

Physicochemical Properties and Activities of Catechins

Subjects: **Dermatology**

Contributor: Soraya Ratnawulan Mita , Patihul Husni , Norisca Aliza Putriana , Rani Maharani , Ryan Proxy Hendrawan , Dian Anggraeni Dewi

Catechins are bioactive polyphenols and are typically isolated from green tea (*Camellia sinensis* L.) and gambir leaves (*Uncaria gambir* Roxb). There are a few types of catechins which possess the flavan-3-ol structure consisting of two benzene rings, a heterocycle dihydropyran, and a hydroxyl.

catechins

flavonoid

antioxidant

cosmeceutical

1. Introduction

Cosmetics are widely used by everyone and can be classified as decorative or skincare cosmetics. They are defined by BPOM Indonesia as materials or preparations intended for use on the external parts of the human body such as the epidermis, hair, nails, lips, and external genital organs, or on the teeth and oral mucous membranes, especially for cleaning, perfuming, changing the appearance and/or improving body odour, or to protect or maintain the body in good condition. Based on the literature, cosmetics are often used for aesthetic and self-care benefits, and that they also intersect with skin health. The skin is the largest human organ and protects the body from external insults, therefore, it is essential to care for the skin according to the unique needs of the individual. The term 'cosmeceutical' refers to a cosmetic product that provides medical effects and contains certain active compounds. These cosmeceutical products are not only for whitening, antiageing, and sunscreen purposes, but also for hyperpigmentation, photoageing, wrinkles, and hair loss ^[1]. It is important to note that cosmeceutical preparations are not intended to be given systematically, rather, they are applied locally/topically, that is, the 'dermal delivery' as the site of action is usually the stratum corneum (SC), the viable epidermis and/or dermis ^[2].

Skin ageing typically starts to manifest when an individual reaches their late 20s to early 30s due to two distinct sources ^[3]: intrinsic ageing, which results from issues within the network of elastin fibres and collagen, or extrinsic ageing due to exposure to environmental factors such as sun radiation. Oxidative stress triggers inflammation, constraining epidermal cell renewal and ultimately leads to a reduction in epidermal thickness and a weakening of the protective barrier ^[4]. Sun radiation triggers the creation of reactive oxygen radicals (ROS) that cause keratinocytes to generate pro-inflammatory cytokines, including tumor necrosis factor- α (TNF- α) and interleukin-8 (IL-8), thereby producing more ROS ^[5]. Excessive ROS leads to the development of wrinkles by causing the breakdown and abnormal interlinking of structural proteins such as glycosaminoglycans, collagen, and elastin fibres in the skin's extracellular matrix. Hence, antioxidants isolated from natural products can be used to suppress ROS production to slow down skin ageing ^[6].

Catechins are natural flavan-3-ols (or flavonols), a type of polyphenolic compound belonging to the flavonoid family. They are present in a variety of fruits, vegetables, and plant-based beverages [7] and are particularly concentrated in tea leaves, red wine, broad beans, rock-rose leaves, apricots, black grapes, and strawberries. Epicatechin is abundant in chocolate, apples, broad beans, pears, black grapes, cherries, and certain types of berries including blackberries and raspberries [8].

Catechins offer numerous health benefits by effectively eliminating free radicals and slowing down the breakdown of the extracellular matrix caused by exposure to ultraviolet (UV) radiation and pollution. They stimulate collagen production while preventing the generation of matrix metalloproteinase enzymes. Due to the presence of hydroxyl in the gallate group, epigallocatechin gallate (EGCG) and epigallocatechin (ECG) can neutralise free radicals, surpassing several antioxidants like trolox, ascorbic acid, and tocopherol [9]. Thus, catechins have the potential to be used in cosmetic and dermatological products [10] and are now commonly included in pharmaceutical, medical, and cosmetic products. For example, a transethosomal gel form of catechins can reduce total cholesterol in mice [11] and *Uncaria gambir* is used to treat diarrhoea, sore throat, spongy gums, dysentery, arteriosclerosis, and obesity [12].

2. Physicochemical Properties of Catechins

Catechins are bioactive polyphenols and are typically isolated from green tea (*Camellia sinensis* L.) and gambir leaves (*Uncaria gambir* Roxb). There are a few types of catechins which possess the flavan-3-ol structure consisting of two benzene rings, a heterocycle dihydropyran, and a hydroxyl [13][14]. The structures are shown in **Figure 1**.

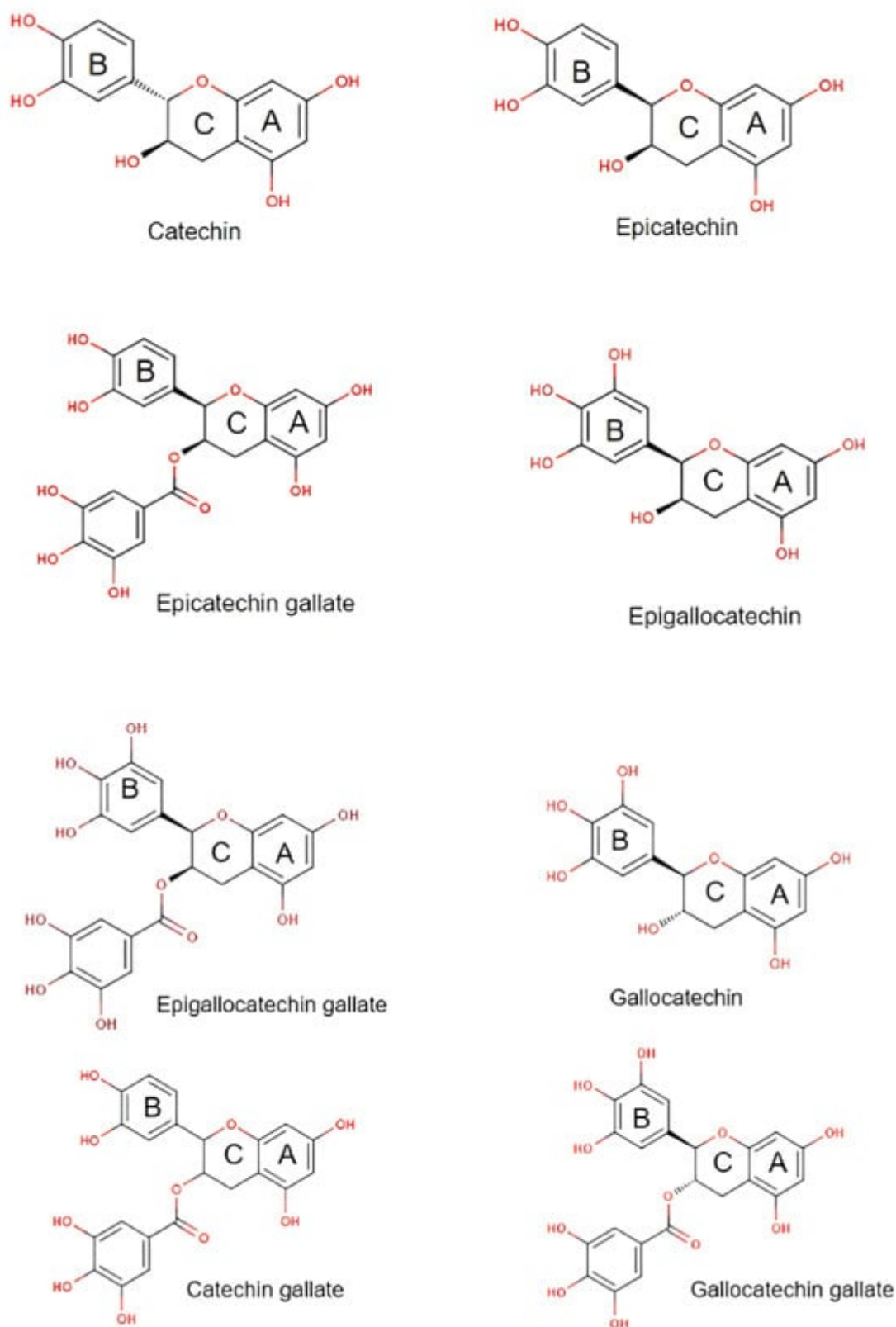


Figure 1. Various type of catechin structures.

Catechins have been utilised in many pharmacological formulations [15][16]. The use of catechins is based on its physicochemical characteristics such as polarisability, dipole moment, molecular weight, surface area, and van der Waals volume, as well as macroscopic traits including solubility, octanol/water partition coefficient, acidity or

basicity in solution, etc. [17]. In the Indonesian standard guide, namely the *Indonesian Herbal Pharmacopoeia*, there are several physicochemical profiles of catechins obtained from gambir plants (*Uncaria gambir* (Hunter) Roxb.) as shown in **Table 1**.

Table 1. Physicochemical parameters of catechins based on the *Pharmacopoeia* handbook.

Physicochemical Parameters	Accepted Value	Reference
Organoleptic	Solid form, it appears as a light brown to dark reddish-brown substance with a distinctive odour. It possesses a chelate taste that is slightly bitter at first but ends with a sweet aftertaste.	[18]
Water Content	Quantity should not exceed 14%	[18]
Ash Content	Quantity should not exceed 0.5%	[18]
Ash, not soluble in acid	Quantity should not exceed 0.1%	[18]
Purity	Contains no less than 90% tannins counted as catechins.	[18]
Identification	The assay was carried out using spectrophotometry with a wavelength of 294 nm.	[18]
Molecular weight	290.27 g/mol	[18]
Solubility	Soluble in water and polar organic solvents; soluble in pressurised hot water between 298.75 to 415.85 K; soluble in mixtures of supercritical carbon dioxide (SC-CO ₂) and ethanol at 313 K and pressures ranging from 80 to 120 bar; soluble in SC-CO ₂ between 313.15 and 343.15 K and pressures ranging from 12 to 26 MPa using ethanol as the co-solvent.	[19]

3. Activities of Catechin

3.1. Antioxidant

Antioxidants scavenge oxygen free radicals (singlet and triplet), ROS, peroxide decomposers, and enzyme inhibitors to protect vital molecules from harm [20]. The use of antioxidants derived from natural ingredients is increasing. Natural antioxidants can be single pure compounds/isolates, combinations of compounds, or plant extracts. The secondary metabolites, polyphenols, are the most common phyto-antioxidants [21]. Polyphenols have benzene rings with attached -OH groups which determine the antioxidant activity based on their number and position [22]. Protein phosphorylation is influenced by phenol groups by inhibiting lipid peroxidation. Flavonoids are the main source of polyphenols, while carotenoids are the most abundant sources of terpenes [23].

Catechin produces and discards free radicals [24] through several key direct and indirect antioxidant mechanisms. The direct mechanism involves the scavenging of ROS, whereas the indirect mechanism occurs through increased

antioxidant enzymes and the inhibition of the pro-enzyme that participates in oxidant stress [8][25]. The phenolic hydroxyl group in catechin is involved in the scavenging of ROS, therefore more hydroxyl groups will improve the antioxidant activity. According to the structure, the hierarchy of antioxidant activity of catechins is EGCG, EGG, EGC, EC, and, lastly, catechin [8].

The structural characteristics of flavan-3-ols, specifically their resorcinol and catechol components, which consist of A and B rings connected by the Pyron ring (C ring), are responsible for their antioxidant properties (Figure 1) [26]. The ability of flavan-3-ols to scavenge radicals primarily relies on the arrangement of hydroxyl groups and their capacity to donate hydrogen atoms [27]. The stability of the phenoxy radical produced after hydrogen atom transfer (HAT) also plays a role in their ability to counteract reactive oxygen radicals [28]. Catechins can exist in four different diastereoisomers, which arise from two chiral centers (2^n) at C2 and C3. These diastereoisomers are referred to as (+) catechin (2R, 3S), (–) epicatechin (2R, 3R), (–) catechin (2S, 3R), and (+) epicatechin (2S, 3S) [29]. Overall, the stereoisomerisms are determined by the positioning of the B ring connected to the C ring at the C2 atom and also the chirality of R1 and R2 attached to the C ring at the C3.

The structures are indicative of a site of a projected bond—R stands for dashed wedge bonds that extend away from the viewer, and S represents solid wedge bonds that project out of the paper towards the viewer, as seen in Figure 2. The degree of polymerization also plays a role in determining the antioxidant properties [30]. The capability of catechin and epicatechin to scavenge radicals is attributed to the dihydroxyl group at C-3' and C-4' on the B ring, a C3-hydroxyl group on the C ring without a 2,3 double bond, and hydroxyl groups at C5 and C7 on ring A. In this context, the catechol ring (B) exhibits greater electron-donating capacity than the other rings because it contains an ortho-dihydroxyl group (detail see Figure 2 and Figure 3) [31]. The scavenging of free radicals by the hydroxyl groups occurs via hydrogen atom transfer and single-electron atom transfer, with catechin undergoing oxidation to form the relatively reactive quinone [32] as could be observed from Figure 4.

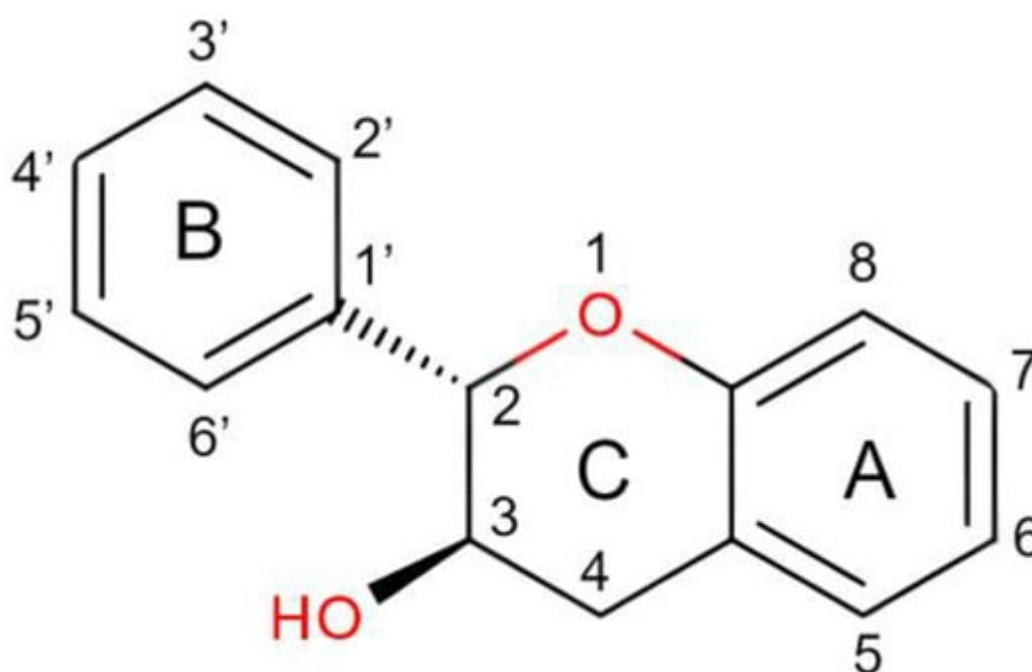
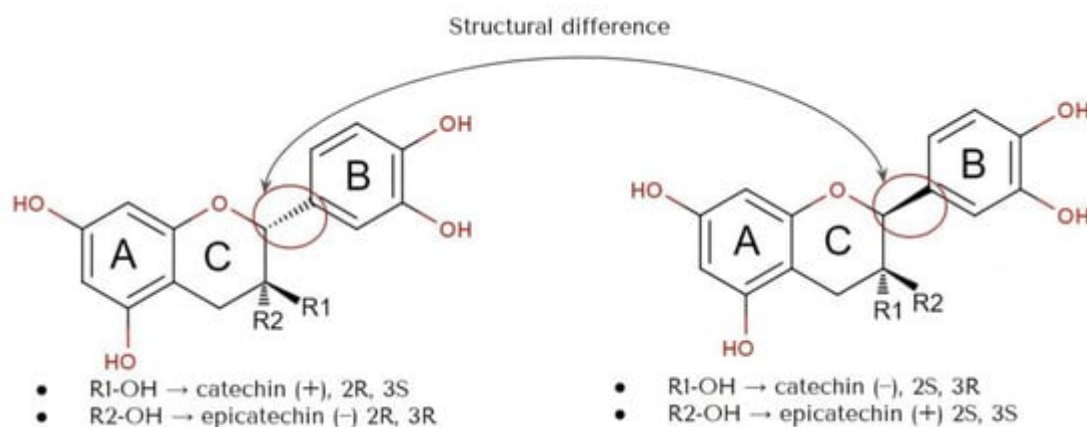
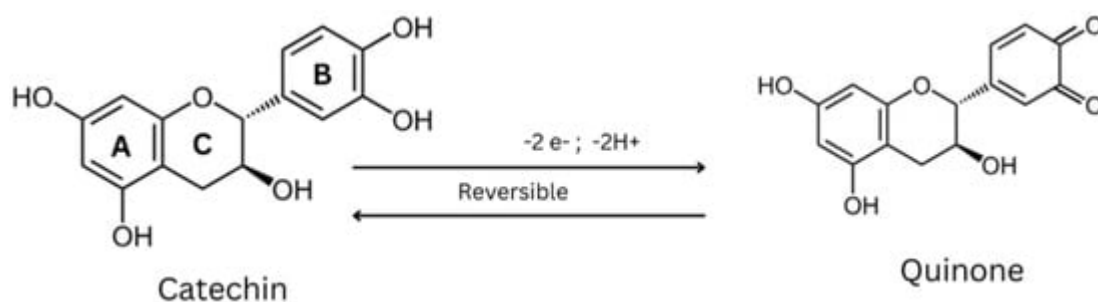


Figure 2. Structure of Catechin in Detail.**Figure 3.** Structure of stereoisomers of catechin (+) and epicatechin (-), also catechin (-) and epicatechin (+).**Figure 4.** Scavenging of free radicals by catechin.

Numerous studies have reported that the primary source of antioxidant activity in CT and ECT is their elevated redox characteristics. The antioxidant properties of catechin and epicatechin, which are found in different plant extracts, were assessed using various experimental antioxidant tests, including ferric reducing antioxidant power (FRAP), 1,1-diphenyl-1-picrylhydrazyl (DPPH), nitric oxide (NO), and 2,2-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) (ABTS⁺), demonstrating that CT and ECT exhibit strong antioxidant capabilities [33]. CT and ECT share a similar molecular structure, but they exhibit distinct chemical reactivity properties because CT's antioxidant capability relies on its planar geometry, whereas ECT's antioxidant activity is due to the interaction of hydrogen bonds at the catechol moiety. The lack of a double bond at C2 = C3 in the C ring significantly affects ECT's antioxidant capacity. Additionally, it has been demonstrated that the reactivity of flavan-3-ols is thermodynamically modified depending on the solvents used [34].

There are many uses of catechins. Aside from being used as pure antioxidant compounds, catechins can be formulated into many products including sunscreen, lip balm, anti-dandruff shampoo, cosmetic cleansers, and cosmetic creams. They have also been extracted from many natural sources, as detailed in **Table 2**.

Table 2. Antioxidant activity of catechins from various sources.

Methods	Sample	Result	Ref.
DPPH Assay	Camellia sinensis	<ul style="list-style-type: none">Solvent Used: Methanol	[35]
		<ul style="list-style-type: none">Green Tea = 67.3%	
		<ul style="list-style-type: none">White Tea = 47.9%	
		<ul style="list-style-type: none">Black Tea = 28.9%	
	Lepisanthes alata (Blume) Leenh	<ul style="list-style-type: none">Solvent Used: Water	[36]
		<ul style="list-style-type: none">Rind: 61.61%	
		<ul style="list-style-type: none">Flesh: 47.93%	
		<ul style="list-style-type: none">Seeds: 48.66%	
		<ul style="list-style-type: none">Whole Fruit: 69.30%	
		<ul style="list-style-type: none">Leaves: 59.35%	
		<ul style="list-style-type: none">Bark: 49.91%	
		<ul style="list-style-type: none">Solvent Used: Methanol	
		<ul style="list-style-type: none">Rind: 86.17%	
		<ul style="list-style-type: none">Flesh: 27.47%	
		<ul style="list-style-type: none">Seeds: 89.58%	
		<ul style="list-style-type: none">Whole Fruit: 78.34%	
		<ul style="list-style-type: none">Leaves: 61.71%	
		<ul style="list-style-type: none">Bark: 87.03%	
		<ul style="list-style-type: none">Solvent Used: Ethanol	
		<ul style="list-style-type: none">Rind: 85.81%	

Methods	Sample	Result	Ref.
		<ul style="list-style-type: none"> Flesh: 21.23% Seeds: 90.12% Whole Fruit: 46.20% Leaves: 79.61% Bark: 87.03% 	
	<i>Sterculia quadrifida</i> R.	Bark: 51.5 µg/mL (50%)	[20]
Malondialdehyde	<i>Uncaria Gambir</i> Roxb	<ul style="list-style-type: none"> Dose of 5 mg/kg: 0.19% Dose of 10 mg/kg: 31.28% Dose of 20 mg/kg: 57.63% Control + (Vit E): 5.55% Control-: -77.79% 	[37]
	<i>Uncaria Gambir</i> Roxb.	Varies from 2.732% to 3.792%	[38]
	Combination of Isolated Catechin and Quercetin	<ul style="list-style-type: none"> Insignificant antioxidant activity Dose of catechin used was 100 µg/mL ($p < 0.05$) 	[39]
[40]			

effects of

Table 3. The various roles of antioxidants on skin.

Antioxidant Function	Mechanism of Action	Reference
Anti-ageing	Antioxidants inhibit the action of superoxide dismutase (SOD) enzymes, which play a role in degrading collagen.	[41]
Skin Brightening	Antioxidants have whitening effects by inhibiting tyrosinase and act as anti-inflammatory agents for hyperpigmentation caused by UV exposure (commonly known as melasma).	[42]
UV Filters	Topically administered antioxidants can enhance the photoprotective capabilities of UV filters by reducing erythema, inhibiting the	[43]

Antioxidant Function	Mechanism of Action	Reference
	development of sunburned skin cells, and causing immunosuppression.	
Skin Hydration and Anti-Hyperpigmentation	Antioxidants suppress the production and secretion of melanin in melanoma cells to enhance skin hydration and improve hyperpigmentation.	[44]

3.2. Anti-Ageing

Elastase, a protein kinase enzyme, cleaves specific polypeptide bonds to reduce elastin levels. Preventing elastase functions within the dermis layer can be utilised to maintain the skin's flexibility, therefore, elastase activity inhibitors can serve as cosmetic ingredients to counteract the signs of skin ageing [45]. Polyphenols found in isolated white tea inhibit collagenase and elastase enzymes, specifically catechin and EGCG (**Figure 5**). Furthermore, given that collagenase is a zinc-containing metalloproteinase, these catechins could potentially attach to the Zn^{2+} ion present in the enzyme, thereby obstructing its ability to bind to the substrate [46].

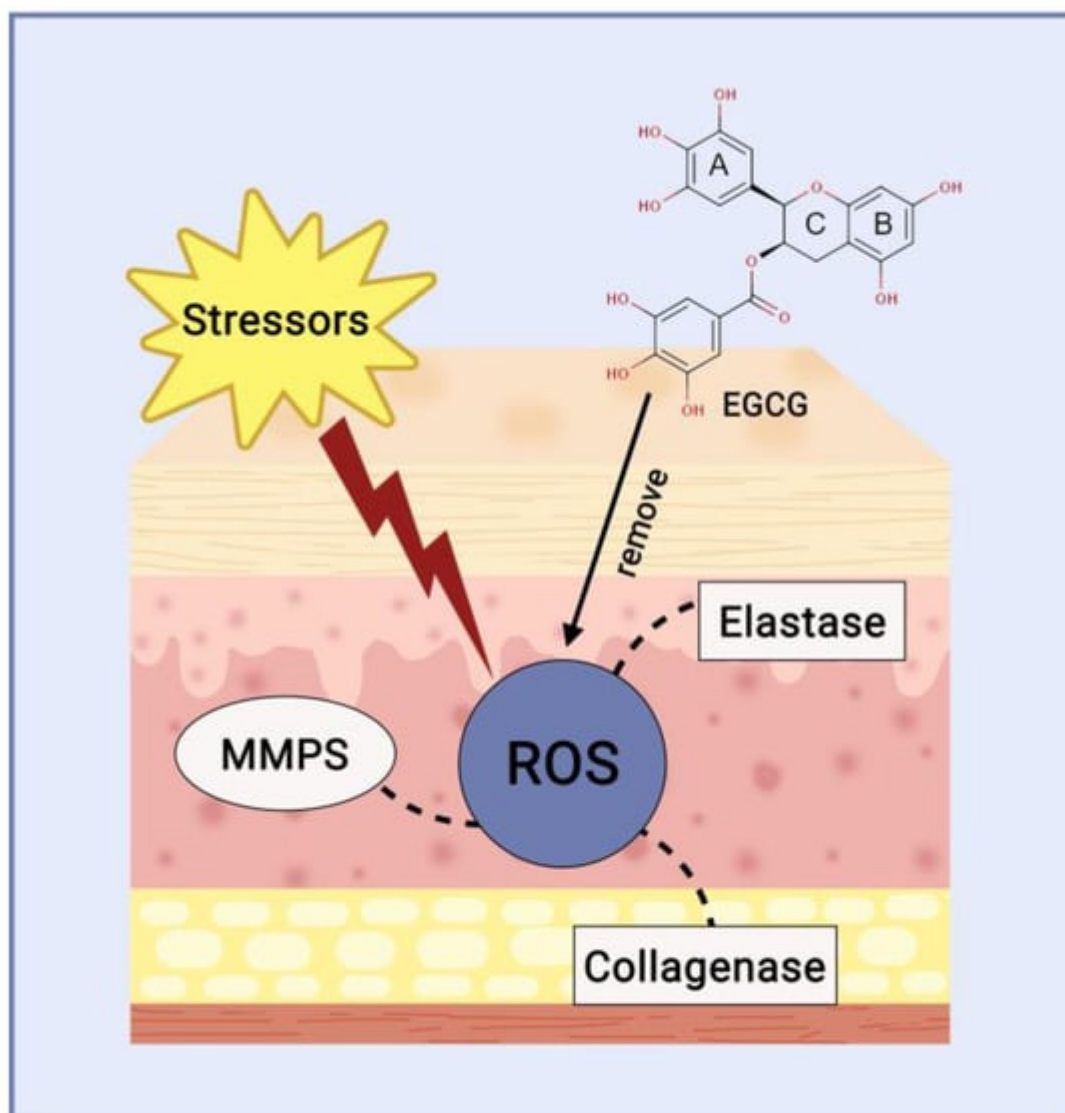


Figure 5. Catechin mechanism of action as anti-ageing agent.

3.3. Skin Brightener

Catechin directly inhibits tyrosinase activity and reduces the expression of tyrosinase [47]. EGCG, GCG, and EGC demonstrate great potential as tyrosinase activity inhibitors [48]. The catechins have a substantial inhibitory effect on tyrosinase activity and melanin production by downregulating the cAMP/CREB/MITF signalling pathway in B16F10 cells (Figure 6), with EGC demonstrating the strongest effect, followed by EGCG and GCG [49]. EGCG also suppresses the production of melanin induced by α -MSH in B16 melanoma cells [47].

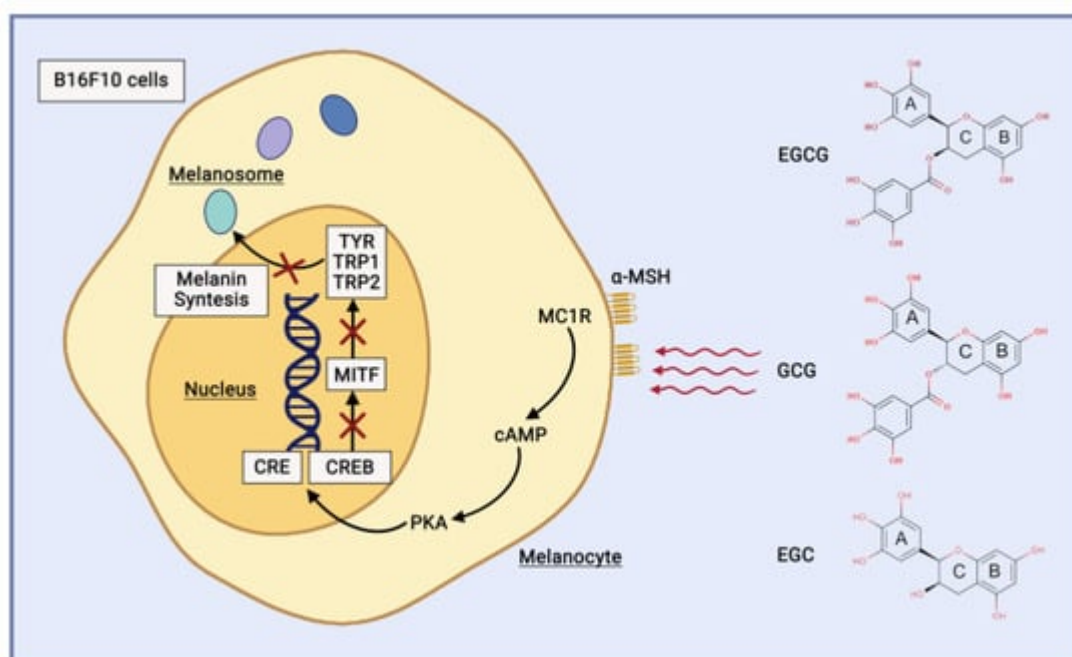


Figure 6. Catechin mechanism of action as anti-hyperpigmentation and brightening agent.

3.4. Anti-Hyperpigmentation

UV radiation results in melanogenesis. The mass production of melanin in the skin minimises UV radiation but causes the skin to slightly darken to a brownish color. Catechin as a depigmentation agent will inhibit melanin formation by inhibiting melanin synthesis through tyrosinase (TYR) and microphthalmia-associated transcription factor (MITF). MITF is a transcription factor that is responsible for melanocyte development in melanogenesis. Catechin will inhibit MITF, thereby preventing melanocytes from producing melanin and hyperpigmentation [50].

3.5. UV-Reduction and Sunscreen

UV radiation is categorised into UV-A (315–400 nm), UV-B (280–315 nm), and UV-C (280–100 nm). Continuous exposure to UV-B radiation can lead to disruptions in the skin caused by free radicals and ROS, which stimulate melanin production and melanocyte proliferation. Tyrosinase facilitates melanin synthesis and represents a pivotal point in melanogenesis. High melanin levels can disrupt pigmentation in human skin, leading to conditions such as

age spots, melasma, malignant melanomas, and freckles. EGCG is acknowledged as a natural antioxidant to neutralise free radicals, modulate the activity of antioxidant enzymes, diminish the effects of oxidative stress, and inhibit tyrosinase activity [51].

Sunscreen is a cosmetic that functions as a skin protector [52], filtering UV to reduce the radiation emitted from the sun [53][54]. Aromatic compounds conjugated to carbonyl groups convert UV energy into minimised UV energy [49], preventing the chemical properties of UV-absorbing potency without the need for significant photodegradation (**Figure 7**). The usage of sunscreen is determined based on the Sun Protection Factor (SPF), which is defined as the ability to supply a minimal erythema dose (MED) on the skin divided by the variable of UV energy that is needed to supply MED on unprotected skin [55].

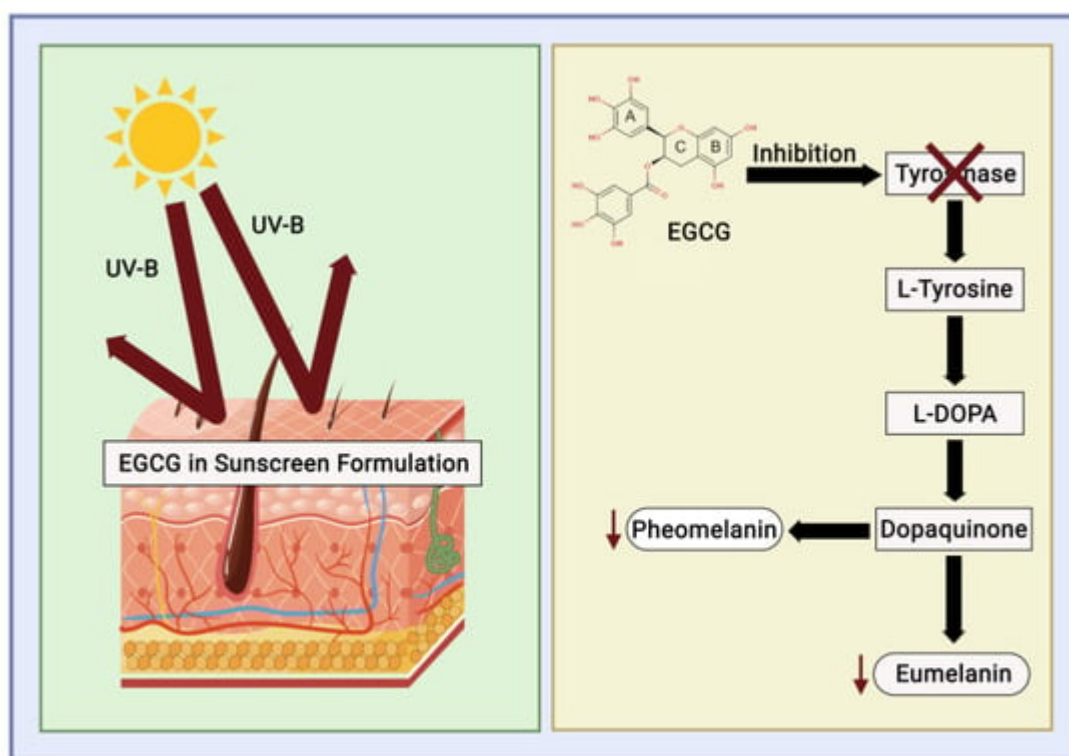


Figure 7. Catechin mechanism of action as a sunscreen.

The higher the SPF, the more protective the sunscreen [56] to prevent sunburn as well as skin cancer in the long term. The SPF can be determined with in vivo or in vitro methods. The in vivo test involves human volunteers while the in vitro is differentiated through two categories: measuring the UV absorption or transmission of the test product or spectrophotometric analysis. Effective sunscreen products at least need to have an absorbance of 290 to 400 nm [57].

3.6. Anti-Acne

Acne is a skin disease caused by the overgrowth of *Staphylococcus epidermidis* and *Propionibacterium acnes*. Some recent research showed that catechin from gambir and green tea could inhibit the growth of acne pathogens.

Recent research showed the MIC of catechin as anti-acne is 0.1%. Another study showed that the inhibition zone of catechin is 18.45 mm for *P. acnes* and 15.68 mm for *S. epidermidis* [58]. The antibacterial mechanism of catechin involves disrupting the cell membrane and inhibiting intracellular enzymes (Figure 8). The mechanism of catechin to counter acne inflammation is through the inhibition of the pro-inflammatory cytokines IL-8 and TNF- α [59].

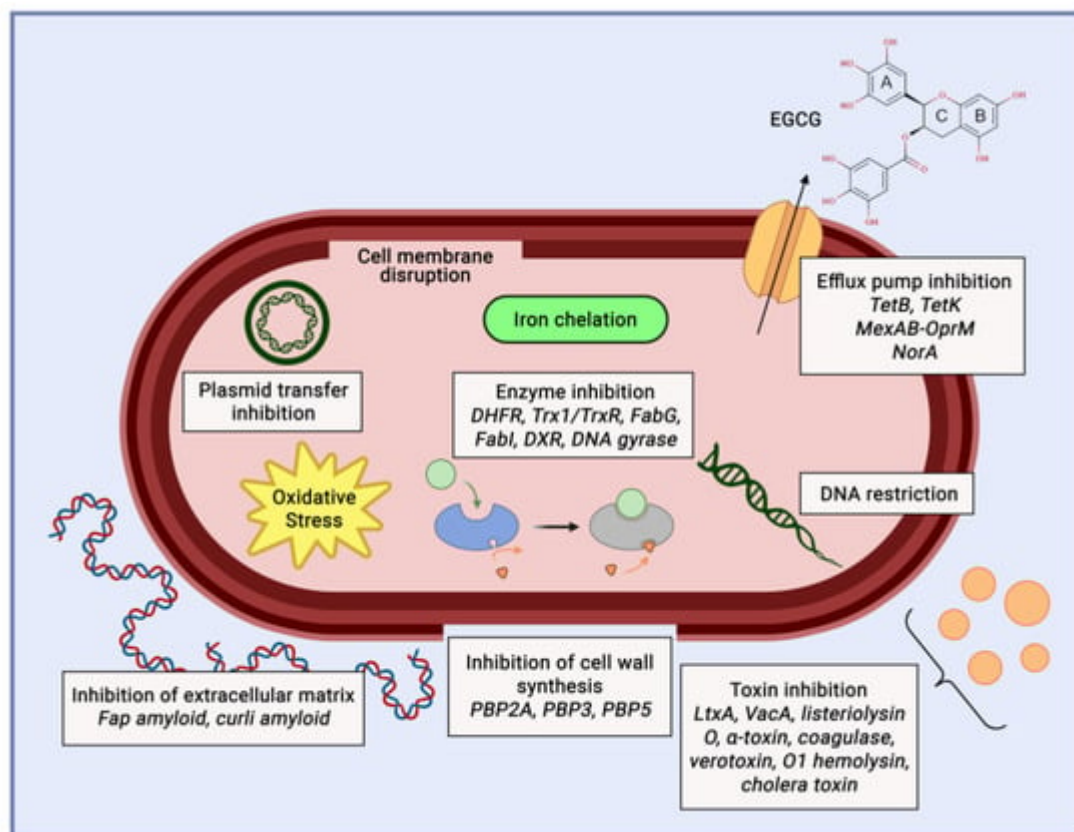


Figure 8. Catechin mechanism of action as anti-acne agent.

References

1. Bellad, K.A.; Nanjwade, B.K.; Kamble, M.S.; Srichana, T.; Idris, N.F. Development of cosmeceuticals. *World J. Pharm. Pharm. Sci.* 2017, 6, 643–691.
2. Yamada, M.; Mohammed, Y.; Prow, T.W. Advances and controversies in studying sunscreen delivery and toxicity. *Adv. Drug Deliv. Rev.* 2020, 153, 72–86.
3. Cavinato, M.; Jansen-Durr, P. Molecular mechanisms of UVB-induced senescence of dermal fibroblasts and its relevance for photoaging of the human skin. *Exp. Gerontol.* 2017, 94, 78–82.
4. Rinnerthaler, M.; Bischof, J.; Streubel, M.K.; Trost, A.; Richter, K. Oxidative stress in aging human skin. *Biomolecules* 2015, 5, 545–589.

5. Chang, H.H.; Chien, C.Y.; Chen, K.H.; Huang, S.C.; Chien, C.T. Catechins Blunt the Effects of oxLDL and its Primary Metabolite Phosphatidylcholine Hydroperoxide on Endothelial Dysfunction Through Inhibition of Oxidative Stress and Restoration of eNOS in Rats. *Kidney Blood Press. Res.* 2017, 42, 919–932.
6. Lee, S.; Yu, J.S.; Phung, H.M.; Lee, J.G.; Kim, K.H.; Kang, K.S. Potential Anti-Skin Aging Effect of (-)-Catechin Isolated from the Root Bark of *Ulmus davidiana* var. *japonica* in Tumor Necrosis Factor- α -Stimulated Normal Human Dermal Fibroblasts. *Antioxidants* 2020, 9, 981.
7. Braicu, C.; Ladamery, M.R.; Chedea, V.S.; Irimie, A.; Berindan-Neagoe, I. The relationship between the structure and biological actions of green tea catechins. *Food Chem.* 2013, 141, 3282–3289.
8. Bernatoniene, J.; Kopustinskiene, D.M. The Role of Catechins in Cellular Responses to Oxidative Stress. *Molecules* 2018, 23, 965.
9. Matsubara, T.; Wataoka, I.; Urakawa, H.; Yasunaga, H. Effect of reaction pH and CuSO₄ addition on the formation of catechinone due to oxidation of (+)-catechin. *Int. J. Cosmet. Sci.* 2013, 35, 362–367.
10. Ferreira-Nunes, R.; da Silva, S.M.M.; de Souza, P.E.N.; de Oliveira Magalhães, P.; Cunha-Filho, M.; Gratieri, T.; Gelfuso, G.M. Incorporation of *Eugenia dysenterica* extract in microemulsions preserves stability, antioxidant effect and provides enhanced cutaneous permeation. *J. Mol. Liq.* 2018, 265, 408–415.
11. Mita, S.R.; Abdassah, M.; Supratman, U.; Shiono, Y.; Rahayu, D.; Sopyan, I.; Wilar, G. Nanoparticulate System for the Transdermal Delivery of Catechin as an Antihypercholesterol: In Vitro and In Vivo Evaluations. *Pharmaceuticals* 2022, 15, 1142.
12. Munggari, I.P.; Kurnia, D.; Deawati, Y.; Juliaha, E. Current Research of Phytochemical, Medicinal and Non-Medicinal Uses of *Uncaria gambir* Roxb.: A Review. *Molecules* 2022, 27, 6551.
13. Bae, J.; Kim, N.; Shin, Y.; Kim, S.-Y.; Kim, Y.-J. Activity of catechins and their applications. *Biomed. Dermatol.* 2020, 4, 1–10.
14. Isemura, M. Catechin in Human Health and Disease. *Molecules* 2019, 24, 528.
15. Syukri, D.; Azima, F.; Aprialldho, R. Study on the Utilization of Catechins from Gambir (*Uncaria Gambir* Roxb) Leaves as Antioxidants Cooking Oil. *Andalasian Int. J. Agric. Nat. Sci.* 2022, 3, 12–25.
16. Zillich, O.; Schweiggert-Weisz, U.; Eisner, P.; Kerscher, M. Polyphenols as active ingredients for cosmetic products. *Int. J. Cosmet. Sci.* 2015, 37, 455–464.
17. Moldoveanu, S.C.; David, V. *Essentials in Modern HPLC Separations*; Elsevier: Amsterdam, The Netherlands, 2022.

18. Hariyati, N. Ministry of Health of the Republic of Indonesia Farmakope Herbal Indonesia, 2nd ed.; Ministry of Health Indonesia: Kota Jakarta, Indonesia, 2017.
19. Cuevas-Valenzuela, J.; González-Rojas, Á.; Wisniak, J.; Apelblat, A.; Pérez-Correa, J.R. Solubility of (+)-catechin in water and water-ethanol mixtures within the temperature range 277.6–331.2 K: Fundamental data to design polyphenol extraction processes. *Fluid Phase Equilibria* 2014, 382, 279–285.
20. Riwu, A.G.; Nugraha, J.; Purwanto, D.A.; Triyono, E.A. Determination of (+)-Catechin and Antioxidant Activity in Faloak (*Sterculia quadrifida* R. Br) Stem Bark Infusion. *Sci. Technol. Indones.* 2023, 8, 59–65.
21. Delarosa, A.; Hendrawan, R.P.; Halimah, E. Screening of *Costus speciosus* and Determination of Antioxidant Potential Using DPPH Method: A Review. *Eur. J. Med. Plants* 2023, 34, 17–28.
22. Chen, Z.; Liu, Q.; Zhao, Z.; Bai, B.; Sun, Z.; Cai, L.; Fu, Y.; Ma, Y.; Wang, Q.; Xi, G. Effect of hydroxyl on antioxidant properties of 2, 3-dihydro-3, 5-dihydroxy-6-methyl-4 H-pyran-4-one to scavenge free radicals. *RSC Adv.* 2021, 11, 34456–34461.
23. Hoang, H.T.; Moon, J.-Y.; Lee, Y.-C.J.C. Natural antioxidants from plant extracts in skincare cosmetics: Recent applications, challenges and perspectives. *Cosmetics* 2021, 8, 106.
24. Grzesik, M.; Naparło, K.; Bartosz, G.; Sadowska-Bartos, I. Antioxidant properties of catechins: Comparison with other antioxidants. *Food Chem.* 2018, 241, 480–492.
25. Sheng, Y.; Sun, Y.; Tang, Y.; Yu, Y.; Wang, J.; Zheng, F.; Li, Y.; Sun, Y. Catechins: Protective mechanism of antioxidant stress in atherosclerosis. *Front. Pharmacol.* 2023, 14, 1144878.
26. Šeruga, M.; Tomac, I. Influence of chemical structure of some flavonols on their electrochemical behaviour. *Int. J. Electrochem. Sci.* 2017, 12, 7616–7637.
27. Hassanpour, S.H.; Doroudi, A. Review of the antioxidant potential of flavonoids as a subgroup of polyphenols and partial substitute for synthetic antioxidants. *Avicenna J. Phytomed.* 2023, 13, 354.
28. Platzer, M.; Kiese, S.; Tybussek, T.; Herfellner, T.; Schneider, F.; Schweiggert-Weisz, U.; Eisner, P. Radical scavenging mechanisms of phenolic compounds: A quantitative structure-property relationship (QSPR) study. *Front. Nutr.* 2022, 9, 882458.
29. Anitha, S.; Krishnan, S.; Senthilkumar, K.; Sasirekha, V. Theoretical investigation on the structure and antioxidant activity of (+) catechin and (–) epicatechin—a comparative study. *Mol. Phys.* 2020, 118, e1745917.
30. Zhou, H.-C.; Tam, N.F.-y.; Lin, Y.-M.; Ding, Z.-H.; Chai, W.-M.; Wei, S.-D. Relationships between degree of polymerization and antioxidant activities: A study on proanthocyanidins from the leaves of a medicinal mangrove plant *Ceriops tagal*. *PLoS ONE* 2014, 9, e107606.

31. Spiegel, M.; Andruniów, T.; Sroka, Z. Flavones' and Flavonols' Antiradical Structure–Activity Relationship—A Quantum Chemical Study. *Antioxidants* 2020, 9, 461.
32. Munteanu, I.G.; Apetrei, C. Assessment of the Antioxidant Activity of Catechin in Nutraceuticals: Comparison between a Newly Developed Electrochemical Method and Spectrophotometric Methods. *Int. J. Mol. Sci.* 2022, 23, 8110.
33. Doshi, P.; Adsule, P.; Banerjee, K.; Oulkar, D. Technology. Phenolic compounds, antioxidant activity and insulinotropic effect of extracts prepared from grape (*Vitis vinifera* L) byproducts. *J. Food Sci. Technol.* 2015, 52, 181–190.
34. Dias, M.C.; Pinto, D.C.; Silva, A.M.S. Plant flavonoids: Chemical characteristics and biological activity. *Molecules* 2021, 26, 5377.
35. Nuryana, I.; Ratnakomala, S.; Fahrurrozi, A.B.J.; Andriani, A.; Putra, F.J.N.; Rezamela, E.; Wulansari, R.; Prawira-Atmaja, M.I.; Lisdiyanti, P. Catechin Contents, Antioxidant and Antibacterial Activities of Different Types of Indonesian Tea (*Camellia Sinensis*). *Ann. Bogor.* 2020, 24, 107.
36. Anggraini, T.; Wilma, S.; Syukri, D.; Azima, F.J.I.J.o.F.S. Total phenolic, anthocyanin, Catechins, DPPH radical scavenging activity, and toxicity of *Lepisanthes alata* (Blume) Leenh. *Int. J. Food Sci.* 2019, 2019, 9703176.
37. Musdja, M.Y.; Rahman, H.A.; Hasan, D.J.L.I.J.H.L.-S. Antioxidant activity of catechins isolate of *Uncaria gambier* Roxb in male rats. *LIFE Int. J. Health Life-Sci.* 2018, 4, 34–46.
38. Rahmi, M.; Rita, R.S.; Yetti, H. Gambir Catechins (*Uncaria gambier* Roxb) Prevent Oxidative Stress in Wistar Male Rats Fed a High-Fat Diet. *Maj. Kedokt. Andalas* 2021, 44, 436–441.
39. Yetuk, G.; Pandir, D.; Bas, H. Protective role of catechin and quercetin in sodium benzoate-induced lipid peroxidation and the antioxidant system in human erythrocytes in vitro. *Sci. World J.* 2014, 2014, 874824.
40. Amber, K.T.; Shiman, M.I.; Badiavas, E.V. The use of antioxidants in radiotherapy-induced skin toxicity. *Integr. Cancer Ther.* 2014, 13, 38–45.
41. Zheng, M.; Liu, Y.; Zhang, G.; Yang, Z.; Xu, W.; Chen, Q. The Applications and Mechanisms of Superoxide Dismutase in Medicine, Food, and Cosmetics. *Antioxidants* 2023, 12, 1675.
42. Boo, Y.C. Arbutin as a Skin Depigmenting Agent with Antimelanogenic and Antioxidant Properties. *Antioxidants* 2021, 10, 1129.
43. Jesus, A.; Mota, S.; Torres, A.; Cruz, M.T.; Sousa, E.; Almeida, I.F.; Cidade, H. Antioxidants in Sunscreens: Which and What For? *Antioxidants* 2023, 12, 138.
44. Nahhas, A.F.; Abdel-Malek, Z.A.; Kohli, I.; Braunberger, T.L.; Lim, H.W.; Hamzavi, I.H. The potential role of antioxidants in mitigating skin hyperpigmentation resulting from ultraviolet and

- visible light-induced oxidative stress. *Photodermatol. Photoimmunol. Photomed.* 2019, 35, 420–428.
45. Andrade, J.M.; Domínguez-Martín, E.M.; Nicolai, M.; Faustino, C.; Rodrigues, L.M.; Rijo, P. Screening the dermatological potential of plectranthus species components: Antioxidant and inhibitory capacities over elastase, collagenase and tyrosinase. *J. Enzyme Inhib. Med. Chem.* 2021, 36, 258–270.
 46. Sonawane, G.B.; Jadhav, S.P.; Patil, C.D.; Kamble, P.R.; Somavanshi, D.B. A Review on the Antioxidant and Antiaging Properties of White Tea. *J. Pharm. Res. Int.* 2021, 33, 129–136.
 47. Wang, W.; Di, T.; Wang, W.; Jiang, H. EGCG, GCG, TFDG, or TSA inhibiting melanin synthesis by downregulating MC1R expression. *Int. J. Mol. Sci.* 2023, 24, 11017.
 48. Zhang, X.; Li, J.; Li, Y.; Liu, Z.; Lin, Y.; Huang, J.-A. Anti-melanogenic effects of epigallocatechin-3-gallate (EGCG), epicatechin-3-gallate (ECG) and galocatechin-3-gallate (GCG) via down-regulation of cAMP/CREB/MITF signaling pathway in B16F10 melanoma cells. *Fitoterapia* 2020, 145, 104634.
 49. Jiang, T.; Qi, Y.; Wu, Y.; Zhang, J. Application of antioxidant and ultraviolet absorber into HDPE: Enhanced resistance to UV irradiation. *e-Polymers* 2019, 19, 499–510.
 50. Laksmiani, N.P.L.; Sanjaya, I.K.N.; Leliqia, N.P.E. The activity of avocado (*Persea americana* Mill.) seed extract containing catechin as a skin lightening agent. *J. Pharm. Pharmacogn. Res.* 2020, 8, 449–456.
 51. Vale, E.P.; dos Santos Morais, E.; de Souza Tavares, W.; de Sousa, F.F.O. Epigallocatechin-3-gallate loaded-zein nanoparticles: Characterization, stability and associated antioxidant, anti-tyrosinase and sun protection properties. *J. Mol. Liq.* 2022, 358, 119107.
 52. Donglikar, M.M.; Deore, S.L. Sunscreens: A review. *Pharmacogn. J.* 2016, 8, 171–179.
 53. Sander, M.; Sander, M.; Burbidge, T.; Beecker, J. The efficacy and safety of sunscreen use for the prevention of skin cancer. *CMAJ* 2020, 192, E1802–E1808.
 54. Dewi, D.A.R. Sunscreen Protection Against Visible Light: Is It Needed? *Malahayati Nurs. J.* 2022, 4, 2527–2536.
 55. Latha, M.; Martis, J.; Shobha, V.; Shinde, R.S.; Bangera, S.; Krishnankutty, B.; Bellary, S.; Varughese, S.; Rao, P.; Kumar, B.N.; et al. Sunscreening agents: A review. *J. Clin. Aesthet. Dermatol.* 2013, 6, 16.
 56. Portilho, L.; Aiello, L.M.; Vasques, L.I.; Bagatin, E.; Leonardi, G.R. Effectiveness of sunscreens and factors influencing sun protection: A review. *Braz. J. Pharm. Sci.* 2023, 58, e20693.
 57. Ebrahimzadeh, M.A.; Enayatifard, R.; Khalili, M.; Ghaffarloo, M.; Saeedi, M.; Charati, J.Y. Correlation between sun protection factor and antioxidant activity, phenol and flavonoid contents

- of some medicinal plants. Iran J. Pharm. Res. 2014, 13, 1041.
58. Warnida, H.; Masliyana, A.; Sapri, S.J. Formulasi Ekstrak Etanol Gambir (*Uncaria gambir* Roxb.) dalam Bedak Anti Jerawat. J. Ilm. Manuntung 2016, 2, 99–106.
59. Messire, G.; Serreau, R.; Berteina-Raboin, S. Antioxidant Effects of Catechins (EGCG), Andrographolide, and Curcuminoids Compounds for Skin Protection, Cosmetics, and Dermatological Uses: An Update. Antioxidants 2023, 12, 1317.
-

Retrieved from <https://encyclopedia.pub/entry/history/show/124731>