; Liu, Z.; Li, Z.; Huang, K.; Wu, Y.; et al. An ultralight, flexible, and biocompatible all-fiber motion sensor for artificial intelligence wearable electronics. *Npj Flex. Electron.* 2022, 6, 27.
160. Sankhala, D.; Sardesai, A.U.; Pali, M.; Lin, K.C.; Jagannath, B.; Muthukumar, S.; Prasad, S. A machine learning-based on-demand sweat glucose reporting platform. *Sci. Rep.* 2022, 12, 2442.
161. Wang, H.L.; Guo, Z.H.; Pu, X.; Wang, Z.L. Ultralight Iontronic Triboelectric Mechanoreceptor with High Specific Outputs for Epidermal Electronics. *Nano-Micro Lett.* 2022, 14, 86.
162. Liu, T.L.; Dong, Y.; Chen, S.; Zhou, J.; Ma, Z.; Li, J. Battery-free, tuning circuit-inspired wireless sensor systems for detection of multiple biomarkers in bodily fluids. *Sci. Adv.* 2022, 8, 7049.
163. Baker, L.B.; Seib, M.S.; Barnes, K.A.; Brown, S.D.; King, M.A.; De Chavez, P.J.D.; Qu, S.; Archer, J.; Wolfe, A.S.; Stofan, J.R.; et al. Skin-Interfaced Microfluidic System with Machine Learning-Enabled Image Processing of Sweat Biomarkers in Remote Settings. *Adv. Mater. Technol.* 2022, 7, 2200249.
164. Upasham, S.; Rice, P.; Shahub, S.; Dhamu, V.N.; Prasad, S. Passive Sweat-Based Pruritic Cytokine Detection and Monitoring System. *ECS Sens. Plus* 2022, 1, 031602.
165. Jagannath, B.; Pali, M.; Lin, K.C.; Sankhala, D.; Naraghi, P.; Muthukumar, S.; Prasad, S. Novel Approach to Track the Lifecycle of Inflammation from Chemokine Expression to Inflammatory Proteins in Sweat Using Electrochemical Biosensor. *Adv. Mater. Technol.* 2022, 7, 2101356.
166. Wang, B.; Zhao, C.; Wang, Z.; Yang, K.A.; Cheng, X.; Liu, W.; Yu, W.; Lin, S.; Zhao, Y.; Cheung, K.M.; et al. Wearable aptamer-field-effect transistor sensing system for noninvasive cortisol monitoring. *Sci. Adv.* 2022, 8, 967.
167. Noushin, T.; Tabassum, S. WRRIST: A wearable, rapid, and real-time infection screening tool for dual-mode detection of inflammatory biomarkers in sweat. In *Microfluidics, BioMEMS, and Medical Microsystems XX*; SPIE: Bellingham, WA, USA, 2022; Volume 11955, p. 1195502.

Retrieved from <https://www.encyclopedia.pub/entry/history/show/72655>